




DATA NOTE

The genome sequence of the Birch Bell, *Epinotia demarniana* (Fischer von Röslerstamm, 1839) [version 1; peer review: awaiting peer review]

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V1 First published: 21 Jun 2023, 8:256
<https://doi.org/10.12688/wellcomeopenres.19574.1>
Latest published: 21 Jun 2023, 8:256
<https://doi.org/10.12688/wellcomeopenres.19574.1>

Abstract

We present a genome assembly from an individual male *Epinotia demarniana* (the Birch Bell; Arthropoda; Insecta; Lepidoptera; Tortricidae). The genome sequence is 735.8 megabases in span. Most of the assembly is scaffolded into 28 chromosomal pseudomolecules, including the Z sex chromosome. The mitochondrial genome has also been assembled and is 15.46 kilobases in length.

Keywords

Epinotia demarniana, Birch Bell, genome sequence, chromosomal, Lepidoptera



This article is included in the [Tree of Life gateway](#).

Open Peer Review

Approval Status AWAITING PEER REVIEW

Any reports and responses or comments on the article can be found at the end of the article.

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Author roles: **Boyes D:** Investigation, Resources; **Hammond J:** Writing – Original Draft Preparation, Writing – Review & Editing;

Competing interests: No competing interests were disclosed.

Grant information: This work was supported by Wellcome through core funding to the Wellcome Sanger Institute (206194) and the Darwin Tree of Life Discretionary Award (218328).

The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

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How to cite this article: Boyes D, Hammond J, University of Oxford and Wytham Woods Genome Acquisition Lab *et al.* **The genome sequence of the Birch Bell, *Epinotia demarniana* (Fischer von Röslerstamm, 1839) [version 1; peer review: awaiting peer review]** Wellcome Open Research 2023, 8:256 <https://doi.org/10.12688/wellcomeopenres.19574.1>

First published: 21 Jun 2023, 8:256 <https://doi.org/10.12688/wellcomeopenres.19574.1>

Species taxonomy

Eukaryota; Metazoa; Eumetazoa; Bilateria; Protostomia; Ecdysozoa; Panarthropoda; Arthropoda; Mandibulata; Pancrustacea; Hexapoda; Insecta; Dicondylia; Pterygota; Neoptera; Endopterygota; Amphiesmenoptera; Lepidoptera; Glossata; Neolepidoptera; Heteroneura; Ditrysia; Apoditrysia; Tortricoidea; Tortricidae; Olethreutinae; Eucosmini; *Epinotia*; *Epinotia demarniana* (Fischer von Röslerstamm, 1839) (NCBI:txid1594294).

Background

Epinotia demarniana (Fischer von Röslerstamm, 1839) is a moth of the Tortricidae family. The adult moths have an attractive pied appearance, showing little variation in colour or pattern (Bradley *et al.*, 1979). In Great Britain, the adults are at large between June and July, flying high around treetops in the evening. By day the adult moths rest high up amongst foliage and can be caught by shaking or kicking suitable trees, then catching the adult moths as they flutter to the ground (Bradley *et al.*, 1979; Elliott *et al.*, 2018). After oviposition larvae begin feeding in September. The larval stages feed within the catkins of *Alnus*, *Salix caprea*, and *Betula* until May, subsequently pupating in a cocoon amongst leaf litter until June (Bradley *et al.*, 1979; Elliott *et al.*, 2018).

This species frequents heaths and open woodland, as well as wetter habitats such as fens, river banks, or boggy moorland (Bradley *et al.*, 1979). The species is broadly distributed across England and into Wales, but is absent from Scotland and Ireland (Elliott *et al.*, 2018). The species is regarded within the British Isles as being local or scarce. Globally the species occurs across northern Eurasia in a possibly continuous distribution stretching from Great Britain at its western extremity to Hokkaido in the east (GBIF Secretariat, 2022; Hancock *et al.*, 2015). The species has also been recorded from eastern Russia and north-eastern China (Byun *et al.*, 2008), suggesting a broad distribution at the eastern edge of its range.

The genome of *Epinotia demarniana* was sequenced as part of the Darwin Tree of Life Project, a collaborative effort to sequence all named eukaryotic species in the Atlantic Archipelago of Britain and Ireland. Here we present a chromosomally complete genome sequence for *E. demarniana*, based on one male specimen from Wytham Woods, Oxfordshire, UK.

Genome sequence report

The genome was sequenced from one male *Epinotia demarniana* (Figure 1) collected from Wytham Woods, Oxfordshire (biological vice-county Berkshire), UK (51.77, -1.34). A total of 30-fold coverage in Pacific Biosciences single-molecule HiFi long reads was generated. Primary assembly contigs were scaffolded with chromosome conformation Hi-C data. Manual assembly curation corrected 22 missing joins or mis-joins and removed 6 haplotypic duplications, reducing the assembly length by 0.2% and the scaffold number by 12.5%.

