



DATA NOTE

# The genome sequence of the Orange Footman, *Eilema sororcula* (Hufnagel, 1766) [version 1; peer review: 2 approved, 1 approved with reservations]

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## Abstract

We present a genome assembly from an individual male *Eilema sororcula* (the Orange Footman; Arthropoda; Insecta; Lepidoptera; Erebididae). The genome sequence is 729.4 megabases in span. Most of the assembly is scaffolded into 30 chromosomal pseudomolecules, including the Z sex chromosome. The mitochondrial genome has also been assembled and is 15.46 kilobases in length. Gene annotation of this assembly on Ensembl identified 21,093 protein coding genes.

## Keywords

*Eilema sororcula*, Orange Footman, genome sequence, chromosomal, Lepidoptera



This article is included in the [Tree of Life gateway](#).

## Open Peer Review

Approval Status   

	1	2	3
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2. **Juan Galarza**, University of Oulu, Oulu, Finland
3. **Marta Vila** , Universidade da Coruna, A Coruña, Spain

Any reports and responses or comments on the article can be found at the end of the article.

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**Author roles:** **Boyes D:** Investigation, Resources; **Lewis OT:** Writing – Original Draft Preparation, Writing – Review & Editing;

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## Species taxonomy

Eukaryota; Metazoa; Eumetazoa; Bilateria; Protostomia; Ecdysozoa; Panarthropoda; Arthropoda; Mandibulata; Pancrustacea; Hexapoda; Insecta; Dicondylia; Pterygota; Neoptera; Endopterygota; Amphimesenoptera; Lepidoptera; Glossata; Neolepidoptera; Heteroneura; Ditrysia; Obtectomera; Noctuoidea; Erebidae; Arctiinae; Lithosiini; *Eilema*; *Eilema sororcula* (Hufnagel, 1766) (NCBI:txid987424).

## Background

The Orange Footman *Eilema sororcula* had a local distribution in the south and east of the UK until the late 20th century, but like many related lichen-feeding Footman species, it has recently spread northwards and increased spectacularly in both its distribution and abundance (Randle *et al.*, 2019). At the time of writing there has been only a handful of records from Ireland (Moths Ireland, 2023), but it has been recorded across much of Europe and east across Eurasia to Korea and China (GBIF Secretariat, 2023).

The preferred habitats for *E. sororcula* are mature woodlands where its larvae feed on algae and lichens growing on trees (Henwood *et al.*, 2020). In Britain and Ireland, the adult moth is mostly observed during May and June, with the peak period of observations occurring a few weeks earlier in the year than in the 1970s (Randle *et al.*, 2019).

Here we present a chromosomally complete genome sequence for *E. sororcula* based on one male specimen from Wytham Woods, Oxfordshire, UK. A genome sequence for *E. sororcula* will facilitate studies into molecular adaptations to lichen-feeding and contribute to a growing data set of resources for understanding lepidopteran biology.

## Genome sequence report

The genome was sequenced from one male *Eilema sororcula* (Figure 1) collected from Wytham Woods, Oxfordshire,



**Figure 1.** Photograph of the *Eilema sororcula* (iEilSoro1) specimen used for genome sequencing.

UK (51.77, -1.34). A total of 43-fold coverage in Pacific Biosciences single-molecule HiFi long reads and 93-fold coverage in 10X Genomics read clouds was generated. Primary assembly contigs were scaffolded with chromosome conformation Hi-C data. Manual assembly curation corrected 81 missing joins or mis-joins and removed 7 haplotypic duplications, reducing the assembly length by 0.15% and the scaffold number by 60.4%, and increasing the scaffold N50 by 5.25%.

The final assembly has a total length of 729.4 Mb in 40 sequence scaffolds with a scaffold N50 of 25.3 Mb (Table 1). Most (99.94%) of the assembly sequence was assigned to 30 chromosomal-level scaffolds, representing 29 autosomes and the Z sex chromosome. Chromosome-scale scaffolds confirmed by the Hi-C data are named in order of size (Figure 2–Figure 5; Table 2). While not fully phased, the assembly deposited is of one haplotype. Contigs corresponding to the second haplotype have also been deposited. The mitochondrial genome was also assembled and can be found as a contig within the multifasta file of the genome submission.

The estimated Quality Value (QV) of the final assembly is 61.3 with *k*-mer completeness of 100%, and the assembly has a BUSCO v5.3.2 completeness of 98.5% (single = 97.8%, duplicated = 0.7%), using the lepidoptera\_odb10 reference set (*n* = 5,286).

Metadata for specimens, spectral estimates, sequencing runs, contaminants and pre-curation assembly statistics can be found at <https://links.tol.sanger.ac.uk/species/987424>.

