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Brief Communication

Rhizosolenia mat diatoms associate with nitrogen-fixing microbes

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Abstract

Some Rhizosolenia diatoms living in oligotrophic marine ecosystems are known to form large, conspicuous mats and are thought to be sources of new nitrogen to surface waters via vertical migration to the nitracline where subsurface nitrate is accessed for growth. These vertically migrating Rhizosolenia mats are chronically under sampled, and both the diatom species comprising the mats and the associated microbiome have not been characterized using modern molecular techniques. Here we present the first DNA-based analysis of Rhizosolenia mats collected in the North Pacific Subtropical Gyre. Using sequencing of 18S rRNA and nifH genes (a proxy for N_2 fixation capacity), we report on the molecular diversity of mat-forming Rhizosolenia species, which include two newly sequenced clades, and an assemblage of associated N_2 -fixing microorganisms that is distinct from the non-mat associated water column assemblage. Our findings advance knowledge of oligotrophic diatom diversity and challenge prevailing views of their nitrogen sources, suggesting these mats may obtain nitrogen through association-based N_2 fixation. Further work is needed to understand the nature of these associations, and whether Rhizosolenia mat communities are a significant unrecognized source of N_2 -fixation-derived new nitrogen to the oligotrophic surface waters.

Keywords: Rhizosolenia; diatoms; diazotrophs; N2 fixation; nifH

Introduction

Rhizosolenia is a cosmopolitan diatom genus, observed across broad latitudinal gradients from tropical to subpolar waters, including inland seas [1]. In the subtropical oligotrophic oceans, some Rhizosolenia sp. are known to have intracellular heterocystforming, filamentous, N2-fixing cyanobacterial symbionts (Richelia intracellularis), while other Rhizosolenia species are known to form large aggregates (or "mats") visible to the naked eye and comprised of multiple morphologically distinct Rhizosolenia sp. [2-4] and a microbiome that includes bacteria and ciliates [5]. Rhizosolenia mats contribute significantly to primary productivity and carbon export fluxes [6-9] and are thought to be sustained by nitrate assimilation at the nitracline accessed through vertical migration [10]. However, reports of N2 fixation in non-Richeliabearing Rhizosolenia and of intracellular bacteria found within the diatoms suggest a symbiotic interaction between matforming species and N2-fixing microbes [3]. Subsequent efforts to measure N2 fixation using acetylene reduction assays have not substantiated these findings [4], though short incubation periods (30-45 min) may have missed the active N2 fixation period or signals may have been undetectable over background

ethylene. Thus, the relative importance of nitrate assimilation vs. diazotrophy in supplying Rhizosolenia mats with nitrogen and the impact of mat diazotrophy on nitrogen biogeochemistry in the oligotrophic oceans remains unresolved.

In June 2022, we encountered extensive fields of large Rhizosolenia mats (ca. 10s of cm in length) in the North Pacific Subtropical Gyre (NPSG). These mats remained visible to the naked eye throughout a 48 h station occupation situated in a frontal region between two eddies (Fig. S1). We gently collected the fragile mats with a bucket (Fig. 1A), and light microscopy revealed that they were a matrix of morphologically distinct Rhizosolenia species including large (>50 μ m) and small (< 10 μ m) diameter chains. These morphotypes were consistent with gross morphological descriptions of Rhizosolenia castracanei, R. imbricata var. shrubsolei, R. formosa, R. debyana, and R. fallax species reported in previously described multi-species mats (Fig. 1B and C) [3, 4]. None of the diatoms were observed to host the heterocyst-forming cyanobacterial symbiont Richelia, and quantitative PCR (qPCR) confirmed that Rhizosolenia-associated Richelia (Het-1) were rare in surface water at this station (6 \times 10² nifH copies L⁻¹). Although detailed characterization of diagnostic frustule features was

