



DATA NOTE

The genome sequence of the Ruddy Flat-body, *Agonopterix subpropinquella* (Stainton, 1849) [version 1; peer review: 2 approved]

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Abstract



We present a genome assembly from an individual male *Agonopterix subpropinquella* (the Ruddy Flat-body; Arthropoda; Insecta; Lepidoptera; Depressariidae). The genome sequence is 667.9 megabases in span. Most of the assembly is scaffolded into 28 chromosomal pseudomolecules, including the Z sex chromosome. The mitochondrial genome has also been assembled and is 16.5 kilobases in length. Gene annotation of this assembly on Ensembl identified 18,796 protein coding genes.

Keywords

Agonopterix subpropinquella, Ruddy Flat-body, genome sequence, chromosomal, Lepidoptera

Open Peer Review

Approval Status  

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Any reports and responses or comments on the article can be found at the end of the article.



This article is included in the [Tree of Life gateway](#).

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Species taxonomy

Eukaryota; Metazoa; Eumetazoa; Bilateria; Protostomia; Ecdysozoa; Panarthropoda; Arthropoda; Mandibulata; Pancrustacea; Hexapoda; Insecta; Dicondylia; Pterygota; Neoptera; Endopterygota; Amphiesmenoptera; Lepidoptera; Glossata; Neolepidoptera; Heteroneura; Ditrysiina; Gelechioidea; Depressariidae; Depressariinae; *Agonopterix*; *Agonopterix subpropinquella* (Stainton, 1849) (NCBI:txid1857958).

Background

Agonopterix subpropinquella, the Ruddy Flat-body, is a small moth of the Depressariidae family. The species has a pan-European, mainly coastal, distribution (GBIF Secretariat, 2023), and is found widely across Britain and Ireland (Harper *et al.*, 2002). Like many species in its genus, the imago is drab-coloured and indistinctly patterned, indeed the species' scientific name likely being a reference to its similarity to other members of the genus (Emmet, 1991). However, the species also shows a distinctive form f. *rhodochrella*, with exaggerated blackish colouring on the head, thorax, and forewing (Harper *et al.*, 2002).

In common with many members of its genus, the species overwinters as an adult (Harper *et al.*, 2002). Adults are on the wing between August and May, hibernating over the winter, and may be disturbed from their hibernation by beating thatch or dense vegetation over the winter (Sterling & Parsons, 2018). Eggs are laid in May on knapweeds (*Centaurea* spp.) or thistles (*Cirsium* spp.) (Harper *et al.*, 2002). The larva is green and initially mines the foodplant before feeding in a silken spinning (Harper *et al.*, 2002). Larvae feed between June and July, pupating from July to August in earth or amongst detritus (Harper *et al.*, 2002).

The genome of the ruddy flat-body, *Agonopterix subpropinquella*, was sequenced as part of the Darwin Tree of Life Project, a collaborative effort to sequence all named eukaryotic species in the Atlantic Archipelago of Britain and Ireland. Here we present a chromosomally complete genome sequence for *Agonopterix subpropinquella*, based on one male specimen from Wytham Woods, Oxfordshire.

Genome sequence report

The genome was sequenced from one male *Agonopterix subpropinquella* (Figure 1) collected from Wytham Woods, Oxfordshire, UK (51.77, -1.34). A total of 28-fold coverage in Pacific Biosciences single-molecule HiFi long reads and 63-fold coverage in 10X Genomics read clouds were generated. Primary assembly contigs were scaffolded with chromosome conformation Hi-C data. Manual assembly curation corrected 56 missing joins or mis-joins and removed 12 haplotypic duplications, reducing the assembly length by 0.43% and the scaffold number by 50.79%.

The final assembly has a total length of 667.9 Mb in 31 sequence scaffolds with a scaffold N50 of 25.1 Mb (Table 1). Most (99.98%) of the assembly sequence was assigned to 28 chromosomal-level scaffolds, representing 27 autosomes



Figure 1. Photograph of the *Agonopterix subpropinquella* (ilAgoSubp1) specimen used for genome sequencing.

and the Z sex chromosome. A summary of the assembly statistics is shown in Figure 2, while the distribution of assembly scaffolds on GC proportion and coverage is shown in Figure 3. The cumulative assembly plot in Figure 4 shows curves for subsets of scaffolds assigned to different phyla. Chromosome-scale scaffolds confirmed by the Hi-C data are named in order of size (Figure 5; Table 2). While not fully phased, the assembly deposited is of one haplotype. Contigs corresponding to the second haplotype have also been deposited. The mitochondrial genome was also assembled and can be found as a contig within the multifasta file of the genome submission.