## Genome annotation report

The *Eilema sororcula* genome assembly (GCA\_914829495.1) was annotated using the Ensembl rapid annotation pipeline (Table 1; [https://rapid.ensembl.org/Eilema\\_sororculum\\_GCA\\_914829495.1/Info/Index](https://rapid.ensembl.org/Eilema_sororculum_GCA_914829495.1/Info/Index)). The resulting annotation includes 21,274 transcribed mRNAs from 21,093 protein-coding genes.

## Methods

### Sample acquisition and nucleic acid extraction

A male *Eilema sororcula* (specimen ID Ox000399, individual iEilSoro1) was collected in Wytham Woods, Oxfordshire (biological vice-county Berkshire), UK (latitude 51.77, longitude -1.34) on 2020-05-22, using a light trap. The specimen was collected and identified by Douglas Boyes (University of Oxford) and preserved on dry ice.

DNA was extracted at the Tree of Life laboratory, Wellcome Sanger Institute (WSI). The iEilSoro1 sample was weighed and dissected on dry ice with tissue set aside for Hi-C sequencing. Tissue from the whole organism was disrupted using a Nippi Powermasher fitted with a BioMasher pestle. High molecular weight (HMW) DNA was extracted using the Qiagen MagAttract HMW DNA extraction kit. Low molecular weight DNA was removed from a 20 ng aliquot of extracted DNA using the 0.8X AMPure XP purification kit prior to 10X Chromium sequencing; a minimum of 50 ng DNA was submitted for 10X sequencing. HMW DNA was sheared into an

**Table 1. Genome data for *Eilema sororcula*, iEilSoro1.1.**

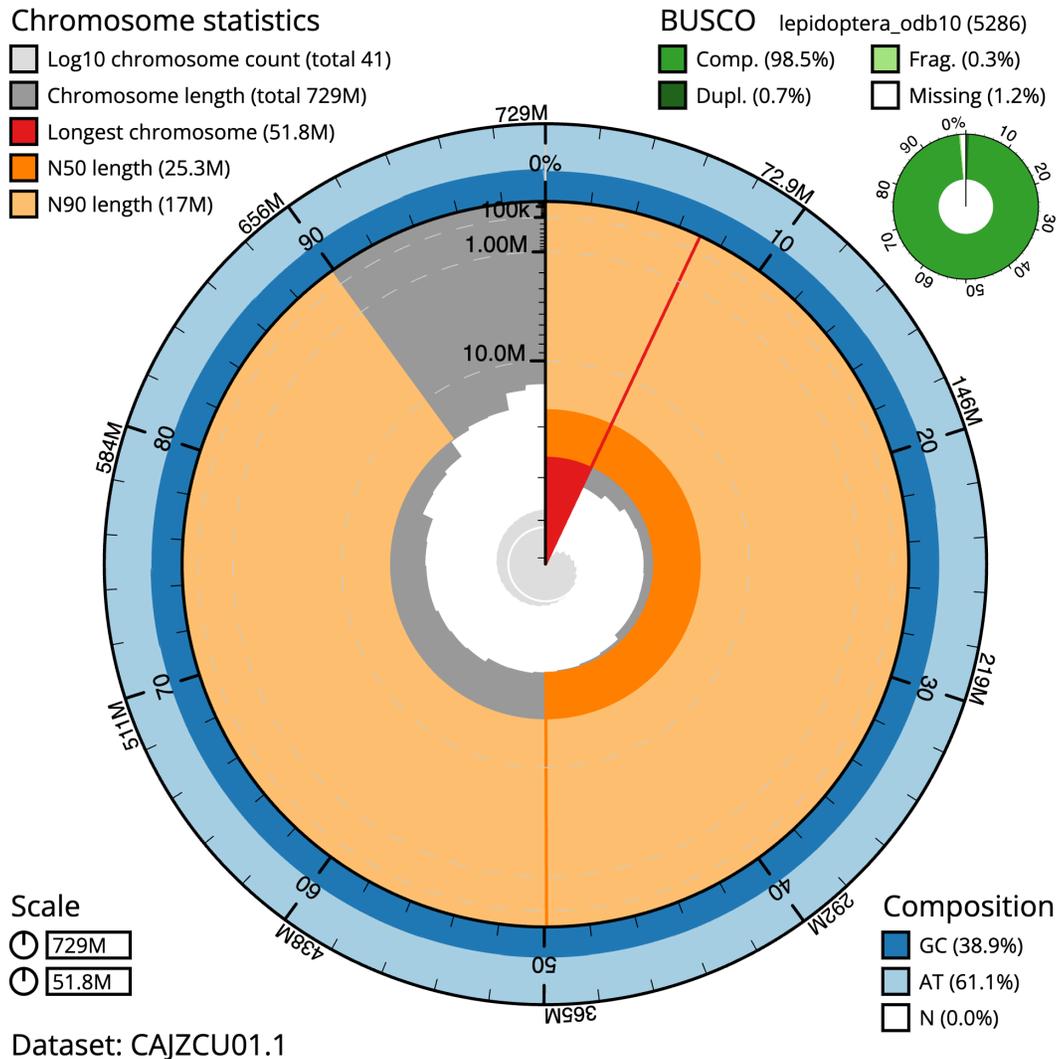
Project accession data		
Assembly identifier	iEilSoro1.1	
Species	<i>Eilema sororcula</i>	
Specimen	iEilSoro1	
NCBI taxonomy ID	987424	
BioProject	PRJEB46305	
BioSample ID	SAMEA7631555	
Isolate information	iEilSoro1, male: whole organism (DNA sequencing and Hi-C scaffolding)	
Assembly metrics*		Benchmark
Consensus quality (QV)	61.3	≥ 50
k-mer completeness	100%	≥ 95%
BUSCO**	C:98.5%[S:97.8%,D:0.7%],F:0.3%,M:1.2%,n:5,286	C ≥ 95%
Percentage of assembly mapped to chromosomes	99.94%	≥ 95%
Sex chromosomes	Z chromosome	localised homologous pairs
Organelles	Mitochondrial genome assembled	complete single alleles
Raw data accessions		
PacificBiosciences SEQUEL II	ERR6807993, ERR6939231	
10X Genomics Illumina	ERR6688449-ERR6688452	
Hi-C Illumina	ERR6688453	
Genome assembly		
Assembly accession	GCA_914829495.1	
Accession of alternate haplotype	GCA_914829255.1	
Span (Mb)	729.4	
Number of contigs	157	
Contig N50 length (Mb)	11.8	
Number of scaffolds	40	
Scaffold N50 length (Mb)	25.3	
Longest scaffold (Mb)	51.8	
Genome annotation		
Number of protein-coding genes	21,093	
Number of gene transcripts	21,274	