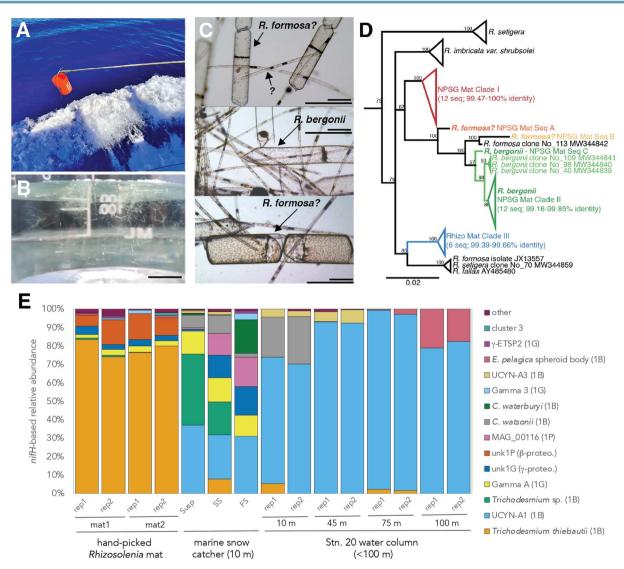


Figure 1. NPSG diatom mats contain diverse Rhizosolenia sp. and distinct diazotroph assemblages. (A) Rhizosolenia mats were bucket-sampled from the deck of the R/V kilo Moana (KM2206) at 27° 13.3' N, 178° 11.2' E on June 19, 2022. (B) Mat samples in a jar (scale bar, 1 cm). (C) Light micrographs of different diatom mat species (scale bars, 50 μ m, 100 μ m). (D) Full-length 18S rRNA gene neighbor-joining consensus phylogenetic tree of NPSG mat sequences with reference Rhizosolenia sp. sequences from Medlin et al. [22] retrieved from the NCBI nucleotide database. All branches or collapsed clusters with color designate sequences retrieved from Rhizosolenia mats in this study. Branch labels show consensus support (%). (E) Relative abundances of diazotroph taxa based on nifH amplicon high throughput sequencing in mat, marine snow catcher and water column samples. Marine snow catcher deployments (10 m) captured three size fractions—Neutrally buoyant (or suspended), slow sinking, and fast sinking particles. Partial nifH fragments were amplified, sequenced and analyzed as described previously [23, 24] and in supplemental materials. MSC—Marine snow catcher; Susp—Suspended particles; SS—Slow sinking particles; FS—Fast sinking particles.

not performed, precluding definitive morphology-based species identification, 18S rRNA gene analyses [11] indicated the presence of at least five distinct Rhizosolenia species or strains belonging to three major clades (Fig. 1D; Supplemental Material). Only one clade (NPSG Mat Clade II) and one additional sequence (NPSG Mat Seq C) showed sufficient similarity (>98%) to be reasonably assigned to R. bergonii, a species not typically associated with mat formation [12]. A single sequence (NPSG Mat Seq B) was identified as the mat-former R. formosa, and no sequences matched R. imbricata var. shrubsolei reference sequences. Two other major clades of NPSG mat-forming diatom sequences (Mat Clade I and Mat Clade III) could not be assigned to the species level due to limited reference sequence information for this genus. These sequences represent the first report of the molecular identities of confirmed mat-forming, non-Richelia-bearing, oligotrophic

Rhizosolenid diatoms. Resolving the taxonomy of this group is particularly important because environmental sequence datasets (e.g. Ocean Barcode Atlas: [13]; CalCOFI: [14]; Northwest Atlantic: [15]) often classify Rhizosolenia only at the genus level, which is insufficient to constrain diatom mat biogeography underscoring the need to improve reference databases.

Diatom mats were associated with a diazotroph assemblage distinct from those in marine snow and ambient water (Fig. 1E). The diazotroph assemblage associated with Rhizosolenia mats was investigated using nifH amplicon sequencing from hand-picked mat samples and compared to particle and water column samples collected from the same site using a marine snow catcher (MSC) and Niskin-Conductivity, Temperature, and Depth (CTD) rosette, respectively (See Supplemental Material for methodological details). A non-cyanobacterial diazotroph (NCD) affiliated with

a putative beta-proteobacterium (nifH cluster 1P) was specifically associated with the Rhizosolenia mats and not found in MSC or water column samples. Additionally, Trichodesmium thiebautii had high relative abundance within the mats, despite not often being observed in microscopic characterizations of the mats. Notably, the dominant mat-associated Trichodesmium sequences were distinct from MSC-associated Trichodesmium, another line of evidence supporting strain-specific mat associations (Fig. 1E). Collectively, these findings show that Rhizosolenia mats contain a distinct and varied community of both cyanobacterial diazotrophs (non-Richelia) and NCDs that does not simply reflect the diazotroph populations present in the MSC or water column samples, as would be expected if mats simply entrained the microbial populations present. Alternatively, mat assemblages may contain rare diazotroph taxa if the mat structure selects against the dominant diazotrophs in the water column or MSC samples. More work is needed to understand the variability of Rhizosolenia mat N2-fixing microbiomes, whether some mat-forming species host endosymbiotic NCDs as previously

