PAPER • OPEN ACCESS

Screening of significant factors affecting pravastatin production by *Penicillium* sp. ESF21P

To cite this article: Norazwina Zainol et al 2020 IOP Conf. Ser.: Mater. Sci. Eng. 736 022087

View the article online for updates and enhancements.

IOP Conf. Series: Materials Science and Engineering 736 (2020) 022087 doi:10.1088/1757-899X/736/2/022087

Screening of significant factors affecting pravastatin production by Penicillium sp. ESF21P

Norazwina Zainol¹, Emine Seydametova^{1,2}, Jailani Salihon¹ and Peter Convey^{1,3}

¹Faculty of Chemical & Natural Resources Engineering, Universiti Malaysia Pahang, 26300 Gambang, Pahang, Malaysia

²Department of Integrative Biotechnology, College of Biotechnology and Bioengineering, Sungkyunkwan University, Gyeonggi-do16419, Republic of Korea ³British Antarctic Survey, Natural Environment Research Council, Cambridge CB3 0ET, UK

Corresponding author: azwina@ump.edu.my

Abstract. Pravastatin is a clinically useful cholesterol-lowering agent. The development of a one-step fermentation process using pravastatin-producing microfungi may be an attractive approach from an economic point of view. To facilitate this, previously 54 fungal cultures were isolated from soil samples. Among them, Penicillium sp. ESF21P was the most active pravastatin producer (196.83 mg/L). The objective of the present study is to determine significant factors affecting pravastatin production by *Penicillium* sp. ESF21P. The method of the 2⁷⁻³ fractional factorial design with seven variables was performed using Design-Expert 6.0.8 software package. The seven factors studied were slant age, spore concentration, inoculum volume, fermentation time, temperature, initial pH of the medium, and agitation rate. The results obtained confirmed that the factorial model was significant. Amongst the tested factors, only four were important: agitation rate, slant age, initial pH of the medium, and fermentation time with a percentage contribution of 25.66%, 11.56%, 9.72%, and 7.69%, respectively. These significant factors will be optimized further using response surface methodology.

1. Introduction

According to the World Health Organization (WHO), cardiovascular diseases are considered as a leading cause of death both in developed and some developing countries. Pravastatin selectively inhibits 3-hydroxy-3-methylglutaryl-coenzyme A reductase, the regulatory enzyme in cholesterol biosynthesis. Currently, large-scale production of this statin is based on a two-step fermentation process: the initial production of compactin and its microbial hydroxylation to pravastatin [1, 2, 3]. The development of a one-step fermentation process for production of pravastatin could be a more attractive approach.

Previously, our research group examined different fungal strains for lovastatin production [4, 5]. Later, to facilitate the development of a fermentation process for direct production of pravastatin, a number of fungal cultures isolated from both uncultivated and cultivated soils in Pahang State (Malaysia) were investigated for pravastatin production [6, 7]. In the present study, among 54 fungal isolates screened earlier, *Penicillium* sp. ESF21P was the most active pravastatin producer (196.83 mg/L). In order for a potential biotechnological product to become a commercial reality, screening of significant factors affecting its production process is very important. The objective of this study is to determine the significant factors affecting pravastatin production by Penicillium sp. ESF21P using a

Content from this work may be used under the terms of the Creative Commons Attribution 3.0 licence. Any further distribution of this work must maintain attribution to the author(s) and the title of the work, journal citation and DOI. Published under licence by IOP Publishing Ltd 1

IOP Conf. Series: Materials Science and Engineering 736 (2020) 022087 doi:10.1088/1757-899X/736/2/022087

two-level factorial method. The seven factors investigated in this study are slant age, spore concentration, inoculum volume, fermentation time, temperature, initial pH of the medium, and agitation rate.

2. Materials and Methods

2.1 Fungal culture

The fungal strain, *Penicillium* sp. ESF21P, isolated from a soil sample collected from oil palm plantation in Yayasan Pahang Plantation Holding Sdn Bhd (Gambang, Pahang State, Malaysia), was used in this study. This strain was selected during screening as the best pravastatin producer amongst the fungal isolates tested.

