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1 **Metal Bioavailability Models: Current Status, Lessons Learned, Considerations for Regulatory Use,**
2 **and the Path Forward**

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5 Abstract

6 Since the early 2000s, biotic ligand models and related constructs have been a dominant paradigm
7 for risk assessment of aqueous metals in the environment. We critically review 1) the evidence for
8 the mechanistic approach underlying metal bioavailability models; 2) considerations for the use and
9 refinement of bioavailability-based toxicity models; 3) considerations for the incorporation of metal
10 bioavailability models into environmental quality standards; and 4) some consensus
11 recommendations for developing or applying metal bioavailability models. We note that models
12 developed to date have been particularly challenged to accurately incorporate pH effects because
13 they are unique with multiple possible mechanisms. As such, we doubt it is ever appropriate to lump
14 algae/plant and animal bioavailability models; however, it is often reasonable to lump bioavailability
15 models for animals, although aquatic insects may be an exception. Other recommendations include
16 that data generated for model development should consider equilibrium conditions in exposure
17 designs, including food items in combined waterborne–dietary matched chronic exposures. Some
18 potentially important toxicity-modifying factors are currently not represented in bioavailability
19 models and have received insufficient attention in toxicity testing. Temperature is probably of
20 foremost importance; phosphate is likely important in plant and algae models. Acclimation may
21 result in predictions that err on the side of protection. Striking a balance between comprehensive,
22 mechanistically sound models and simplified approaches is a challenge. If empirical bioavailability
23 tools such as multiple-linear regression models and look-up tables are employed in criteria, they
24 should always be informed qualitatively and quantitatively by mechanistic models. If bioavailability
25 models are to be used in environmental regulation, ongoing support and availability for use of the
26 models in the public domain are essential.

27 EVIDENCE FOR THE MECHANISTIC APPROACH

28 In 1996, the Society of Environmental Toxicology and Chemistry (SETAC) held a workshop to evaluate
29 how regulatory criteria for protecting aquatic life could better reflect the science of metal
30 bioavailability and toxicology (Bergman and Dorward-King 1997). This was followed by an irruption
31 of publications on biotic ligand models (BLMs), related mechanistic or quasi-mechanistic models, and
32 simpler empirical approaches such as multiple linear regression (MLR) models. Although some of
33 these bioavailability models have been incorporated into regulatory frameworks, many jurisdictions
34 retain 1980s vintage criteria. In December 2017, SETAC sponsored a follow-up workshop titled
35 Bioavailability-Based Aquatic Toxicity Models for Metals in Pensacola, Florida, USA. The purpose of
36 the workshop was to consider the status of different modeling approaches for predicting the
37 bioavailability and toxicity of metals in freshwaters and their incorporation into regulatory water
38 quality criteria. This is one of 5 articles that evaluated the performance of the models and sought to
39 identify best practices in the use of these models for developing and applying bioavailability-based
40 criteria, benchmarks, or guidelines for metals that are intended to protect aquatic life (Adams et al.
41 2020; Brix et al. 2020; Garman et al. 2020; Van Genderen et al. 2020).

42 The concept of mechanistic models incorporating metal bioavailability as a key factor governing
43 toxicity (Paquin et al. 2002; Niyogi and Wood 2004) can be traced back to early experimental studies.
44 These studies established that toxicity could vary considerably according to the water chemistry,

45 reflecting influences of factors such as salinity, pH, hardness (i.e., Ca + Mg concentration), alkalinity,
46 and dissolved organic matter (DOM; Jones 1938; Holm-Jensen 1948; Lloyd and Herbert 1962; Zitko
47 et al. 1973). Today we recognize that these factors reflect competition by naturally occurring cations
48 (e.g., Ca^{2+} , Mg^{2+} , Na^+ , H^+) for the binding of free metal cations (Me^{n+}) to ligands on target organisms
49 and complexation of these Me^{n+} ions by waterborne anions (e.g., HCO_3^- , Cl^- , and most importantly
50 DOM). In both cases, the binding of the metal to ligands on the organism is decreased, thereby
51 offering protection. The first regulatory tools incorporating this bioavailability concept proposed
52 different ambient water quality criteria (AWQC) for freshwater and sea water and, in freshwater,
53 applied hardness as the key factor modifying metal toxicity (e.g., Alabaster and Lloyd 1980; US
54 Environmental Protection Agency 1986; Canadian Council of Ministers of the Environment 2007).
55 These approaches were empirically based and either proposed different AWQC for waters in
56 different hardness ranges or else used equations to adjust AWQC for hardness (US Environmental
57 Protection Agency 1986). The latter were forerunners to the current MLR models that incorporate
58 multiple toxicity-modifying factors (Brix et al. 2017a, 2020). In hindsight, it is now clear that hardness
59 was often a surrogate for other water chemistry variables (e.g., alkalinity, specific ions, pH), which
60 may have been equally or more important in the regression data sets that were used to derive these
61 AWQCs and that another crucial water chemistry variable (DOM) was completely overlooked.

62 Pagenkopf (1983) presented the first mechanistic model, the gill surface interaction model (GSIM).
63 This recognized that metals could bind to biological ligands on the respiratory surfaces of target
64 organisms, thereby causing toxicity. The GSIM postulated that toxicity was attributable to free metal
65 ions (Me^{n+}) and used trace metal speciation, gill surface interaction, and competitive inhibition to
66 explain the protective effect of water hardness. The model also recognized that pH and alkalinity
67 influenced metal speciation and that inorganic anions (the role of DOM was curiously discounted)
68 could complex metals, decreasing their bioavailability. These reactions, including those at the gills,
69 were assigned conditional equilibrium constant ($\log K$) values, and steady-state conditions were
70 assumed, allowing prediction of toxicity through equilibrium modeling. Almost simultaneously,
71 Morel (1983) formulated the free ion activity model (FIAM), which focused on algae and made very
72 similar assumptions to the GSIM but in addition recognized the importance of DOM in complexation
73 reactions. Again, a similar geochemical modeling framework was used, and chemical equilibrium was
74 assumed. In both the GSIM and the FIAM, the degree of toxic response was related to the fraction of
75 sites to which Me^{n+} was bound, a concept that became a key component of future models. The GSIM
76 and the FIAM can be considered the parents of modern bioavailability models such as the BLM. In
77 this same era, the advent of geochemical modeling programs (e.g., MINEQL+, MINTEQA2) facilitated
78 further progress. Subsequently, the development of the Windermere Humic Aqueous Model
79 (WHAM) (Tipping 1994) incorporated multisite binding to deal with metal interactions with DOM, an
80 important breakthrough.