The final assembly has a total length of 735.8 Mb in 42 sequence scaffolds with a scaffold N50 of 26.9 Mb (Table 1). Most (99.85%) of the assembly sequence was assigned to 28



Figure 1. Photograph of the *Epinotia demarniana* (ilEpiDema1) specimen used for genome sequencing.

chromosomal-level scaffolds, representing 27 autosomes and the Z sex chromosome. Chromosome-scale scaffolds confirmed by the Hi-C data are named in order of size (Figure 2–Figure 5; Table 2). While not fully phased, the assembly deposited is of one haplotype. Contigs corresponding to the second haplotype have also been deposited. The mitochondrial genome was also assembled and can be found as a contig within the multifasta file of the genome submission.

The estimated Quality Value (QV) of the final assembly is 63.6 with *k*-mer completeness of 100%, and the assembly has a BUSCO v5.3.2 completeness of 98.4% (single = 97.7%, duplicated = 0.7%), using the lepidoptera_odb10 reference set (*n* = 5,286).

Metadata for specimens, spectral estimates, sequencing runs, contaminants and pre-curation assembly statistics can be found at <https://links.tol.sanger.ac.uk/species/1594294>.

Methods

Sample acquisition and nucleic acid extraction

Two *Epinotia demarniana* specimens (specimen ID Ox001904, ilEpiDema1 and specimen ID Ox001904, ilEpiDema2) were collected from Wytham Woods, Oxfordshire (biological vice-county Berkshire), UK (latitude 51.77, longitude -1.34) on 2021-06-16 using a light trap. The specimens were collected and identified by Douglas Boyes (University of Oxford) and were then preserved on dry ice.

DNA was extracted at the Tree of Life laboratory, Wellcome Sanger Institute (WSI). The ilEpiDema1 sample was weighed and dissected on dry ice with tissue set aside for Hi-C sequencing. Whole organism tissue was disrupted using a Nippi Powermasher fitted with a BioMasher pestle. High molecular weight (HMW) DNA was extracted using the Qiagen MagAttract HMW DNA extraction kit. HMW DNA was sheared into an average fragment size of 12–20 kb in a Megaruptor 3 system with speed setting 30. Sheared DNA was purified by solid-phase reversible immobilisation using AMPure PB beads with

Table 1. Genome data for *Epinotia demarniana*, ilEpiDema1.1.

| Project accession data | | |
|--|--|-----------------------------------|
| Assembly identifier | ilEpiDema1.1 | |
| Species | <i>Epinotia demarniana</i> | |
| Specimen | ilEpiDema1 | |
| NCBI taxonomy ID | 1594294 | |
| BioProject | PRJEB54049 | |
| BioSample ID | SAMEA10979166 | |
| Isolate information | ilEpiDema1, male, whole organism (DNA sequencing) ilEpiDema2 (Hi-C scaffolding) | |
| Assembly metrics* | | Benchmark |
| Consensus quality (QV) | 63.6 | ≥ 50 |
| <i>k</i> -mer completeness | 100% | ≥ 95% |
| BUSCO** | C:98.4%[S:97.7%,D:0.7%], F:0.4%,M:1.2%,n:5,286 | C ≥ 95% |
| Percentage of assembly mapped to chromosomes | 99.85% | ≥ 95% |
| Sex chromosomes | Z chromosome | <i>localised homologous pairs</i> |
| Organelles | Mitochondrial genome assembled | <i>complete single alleles</i> |
| Raw data accessions | | |
| PacificBiosciences SEQUEL II | ERR9924611 | |
| Hi-C Illumina | ERR9930686 | |
| Genome assembly | | |
| Assembly accession | GCA_945867215.1 | |
| Accession of alternate haplotype | GCA_945869435.1 | |
| Span (Mb) | 735.8 | |
| Number of contigs | 109 | |
| Contig N50 length (Mb) | 13.5 | |
| Number of scaffolds | 42 | |
| Scaffold N50 length (Mb) | 26.9 | |
| Longest scaffold (Mb) | 64.2 | |

* Assembly metric benchmarks are adapted from column VGP-2020 of “Table 1: Proposed standards and metrics for defining genome assembly quality” from (Rhie *et al.*, 2021).

** BUSCO scores based on the lepidoptera_odb10 BUSCO set using v5.3.2. C = complete [S = single copy, D = duplicated], F = fragmented, M = missing, n = number of orthologues in comparison. A full set of BUSCO scores is available at <https://blobtoolkit.genomehubs.org/view/ilEpiDema1.1/dataset/CAMBMD01/>

a 1.8X ratio of beads to sample to remove the shorter fragments and concentrate the DNA sample. The concentration of the sheared and purified DNA was assessed using a Nanodrop spectrophotometer and Qubit Fluorometer and Qubit dsDNA High Sensitivity Assay kit. Fragment size distribution was evaluated by running the sample on the FemtoPulse system.