The estimated Quality Value (QV) of the final assembly is 56.5 with *k*-mer completeness of 99.99%, and the assembly has a BUSCO v5.3.2 completeness of 98.7% (single = 98.0%, duplicated = 0.7%), using the lepidoptera_odb10 reference set ($n = 5,286$).

Metadata for specimens, spectral estimates, sequencing runs, contaminants and pre-curation assembly statistics can be found at <https://links.tol.sanger.ac.uk/species/1857958>.

Genome annotation report

The *Agonopterix subpropinquella* genome assembly (GCA_922987775.1) was annotated using the Ensembl rapid annotation pipeline (Table 1; https://rapid.ensembl.org/Agonopterix_subpropinquella_GCA_922987775.1/Info/Index). The resulting annotation includes 18,967 transcribed mRNAs from 18,796 protein-coding genes.

Methods

Sample acquisition and nucleic acid extraction

A male *Agonopterix subpropinquella* (specimen ID Ox000822, ToLID ilAgoSubp1) was collected from Wytham Woods, Oxfordshire (biological vice-county Berkshire), UK (latitude 51.77, longitude -1.34) on 2020-08-01 using a light trap. The specimen was collected and identified by Douglas Boyes (University of Oxford) and preserved on dry ice.

DNA was extracted at the Tree of Life laboratory, Wellcome Sanger Institute (WSI). The ilAgoSubp1 sample was weighed and dissected on dry ice with tissue set aside for Hi-C

Table 1. Genome data for *Agonopterix subpropinquella*, ilAgoSubp1.1.

Project accession data		
Assembly identifier	ilAgoSubp1.1	
Assembly release date	2021-12-19	
Species	<i>Agonopterix subpropinquella</i>	
Specimen	ilAgoSubp1	
NCBI taxonomy ID	1857958	
BioProject	PRJEB47465	
BioSample ID	SAMEA7746629	
Isolate information	ilAgoSubp1, male: whole organism (DNA sequencing and Hi-C data)	
Assembly metrics*		Benchmark
Consensus quality (QV)	56.5	≥ 50
k-mer completeness	99.99%	≥ 95%
BUSCO**	C:98.7%[S:98.0%,D:0.7%], F:0.2%,M:1.1%,n:5,286	C ≥ 95%
Percentage of assembly mapped to chromosomes	99.98%	≥ 95%
Sex chromosomes	Z chromosome	localised homologous pairs
Organelles	Mitochondrial genome assembled	complete single alleles
Raw data accessions		
PacificBiosciences SEQUEL II	ERR6808069, ERR6939284	
10X Genomics Illumina	ERR6747928, ERR6747929, ERR6747930, ERR6747931	
Hi-C Illumina	ERR6747927	
Genome assembly		
Assembly accession	GCA_922987775.1	
Accession of alternate haplotype	GCA_922987765.1	
Span (Mb)	667.9	
Number of contigs	95	
Contig N50 length (Mb)	11.6	
Number of scaffolds	31	
Scaffold N50 length (Mb)	25.1	
Longest scaffold (Mb)	69.3	
Genome annotation		
Number of protein-coding genes	18,796	
Number of gene transcripts	18,967	

* Assembly metric benchmarks are adapted from column VGP-2020 of "Table 1: Proposed standards and metrics for defining genome assembly quality" from (Rhie *et al.*, 2021).

** BUSCO scores based on the lepidoptera_odb10 BUSCO set using v5.3.2. C = complete [S = single copy, D = duplicated], F = fragmented, M = missing, n = number of orthologues in comparison. A full set of BUSCO scores is available at <https://blobtoolkit.genomehubs.org/view/Agonopterix%20subpropinquella/dataset/CAKLPO01.1/busco>.