2.2 Experimental setup for two-level factorial analysis

For the first step of the optimization study, the method of the 2⁷⁻³ fractional factorial experiments with seven variables, involving a total of 16 experiments, was performed. The seven factors studied were slant age (A), spore concentration (B), inoculum volume (C), fermentation time (D), temperature (E), initial pH of the medium (F), and agitation rate (G). These variables and their lower and upper levels were chosen based on the previous literature data [8-10]. All runs were performed in a random order (overall randomization). The response variable in these screening experiments was the concentration of pravastatin produced by *Penicillium* sp. ESF21P (in mg/L). The Design-Expert 6.0.8 software package (Stat-Ease Inc., USA) was used to analyze the experimental data.

2.3 Fermentation procedure

A spore suspension was prepared by suspending spores from the slant of appropriate age (6-12 d) in 10 mL of sterile distilled water containing 0.01% (v/v) Tween-80. Then, a spore suspension of appropriate value $(10^4-10^8 \text{ spores/mL})$ was added as inoculum, in accordance with the specification in experimental design, into sterile medium reported by Konya et al. [11]: 3% (w/v) glucose, 3% (w/v) glycerol, 0.4% (w/v) peptone, 0.2% (w/v) NaNO₃, 0.1% (w/v) MgSO₄·7H₂O. During the cultivation, pH was not controlled but the initial pH of the medium was set at different values (6.0-8.0) using 10% HCl or 10% NaOH before sterilization. The temperature and agitation rate were set using the control panel of the Ecotron microbiological incubator (Infors HT, Switzerland). The fermentations with medium total volumes of 50 mL were carried out in 250-mL conical flasks at conditions as specified in the experimental design.

2.4 Analytical determination of pravastatin

Ethyl acetate extracts from the whole cell broths obtained according to the extraction procedure of Manzoni et al. [12] were used for HPLC analysis of pravastatin (Agilent 1200; Agilent Technologies, USA). HPLC grade pravastatin (\geq 98% purity, Sigma, USA) was used as a standard.

2.5 Statistical analysis

All the experiments were conducted in triplicate to ensure reproducibility of the results. An analysis of variance (ANOVA) was performed using Design-Expert 6.0.8 software. The percentage contribution was considered in determining the main variables that have a significant influence on pravastatin production (defined as the percentage contribution of a given variable's effect being $\geq 5\%$).

3. Results and Discussion

3.1 Effects of factors on pravastatin accumulation

The goals of the current screening experiments were the investigations of the variables under study that influencing pravastatin production and selection of the important ones on the basis of their effects. Based on literature data, seven factors were chosen as most likely to influence fungal growth and statin production [8-10]. The design comprised of 16 runs without center points, which is acceptable. The levels of variables corresponding to the design of the 2^{33} fractional factorial experiments together

IOP Conf. Series: Materials Science and Engineering 736 (2020) 022087 doi:10.1088/1757-899X/736/2/022087

with the observed response are listed in Table 1. The maximum pravastatin production was found in experimental run 5 and the minimum in experimental run 12.

Run	A:	B:	C:	D:	E:	F:	G:		astatin
	SA	SC	IV	FT	T ° C	pН	AR	concentration	
	(d)	(spores/	(%)	(d)	(°C)		(rpm)	(mg/L)	
		mL)						Actual	Predicted
1	12(+1)	$10^{4}(-1)$	3(-1)	16(+1)	29(+1)	8(+1)	210(-1)	4.92	4.91
2	12(+1)	$10^{4}(-1)$	5(+1)	16(+1)	25(-1)	6(-1)	260(+1)	8.55	9.45
3	6(-1)	$10^{8}(+1)$	3(-1)	16(+1)	29(+1)	6(-1)	260(+1)	5.59	4.69
4	12(+1)	$10^{8}(+1)$	3(-1)	16(+1)	25(-1)	6(-1)	210(-1)	22.31	22.02
5	6(-1)	$10^{4}(-1)$	5(+1)	16(+1)	29(+1)	6(-1)	210(-1)	25.58	25.87
6	6(-1)	$10^{4}(-1)$	3(-1)	16(+1)	25(-1)	8(+1)	260(+1)	3.20	3.80
7	6(-1)	$10^{8}(+1)$	3(-1)	8(-1)	29(+1)	8(+1)	210(-1)	16.12	15.94
8	6(-1)	$10^{4}(-1)$	5(+1)	8(-1)	29(+1)	8(+1)	260(+1)	9.02	8.59
9	6(-1)	$10^{4}(-1)$	3(-1)	8(-1)	25(-1)	6(-1)	210(-1)	13.30	13.17
10	6(-1)	$10^{8}(+1)$	5(+1)	16(+1)	25(-1)	8(+1)	210(-1)	13.51	13.52
11	6(-1)	$10^{8}(+1)$	5(+1)	8(-1)	25(-1)	6(-1)	260(+1)	5.24	5.97
12	12(+1)	$10^{4}(-1)$	5(+1)	8(-1)	25(-1)	8(+1)	210(-1)	1.53	1.71
13	12(+1)	$10^{8}(+1)$	3(-1)	8(-1)	25(-1)	8(+1)	260(+1)	2.55	2.98
14	12(+1)	$10^{4}(-1)$	3(-1)	8(-1)	29(+1)	6(-1)	260(+1)	5.85	5.12
15	12(+1)	$10^{8}(+1)$	5(+1)	8(-1)	29(+1)	6(-1)	210(-1)	3.57	3.69
16	12(+1)	$10^{8}(+1)$	5(+1)	16(+1)	29(+1)	8(+1)	260(+1)	4.41	3.81