Sequencing

Pacific Biosciences HiFi circular consensus and DNA sequencing libraries were constructed according to the manufacturers’ instructions. DNA sequencing was performed by the Scientific Operations core at the WSI on a Pacific Biosciences SEQUEL II (HiFi) instrument. Hi-C data were also generated from

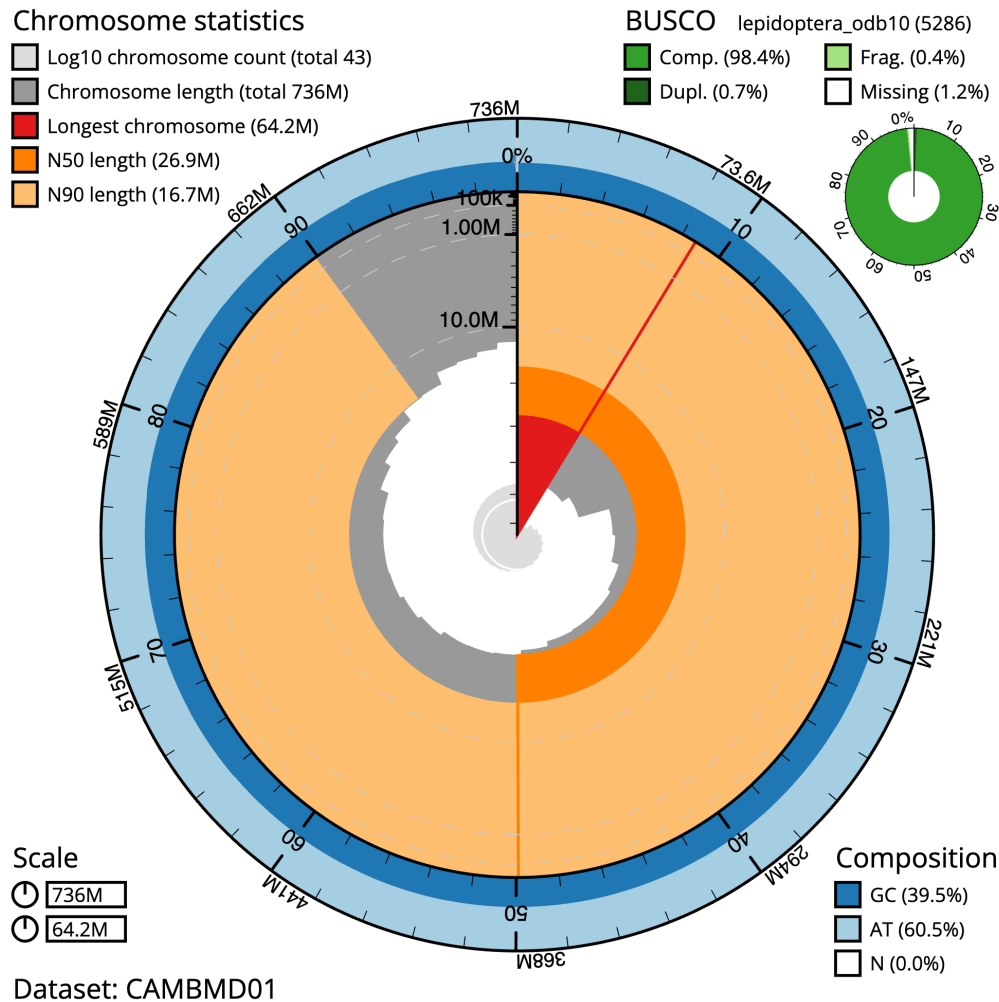


Figure 2. Genome assembly of *Epinotia demarniana*, iEpiDema1.1: metrics. The BlobToolKit Snailplot shows N50 metrics and BUSCO gene completeness. The main plot is divided into 1,000 size-ordered bins around the circumference with each bin representing 0.1% of the 735,811,797 bp assembly. The distribution of scaffold lengths is shown in dark grey with the plot radius scaled to the longest scaffold present in the assembly (64,174,388 bp, shown in red). Orange and pale-orange arcs show the N50 and N90 scaffold lengths (26,883,300 and 16,660,370 bp), respectively. The pale grey spiral shows the cumulative scaffold count on a log scale with white scale lines showing successive orders of magnitude. The blue and pale-blue area around the outside of the plot shows the distribution of GC, AT and N percentages in the same bins as the inner plot. A summary of complete, fragmented, duplicated and missing BUSCO genes in the lepidoptera_odb10 set is shown in the top right. An interactive version of this figure is available at <https://blobtoolkit.genomehubs.org/view/iEpiDema1.1/dataset/CAMBMD01/snail>.

whole organism tissue of iEpiDema2 using the Arima2 kit and sequenced on the Illumina NovaSeq 6000 instrument.