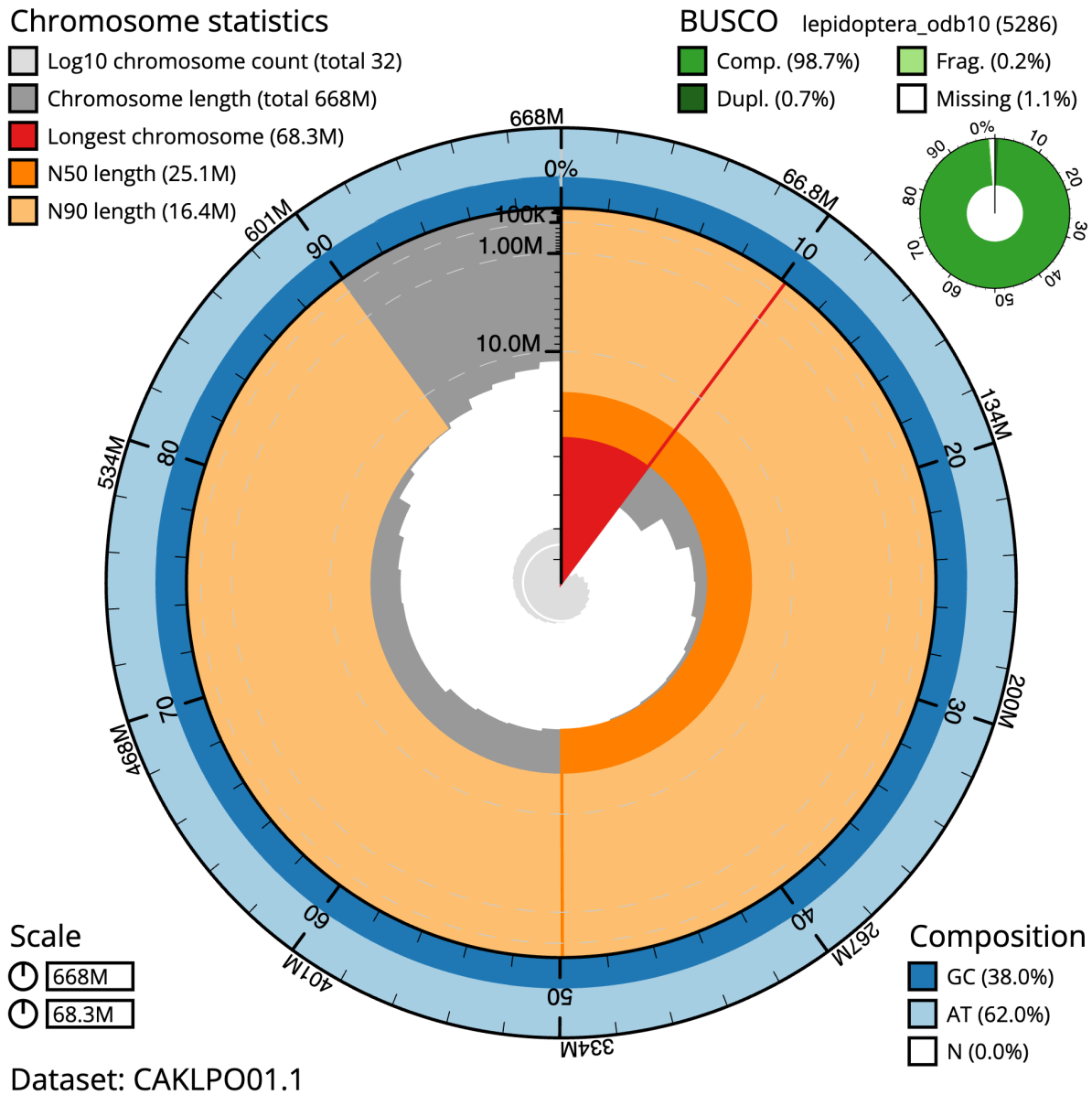


Figure 2. Genome assembly of *Agonopterix subpropinquella*, ilAgoSubp1.1: metrics. The BlobToolKit Snailplot shows N50 metrics and BUSCO gene completeness. The main plot is divided into 1,000 size-ordered bins around the circumference with each bin representing 0.1% of the 667,905,724 bp assembly. The distribution of scaffold lengths is shown in dark grey with the plot radius scaled to the longest scaffold present in the assembly (68,253,362 bp, shown in red). Orange and pale-orange arcs show the N50 and N90 scaffold lengths (25,090,608 and 16,437,072 bp), respectively. The pale grey spiral shows the cumulative scaffold count on a log scale with white scale lines showing successive orders of magnitude. The blue and pale-blue area around the outside of the plot shows the distribution of GC, AT and N percentages in the same bins as the inner plot. A summary of complete, fragmented, duplicated and missing BUSCO genes in the lepidoptera_odb10 set is shown in the top right. An interactive version of this figure is available at <https://blobtoolkit.genomehubs.org/view/Agonopterix%20subpropinquella/dataset/CAKLPO01.1/snail>.