Table 1. Experimental design table for a two-level factorial method.

Note: SA, slant age; SC, spore concentration; IV, inoculum volume; FT, fermentation time; T, temperature; pH, initial pH of the medium; AR, agitation rate.

Coded levels of variables are given in the brackets.

Results of pravastatin concentration are presented as mean from three replicates.

Variable	Code	Sum of squares	% Contribution of effect
Slant age	А	89.63	11.56
Spore concentration	В	0.11	0.015
Inoculum volume	С	0.37	0.048
Fermentation time	D	59.64	7.69
Temperature	Е	1.48	0.19
Initial pH of the medium	F	75.39	9.72
Agitation rate	G	199.02	25.66
Interaction AB	AB	32.01	4.13
Interaction AC	AC	66.87	8.62
Interaction AD	AD	31.61	4.08
Interaction AE	AE	86.72	11.18
Interaction AF	AF	22.59	2.91
Interaction AG	AG	74.35	9.59
Interaction ABD	ABD	33.73	4.35

Table 2. The percentage contributions of the main effect of each variable and of the interactive effects between variables on pravastatin accumulation.

Energy Security and Chemical Engineering Congress

IOP Conf. Series: Materials Science and Engineering 736 (2020) 022087 doi:10.1088/1757-899X/736/2/022087

From the experimental data presented in Table 1, the percentage contribution of the main effect of each variable as well as the percentage contribution of the interactive effects between variables were calculated by the Design-Expert 6.0.8 software using Yates' method [13] and are presented in Table 2. As shown in Table 2, among the analyzed variables four were found to be significant: agitation rate, slant age, initial pH of the medium, and fermentation time. Among these four factors, the agitation rate had the highest percentage contribution (25.66%), while the fermentation time had the lowest one (7.69%). The interaction between slant age and temperature (11.18%) also had a considerable influence on pravastatin accumulation.

3.2 Statistical modeling and ANOVA

The pravastatin concentration data (Table 1) as the dependent variable were fitted as a function of the experimental variables using Design-Expert 6.0.8 software in a reduced 3-factor interaction (3FI) factorial model by regression. Based on the experimental data obtained from the 2^{73} fractional factorial experiments, the following equation (1) in terms of the coded variables for pravastatin production was derived:

Pravastatin concentration (mg/L) = $9.08 - 2.37 \times A + 1.93 \times D - 2.17 \times F - 3.53 \times G + 1.41 \times A \times B - 2.04 \times A \times C + 1.41 \times A \times D - 2.33 \times A \times E - 1.19 \times A \times F + 2.16 \times A \times G + 1.45 \times A \times B \times D$ (1),

where: A - slant age; B - spore concentration; C - inoculum volume; D - fermentation time; E - temperature; F - initial pH of the medium; G - agitation rate.

The results of the evaluation of the factorial model by ANOVA are presented in Table 3. The Fvalue of the model (69.10) confirms that it is significant. The data presented in Table 3 justify that amongst the tested variables only four were significant having p < 0.05: slant age (A), fermentation time (D), initial pH of the medium (F), and agitation rate (G). The predicted R^{2} of 0.9162 was in reasonable agreement with the adjusted R^{2} of 0.9804. The adequate precision of 27.695 confirms that the model could be used to navigate the design space. The results of the analysis demonstrated that all model statistics and diagnostic plots are acceptable.