Genome assembly, curation and evaluation

Assembly was carried out with Hifiasm (Cheng *et al.*, 2021) and haplotypic duplication was identified and removed with purge_dups (Guan *et al.*, 2020). The assembly was then scaffolded with Hi-C data (Rao *et al.*, 2014) using YaHS (Zhou *et al.*, 2023). The assembly was checked for contamination and corrected as described previously (Howe *et al.*, 2021). Manual curation was performed using HiGlass (Kerpedjiev *et al.*, 2018) and Pretext (Harry, 2022). The mitochondrial genome was

assembled using MitoHiFi (Uliano-Silva *et al.*, 2022), which runs MitoFinder (Allio *et al.*, 2020) or MITOS (Bernt *et al.*, 2013) and uses these annotations to select the final mitochondrial contig and to ensure the general quality of the sequence.

A Hi-C map for the final assembly was produced using bwa-mem2 (Vasimuddin *et al.*, 2019) in the Cooler file format (Abdennur & Mirny, 2020). To assess the assembly metrics, the *k*-mer completeness and QV consensus quality values were calculated in Merqury (Rhie *et al.*, 2020). This work was done using Nextflow (Di Tommaso *et al.*, 2017) DSL2 pipelines “sanger-tol/readmapping” (Surana *et al.*, 2023a) and

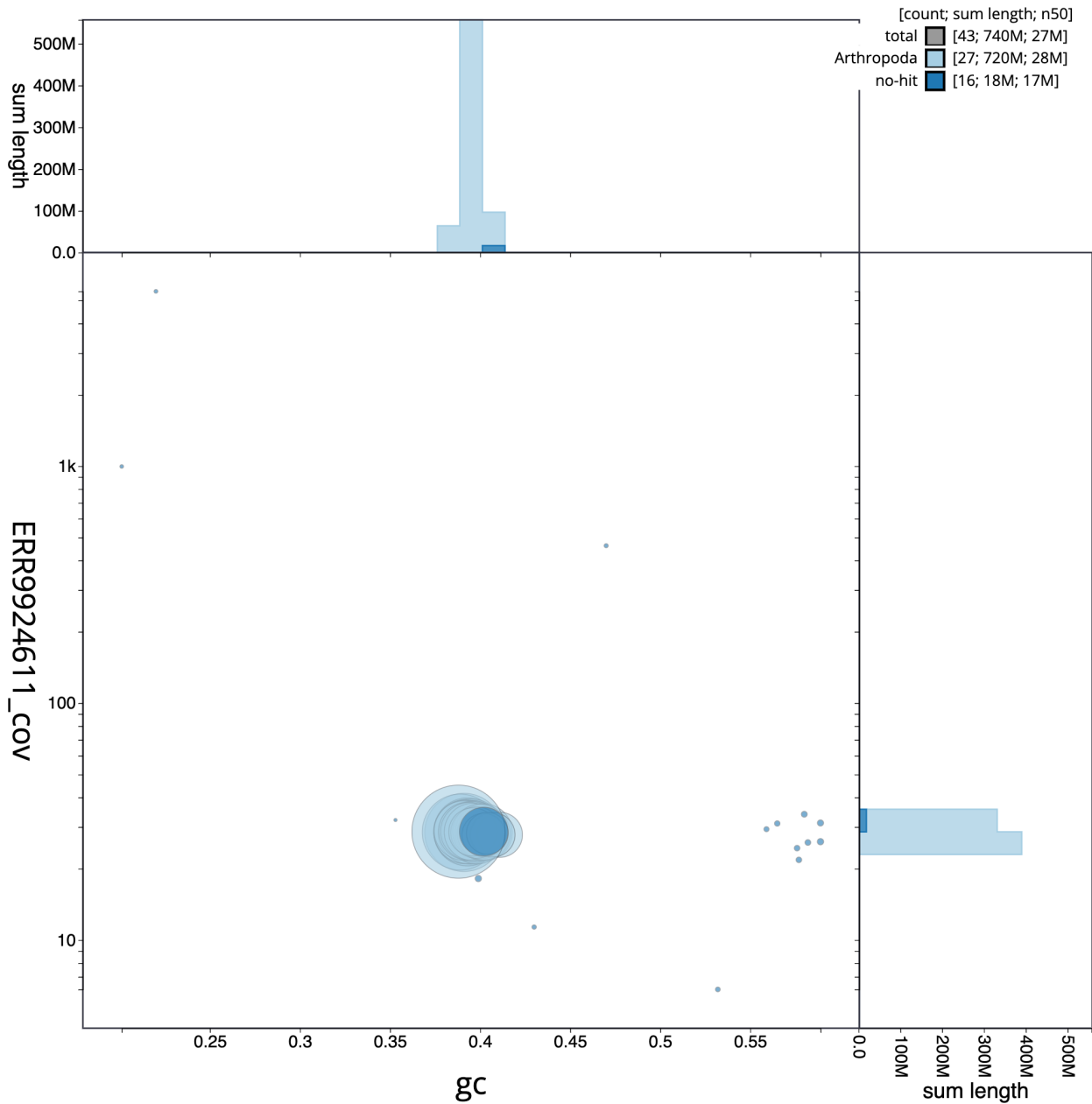


Figure 3. Genome assembly of *Epinotia demarniana*, ilEpiDema1.1: BlobToolKit GC-coverage plot. Scaffolds are coloured by phylum. Circles are sized in proportion to scaffold length. Histograms show the distribution of scaffold length sum along each axis. An interactive version of this figure is available at <https://blobtoolkit.genomehubs.org/view/ilEpiDema1.1/dataset/CAMBMD01/blob>.