sequencing. Tissue from the whole organism was disrupted using a Nippi Powermasher fitted with a BioMasher pestle. High molecular weight (HMW) DNA was extracted using the Qiagen MagAttract HMW DNA extraction kit. Low molecular weight DNA was removed from a 20 ng aliquot of extracted DNA using the 0.8X AMPure XP purification kit prior to 10X Chromium sequencing; a minimum of 50 ng DNA was

submitted for 10X sequencing. HMW DNA was sheared into an average fragment size of 12–20 kb in a Megaruptor 3 system with speed setting 30. Sheared DNA was purified by solid-phase reversible immobilisation using AMPure PB beads with a 1.8X ratio of beads to sample to remove the shorter fragments and concentrate the DNA sample. The concentration of the sheared and purified DNA was assessed using a

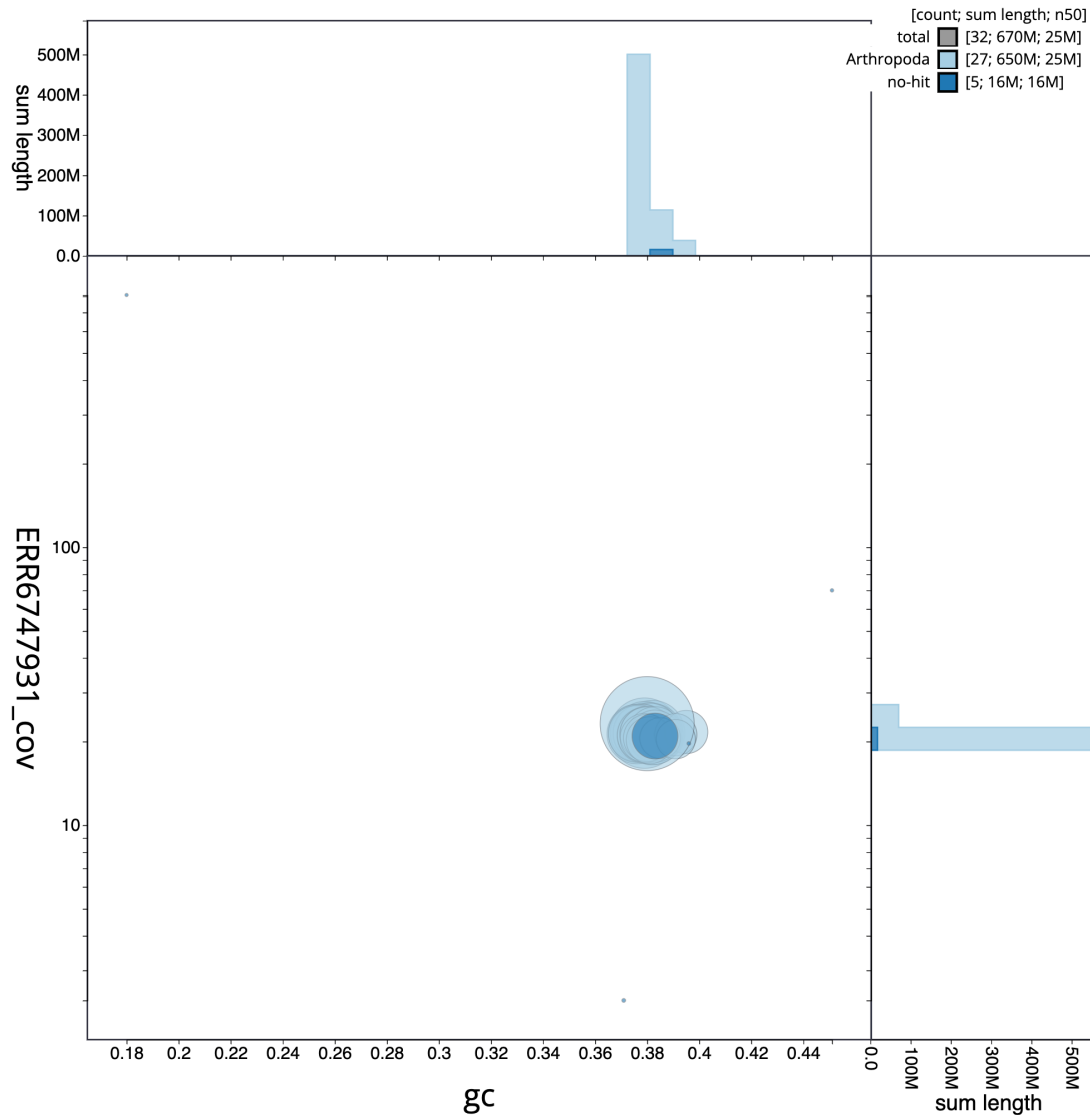


Figure 3. Genome assembly of *Agonopterix subpropinquella*, ilAgoSubp1.1: BlobToolKit GC-coverage plot. Scaffolds are coloured by phylum. Circles are sized in proportion to scaffold length. Histograms show the distribution of scaffold length sum along each axis. An interactive version of this figure is available at <https://blobtoolkit.genomehubs.org/view/Agonopterix%20subpropinquella/dataset/CAKLP001.1/blob>.