81 Pagenkopf (1983) had proposed that the cause of lethality when Me^{n+} bound to critical sites on the
82 gill surface was respiratory toxicity. However, many studies over the next 2 decades demonstrated
83 that the proximate cause of lethality was interference with the active branchial uptake of either Na
84 (Cu, Ag) or Ca (Zn, Cd, Co, Pb) from the water, at least for fish at metal levels causing acute toxicity
85 (Paquin et al. 2002; Niyogi and Wood 2004). These were associated with inhibition of basolateral
86 Na^+ , K^+ -ATPase (for Na uptake) and Ca^{2+} -ATPase (for Ca uptake), as well as blockade of apical Na^+ and
87 Ca^{2+} channels, and were compounded by increased diffusive losses of these major nutrient ions at
88 higher metal concentrations. Thus, different metals targeted different specific sites (transport
89 proteins) on the gills, and measurements of net Na or Ca loss rates to the water or net decreases in
90 plasma or whole-body concentrations provided physiological evidence for this mechanism of
91 toxicity.

92 Subsequently, Playle and colleagues made a major conceptual breakthrough based on experiments
93 with fathead minnow and trout exposed to Cu or Ag (targeting Na transport sites) and Cd (targeting
94 Ca transport sites) in ion-poor synthetic soft water of defined composition (Playle et al. 1993a,
95 1993b; Janes and Playle 1995). Natural DOM decreased the binding of metals to the gills. Through
96 the analysis of gill metal burdens, the use of competitive waterborne ligands with known log K
97 values, Langmuir isotherm analysis, and a geochemical modeling program (MINEQL+), they were
98 able to estimate distinct log K (affinity) and Bmax (site density = capacity) values for these metals at
99 the gills in short-term exposures (2–3 h). Calculated metal accumulation on gills correlated well with
100 measured gill metal concentrations and adverse physiological effects (e.g., Na loss attributable to
101 Ag) in a number of different field-collected waters. These gill accumulation experiments likely reflect
102 correlates to the true accumulation on the “biotic ligand,” for there is a wide range of possible
103 binding sites on a gill surface (including excreted mucus), with a wide spectrum of affinities for
104 different metals. Although it is incorrect to think that by “titrating” a gill surface one can expect to
105 probe and characterize the “biotic ligand,” the strong relationships between short-term gill metal
106 accumulation and toxicity gave operational support to gill-binding modeling.

107 The gill-binding model for Ag (Janes and Playle 1995) was later transformed into a physiologically
108 based BLM by relating the gill Ag burden on trout to the fractional inhibition of gill Na⁺, K⁺-ATPase
109 activity associated with 96-h mortality (McGeer et al. 2000). This work provided the mechanistic step
110 from short-term gill metal accumulation to the proximate cause of acute toxicity. Similar studies by
111 MacRae et al. (1999) with trout more rigorously demonstrated that short-term gill metal
112 accumulation (in this case Cu at 24 h of exposure) was a constant predictor of acute toxicity (in this
113 case percentage of mortality at 120 h) among a range of test media containing different copper-
114 binding ligands. In fathead minnow, Meyer et al. (1999) demonstrated elegantly that the gill burden
115 of Ni at 24 h associated with 50% mortality at 96 h was constant over a range of water qualities,
116 even though the concentration of the free Ni²⁺ ion associated with 50% mortality was not. From
117 these studies arose the concept of the LA50, the short-term accumulation at the biotic ligand that is
118 predictive of 50% mortality at a later time. This is a key component of all BLMs, now more commonly
119 known as the intrinsic sensitivity parameter, which can be varied in model fitting to compensate for
120 differences in sensitivity among species, strains, and clones. The gill log K, Bmax, and LA50 concepts
121 still underpin all modern BLMs. The terminology varies, with the gill accumulation log K values often
122 referred to as biotic ligand log K values because gills are not the sole site of ion exchange in small
123 animals such as cladocerans, and plants obviously do not have gills. Likewise, LA50s can be related to
124 fractional effects other than 50% or to sublethal endpoints, and thus the LA50 term is often replaced
125 by the more general term for a critical accumulation associated with x% effects (CAx). But despite
126 the varied (and sometimes confusing) terminology, the underlying concepts are fundamentally
127 similar.

128 Building on these early results, subsequent investigations have successfully correlated short-term gill
129 metal accumulation with toxic effects in longer-term exposures for a variety of metals (Table 1). For
130 example, Figure 1 shows how measured gill Ag accumulation (first bar in each pair) can be predicted
131 in 2 forms (Ag⁺ and AgCl; second bar) as a function of water chemistry (dissolved organic carbon
132 [DOC], Cl, Na, Ca, pH) using a chemical equilibrium model (Paquin and Di Toro 2008). Toxicity data
133 were then used to evaluate the expected LA50, and the derived BLM could then be used in
134 combination with the LA50 to predict dissolved Ag median lethal concentrations (LC50s) over a wide
135 range of water quality characteristics (Figure 2).

136 *Moving from models based on physiological mechanisms and gill metal burdens to models based on*
137 *toxicity only*

138 A SETAC Pellston Workshop in Pensacola in 1996, and the subsequent book that arose from it
139 (Bergman and Dorward-King 1997), greatly accelerated the pace of BLM development. Thereafter,
140 landmark publications by Paquin et al. (2000), Di Toro et al. (2001), and Santore et al. (2001) laid out
141 the formal technical framework for the BLM and demonstrated its utility in predicting acute toxicity
142 of Cu and Ag to fish (e.g., Figure 2) and invertebrates in a range of natural waters. Although these
143 papers were firmly rooted in the concept that the short-term metal burden on the biotic ligand was
144 the key factor causing longer-term toxicity, they showed that this quantity did not have to be
145 measured but rather could be back-calculated (if required) from toxicity data for the purpose of
146 model generation. The extensive results of Erickson et al. (1996) on acute Cu toxicity to fathead
147 minnow where single water chemistry parameters were varied, one at a time, provided the key data
148 used to illustrate this principle.

149 Relative to the number of BLMs developed since that time, there are relatively few studies where
150 the physiological mechanisms (e.g., gill enzyme inhibition, ion loss) and/or surrogates for the actual
151 metal burden at the biotic ligand (e.g., gill metal concentration) have been measured (see summary
152 in Table 1). This move to model fitting to toxicity data only maintains the conceptual mechanistic
153 framework (that a theoretical critical metal burden at the biotic ligand causes a critical level of
154 toxicity); it has arisen partly as a matter of convenience and partly as a matter of necessity. The
155 former reflects the time-consuming, technically demanding, and costly nature of the measurements,
156 whereas the latter reflects the fact that in many cases the measurements simply cannot be done.
157 Even when metal concentrations in a tissue or an organ are measured, the concentration of metal at
158 the site of toxic action most likely is not the only accumulation being measured. Instead, it is usually
159 assumed that the measured concentration in a tissue or organ is proportional to the currently
160 unmeasurable concentration at the site of action. With Ni, the mechanism of chronic toxicity
161 remains unresolved (Brix et al. 2017b); and indeed for most metals, the mechanism(s) of chronic
162 toxicity, and therefore the target biotic ligands, remains poorly understood. Furthermore, the most
163 sensitive organisms which “drive” AWQC are usually very small (e.g., daphnids, snails, algae), in
164 which the biotic ligands are unknown and the metal burdens difficult to measure. However in
165 daphnids, acute Ag toxicity, active Na uptake inhibition, whole-body Na decrement, whole-body Ag
166 burden, and whole-body Na⁺, K⁺-ATPase inhibition were well correlated (Bianchini and Wood 2003);
167 and chronic Ag toxicity seemed to result from a failure of Na regulation in both fish (Naddy et al.
168 2007) and daphnids (Bianchini and Wood 2002). These data provide some confidence that the
169 conceptual framework remains valid.