Source	Sum of	Degree of	Mean	F-value	p-value	
	squares	freedom	square		(PROB>F)	
Model	771.55	11	70.14	69.10	0.0005	
А	89.63	1	89.63	88.30	0.0007	
D	59.64	1	59.64	58.75	0.0016	
F	75.39	1	75.39	74.26	0.0010	
G	199.02	1	199.02	196.05	0.0002	
AB	32.01	1	32.01	31.53	0.0049	
AC	66.87	1	66.87	65.87	0.0013	
AE	86.72	1	86.72	85.43	0.0008	
AG	74.35	1	74.35	73.24	0.0010	
ABD	33.73	1	33.73	33.22	0.0045	
Residual	4.06	4	1.02			
Total	775.61	15				
Standard deviation	1.01	R-squared			0.9948	
Mean	9.08	Adjusted R-s	quared		0.9804	
Adequate precision	27.695	Predicted R-s	quared		0.9162	

Table 3. Evaluation of the factorial model using ANOVA.

Note: A, slant age; B, spore concentration; C, inoculum volume; D, fermentation time; E, temperature; F, initial pH of the medium; G, agitation rate.

Energy Security and Chemical Engineering CongressIOP PublishingIOP Conf. Series: Materials Science and Engineering 736 (2020) 022087doi:10.1088/1757-899X/736/2/022087

The obtained results were in a good agreement with literature data reported by other researchers. The speed of agitation is one of the main factors affecting the performance of microbial cells and their productivity [14]. Moreover, according to Atalla *et al.* [15], the agitation rate is a key factor for statin production affecting oxygen supply during submerged fermentation of microbial producer.

From the factorial model derived here (equation 1), it was predicted that decreasing the slant age, initial pH of the medium and agitation rate, and increasing the fermentation time, should enhance pravastatin production. Therefore, the new slant age range chosen was 4-8 d with a new initial pH of 5.0-7.0, while the new agitation rate was in a range of 180-240 rpm. However, the fermentation time should be kept in an initial range of 8-16 d as our preliminary experimental results on the time course of pravastatin production (data not shown) demonstrated that pravastatin production considerably decreased by 16 d of fermentation.

4. Conclusion

The two-level factorial analysis was able to determine significant variables influencing pravastatin production by *Penicillium* sp. ESF21P. It was found that the agitation rate had the highest level of significance for pravastatin accumulation (25.66%) followed by slant age (11.56%), initial pH of the medium (9.72%) and fermentation time (7.69%). These four important factors will be optimized further using response surface methodology.

Acknowledgments

The author wishes to acknowledge the Universiti Malaysia Pahang for funding the project under grant RDU1803119.

References

[1] Manzoni M and Rollini M 2002 Appl. Microbiol. Biotechnol. 58(5) 555–64

[2] Barrios-González J and Miranda RU 2010 Appl. Microbiol. Biotechnol. 85(4) 869-83

[3] Syed MB and Ponnusamy T 2018 Biocatal. Agric.Biotechnol. 13 62-74

[4] Seydametova E, Kambol R and Zainol N **2010** Universiti Malaysia Sabah Biotechnology Symp. IV (Kota Kinabalu) pp 210-215

[5] Seydametova E, Salihon J, Zainol N and Convey P 2012 Int. J. Chem. Eng. Appl. 3(5) 337-339

[6] Seydametova E, Zainol N, Salihon J, and Convey P 2015 Int. J. Ext. Res. 5 79-87

[7] Seydametova E 2015 Int. J. Biosci. Biochem. Bioinforma. 5(2) 80-90

[8] Chakravarti R and Sahai V 2002 Biotechnol. Lett. 24 527-30

[9] Chakravarti R and Sahai V 2002 Process Biochem. 38 481-86

[10] Shaligram S, Singh SK, Singhal RS, Szakacs G and Pandey A **2009** *J. Microbiol. Biotechnol.* **19(7)** 690-97

[11] Konya A, Jekkel A, Suto J and Salat J **1998** *J. Ind. Microbiol. Biotechnol.* **20** 150-52

[12] Manzoni M, Bergomi S, Rollini M and Cavazzoni V 1999 Biotechnol. Lett. 21 253-57

[13] Yates F **1937** *The design and analysis of factorial experiments* (Harpenden: Imperial Bureau of Soil Science) p 95

[14] Shuler ML and Kargi F **2002** *Bioprocess engineering: basic concepts* (New Jersey: Prentice Hall PTR) p 500

[15] Atalla MM, Hamed ER and El-Shami AR 2008 Malays. J. Microbiol. 4(2) 6-10