Nanodrop spectrophotometer and Qubit Fluorometer and Qubit dsDNA High Sensitivity Assay kit. Fragment size distribution was evaluated by running the sample on the FemtoPulse system.

Sequencing

Pacific Biosciences HiFi circular consensus and 10X Genomics read cloud DNA sequencing libraries were constructed according to the manufacturers' instructions. DNA sequencing was performed by the Scientific Operations core at the WSI on Pacific Biosciences SEQUEL II (HiFi) and Illumina NovaSeq 6000 (10X) instruments. Hi-C data were also generated from remaining tissue of ilAgoSubp1 using the Arima2 kit and sequenced on the Illumina NovaSeq 6000 instrument.

Genome assembly, curation and evaluation

Assembly was carried out with Hifiasm (Cheng *et al.*, 2021) and haplotypic duplication was identified and removed with purge_dups (Guan *et al.*, 2020). One round of polishing was performed by aligning 10X Genomics read data to the assembly with Long Ranger ALIGN, calling variants with FreeBayes (Garrison & Marth, 2012). The assembly was then scaffolded with Hi-C data (Rao *et al.*, 2014) using SALSA2 (Ghurye *et al.*, 2019). The assembly was checked for contamination and corrected as described previously (Howe *et al.*, 2021). Manual curation was performed using HiGlass (Kerpedjiev *et al.*, 2018) and Pretext (Harry, 2022). The mitochondrial genome was assembled using MitoHiFi (Uliano-Silva *et al.*, 2023), which