170 In approximately 2001, the rate of BLM development was greatly accelerated by the shift from
171 modeling based on gill metal burden/physiological effects to modeling based on toxicity data alone.
172 De Schamphelaere, Janssen, and colleagues have exploited this approach to the greatest extent,
173 particularly for chronic BLMs, which have been in high demand for European regulations (De
174 Schamphelaere and Janssen 2002, 2004a; De Schamphelaere et al. 2005a, 2005b). The chemical
175 submodel remains identical and mechanistic, whereas on the biological/ toxicity side of the model,
176 the implicit mechanistic assumption is that toxicity results faithfully reflect an imaginary metal
177 burden at an imaginary biotic ligand. Studies on chronic (30-d) toxicity of Cu to trout represent one
178 of the rare cases where this assumption has been tested (Ng et al. 2010; Crémazy et al. 2017). These
179 studies concluded that the 24-h gill LA50 predictive of 30-d mortality remained constant from pH 6.0
180 to 8.0 but not at extreme pH values (5.5, 8.5). However, De Schamphelaere et al. (2005b) reported
181 that both surface-bound Cu and internal Cu concentrations were relatively good predictors of
182 chronic toxicity (48–72 h) in algal growth tests across a slightly smaller pH range (5.9–8.5). Possible
183 scenarios for such anomalous effects at extreme pH values are explored in the section *Dealing with*
184 *extreme waters*. Regardless, if the model is calibrated with data in the water chemistry range of

185 interest (and hybrid model strategies for doing so are outlined in the section *Complexity versus*
186 *simplicity: Use of mechanistic and hybrid models to inform development of simpler models*), the
187 model predictions should be reliable.

188 *Incorporating dietary metal exposure into bioavailability models: The importance of equilibration*

189 The question of whether chronic toxicity of metals in aquatic environments is the result of
190 waterborne, dietary, or combined exposures has generated much study and extensive reviews
191 (Clearwater et al. 2002; Meyer et al. 2005; DeForest and Meyer 2015). In nature, ingestion may be a
192 significant route of metal uptake, and for nutrient metals (e.g., Cu, Fe, Zn) it is undoubtedly the
193 major route. Acute models do not take this into account because testing protocols dictate that the
194 organisms must be fasted during the exposure. Because it is almost impossible to envisage a natural
195 situation where dietary metal would cause acute toxicity to aquatic organisms, this is not an issue of
196 concern. However, during chronic exposures, dietary metal may contribute to toxicity or acclimation,
197 and this may occur by both direct (metal poisoning) and indirect (metals affecting the nutritional
198 quality of the diet or causing food aversion) routes (e.g., Irving et al. 2003; Niyogi and Wood 2003;
199 Besser et al. 2005; De Schamphelaere et al. 2007; Golding et al. 2013; Tomczyk et al. 2018). Hook
200 and Fisher (2001, 2002) reported extremely low waterborne effect levels for reproductive
201 impairment when metal-exposed algae (Ag, Hg, Cd, Mn, and Zn) were fed to zooplankton; for Ag,
202 the threshold was below the chronic AWQC. These notable results stimulated subsequent
203 investigations, which confirmed that, in some settings, algae could accumulate metals to harmful
204 levels from low waterborne concentrations (Bielmyer et al. 2006) but, in other settings, metal
205 bioaccumulation occurred without obvious adverse effects (Kolts et al. 2009). Similarly, discordant
206 results were obtained with mayflies fed Cd-exposed algae in repeated experiments (Xie et al. 2010).
207 The reasons for the differing responses are unclear. Further, the literature is not consistent on
208 whether metals incorporated into natural diets by chronic waterborne exposure of the prey
209 organisms are more or less bioavailable than metal salts or prey dipped in metals (DeForest and
210 Meyer 2015).

211 In the real world, we expect that a natural diet will be in some sort of dynamic equilibrium with the
212 metal in the water column. Direct evidence for this is sparse, and metal accumulation studies have
213 shown that time to reach constant tissue burdens ranges from hours for algae to >28 d for predatory
214 insects and oligochaete worms (Timmermans et al. 1992; Stephenson and Turner 1993; Roy and
215 Hare 1999; Meylan et al. 2003). Nevertheless, DeForest and Meyer (2015) argued that “exposure of
216 test organisms to matched water-borne and diet-borne metal concentrations is perhaps the most
217 relevant for evaluating the protectiveness of water-borne metal guidelines.” In this context,
218 “matched” means that the test organisms were exposed to the same waterborne-metal
219 concentration to which its food was exposed. This was also a key recommendation of the 2002
220 SETAC Pellston workshop on this topic (Meyer et al. 2005). However, this has rarely been done.
221 There are 2 aspects to this equilibration: 1) physicochemical equilibration reflecting the slow kinetics
222 of diffusion into and sorption onto the food item, which is mainly a concern when dead organisms or
223 artificial food (e.g., trout pellets) are used as the diet—in this case, the same kinetic constraints as
224 for equilibration of metal with DOM will likely apply, a process that can take 24 h or more (see
225 section Equilibrium issues), and 2) biodynamic equilibration (achievement of constant
226 concentrations) with a live diet, where the prey organisms may concentrate the metal many fold
227 above that in the water column, which may take days to weeks.

228 Two recent studies assessed dietary impacts of Pb with matched waterborne and diet-borne metal
229 exposure concentrations. These examined the interactive effects of waterborne and dietary Pb
230 exposure in daphnids (*Ceriodaphnia dubia*; Nys et al. 2013) and rainbow trout (Alsop et al. 2016) and

231 concluded that dietary Pb exposure to these freshwater organisms may not be of concern under the
232 scenarios tested.