“sanger-tol/genomenote” (Surana *et al.*, 2023b). The genome was analysed within the BlobToolKit environment (Challis *et al.*, 2020) and BUSCO scores (Manni *et al.*, 2021; Simão *et al.*, 2015) were calculated.

Table 3 contains a list of relevant software tool versions and sources.

Wellcome Sanger Institute – Legal and Governance
 The materials that have contributed to this genome note have been supplied by a Darwin Tree of Life Partner. The submission of materials by a Darwin Tree of Life Partner is subject to the ‘Darwin Tree of Life Project Sampling Code of Practice’, which can be found in full on the Darwin Tree of Life website [here](#). By agreeing with and signing up to the

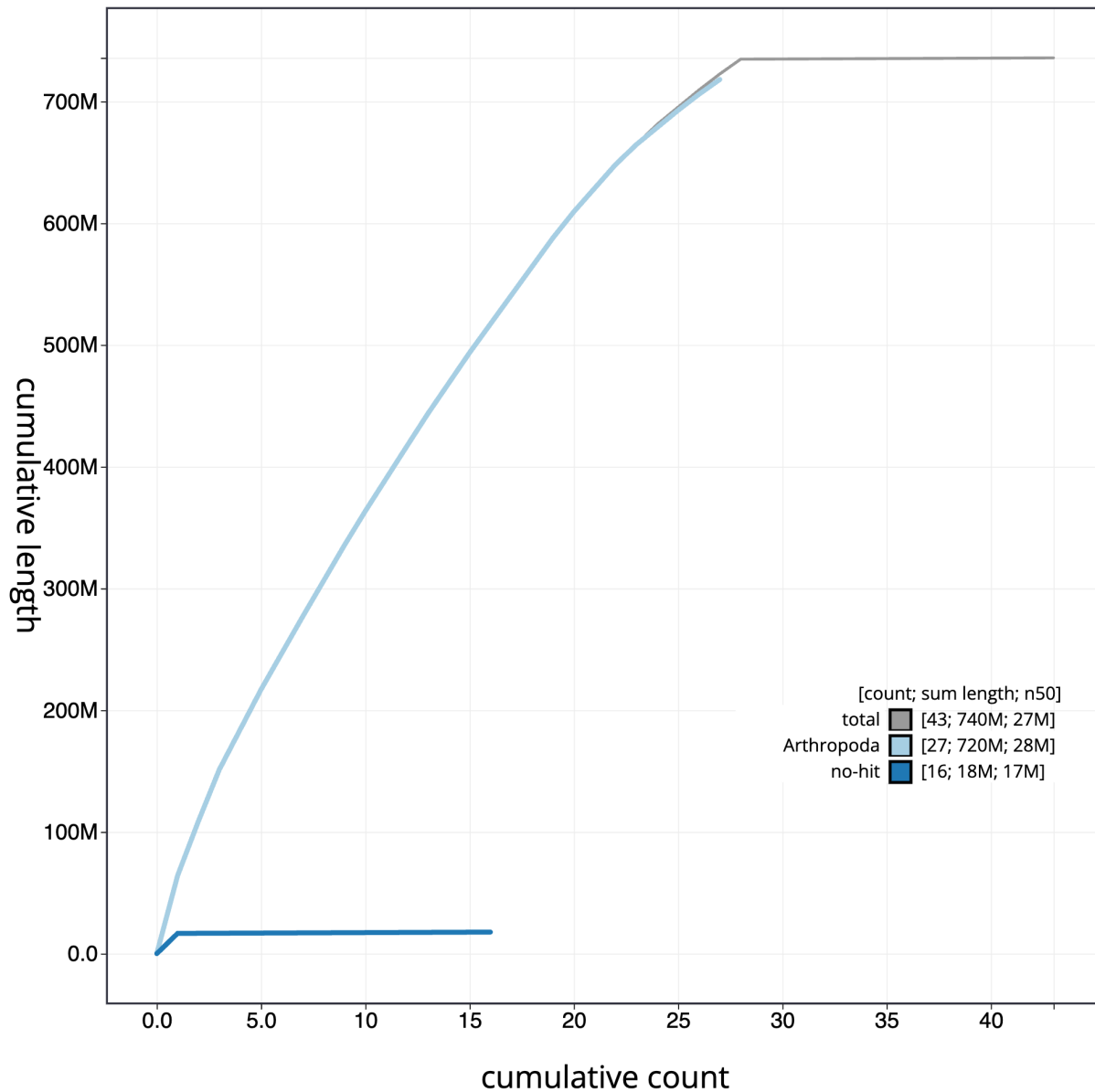


Figure 4. Genome assembly of *Epinotia demarniana*, ilEpiDema1.1: BlobToolKit cumulative sequence plot. The grey line shows cumulative length for all scaffolds. Coloured lines show cumulative lengths of scaffolds assigned to each phylum using the buscodegenes taxrule. An interactive version of this figure is available at <https://blobtoolkit.genomehubs.org/view/ilEpiDema1.1/dataset/CAMBMD01/cumulative>.