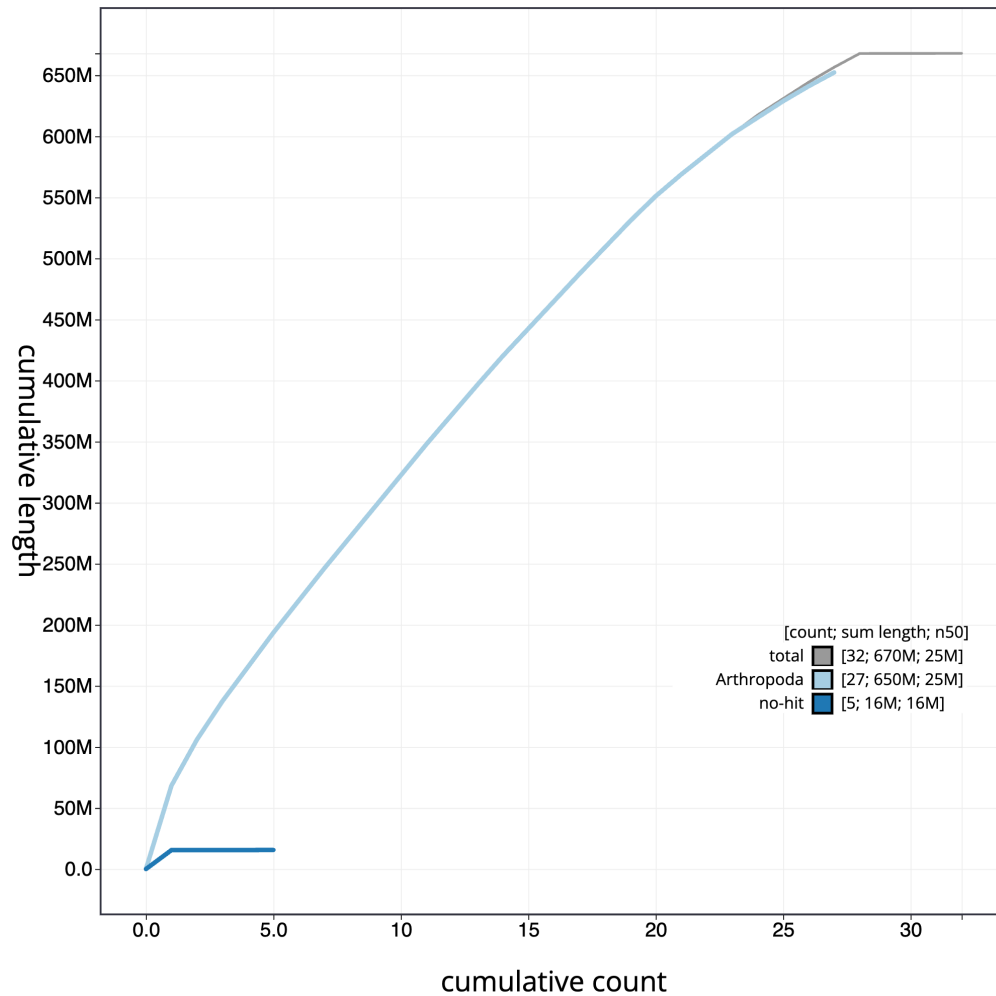


Figure 4. Genome assembly of *Agonopterix subpropinquella*, ilAgoSubp1.1: BlobToolKit cumulative sequence plot. The grey line shows cumulative length for all scaffolds. Coloured lines show cumulative lengths of scaffolds assigned to each phylum using the buscogenes taxrule. An interactive version of this figure is available at <https://blobtoolkit.genomehubs.org/view/Agonopterix%20subpropinquella/dataset/CAKLPO01.1/cumulative>.

runs MitoFinder (Allio *et al.*, 2020) or MITOS (Bernt *et al.*, 2013) and uses these annotations to select the final mitochondrial contig and to ensure the general quality of the sequence.

A Hi-C map for the final assembly was produced using bwa-mem2 (Vasimuddin *et al.*, 2019) in the Cooler file format (Abdennur & Mirny, 2020). To assess the assembly metrics, the *k*-mer completeness and QV consensus quality values were calculated in Merqury (Rhie *et al.*, 2020). This work was done using Nextflow (Di Tommaso *et al.*, 2017) DSL2 pipelines “sanger-tol/readmapping” (Surana *et al.*, 2023a) and “sanger-tol/genomenote” (Surana *et al.*, 2023b). The genome was analysed within the BlobToolKit environment (Challis *et al.*, 2020) and BUSCO scores (Manni *et al.*, 2021; Simão *et al.*, 2015) were calculated.

Table 3 contains a list of relevant software tool versions and sources.

Genome annotation

The BRAKER2 pipeline (Brůna *et al.*, 2021) was used in the default protein mode to generate annotation for the *Agonopterix subpropinquella* assembly (GCA_922987775.1) in Ensembl Rapid Release.

Wellcome Sanger Institute – Legal and Governance

The materials that have contributed to this genome note have been supplied by a Darwin Tree of Life Partner. The submission of materials by a Darwin Tree of Life Partner is subject to the ‘**Darwin Tree of Life Project Sampling Code of Practice**’, which can be found in full on the Darwin Tree of Life website [here](#). By agreeing with and signing up to the Sampling Code of Practice, the Darwin Tree of Life Partner agrees they will meet the legal and ethical requirements and standards set out within this document in respect of all samples acquired for, and supplied to, the Darwin Tree of Life Project.