233 At present, there is insufficient evidence to conclude that chronic bioavailability models would be
234 underprotective if based on waterborne-only exposures or on combined exposures with insufficient
235 equilibration. Therefore, this should not be a reason for rejecting the large amount of otherwise
236 high-quality data available for use in model generation or for current models that exist based on
237 such data. However, we recommend, for best practice in the future, that during chronic tests
238 combined waterborne and dietary matched exposures should be performed. These should be based
239 on natural live diets that have undergone full biological equilibration with the waterborne metal
240 through pre-exposure. If it becomes apparent during such tests that whole-body and target organ-
241 specific metal concentrations are not at equilibrium with ambient metal concentrations in water and
242 food, a biodynamic modeling framework that incorporates uptake (via water and food) and
243 elimination kinetics may be needed.

244 *Incorporating behavioral endpoints (e.g., olfaction, mechanoreception) into bioavailability models*

245 In most jurisdictions, mortality, growth inhibition, and reproductive inhibition are the only toxicity
246 endpoints that can be used in a regulatory framework. Nevertheless, there is increasing evidence
247 that disruptions of behavior caused by metal exposure may be equally or more sensitive endpoints
248 and that these disruptions are mediated by disturbances in olfaction and/or mechanoreception
249 (reviewed for many metals in Wood et al. 2012a, 2012b). If organisms cannot navigate properly,
250 sense predators or prey, maintain social hierarchies, or find mates, population impacts will likely
251 occur. Furthermore, the limited information available suggests that mechanisms governing olfactory
252 toxicity are rather different from those governing toxicity for other endpoints. For example, for
253 waterborne Cu, inhibitory effects are almost immediate, Ca provides little protection, the log *K* value
254 for Cu at the olfactory rosette is lower than at the gill, and there is evidence of recovery/acclimation
255 from olfactory inhibition during chronic exposure (e.g., McIntyre et al. 2008; Mirza et al. 2009; Green
256 et al. 2010; Dew et al. 2012). Furthermore, there are different viewpoints (e.g., Green et al. 2010 and
257 Dew et al. 2012 vs Meyer and Adams 2010, DeForest et al. 2011, and Meyer and DeForest 2018) on
258 whether or not mechanistic bioavailability models based on concepts of ionoregulatory disturbance,
259 such as the BLM-based US Cu criterion (US Environmental Protection Agency 2007), are protective
260 against olfactory effects such as behavioral disturbance. If they are protective, it would appear that
261 this is because highly sensitive taxa are included (e.g., cladocerans) for criteria derivation, not
262 because the bioavailability models are mechanistically correct for behavioral endpoints (i.e., the
263 comparison is of apples vs oranges). The matter remains unresolved, but moving forward, as argued
264 by Pyle and Wood (2007), we recommend that mechanistically based bioavailability models for
265 behavioral toxicity should be developed. These should be built from the ground up using behavioral
266 endpoints, rather than by adjusting the intrinsic sensitivity parameter in existing BLMs. The areas of
267 agreement and disagreement with models built on traditional endpoints will then be highly
268 informative, and there will be a stronger foundation for deciding whether models based on
269 behavioral endpoints should be used in environmental regulation.

270 CONSIDERATIONS FOR THE USE AND REFINEMENT OF BIOAVAILABILITY-BASED TOXICITY MODELS

271 *Types of bioavailability-based models currently available*

272 Table 2 presents a representative summary of available models, but a thorough listing of all models
273 would be beyond the scope of this article. The models include classic BLMs that predict acute
274 toxicity based on measured accumulations (Table 1); models fitted to acute and chronic toxicity

275 data; models predicting toxicity using humic acid or surfaces as surrogates for biotic ligands; and
276 “generalized bioavailability models,” which may be as simple as a single-variable regression such as
277 pH against free metal ion toxicity (Table 2). Metal bioavailability models directly fitted to acute or
278 chronic toxicity data such as those in Table 2 are often used to normalize single-species toxicity data
279 to a target water chemistry prior to inputting such data into species-sensitivity distributions for
280 guideline development. The problem of relying on acute models to predict chronic effects is further
281 explored in the Supplemental Data. Many recent models have also extended single-metal
282 approaches to mixtures (Table 2).

283 Some themes become apparent from inspecting different models. All include chemical speciation
284 calculations, which require as inputs at least major ion chemistry (e.g., Ca, Mg, Na, K, Cl, SO₄, and
285 alkalinity or dissolved inorganic carbon), DOC, pH, and temperature. For shorthand, we refer to
286 these as the “BLM” inputs. Some models, in addition, include Al and Fe. Thus, even the generalized
287 bioavailability models (gBAMs), which predict free metal ion toxicity as a function of pH and/or free
288 major cation activities, require the full BLM water chemistry to compute free ion activities. Further,
289 pH is consistently incorporated as an important toxicity-modifying factor, but the direction of
290 responses (i.e., whether an increase or a decrease in pH would increase or decrease the effect
291 concentrations of dissolved metal) often differ between plant and animal models (Figure 4). In
292 addition to affecting speciation, pH affects the bioavailability and toxicity of metals to plants by
293 changing membrane permeability (Boullemant et al. 2009; Lavoie et al. 2012). These are key reasons
294 why it will likely never be feasible to combine plant and animal bioavailability models into a single
295 model. The important but complex role of pH in bioavailability models is discussed in the section
296 *Complexity versus simplicity: Use of mechanistic and hybrid models to inform development of simpler*
297 *models.*

298 *Alternatives to the conventional single-site unidentate BLM*

299 When the BLM was first formulated (Di Toro et al. 2001), the biotic ligand was considered a single
300 unidentate binding site, for reasons of simplicity and lack of support for a more complicated
301 formulation. It was later shown that this formulation dictates a linear relationship between the
302 chemical activity of a competing cation and the x% effect concentration (EC_x) of the toxic metal
303 expressed as free metal ion activity (i.e., EC_x Meⁿ⁺) and that stability constants of competitive
304 cations could be estimated directly from a linear regression (De Schamphelaere and Janssen 2002).
305 This prompted an explosion of studies (both acute and chronic) that estimated log KBL values for
306 various organisms and metals directly from toxicity data, using univariate test designs.

307 However, observed relationships often deviated from perfect linearity. Various potential mechanistic
308 hypotheses have been put forward to explain these deviations, including binding of metal species
309 (e.g., MeOH⁺, MeCO₃) other than the free metal ion (De Schamphelaere et al. 2004), differences
310 between pH in bulk solutions and the organism microenvironment (Playle and Wood 1989), multiple
311 biotic ligand sites relating to multiple uptake sites or multiple simultaneous mechanisms of toxicity
312 (Peters et al. 2011), bidentate biotic ligand sites (Farley and Meyer 2015), and the influence of
313 plasma membrane potential on free metal ion activity at the biotic ligand (Kinraide 2006). In quite a
314 few cases, researchers (including some authors of the present study) have generated models
315 providing good fits and predictive capacities but at the expense of unrealistic parameter estimates
316 (e.g., assuming that MeOH⁺ species are almost equally as bioavailable as the free metal ion [De
317 Schamphelaere and Janssen 2004b; De Schamphelaere et al. 2004]).