Oligotrophic Rhizosolenia mat diatoms are thought to migrate to the nitracline to access subsurface nitrate to support their nitrogen demands [7]. However, uncertainty persists over whether some mat-forming species obtain nitrogen from diazotrophs. Reports of N₂ fixation potential in Rhizosolenia mats have varied, perhaps since the specific Rhizosolenia species involved were not clearly identified, highlighting the importance of voucher specimens and a more precisely resolved Rhizosolenia phylogeny moving forward. However, our data provides evidence that matforming Rhizosolenia species, and possibly the associated mat microbiome, may obtain at least some of their nitrogen through a distinct assemblage of associated cyanobacterial diazotrophs and/or NCDs. If Rhizosolenia mats acquire nitrogen through N2 fixation, it represents a potentially significant and currently overlooked source of new nitrogen to the oligotrophic ocean. Current estimates indicate that N₂ fixation in the oligotrophic waters of the North Pacific fuels ~50% of new production [16, 17]. Likewise, previously described associations between specific diatom hosts and heterocyst-forming cyanobacteria, including Richelia intracellularis associated with Rhizosolenia, and Richelia euintracellularis associated with Hemiaulus [18], are believed to support a significant portion of carbon export [16, 19]. Outside the predictable summertime export events, yearly N2 fixation is generally associated with smaller unicellular cyanobacterial diazotrophs (<10 μ m; [20]). The most common approach for measuring N₂ fixation uses small volume (~4 L) ¹⁵N₂ tracer incubations of whole water, typically collected with Niskin bottles on a CTD rosette, and likely underestimates contributions from large and heterogeneously distributed diazotrophs (>20 μ m; [21]). Thus, the magnitude of N₂ fixed by heterogeneous and delicate Rhizosolenia mat communities likely remains uncaptured in current estimates, yet our work suggests N2 fixation within Rhizosolenia mats may add to diazotrophic support of carbon export. Determining if Rhizosolenia mats are an important and overlooked niche for diazotrophs and rely, even in part, on N2 fixation to supply their nitrogen demand has broad implications for prior estimates of their contribution to primary productivity and carbon export flux in oligotrophic ecosystems. Further work is necessary to clarify the nature of these associations using modern molecular techniques and directly measure Rhizosolenia mat associated N2 fixation. It is clearly time to revisit the old paradigm.

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Supplementary material

Supplementary material is available at ISME Communications online.

Conflicts of interest

None declared.

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Data availability

All Rhizosolenia 18S rRNA sequences are available in the nr database at NCBI Genbank under Accession numbers PV187244-PV187289. Raw nifH read data is available at NCBI SRA under BioProject # PRJNA1229115.

References

- 1. Malviya S, Scalco E, Audic S. et al. Insights into global diatom distribution and diversity in the world's ocean. Proc Natl Acad Sci USA 2016;113:E1516-25. https://doi.org/10.1073/ pnas.1509523113
- 2. Carpenter EJ, Harbison GR, Madin LP. et al. Rhizosolenia mats. Limnol Oceanogr 1977;22:739-41. https://doi.org/10.4319/ lo.1977.22.4.0739
- 3. Martínez L, Silver MW, King JM. et al. Nitrogen fixation by floating diatom mats: a source of new nitrogen to oligotrophic ocean waters. Science. 1983;221:152-4. https://doi.org/10.1126/ science.221.4606.152
- 4. Villareal TA, Carpenter EJ. Nitrogen fixation, suspension characteristics and chemical composition of Rhizosolenia mats in the central North Pacific gyre. Biol Oceanogr 1989;6:387-405.
- 5. Caron DA, Davis PG, Madin LP. et al. Heterotrophic bacteria and bacterivorous protozoa in oceanic macroaggregates. Science. 1982;**218**:795–7. https://doi.org/10.1126/science.218.4574.795
- 6. Villareal TA, Altabet MA, Culver-Rymzsa K. Nitrogen transport by vertically migrating diatom mats in the North Pacific Ocean. Nature. 1993;363:709-12. https://doi.org/10.1038/363709a0
- 7. Villareal TA, Woods S, Moore JK. et al. Vertical migration of Rhizosolenia mats and their significance to NO₃- fluxes in the