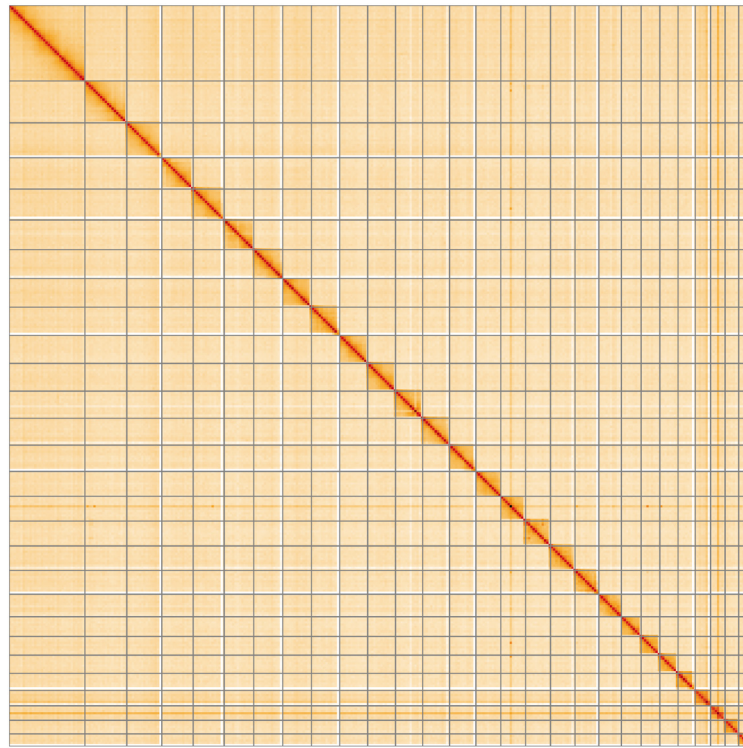


Figure 5. Genome assembly of *Agonopterix subpropinquella*, ilAgoSubp1.1: Hi-C contact map of the ilAgoSubp1.1 assembly, visualised using HiGlass. Chromosomes are shown in order of size from left to right and top to bottom. An interactive version of this figure may be viewed at <https://genome-note-higlass.tol.sanger.ac.uk/l/?d=RQh-QoTJRnastq9ymSznYA>.

Table 2. Chromosomal pseudomolecules in the genome assembly of *Agonopterix subpropinquella*, ilAgoSubp1.

INSDC accession	Chromosome	Length (Mb)	GC%
OV277694.1	1	37.69	38.0
OV277695.1	2	31.47	38.0
OV277696.1	3	28.22	38.0
OV277697.1	4	27.9	38.0
OV277698.1	5	26.69	37.5
OV277699.1	6	26.05	38.0
OV277700.1	7	25.78	38.0
OV277701.1	8	25.49	37.5
OV277702.1	9	25.12	38.0
OV277703.1	10	25.09	38.0
OV277704.1	11	24.47	37.5
OV277705.1	12	24.19	37.5
OV277706.1	13	23.65	38.0

INSDC accession	Chromosome	Length (Mb)	GC%
OV277707.1	14	22.38	38.0
OV277708.1	15	22.37	38.0
OV277709.1	16	22.35	38.0
OV277710.1	17	22.07	38.0
OV277711.1	18	21.46	38.5
OV277712.1	19	20.32	38.5
OV277713.1	20	17.76	38.0
OV277714.1	21	16.71	38.0
OV277715.1	22	16.44	38.0
OV277716.1	23	15.51	38.5
OV277717.1	24	13.79	39.5
OV277718.1	25	13.08	39.0
OV277719.1	26	12.23	38.5
OV277720.1	27	11.25	39.0
OV277693.1	Z	68.25	38.0
OV277721.1	MT	0.02	18.0

Table 3. Software tools: versions and sources.

Software tool	Version	Source
BlobToolKit	4.0.7	https://github.com/blobtoolkit/blobtoolkit
BUSCO	5.3.2	https://gitlab.com/ezlab/busco
FreeBayes	1.3.1-17-gaa2ace8	https://github.com/freebayes/freebayes
Hifiasm	0.15.3-r339	https://github.com/chhylp123/hifiasm
HiGlass	1.11.6	https://github.com/higlass/higlass
Long Ranger ALIGN	2.2.2	https://support.10xgenomics.com/genome-exome/software/pipelines/latest/advanced/other-pipelines
Merqury	MerquryFK	https://github.com/thegenemyers/MERQURY.FK
MitoHiFi	2	https://github.com/marcelauliano/MitoHiFi
PretextView	0.2	https://github.com/wtsi-hpag/PretextView
purge_dups	1.2.3	https://github.com/dfguan/purge_dups
SALSA	2.2	https://github.com/salsa-rs/salsa
sanger-tol/genomenote	v1.0	https://github.com/sanger-tol/genomenote
sanger-tol/readmapping	1.1.0	https://github.com/sanger-tol/readmapping/tree/1.1.0