318 It is always possible to perform a linear regression and derive classical log K values, often with still
319 reasonably accurate representation of observed bioavailability relations. Yet, an increasing number

320 of observations have shown such strong deviations from linearity that alternative modeling
321 approaches have been pursued. Indeed, some of them have already been implemented in European
322 Union regulations, whereas others have not been implemented. Deviations from linearity appear to
323 be greatest for H^+ ions (i.e., pH), with some very obvious examples where log-linear relationships
324 describe the observations much better than linear regressions (e.g., algae–Cu [De Schampelaere
325 and Janssen 2006]; Daphnia–Ni [Deleebeeck et al. 2008]; Daphnia–Cu [Van Regenmortel et al.
326 2015]). These observations have led to the formulation of “hybrid models,” or gBAMs, that combine
327 a log-linear pH effect (e.g., Figure 5) with the classic competition for other cations. Although this
328 practice can be perceived as less mechanistic than the classic unidentate single-site BLM, it should
329 be emphasized that 1) the BLM is also just a fitted regression when the link between accumulation
330 at a critical site and toxicity is not made (e.g., Figure 3) and 2) various alternative mechanisms may
331 lead to log-linear pH effects on free metal ion toxicity (e.g., Figure 3).

332 The overall message of Figure 3 is that different mechanistic theories and model formulations can
333 generate either approximately linear or log-linear relations. In addition, assuming, for instance, a
334 bidentate binding model, the pKa values can generate a wide range of pH–ECx Me^{n+} slopes (on a log
335 scale) In the absence of mechanistic evidence, such relationships are equivalent to MLRs.
336 Nevertheless, hybrid models which incorporate such regressions into a BLM framework are useful
337 for regulatory purposes, as long as they accurately predict toxicity over a wide range of conditions.

338 A consequence of approaching the BLM as just equations to be solved by using optimization routines
339 to find the best values for unknown parameters is that the fitted solutions may be disconnected
340 from the BLMs' mechanistic foundations. Prominently, a fundamental BLM tenet is that toxicity
341 follows accumulation on the biotic ligand (Table 1). In model construction, if the speciation model
342 and log K values used successfully reproduce measured accumulation values, then the model is
343 grounded in reality. However, if this step is bypassed, such as when appropriate accumulation data
344 are not available, it is possible to successfully fit BLMs to toxicity values using binding constants or
345 LA50 values that appear to be chemically or biologically unrealistic.

346 With Cu and Na, the apparent protective effect of Na^+ against Cu toxicity has been incorporated into
347 BLMs as a competition between Na^+ and Cu^{2+} for binding to the biotic ligand. The binding constant
348 for Na^+ that can be extracted from toxicity experiments tends to produce log K (biotic ligand–Na)
349 values of 2.5 to 3.5 in various BLMs. This agrees well with the common observation that the binding
350 affinity (K_m) values for unidirectional active Na^+ uptake values in most freshwater organisms are in
351 the range of 10–3 M. It is curious therefore that, in contrast, log K (organic acid–Na) stability
352 constants for various organic acids and Na are mostly between 0.7 and 1.9 (Stumm and Morgan
353 1996). This illustrates that organismal biology is more complicated than simple chemistry. Figure 5
354 illustrates a further example, comparing measured and predicted Cd LC50s with rainbow trout from
355 diverse studies using 2 BLM constructs. In Figure 5, model A, the log K Cd–biotic ligand binding
356 affinity coefficient of 8.1 is similar to those derived from gill accumulation experiments (e.g., 8.6
357 [Playle et al. 1993a]; 8.0 [Niyogi et al. 2008]). Yet in Figure 5, model B, even when the Cd–biotic
358 ligand binding affinity is decreased 2 full orders of magnitude (2 log units), the fit of the toxicity data
359 is just as good, so long as the LA50 is reciprocally lowered 2 orders of magnitude to 0.03% (see
360 online Supplemental Data SI-2 in Farley et al. 2015). However, fit aside, both of these LA50 values
361 are lower than experimentally determined values, and an LA50 of 0.03% seems implausible,
362 assuming that binding to the site of toxic action is proportional to binding sites on the gill. An LA50
363 of 0.03% implies that Ca regulation would be fatally compromised with channels that are 99.97%
364 intact, which does not seem physiologically plausible. For comparison, experimentally derived LA50

365 values for Cd in rainbow trout studies have been approximately 10 to 30% of the strong binding sites
366 on the gill (Birceanu et al. 2008; Niyogi et al. 2008).

367 The reason for these modeling manipulations was to try to find a combination of parameters that
368 would mimic tests that showed that Cd toxicity was reduced by adding Cu, implying that Cu may
369 have a higher affinity than Cd to the biotic ligand. Although ungrounded from accumulation, such
370 manipulations have shown practical success across complex and varied exposure conditions (Farley
371 et al. 2015). The capacity of the BLM structure to use widely available toxicity data to predict
372 responses over a wide combination of waters, metals, and organisms is a major strength of the
373 approach. However, too much flexibility from many adjustable parameters can lead to
374 unconstrained models that would more accurately be described as “mechanistically inspired”
375 models, rather than “mechanistic” models.

376 *Complexity versus simplicity: Use of mechanistic and hybrid models to inform development of simpler*
377 *models*

378 The broad influence of metal bioavailability models within science communities has not always
379 translated to their broad adoption by regulatory authorities (e.g., Wood et al. 2012a, 2012b). The
380 necessity of determining the full composition of each water sample and the complexity underlying
381 bioavailability-based models has proven to be a limitation for their wide application. To broaden
382 access to potential users, various functional interfaces and simplified versions have been developed.
383 In the early 2000s, Robert Santore and colleagues developed and freely shared an intuitive,
384 spreadsheet-style software interface that executed the BLM structure developed by Di Toro et al.
385 (2001) and Santore et al. (2001). The software provided users a flexible platform to explore toxicity
386 data and water chemistry and to expand on the work of the developers with new models. The
387 influence and utility of this modeling platform are evidenced by approximately 300 literature
388 citations to the software to date and by its incorporation into the US Environmental Protection
389 Agency's national recommended aquatic life criteria for copper (US Environmental Protection
390 Agency 2007). At the time of writing, the software was on its fourth major version. In the European
391 Union, environmental quality standards under the Water Framework Directive and risk assessments
392 under the policy Registration, Evaluation, Authorisation and Restriction of Chemicals employ
393 computationally intensive applications of multiple BLMs developed to protect algae/plants,
394 invertebrates, and fish (see Nys et al. (2016) for Ni; Van Regenmortel et al. (2017) for Cu and Zn; and
395 Van Sprang et al. (2016) for Pb). Various simplified proxies have been developed including Bio-Met,
396 which employs look-up tables that approximate BLM calculations for Cu, Ni, Zn, or Pb with over 20
397 000 values covering a wide range of environmentally relevant water chemistries, using fewer
398 parameters (Ca, pH, and DOC) than the full BLMs. Similar algorithm-based approaches that simplify
399 inputs and user calculations include the Metals Bioavailability Tool and PNEC-Pro (Peters et al. 2016;
400 Verschoor et al. 2017). For Pb, a separate tool simplifying speciation and toxicity predictions is also
401 available (Van Sprang et al. 2016). All these resources can readily be found through internet
402 searches.