- central North Pacific gyre. J Plankton Res 1996; 18:1103–21. https://doi.org/10.1093/plankt/18.7.1103
- 8. Pilskaln C, Villareal T, Dennett M. et al. High concentrations of marine snow and diatom algal mats in the North Pacific subtropical gyre: implications for carbon and nitrogen cycles in the oligotrophic ocean. *Deep-Sea Res I Oceanogr Res Pap* 2005;**52**: 2315–32. https://doi.org/10.1016/j.dsr.2005.08.004
- Villareal TA, Pilskaln CH, Montoya JP. et al. Upward nitrate transport by phytoplankton in oceanic waters: balancing nutrient budgets in oligotrophic seas. *PeerJ.* 2014;2:e302. https://doi. org/10.7717/peerj.302
- Singler H, Villareal T. Nitrogen inputs into the euphotic zone by vertically migrating Rhizosolenia mats. J Plankton Res 2005;27: 545–56. https://doi.org/10.1093/plankt/fbi030
- Moon-van der Staay SY, De Wachter R, Vaulot D. Oceanic 18S rDNA sequences from picoplankton reveal unsuspected eukaryotic diversity. Nature. 2001;409:607–10. https://doi. org/10.1038/35054541
- 12. Boonprakob A, Lundholm N, Medlin LK. et al. The morphology and phylogeny of the diatom genera Rhizosolenia, Proboscia, Pseudosolenia and Neocalyptrella from gulf of Thailand and the Andaman Sea, with description of Rhizosolenia loanicola sp. nov., Proboscia siamensis sp. nov. and Probosciales Ord. Nov. Diatom Res 2021;36:143–84. https://doi.org/10.1080/0269249X.2021.1957719
- Vernette C, Henry N, Lecubin J. et al. The ocean barcode atlas: a web service to explore the biodiversity and biogeography of marine organisms. Mol Ecol Resour 2021;21:1347–58. https://doi. org/10.1111/1755-0998.13322
- James CC, Barton AD, Allen LZ. et al. Influence of nutrient supply on plankton microbiome biodiversity and distribution in a coastal upwelling region. Nat Commun 2022;13:2448. https:// doi.org/10.1038/s41467-022-30139-4
- 15. Setta SP, Lerch S, Jenkins BD. *et al.* Oligotrophic waters of the Northwest Atlantic support taxonomically diverse diatom communities that are distinct from coastal waters. *J Phycol* 2023;**59**: 1202–16. https://doi.org/10.1111/jpy.13388

- Karl DM, Church MJ, Dore JE. et al. Predictable and efficient carbon sequestration in the North Pacific Ocean supported by symbiotic nitrogen fixation. Proc Natl Acad Sci 2012;109:1842–9. https://doi.org/10.1073/pnas.1120312109
- Böttjer D, Dore JE, Karl DM. et al. Temporal variability of nitrogen fixation and particulate nitrogen export at station ALOHA. Limnol Oceanogr 2017;62:200–16. https://doi.org/10.1002/lno.10386
- 18. Foster RA, Villareal TA, Lundin D. Richelia. In: Trujillo ME, Dedysh S, Devos P. et al. (eds.), Bergey's Manual of Systematics of Archaea and Bacteria. John Wiley & Sons, Inc., in association with Bergey's Manual Trust.
- Poff KE, Leu AO, Eppley JM. et al. Microbial dynamics of elevated carbon flux in the open ocean's abyss. Proc Natl Acad Sci USA 2021;118:e2018269118. https://doi.org/10.1073/ pnas.2018269118
- Church MJ, Mahaffey C, Letelier RM. et al. Physical forcing of nitrogen fixation and diazotroph community structure in the North Pacific subtropical gyre. Glob Biogeochem Cycles 2009;23:GB2020. https://doi.org/10.1029/2008GB003418
- Follett CL, White AE, Wilson ST. et al. Nitrogen fixation rates diagnosed from diurnal changes in elemental stoichiometry. Limnol Oceanogr 2018;63:1911–23. https://doi.org/10.1002/ lno.10815
- Medlin LK, Boonprakob A, Lundholm N. et al. On the morphology and phylogeny of the diatom species Rhizosolenia setigera: comparison of the type material to modern cultured strains, and a taxonomic revision. Nova Hedwigia Beihefte 2021;151:223–47. https://doi.org/10.1127/nova-suppl/2021/223
- Turk-Kubo KA, Henke BA, Gradoville MR. et al. Seasonal and spatial patterns in diazotroph community composition at station ALOHA. Front Mar Sci 2023;10:1130158. https://doi.org/10.3389/fmars.2023.1130158
- 24. Morando M, Magasin J, Cheung S. et al. Global biogeography of N₂-fixing microbes: nifH amplicon database and analytics workflow. Earth Syst Sci Data Discus 2024; 1–39.