\* Assembly metric benchmarks are adapted from column VGP-2020 of "Table 1: Proposed standards and metrics for defining genome assembly quality" from (Rhie *et al.*, 2021).

\*\* BUSCO scores based on the lepidoptera\_odb10 BUSCO set using v5.3.2. C = complete [S = single copy, D = duplicated], F = fragmented, M = missing, n = number of orthologues in comparison. A full set of BUSCO scores is available at <https://blobtoolkit.genomehubs.org/view/iEilSoro1.1/dataset/CAJZCU01.1/busco>.

average fragment size of 12–20 kb in a Megaruptor 3 system with speed setting 30. Sheared DNA was purified by solid-phase reversible immobilisation using AMPure PB beads

with a 1.8X ratio of beads to sample to remove the shorter fragments and concentrate the DNA sample. The concentration of the sheared and purified DNA was assessed using a



**Figure 2. Genome assembly of *Eilema sororcula*, iEilSoro1.1: metrics.** The BlobToolKit Snailplot shows N50 metrics and BUSCO gene completeness. The main plot is divided into 1,000 size-ordered bins around the circumference with each bin representing 0.1% of the 729,417,332 bp assembly. The distribution of scaffold lengths is shown in dark grey with the plot radius scaled to the longest scaffold present in the assembly (51,764,148 bp, shown in red). Orange and pale-orange arcs show the N50 and N90 scaffold lengths (25,298,203 and 16,955,058 bp), respectively. The pale grey spiral shows the cumulative scaffold count on a log scale with white scale lines showing successive orders of magnitude. The blue and pale-blue area around the outside of the plot shows the distribution of GC, AT and N percentages in the same bins as the inner plot. A summary of complete, fragmented, duplicated and missing BUSCO genes in the lepidoptera\_odb10 set is shown in the top right. An interactive version of this figure is available at <https://blobtoolkit.genomehubs.org/view/iEilSoro1.1/dataset/CAJZCU01.1/snail>.

Nanodrop spectrophotometer and Qubit Fluorometer and Qubit dsDNA High Sensitivity Assay kit. Fragment size distribution was evaluated by running the sample on the FemtoPulse system.