403 Because mechanistic and hybrid models typically integrate the effects of water chemistry on
404 geochemical speciation as well as interactions of toxic metal species (mostly Me^{n+}) with the
405 organism, obvious roles of these models are to populate look-up tables and to inform the
406 development of MLRs (Brix et al. 2017a; DeForest et al. 2018). For example, a statistical approach to
407 developing MLRs using stepwise automated routines to maximize partial regression coefficients may
408 yield a good fit between effects and predictor variables. Yet, variables that are only important in a
409 subset of the data or that have subtle effects may be missed, and a statistical approach alone cannot
410 distinguish between causative and correlative variables. In the case of MLRs for Cu and Al, the choice

411 of potential toxicity predictor variables (DOC, pH, and water hardness or Ca) was informed by
412 associated BLMs (e.g., Santore et al. 2001, 2018), not by statistical explorations.

413 To further illustrate how mechanistic bioavailability models can inform simpler approaches, we have
414 used a chronic gBAM for fish (De Schamphelaere 2018) to predict how the 30-d LC20(dissolved) of
415 Cu in soft water (hardness ~10 mg/L) for fish varies as a function of the DOC, species sensitivity, and
416 pH (Figure 6). This gBAM accounts for geochemical speciation effects, competition of Cu^{2+} with Ca^{2+}
417 and Mg^{2+} , and effects of pH on Cu^{2+} ion toxicity. The simulations provide relationships between ECx
418 and DOC that “emerge” from the joint effects of these 3 processes in the hybrid gBAM model. These
419 simulations illustrate 3 important points: 1) the relation between ECx and DOC is nearly perfectly
420 linear, on both linear and logarithmic scales; 2) on a linear scale, the slope of the ECx versus DOC
421 relation is higher for less sensitive organisms and at higher pH; and 3) on a logarithmic scale, slopes
422 appear nearly independent of species sensitivity or pH. This indicates that an MLR on a linear scale
423 should contain not only a linear DOC term but also an interaction term between pH and DOC and
424 furthermore that an MLR slope derived on the basis of an insensitive species should not be
425 extrapolated to a more sensitive species. Similar simulations can be performed for other species,
426 water quality variables, and metals, resulting in specific recommendations for MLR construction.

427 *Equilibrium issues*

428 From both chemical and biological perspectives, BLMs and related constructs all assume equilibrium
429 conditions. If this is not the case, the predictive abilities of the model may be compromised.
430 Instances where equilibrium may not occur include laboratory exposures with incompletely
431 equilibrated diet and/or DOC and environmental exposures such as pulse exposures from
432 stormwater or meltwater runoff (Kayhanian et al. 2008; Nimick et al. 2011; Balistreri et al. 2012;
433 Figure 7) and exposures within mixing zones downstream of point-source releases (Vandenberg et
434 al. 2005). Nonequilibrium conditions can cause metals which equilibrium-based speciation
435 calculations assign to complexes to actually be bioavailable, whereas equilibrium models such as
436 BLMs generally assume that complexed metals are not bioavailable (Zhao et al. 2016).

437 Reactions of metal ions with dissolved inorganic ligands typically reach equilibrium in seconds to
438 minutes. However, equilibration of metals with DOC can take hours to days. This issue was initially
439 manifested in fathead minnow Cu toxicity data reported by Erickson et al. (1996). In nonrenewal
440 static trials, the 96-h test duration was sufficient for any disequilibrium associated with Cu–DOC
441 complexation to be effectively eliminated during the early part of the exposure. However, in flow-
442 through tests, a Cu stock solution was mixed with unamended Lake Superior water in the diluter
443 head tank just prior to entering the 45-min residence time test chamber. In this case free Cu was
444 continuously elevated relative to equilibrium conditions, and toxicity was increased relative to static
445 tests at the same dissolved Cu concentration. Model-predicted LC50s were almost identical in the 2
446 flow regimes, but in the flow-through tests, they were consistently higher than observed values (i.e.,
447 toxicity was underestimated by the model), whereas in the static tests they were consistently lower
448 than observed values (i.e., toxicity was overestimated), under otherwise similar conditions (Figure
449 8A,B; Santore et al. 2001). This interpretation is consistent with tests that found that *Ceriodaphnia*
450 *dubia* Cu LC50s were directly related to equilibration time with DOC (Kim et al. 1999; Ma et al. 1999).
451 A caution, however, is that DOC tends to increase over time during static tests, as a result of
452 accretion of organic carbon from the test organisms. For Ag and Cu, metals with strong affinities for
453 DOC, small increases in DOC in the range of 0.2 to 0.5 mg/L can produce noticeable changes in
454 modeled or measured toxicity (Erickson et al. 1998; Welsh et al. 2008). Thus, the pattern of greater
455 toxicity of Cu in flow-through tests than static tests could be influenced by both incomplete
456 equilibration in the former and increasing DOC over the course of the latter tests.

457 At high humic acid (>5 mg/L DOC) and Cu (>1 μ M) concentrations, up to 30 h were required to reach
458 equilibrium (Ma et al. 1999). Under more dilute conditions with DOC <1 mg/L, Cu more rapidly
459 equilibrated in 0.1 to 4 h (Louis et al. 2009; Meyer and Adams 2010). As an example, the kinetic data
460 of Ma et al. (1999) were used to calculate that at the start of a static exposure the free Cu (Cu^{2+})
461 concentrations might be elevated as much as 10-fold relative to the concentration at equilibrium
462 (Figure 9A). After the first 24 h, the deviation from equilibrium is small; and after 30 h, equilibrium is
463 achieved (Figure 9B). Similarly, in static-renewal tests, the solution is refreshed at regular intervals,
464 resulting in elevated free Cu with each renewal unless pre-equilibration is used (Figure 9C). Most
465 seriously, in flow-through tests with a short hydraulic residence, free Cu will be elevated throughout
466 the test (Figure 9D). Similarly, Meyer and DeForest (2018) invoked the Ma et al. (1999) kinetic model
467 to argue that a lack of adequate equilibration could explain the apparently very low threshold effect
468 concentrations (<2 μ g Cu/L) reported by Dew et al. (2012) for olfactory impairment in fathead
469 minnows exposed to Cu for short durations (1–24 h), as an alternative to the damage-repair
470 hypothesis proposed by Dew et al. (2012) for explaining the decrease of olfactory impairment as
471 exposure time increased.