Further, the Wellcome Sanger Institute employs a process whereby due diligence is carried out proportionate to the nature of the materials themselves, and the circumstances under which they have been/are to be collected and provided for use. The purpose of this is to address and mitigate any potential legal and/or ethical implications of receipt and use of the materials as part of the research project, and to ensure that in doing so we align with best practice wherever possible. The overarching areas of consideration are:

- Ethical review of provenance and sourcing of the material
- Legality of collection, transfer and use (national and international)

Each transfer of samples is further undertaken according to a Research Collaboration Agreement or Material Transfer Agreement entered into by the Darwin Tree of Life Partner, Genome Research Limited (operating as the Wellcome Sanger Institute), and in some circumstances other Darwin Tree of Life collaborators.

Data availability

European Nucleotide Archive: *Agonopterix subpropinquella* (ruddy flat-body). Accession number PRJEB47465; <https://identifiers.org/ena.embl/PRJEB47465>. (Wellcome Sanger Institute, 2021) The genome sequence is released openly for reuse.

The *Agonopterix subpropinquella* genome sequencing initiative is part of the Darwin Tree of Life (DTOL) project. All raw sequence data and the assembly have been deposited in INSDC databases. Raw data and assembly accession identifiers are reported in Table 1.

Author information

Members of the University of Oxford and Wytham Woods Genome Acquisition Lab are listed here: <https://doi.org/10.5281/zenodo.4789928>.

Members of the Darwin Tree of Life Barcoding collective are listed here: <https://doi.org/10.5281/zenodo.4893703>.

Members of the Wellcome Sanger Institute Tree of Life programme are listed here: <https://doi.org/10.5281/zenodo.4783585>.

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Members of the Tree of Life Core Informatics collective are listed here: <https://doi.org/10.5281/zenodo.5013541>.

Members of the Darwin Tree of Life Consortium are listed here: <https://doi.org/10.5281/zenodo.4783558>.

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Reviewer Report 06 May 2024

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Naciye Sena Çağatay 

University of Liverpool, Liverpool, England, UK

The complete genome of the Ruddy Flat-body *Agonopterix subpropinquella* was assembled with Boyes and Hammond using PacBio HiFi sequencing, 10X Genomics and Hi-C Illumina sequencing techniques. The analysis were explained very well in each method. The genome assembly is of high quality and the figures shown are informative. I have a minor revision below.

Background

Can you check these words.

Like many species in its genus, the imago is "image"

Eggs are lain in May on knapweeds "laid"

Q1:

I was just wondering, there is no information about morphological identification. How do you decide this species? Why are you using male sample?

Is the rationale for creating the dataset(s) clearly described?

Yes

Are the protocols appropriate and is the work technically sound?

Yes

Are sufficient details of methods and materials provided to allow replication by others?

Yes

Are the datasets clearly presented in a useable and accessible format?

Yes

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: Molecular entomology

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Reviewer Report 27 March 2024

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Simo Njabulo Maduna 

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The near-complete genome of the Ruddy Flat-body *Agonopterix subpropinquella* was assembled by Boyes and Hammond through the utilization of BacBio HiFi sequencing, as well as 10X Genomics and Hi-C Illumina sequencing techniques. The authors present the initial and superior reference genome for the Ruddy Flat-body. Their manuscript is skilfully composed and includes an elaborate bioinformatics pipeline and succinct findings, accompanied by genome assembly metric benchmarks to facilitate comprehension. Below I present minor revisions.

ABSTRACT

Include a sentence on the sequencing strategy employed to generate the reported chromosomal-level genome assembly for the target species.

BACKGROUND

Kindly incorporate a sentence regarding the genome sizes of moths, accompanied by a few examples, to provide the reader with an understanding of the average genome size of moths.

Change "...Like many species in its genus..." to "...Like many of its congeners..."

Question: What was the rationale behind selecting a homogametic male for sequencing rather than a heterogametic female? The reviewer is merely inquisitive given that the research gap is now on the assembly of the W sex chromosome.

Is the rationale for creating the dataset(s) clearly described?

Yes

Are the protocols appropriate and is the work technically sound?

Yes

Are sufficient details of methods and materials provided to allow replication by others?

Yes

Are the datasets clearly presented in a useable and accessible format?

Yes

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: Genome Assembly; Ecological Genomics; Conservation Genomics; Invasive Genomics; Population Genomics; Phylogenomics; Molecular Phylogenetics

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.