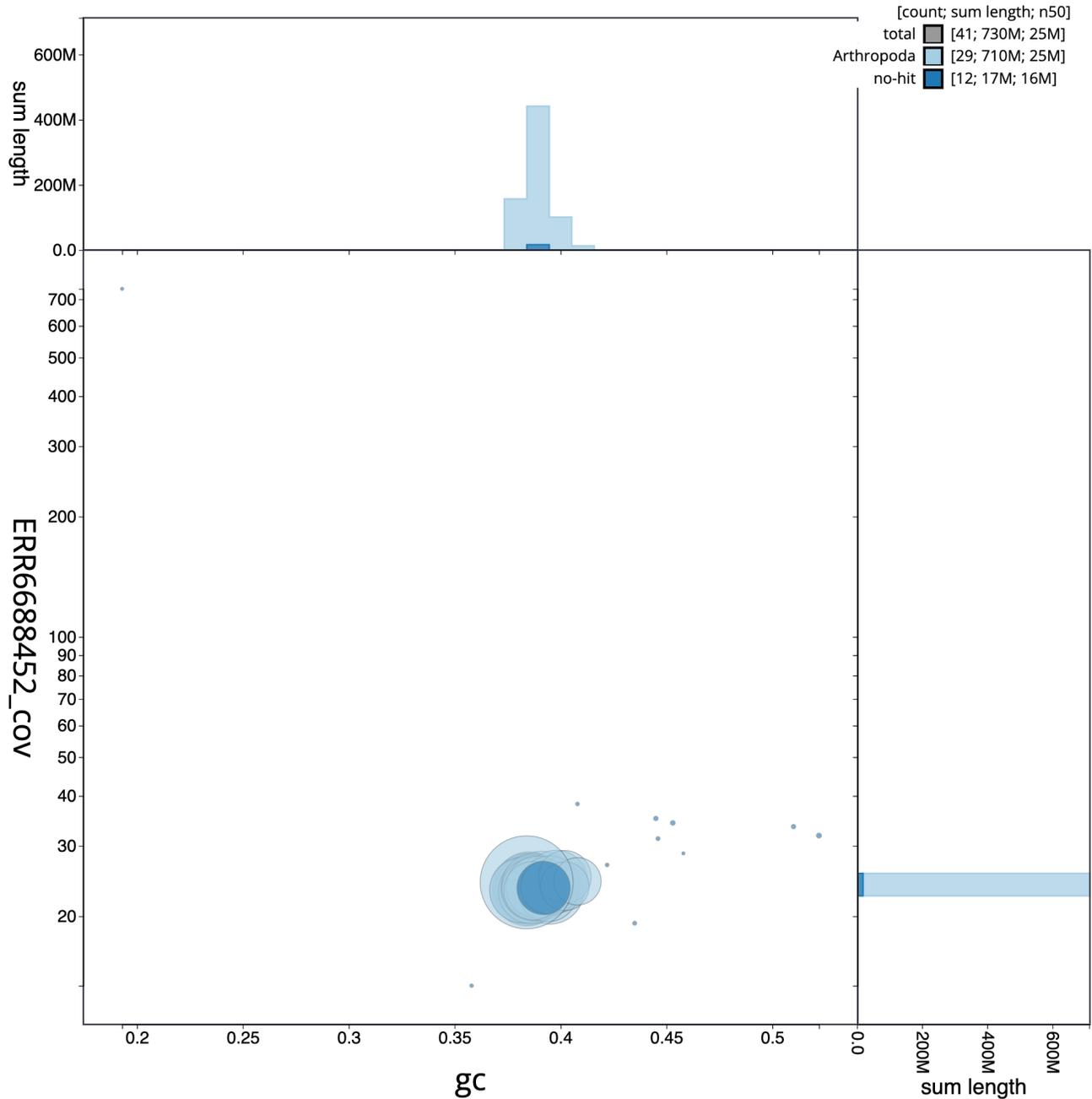
### Sequencing

Pacific Biosciences HiFi circular consensus and 10X Genomics read cloud DNA sequencing libraries were constructed according to the manufacturers' instructions. DNA sequencing was performed by the Scientific Operations core at the WSI on

Pacific Biosciences SEQUEL II (HiFi) and Illumina NovaSeq 6000 (10X) instruments. Hi-C data were also generated from tissue of iEilSoro1 using the Arima2 kit and sequenced on the Illumina NovaSeq 6000 instrument.

### Genome assembly, curation and evaluation

Assembly was carried out with Hifiasm (Cheng *et al.*, 2021) and haplotypic duplication was identified and removed with purge\_dups (Guan *et al.*, 2020). One round of polishing was performed by aligning 10X Genomics read data to the