Sampling Code of Practice, the Darwin Tree of Life Partner agrees they will meet the legal and ethical requirements and standards set out within this document in respect of all samples acquired for, and supplied to, the Darwin Tree of Life Project.

Further, the Wellcome Sanger Institute employs a process whereby due diligence is carried out proportionate to the nature of the materials themselves, and the circumstances under which

they have been/are to be collected and provided for use. The purpose of this is to address and mitigate any potential legal and/or ethical implications of receipt and use of the materials as part of the research project, and to ensure that in doing so we align with best practice wherever possible. The overarching areas of consideration are:

- Ethical review of provenance and sourcing of the material

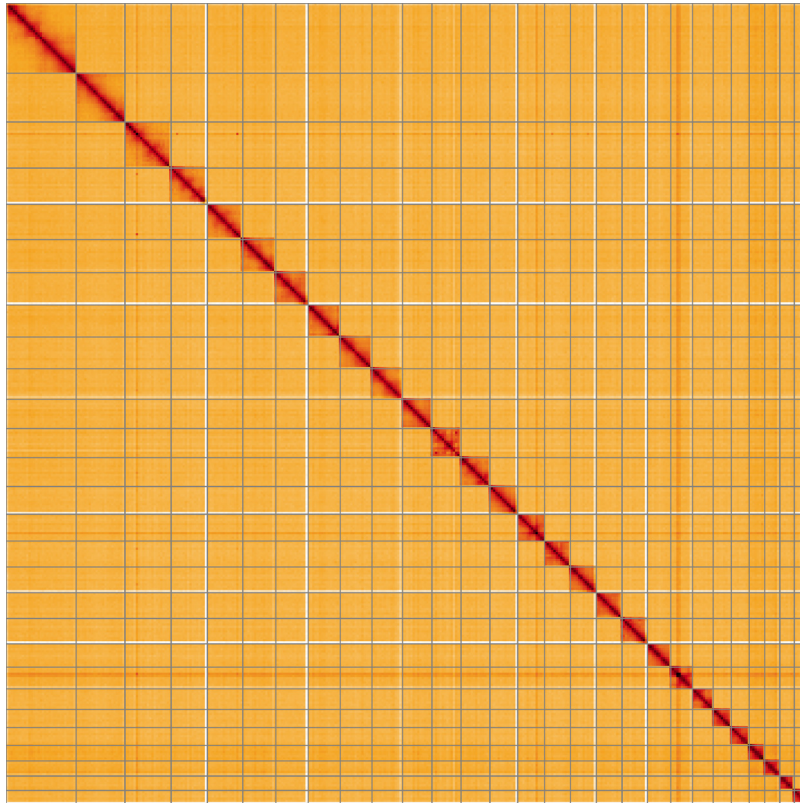


Figure 5. Genome assembly of *Epinotia demarniana*, ilEpiDema1.1: Hi-C contact map of the ilEpiDema1.1 assembly, visualised using HiGlass. Chromosomes are shown in order of size from left to right and top to bottom. An interactive version of this figure may be viewed at <https://genome-note-higlass.tol.sanger.ac.uk/?d=ItZbq6kISSzVWjNKIciQ>.

Table 2. Chromosomal pseudomolecules in the genome assembly of *Epinotia demarniana*, ilEpiDema1.