472 Reports on the importance of pre-equilibration as a factor modifying metal toxicity have been
473 inconsistent. For instance, although Glover et al. (2005) found that Ag was more toxic to *Daphnia*
474 magna (lower 24-h LC50s) in tests initiated after 3-h metal–DOM contact time than in tests initiated
475 after 24 h contact time (Figure 10), Erickson et al. (1998) found little effect on Ag toxicity from aging
476 solutions for 72 h before testing. Further, in contrast to the Ma et al. (1999) results, Wang et al.
477 (2011) found little differences in Cu toxicity between tests initiated with freshly mixed exposures
478 versus solutions that had been aged for 24 h.

479 Equilibrium models are a reasonable simplification of real-world systems in many cases, such that
480 simple relationships have been obtained that relate metal speciation to biological effects (e.g., the
481 present review; Zhao et al. 2016). This simplification is probably necessary when using bioavailability
482 models, whether mechanistic or empirical, to inform regulatory applications. Nevertheless, the
483 controversies over equilibrium assumptions and contrasting results preclude conclusive
484 generalizations to resolve the apparent equilibration dilemma. Still, unless time-varying conditions
485 are the focus of testing, we suggest a 24-h pre-equilibration period in experimental designs to allay
486 concerns of nonequilibrium.

487 *Toxicity test design considerations for developing mechanistic and hybrid bioavailability models*

488 Historically, mechanistic and hybrid metal bioavailability models have been developed from 1)
489 multiple univariate toxicity experiments, where each factor (e.g., Ca, Na, DOC, pH) is varied alone; 2)
490 full-factorial test designs (which can be considered a series of univariate experiments at various
491 conditions); 3) multivariate toxicity experiments, where factors are varied in various combinations;
492 or 4) a combination of 1) and 2). Regardless of the design, well-informed selection of the water
493 chemistry variables is crucial. This selection should be based on prior knowledge of suspected
494 influential variables, using information on other organisms, other metals, previous metal uptake and
495 toxicity data, and physiological understanding.

496 Multivariate experiments are particularly useful as a first step to discriminate the more from the less
497 important toxicity-modifying variables. However, on their own, they are not necessarily the most
498 useful for mechanistic model development, as illustrated in the following example. De
499 Schampelaere and Janssen (2004b) performed chronic *D. magna* toxicity experiments with Cu in a
500 multivariate design with pH, DOC, and hardness as the factors. They found that DOC and pH were
501 significant factors but that hardness was not. They subsequently used the data to calibrate a chronic

502 Cu-BLM for *Daphnia*, in which Ca or Mg competition was not included. Also, Na was included in the
503 model because of evidence from a parallel univariate experiment with Na that showed a correlation
504 with Cu toxicity. Later, Rodriguez et al. (2012) performed univariate experiments and did find
505 protective effects of Ca and Mg. Van Regenmortel et al. (2015) developed an optimized chronic Cu
506 bioavailability model by reformulating it as a gBAM and including biotic ligand constants for Ca and
507 Mg. This optimized model accurately predicted the toxicity observed in both data sets.

508 This example indicates that non-full-factorial multivariate test designs can “miss” toxicologically
509 significant modifying factors, especially when the design is run over multiple test series where
510 between-batch variability may play a role (see Supplemental Data). On the positive side, the
511 example illustrates that, over time, existing models can be improved if new data become available
512 and that data sets from different sources can be used jointly for model calibration. It also suggests
513 that univariate test designs are best for calibration of individual model parameters (e.g., biotic ligand
514 stability constants for competing cations in BLMs or pH slopes in gBAMs). Furthermore, the
515 mathematical method to estimate log K values is well known and relatively straightforward (De
516 Schampelaere et al. 2002). Full-factorial designs (the special case of multiple univariate
517 experiments) have been rarely used in generating bioavailability models because they are the most
518 costly and labor-intensive, but they are particularly useful to detect interactive effects. Using this
519 design, Deleebeeck et al. (2009) were able to show no interactions between pH and Mg on chronic
520 Ni toxicity to algae, allowing them to formulate a model that was successful at predicting Ni toxicity
521 in a range of spiked field waters. In summary, all sorts of designs can help in initial model
522 development, but univariate or full-factorial designs are the best for calibration of individual model
523 parameters.

524 *Consideration of other toxicity-modifying factors*

525 Although pH, DOC, and major ions have been incorporated into most bioavailability models (Table
526 2), other factors have received much less attention by bioavailability model developers. We are not
527 aware of any current regulatory framework that explicitly incorporates effects of other metal toxicity
528 factors such as acclimation to prior metal exposure, temperature, nutrients, suspended solids, or
529 iron hydroxides in the assessment of metal toxicity. Yet we now know that such factors may strongly
530 affect metal toxicity, and in the case of nutrients and temperature, for example, a wealth of
531 empirical data is available.

532 Acclimation to chronic metal exposures can have major effects on responses to subsequent acute
533 exposures, through acquired tolerance. During long-term, low-level exposures, gill metal burdens
534 may increase above the concentrations usually associated with acute toxicity (reviewed by Niyogi
535 and Wood 2003). This has been explained by the “damage-repair hypothesis,” whereby the repair
536 processes increase the tolerance of the organism to a particular metal burden (McDonald and Wood
537 1993). Likewise, the log K and Bmax conditional binding constants of the metal biotic ligand and of
538 the various cation–biotic ligand complexes will be markedly changed following acquired tolerance,
539 yet these constants are considered to be unchangeable with the changes in environmental
540 conditions in BLMs (Niyogi and Wood 2003). Although acquired tolerance from exposure to elevated
541 metals has long been recognized (Chapman 1985), acclimation to soft or hard water, or differences
542 in the composition of the diet (e.g., high calcium or sodium content) may also have profound effects
543 on metal tolerance and modeling (e.g., Niyogi and Wood 2003; Franklin et al. 2005; Todd et al. 2009;
544 Mebane et al. 2010). In criteria development, acclimation is considered a confounding, false
545 protection to be guarded against because acquired protections may not be persistent and,
546 furthermore, the energetic costs of tolerance may lead to other adverse effects (Stephan et al. 1985;
547 Brinkman and Woodling 2014). However, the implications of variations in binding constants

548 depending on earlier metal exposure, water hardness, diet, and other environmental or
549 physiological conditions seem to be underrecognized in the development and application of metal
550 bioavailability models (Niyogi and Wood 2003).