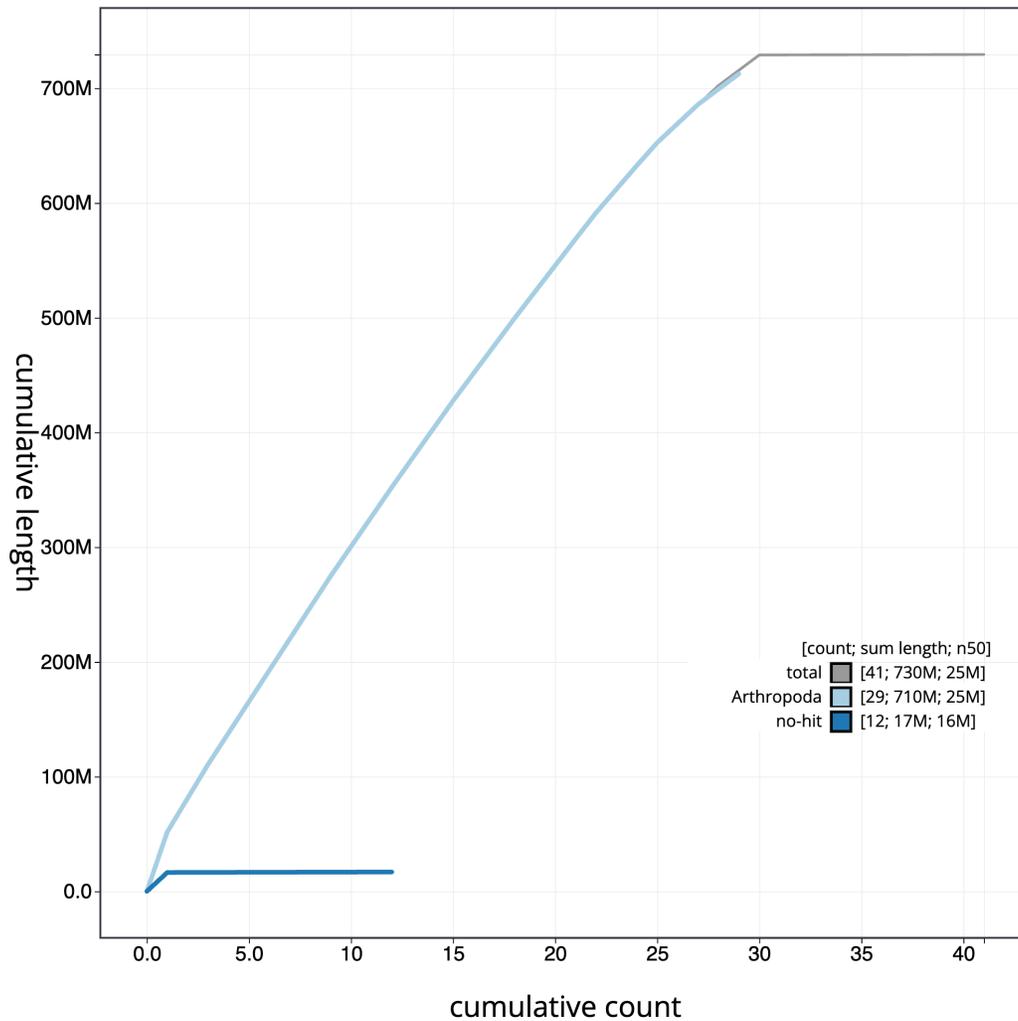


**Figure 3. Genome assembly of *Eilema sororcula*, ilEiSoro1.1: BlobToolKit GC-coverage plot.** Scaffolds are coloured by phylum. Circles are sized in proportion to scaffold length. Histograms show the distribution of scaffold length sum along each axis. An interactive version of this figure is available at <https://blobtoolkit.genomehubs.org/view/ilEiSoro1.1/dataset/CAJZCU01.1/blob>.

assembly with Long Ranger ALIGN, calling variants with FreeBayes (Garrison & Marth, 2012). The assembly was then scaffolded with Hi-C data (Rao *et al.*, 2014) using SALSA2 (Ghurye *et al.*, 2019). The assembly was checked for contamination and corrected as described previously (Howe *et al.*, 2021). Manual curation was performed using HiGlass (Kerpedjiev *et al.*, 2018) and Pretext (Harry, 2022). The mitochondrial genome was assembled using MitoHiFi

(Uliano-Silva *et al.*, 2022), which runs MitoFinder (Allio *et al.*, 2020) or MITOS (Bernt *et al.*, 2013) and uses these annotations to select the final mitochondrial contig and to ensure the general quality of the sequence.

A Hi-C map for the final assembly was produced using bwa-mem2 (Vasimuddin *et al.*, 2019) in the Cooler file format (Abdennur & Mirny, 2020). To assess the assembly metrics,



**Figure 4. Genome assembly of *Eilema sororcula*, iEilSoro1.1: BlobToolKit cumulative sequence plot.** The grey line shows cumulative length for all scaffolds. Coloured lines show cumulative lengths of scaffolds assigned to each phylum using the buscogenes taxrule. An interactive version of this figure is available at <https://blobtoolkit.genomehubs.org/view/iEilSoro1.1/dataset/CAJZCU01.1/cumulative>.

the *k*-mer completeness and QV consensus quality values were calculated in Merqury (Rhie *et al.*, 2020). This work was done using Nextflow (Di Tommaso *et al.*, 2017) DSL2 pipelines “sanger-tol/readmapping” (Surana *et al.*, 2023a) and “sanger-tol/genomenote” (Surana *et al.*, 2023b). The genome was analysed within the BlobToolKit environment (Challis *et al.*, 2020) and BUSCO scores (Manni *et al.*, 2021; Simão *et al.*, 2015) were calculated.