| INSDC accession | Chromosome | Length (Mb) | GC% |
|-----------------|------------|-------------|------|
| OX244260.1 | 1 | 44.8 | 39.0 |
| OX244261.1 | 2 | 42.27 | 39.0 |
| OX244262.1 | 3 | 33.22 | 39.5 |
| OX244263.1 | 4 | 32.51 | 39.5 |
| OX244264.1 | 5 | 30.44 | 39.5 |
| OX244265.1 | 6 | 29.56 | 39.0 |
| OX244266.1 | 7 | 29.39 | 39.5 |
| OX244267.1 | 8 | 29.17 | 39.0 |
| OX244268.1 | 9 | 27.99 | 39.5 |
| OX244269.1 | 10 | 26.88 | 39.5 |
| OX244270.1 | 11 | 26.63 | 40.0 |
| OX244271.1 | 12 | 26.52 | 39.5 |
| OX244272.1 | 13 | 25.44 | 39.0 |
| OX244273.1 | 14 | 24.64 | 39.5 |

| INSDC accession | Chromosome | Length (Mb) | GC% |
|-----------------|------------|-------------|------|
| OX244274.1 | 15 | 23.82 | 39.5 |
| OX244275.1 | 16 | 23.64 | 39.5 |
| OX244276.1 | 17 | 23.61 | 39.5 |
| OX244277.1 | 18 | 23.32 | 40.0 |
| OX244278.1 | 19 | 21.41 | 40.0 |
| OX244279.1 | 20 | 19.94 | 40.0 |
| OX244280.1 | 21 | 18.8 | 40.0 |
| OX244281.1 | 22 | 16.66 | 40.0 |
| OX244282.1 | 23 | 16.44 | 39.5 |
| OX244283.1 | 24 | 14.23 | 41.0 |
| OX244284.1 | 25 | 13.91 | 40.5 |
| OX244285.1 | 26 | 13.19 | 40.0 |
| OX244286.1 | 27 | 12.14 | 40.5 |
| OX244259.1 | Z | 64.17 | 39.0 |
| OX244287.1 | MT | 0.02 | 20.0 |

Table 3. Software tools: versions and sources.

| Software tool | Version | Source |
|------------------------|------------------|---|
| BlobToolKit | 4.0.7 | https://github.com/blobtoolkit/blobtoolkit |
| BUSCO | 5.3.2 | https://gitlab.com/ezlab/busco |
| Hifiasm | 0.16.1-r375 | https://github.com/chhylp123/hifiasm |
| HiGlass | 1.11.6 | https://github.com/higlass/higlass |
| Merqury | MerquryFK | https://github.com/thegenemyers/MERQURY.FK |
| MitoHiFi | 2 | https://github.com/marcelauliano/MitoHiFi |
| PretextView | 0.2 | https://github.com/wtsi-hpag/PretextView |
| purge_dups | 1.2.3 | https://github.com/dfguan/purge_dups |
| sanger-tol/genomenote | v1.0 | https://github.com/sanger-tol/genomenote |
| sanger-tol/readmapping | 1.1.0 | https://github.com/sanger-tol/readmapping/tree/1.1.0 |
| YaHS | yahs-1.1.91eebc2 | https://github.com/c-zhou/yahs |

- Legality of collection, transfer and use (national and international).

Each transfer of samples is further undertaken according to a Research Collaboration Agreement or Material Transfer Agreement entered into by the Darwin Tree of Life Partner, Genome Research Limited (operating as the Wellcome Sanger Institute), and in some circumstances other Darwin Tree of Life collaborators.

Data availability

European Nucleotide Archive: *Epinotia demarniana* (birch bell). Accession number [PRJEB54049](https://identifiers.org/ena.embl/PRJEB54049); <https://identifiers.org/ena.embl/PRJEB54049>. (Wellcome Sanger Institute, 2022)

The genome sequence is released openly for reuse. The *Epinotia demarniana* genome sequencing initiative is part of the Darwin Tree of Life (DTOL) project. All raw sequence data and the assembly have been deposited in INSDC databases. The genome will be annotated using available RNA-Seq data and presented through the [Ensembl](#) pipeline at the European

Bioinformatics Institute. Raw data and assembly accession identifiers are reported in [Table 1](#).

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Members of the Darwin Tree of Life Consortium are listed here: <https://doi.org/10.5281/zenodo.4783558>.

References

- Abdennur N, Mirny LA: **Cooler: Scalable storage for Hi-C data and other genomically labeled arrays.** *Bioinformatics.* 2020; **36**(1): 311–316. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- Allio R, Schomaker-Bastos A, Romiguier J, et al.: **MitoFinder: Efficient automated large-scale extraction of mitogenomic data in target enrichment phylogenomics.** *Mol Ecol Resour.* 2020; **20**(4): 892–905. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- Bernt M, Donath A, Jühling F, et al.: **MITOS: Improved *de novo* metazoan mitochondrial genome annotation.** *Mol Phylogenet Evol.* 2013; **69**(2): 313–319. [PubMed Abstract](#) | [Publisher Full Text](#)
- Bradley JD, Tremewan WD, Smith A: **British Tortricoid Moths – Tortricidae: Olethreutinae.** The Ray Society, 1979. [Reference Source](#)
- Byun BK, Yan S, Lee BW, et al.: **Four newly recorded species of the genus *Epinotia* Hübner (Lepidoptera: Tortricidae) from Northeast China.** *J Asia Pac Entomol.* 2008; **11**(4): 181–184. [Publisher Full Text](#)
- Challis R, Richards E, Rajan J, et al.: **BlobToolKit - Interactive Quality Assessment of Genome Assemblies.** *G3 (Bethesda).* 2020; **10**(4): 1361–1374. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- Cheng H, Concepcion GT, Feng X, et al.: **Haplotype-resolved *de novo* assembly using phased assembly graphs with hifiasm.** *Nat Methods.* 2021;

18(2): 170–175.