Table 3 contains a list of relevant software tool versions and sources.

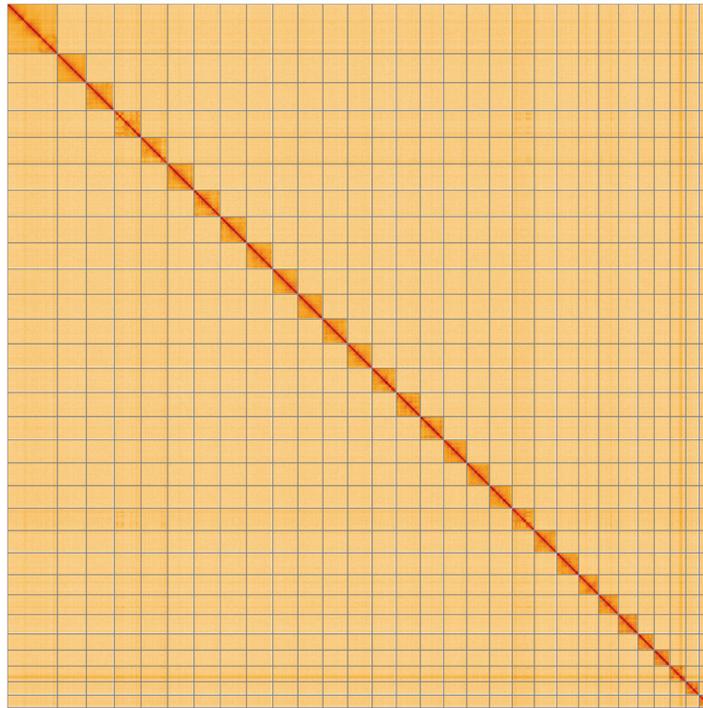
#### Genome annotation

The BRAKER2 pipeline (Brůna *et al.*, 2021) was used in the default protein mode to generate annotation for the *Eilema*

*sororcula* assembly (GCA\_914829495.1) in Ensembl Rapid Release.

#### Wellcome Sanger Institute – Legal and Governance

The materials that have contributed to this genome note have been supplied by a Darwin Tree of Life Partner. The submission of materials by a Darwin Tree of Life Partner is subject to the ‘**Darwin Tree of Life Project Sampling Code of Practice**’, which can be found in full on the Darwin Tree of Life website [here](#). By agreeing with and signing up to the Sampling Code of Practice, the Darwin Tree of Life Partner agrees they will meet the legal and ethical requirements and standards set out within this document in respect of all samples acquired for, and supplied to, the Darwin Tree of Life Project.



**Figure 5. Genome assembly of *Eilema sororcula*, iEilSoro1.1: Hi-C contact map of the iEilSoro1.1 assembly, visualised using HiGlass.** Chromosomes are shown in order of size from left to right and top to bottom. An interactive version of this figure may be viewed at <https://genome-note-higlass.tol.sanger.ac.uk/l/?d=IiePitoAQqCad3afW9wBMg>.

**Table 2. Chromosomal pseudomolecules in the genome assembly of *Eilema sororcula*, iEilSoro1.**

INSDC accession	Name	Size (Mb)	GC%
OU618533.1	1	30.01	38.5
OU618534.1	2	28.88	38.6
OU618535.1	3	27.72	39.1
OU618536.1	4	27.46	39.5
OU618537.1	5	27.37	38.5
OU618538.1	6	27.23	38.4
OU618539.1	7	27.15	38.4
OU618540.1	8	27.12	38.5
OU618541.1	9	26.1	38.7
OU618542.1	10	25.84	38.5
OU618543.1	11	25.59	38.2
OU618544.1	12	25.3	38.7
OU618545.1	13	25.05	38.2
OU618546.1	14	24.98	38.6
OU618547.1	15	24.1	38.7

INSDC accession	Name	Size (Mb)	GC%
OU618548.1	16	23.88	38.8
OU618549.1	17	23.65	38.9
OU618550.1	18	23.21	38.7
OU618551.1	19	23.2	39.8
OU618552.1	20	23.01	38.7
OU618553.1	21	22.79	38.9
OU618554.1	22	20.73	39.4
OU618555.1	23	20.53	39.5
OU618556.1	24	19.97	38.7
OU618557.1	25	16.96	39.4
OU618558.1	26	16.45	39.2
OU618559.1	27	16.3	40.2
OU618560.1	28	13.71	40.2
OU618561.1	29	12.97	40.8
OU618532.1	Z	51.76	38.4
OU618562.1	MT	0.02	19.5
-	-	0.4	45.8

**Table 3. Software tools: versions and sources.**

Software tool	Version	Source
BlobToolKit	4.0.7	<a href="https://github.com/blobtoolkit/blobtoolkit">https://github.com/blobtoolkit/blobtoolkit</a>
BUSCO	5.3.2	<a href="https://gitlab.com/ezlab/busco">https://gitlab.com/ezlab/busco</a>
FreeBayes	1.3.1-17-gaa2ace8	<a href="https://github.com/freebayes/freebayes">https://github.com/freebayes/freebayes</a>
Hifiasm	0.15.3	<a href="https://github.com/chhylp123/hifiasm">https://github.com/chhylp123/hifiasm</a>
HiGlass	1.11.6	<a href="https://github.com/higlass/higlass">https://github.com/higlass/higlass</a>
Long Ranger ALIGN	2.2.2	<a href="https://support.10xgenomics.com/genome-exome/software/pipelines/latest/advanced/other-pipelines">https://support.10xgenomics.com/genome-exome/software/pipelines/latest/advanced/other-pipelines</a>
Mercury	MercuryFK	<a href="https://github.com/thegenemyers/MERQURY.FK">https://github.com/thegenemyers/MERQURY.FK</a>
MitoHiFi	2	<a href="https://github.com/marcelauliano/MitoHiFi">https://github.com/marcelauliano/MitoHiFi</a>
PretextView	0.2	<a href="https://github.com/wtsi-hpag/PretextView">https://github.com/wtsi-hpag/PretextView</a>
purge_dups	1.2.3	<a href="https://github.com/dfguan/purge_dups">https://github.com/dfguan/purge_dups</a>
SALSA	2.2	<a href="https://github.com/salsa-rs/salsa">https://github.com/salsa-rs/salsa</a>
sanger-tol/genomenote	v1.0	<a href="https://github.com/sanger-tol/genomenote">https://github.com/sanger-tol/genomenote</a>
sanger-tol/readmapping	1.1.0	<a href="https://github.com/sanger-tol/readmapping/tree/1.1.0">https://github.com/sanger-tol/readmapping/tree/1.1.0</a>

Further, the Wellcome Sanger Institute employs a process whereby due diligence is carried out proportionate to the nature of the materials themselves, and the circumstances under which they have been/are to be collected and provided for use. The purpose of this is to address and mitigate any potential legal and/or ethical implications of receipt and use of the materials as part of the research project, and to ensure that in doing so we align with best practice wherever possible. The overarching areas of consideration are:

- Ethical review of provenance and sourcing of the material
- Legality of collection, transfer and use (national and international)

Each transfer of samples is further undertaken according to a Research Collaboration Agreement or Material Transfer Agreement entered into by the Darwin Tree of Life Partner, Genome Research Limited (operating as the Wellcome Sanger Institute), and in some circumstances other Darwin Tree of Life collaborators.

### Data availability

European Nucleotide Archive: *Eilema sororcula* (orange footman). Accession number PRJEB46305; <https://identifiers.org/ena.embl/PRJEB46305>. (Wellcome Sanger Institute, 2021)

The genome sequence is released openly for reuse. The *Eilema sororcula* genome sequencing initiative is part of the Darwin Tree of Life (DToL) project. All raw sequence data and the assembly have been deposited in INSDC databases. Raw data and assembly accession identifiers are reported in Table 1.

### Author information

Members of the University of Oxford and Wytham Woods Genome Acquisition Lab are listed here: <https://doi.org/10.5281/zenodo.4789928>.

Members of the Darwin Tree of Life Barcoding collective are listed here: <https://doi.org/10.5281/zenodo.4893703>.

Members of the Wellcome Sanger Institute Tree of Life programme are listed here: <https://doi.org/10.5281/zenodo.4783585>.

Members of Wellcome Sanger Institute Scientific Operations: DNA Pipelines collective are listed here: <https://doi.org/10.5281/zenodo.4790455>.

Members of the Tree of Life Core Informatics collective are listed here: <https://doi.org/10.5281/zenodo.5013541>.

Members of the Darwin Tree of Life Consortium are listed here: <https://doi.org/10.5281/zenodo.4783558>.

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# Open Peer Review

Current Peer Review Status:   

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## Version 1

Reviewer Report 28 February 2024

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 **Marta Vila** 

Universidade da Coruna, A Coruña, Galicia, Spain

This is a fine "genome note". I hope the following comments will help the authors to improve the manuscript.

### BACKGROUND

1. I think many readers will benefit from a sentence where the authors clarify why they use the name *Eilema sororcula* and not *Wittia sororcula* (e.g. Bayarsaikhan et al. 2016).
2. Please indicate that future studies should complement the present work by sequencing a female -preferably from the same population- so that information of the W chromosome is produced.
3. Please indicate if the karyotype of the species was known (if so, does your inferred number of chromosomes match?) or if you are the first researchers to indicate the euploid number of this species. In any case, the manuscript would benefit from briefly discussing how  $2n = 30$  fits within the available karyotypes for *Eilema*/Arctiinae.