[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)

Di Tommaso P, Chatzou M, Floden EW, *et al.*: **Nextflow enables reproducible computational workflows.** *Nat Biotechnol.* 2017; **35**(4): 316–319.

[PubMed Abstract](#) | [Publisher Full Text](#)

Elliott B, Langmaid JR, Palmer RM, *et al.*: **Tortricidae.** In: Langmaid, J. R., Palmer, S. M., and Young, M. R. (eds.) *A Field Guide to the Smaller Moths of Great Britain and Ireland.* The British Entomological and Natural History Society, 2018; **279**.

GBIF Secretariat: **Epinotia demarniana (Fischer von Röslerstamm, 1839).** Checklist dataset, *GBIF Backbone Taxonomy, 2022*; [Accessed 14 December 2022].

[Publisher Full Text](#)

Guan D, McCarthy SA, Wood J, *et al.*: **Identifying and removing haplotypic duplication in primary genome assemblies.** *Bioinformatics.* 2020; **36**(9): 2896–2898.

[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)

Hancock F, Bland KP, Razowski J: **The Moths and Butterflies of Great Britain and Ireland, Volume 5 (Part 2).** Leiden: Brill, 2015.

[Reference Source](#)

Harry E: **PretextView (Paired REad TEXTure Viewer): A desktop application for viewing pretext contact maps.** 2022; [Accessed 19 October 2022].

[Reference Source](#)

Howe K, Chow W, Collins J, *et al.*: **Significantly improving the quality of genome assemblies through curation.** *GigaScience.* Oxford University Press, 2021; **10**(1): gaa153.

[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)

Kerpedjiev P, Abdennur N, Lekschas F, *et al.*: **HiGlass: Web-based visual exploration and analysis of genome interaction maps.** *Genome Biol.* 2018; **19**(1): 125.

[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)

Manni M, Berkeley MR, Seppely M, *et al.*: **BUSCO Update: Novel and Streamlined Workflows along with Broader and Deeper Phylogenetic Coverage for Scoring of Eukaryotic, Prokaryotic, and Viral Genomes.** *Mol Biol Evol.* 2021; **38**(10): 4647–4654.

[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)

Rao SSP, Huntley MH, Durand NC, *et al.*: **A 3D map of the human genome at kilobase resolution reveals principles of chromatin looping.** *Cell.* 2014; **159**(7): 1665–1680.

[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)

Rhie A, McCarthy SA, Fedrigo O, *et al.*: **Towards complete and error-free genome assemblies of all vertebrate species.** *Nature.* 2021; **592**(7856): 737–746.

[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)

Rhie A, Walenz BP, Koren S, *et al.*: **Mercury: Reference-free quality, completeness, and phasing assessment for genome assemblies.** *Genome Biol.* 2020; **21**(1): 245.

[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)

Simão FA, Waterhouse RM, Ioannidis P, *et al.*: **BUSCO: assessing genome assembly and annotation completeness with single-copy orthologs.** *Bioinformatics.* 2015; **31**(19): 3210–3212.

[PubMed Abstract](#) | [Publisher Full Text](#)

Surana P, Muffato M, Qi G: **sanger-tol/readmapping: sanger-tol/readmapping v1.1.0 - Hebridean Black (1.1.0).** Zenodo. 2023a.

[Publisher Full Text](#)

Surana P, Muffato M, Sadasivan Baby C: **sanger-tol/genomenote v1.0.dev (v1.0.dev).** Zenodo. 2023b; [Accessed 17 April 2023].

[Publisher Full Text](#)

Uliano-Silva M, Ferreira JGRN, Krashenninnikova K, *et al.*: **MitoHiFi: a python pipeline for mitochondrial genome assembly from PacBio High Fidelity reads.** *BioRxiv.* 2022.

[Publisher Full Text](#)

Vasimuddin Md, Misra S, Li H, *et al.*: **Efficient Architecture-Aware Acceleration of BWA-MEM for Multicore Systems.** In: *2019 IEEE Int Parallel Distrib Process Symp (IPDPS).* IEEE, 2019; 314–324.

[Publisher Full Text](#)

Wellcome Sanger Institute: **The genome sequence of the Birch Bell, *Epinotia demarniana* (Fischer von Röslerstamm, 1839).** European Nucleotide Archive. [dataset], accession number PRJEB54049, 2022.

Zhou C, McCarthy SA, Durbin R: **YaHS: yet another Hi-C scaffolding tool.** *Bioinformatics.* 2023; **39**(1): btac808.

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