References:[1]

### References

1. Bayarsaikhan U, Ju Y, Bae Y: Review of the *Eilema* group of lichen moths from South Korea, with description of one new species (Lepidoptera: Erebidae: Arctiinae: Lithosiini). *Journal of Forestry Research*. 2016; **27** (2): 407-417 [Publisher Full Text](#)

**Is the rationale for creating the dataset(s) clearly described?**

Yes

**Are the protocols appropriate and is the work technically sound?**

Yes

**Are sufficient details of methods and materials provided to allow replication by others?**

Yes

**Are the datasets clearly presented in a useable and accessible format?**

Yes

**Competing Interests:** No competing interests were disclosed.**Reviewer Expertise:** Genetics, Lepidoptera**I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard, however I have significant reservations, as outlined above.**

Reviewer Report 27 February 2024

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**Juan Galarza**

University of Oulu, Oulu, Finland

The study presents the chromosomally complete genome sequence of the Orange Footman moth (*Eilema sororcula*) based on a male specimen collected from Wytham Woods, Oxfordshire, UK. This species historically had a local distribution in the south and east of the UK until the late 20th century, but recent observations indicate a northward spread and a significant increase in both distribution and abundance. The species has also been recorded across Europe and eastwards to Korea and China.

*E. sororcula* is known for its preference for mature woodlands, where its larvae feed on algae and lichens growing on trees.

The chromosomally complete genome sequence was obtained from one male specimen using Pacific Biosciences single-molecule HiFi long reads and 10X Genomics read clouds, with additional scaffolding through chromosome conformation Hi-C data. Manual assembly curation addressed missing joins, mis-joins, and haplotypic duplications. The final assembly comprises 40 sequence scaffolds with a total length of 729.4 Mb, and 99.94% of the sequence assigned to 30 chromosomal-level scaffolds.

The study emphasizes that the genome sequence of *E. sororcula* will contribute to molecular adaptation studies related to lichen-feeding. It adds to the growing dataset of genomic resources for understanding lepidopteran biology. The comprehensive methods employed in the study, including sequencing, assembly, and curation, contribute to the reliability of the genomic data presented. This resource has the potential to further scientific investigations into the ecological and molecular aspects of *E. sororcula* and enhance our understanding of its expanding distribution and ecological adaptations.

The methods employed in this study are appropriate and technically sound. The researchers have followed established protocols, used recognized tools, and provided comprehensive documentation throughout the experimental procedures. The paper adheres to the standards of

genomics research, and the detailed methods section facilitates potential replication by other researchers.

**Is the rationale for creating the dataset(s) clearly described?**

Yes

**Are the protocols appropriate and is the work technically sound?**

Yes

**Are sufficient details of methods and materials provided to allow replication by others?**

Yes

**Are the datasets clearly presented in a useable and accessible format?**

Yes

**Competing Interests:** No competing interests were disclosed.

**Reviewer Expertise:** Bioinformatics, Molecular Ecology

**I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.**

Reviewer Report 30 June 2023

<https://doi.org/10.21956/wellcomeopenres.21742.r61541>

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**Michael Hiller** 

LOEWE-Centre for Translational Biodiversity Genomics (TBG), Senckenberg Nature Research Society, Frankfurt Am Main, Germany

This data note reports a reference-quality genome assembly of *Eilema sororcula*, together with a gene annotation produced by Ensembl. The generated data and methods used are according to the latest standards. The resulting assembly has a very high quality.

I have 3 minor comments:

1. typo: collected from | Wytham Woods
2. "Most (99.94%) of the assembly sequence was assigned to 30 chromosomal-level scaffolds" - Although I understand that these sentences are generated by filling placeholders with the real values, I think the word 'most' does not capture that essentially the entire assembly (99.94%) is in chrom-level scaffolds. I would suggest to write "The vast majority of the assembly" or something similar, here and in the abstract.

3. "While not fully phased, the assembly deposited is of one haplotype. Contigs corresponding to the second haplotype have also been deposited. " - I don't understand this point. The authors likely produced a primary assembly (and provide alternative contigs in the 'alt assembly'), although this is not precisely mentioned in the methods (HiFiasm has different assembly modes). If so, the primary is likely a mix of both haplotypes, and not 'one haplotype' as stated. This should please be clarified.

**Is the rationale for creating the dataset(s) clearly described?**

Yes

**Are the protocols appropriate and is the work technically sound?**

Yes

**Are sufficient details of methods and materials provided to allow replication by others?**

Yes

**Are the datasets clearly presented in a useable and accessible format?**

Yes

**Competing Interests:** No competing interests were disclosed.

**Reviewer Expertise:** Genome assembly / comparative genomics

**I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.**

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