551 Most studies on temperature effects have investigated acute toxicity. These have reported both
552 increasing metal toxicity with increasing temperature (Rao and Khan 2000; Heugens et al. 2003) and
553 increasing toxicity with decreasing temperature (Hodson and Sprague 1975; Hansen et al. 2002).
554 Much more limited data are available on chronic metal toxicity, but a recent study showed that
555 chronic metal toxicity to *D. magna* varied by approximately 2-fold, with a clear pattern of higher
556 toxicity at lower temperature, opposite to what is most commonly expected for acute toxicity
557 (Pereira et al. 2017). Interestingly, for Cu, the response pattern could be explained by the computed
558 effect of temperature on speciation but not for Ni and Zn. Until now, most BLMs and hybrid models
559 do include temperature as an input parameter, but they only compute the effect of temperature on
560 inorganic speciation, whereas DOC interactions, which rely on WHAM V, VI, or VII, are not
561 temperature-adjusted. Most importantly, any temperature effects at the biological receptor are not
562 incorporated. More empirical and mechanistic research is clearly needed to better integrate
563 temperature into bioavailability models and AWQC.

564 Effects of nutrient concentrations, especially phosphate, on metal toxicity have been studied
565 extensively and can be relatively strong but are inconsistent across various studies. Notably,
566 phosphate concentrations in natural freshwaters (European annual mean in rivers as of 2012 was
567 $65 \mu\text{g PO}_4\text{-P L}^{-1}$ [European Environment Agency 2018]) are typically considerably lower than those
568 applied in standard algae toxicity test protocols ($310\text{--}1550 \mu\text{g PO}_4\text{-P L}^{-1}$ [Organisation for Economic
569 Co-operation and Development 2011; US Environmental Protection Agency 2002]). Thus, a better
570 mechanistic understanding of nutrient effects on metal toxicity, and associated incorporation into
571 models, is urgently needed to improve laboratory-to-field extrapolation. Gao et al. (2016), for
572 example, developed a mechanistic dynamic model that integrated effects of external phosphate,
573 algal cell P content, and Zn on instantaneous algal growth rate. Their model was able to explain why
574 different exposure scenarios (duration, phosphate supply, and initial P content of algae) can lead to
575 opposite apparent effects of phosphate on Zn-induced declines of algal biomass in standard toxicity
576 tests. As bioavailability modeling expands into metalloids and plants, we expect nutrients will have
577 an important role as a modifying factor, especially with pairs such as phosphate and arsenate (Zhao
578 et al. 2016).

579 The role of organic ligands that bind metals and form lipophilic complexes has received little
580 attention in metal bioavailability modeling. Similarly, the potential role of low-molecular weight
581 metabolites such as thiosulfate or citrate (which can bind metals and then transport them across
582 epithelial surfaces as the intact metal-ligand complex via anion transporters) has received little
583 attention (Zhao et al. 2016). Although BLMs and related constructs usually treat complexed metals
584 as nontoxic, attributing toxicity only to the free ion metals, we now know that complexed metals can
585 be bioavailable and toxic under some circumstances (Erickson et al. 1996; Zhao et al. 2016).

586 Other trace metals, such as Fe and Al, may also influence metal toxicity, notably via 1) competition
587 with the toxic metal ion for binding on DOC, and 2) by providing an adsorption phase when in the
588 colloidal hydroxide-precipitated form (Cain et al. 2016). It would be worthwhile to explore how
589 effects of these and other bioavailability-modifying factors could be added to metal bioavailability
590 models.

591 *The choice of speciation modeling platforms for BLMs*

592 Given the importance of DOM (natural organic matter, DOC) in metal speciation, the speciation
593 modeling component of a mechanistic bioavailability model must be capable of accurately predicting
594 metal–DOM complexation. Several mechanistic models exist for computing metal binding to humic
595 and fulvic acid, which are the dominant components of DOM in freshwaters. These include the
596 humic ion–binding family (Models V, VI, and VII; Tipping and Hurley 1992; Tipping 1998; Tipping et
597 al. 2011), the nonideal competitive absorption (NICA)-Donnan model (Kinniburgh et al. 1996) and
598 the Stockholm humic model (Gustafsson 2001). These are combined with inorganic speciation codes
599 in tools such as WHAM, Visual MINTEQ, and the Hydroqual/Windward BLM software. These
600 inorganic speciation models differ somewhat in the binding affinity values used and even the
601 presence of equilibrium constants in their code. Some speciation models do not account for
602 precipitation/dissolution reactions. The absence of the relevant equilibrium constants can lead to
603 the use of unstable metal exposure regimes in experiments, where the metal's solubility limit is
604 exceeded.

605 A number of speciation models have been used in BLM development. Di Toro et al. (2001)
606 implemented WHAM V within the CHESS framework, and many subsequent studies (e.g., De
607 Schamphelaere and Janssen 2002; Heijerick et al. 2005) also used WHAM V. Others have used
608 WHAM VI (e.g., Deleebeek et al. 2008; Peters et al. 2011), WHAM VII (Vukov et al. 2016), or NICA-
609 Donnan (Van Sprang et al. 2016). The choice of speciation model platform may be informed by a
610 number of factors, particularly the availability of binding constants for the metal of interest. Derived
611 BLM binding constants (i.e., log K values) are conditional on the choice of speciation model, and
612 different BLM binding constants can be obtained from the same data set using different speciation
613 models. This has implications for metal-mixture BLMs—here, a single speciation model should be
614 used for all metals to account for intermetal competition on DOM binding sites, and the provenance
615 of any binding constants used should be carefully evaluated. We did not reach consensus whether
616 any single speciation model for either DOM binding or inorganic speciation could be considered
617 optimal. The choice of models may be informed by a number of factors, particularly the availability
618 of binding constants for the metal of interest.

619 *Specification of DOM (DOC) in models*

620 All of the DOM-binding models listed require concentrations of humic and/or fulvic acid to be
621 specified. This requires the use of “activity factors”—the ratio of measured DOC concentration to
622 the concentrations of humic and/or fulvic acid that reproduce the metal-binding properties of that
623 DOC. The activity factor is composed of 3 components, each of which may be assumed or estimated:
624 the carbon content of DOM, the metal-binding properties of that DOM, and the attribution of the
625 binding properties of that DOM to humic and/or fulvic acid. Development of a BLM has usually used
626 a global activity factor, sometimes derived from metal–DOM binding studies (Dwane and Tipping
627 1998; Bryan et al. 2002). Early developments (e.g., Di Toro et al. 2001) assumed DOM to be 50%
628 carbon, with 100% activity and comprising 10% humic and 90% fulvic acid. Because the activity
629 factor is in reality water-specific, some have researched whether the activity correlates to
630 measurable DOM properties. De Schamphelaere et al. (2004) found that optimizing water-specific
631 BLM predictions of acute Cu toxicity to *D. magna* produced activities (as fulvic acid only) that
632 correlated significantly with the specific absorbances of the DOM samples at 350 nm (SAC350). Al-
633 Reasi et al. (2012) found significant correlations between acute Cu toxicity to *D. magna* and a
634 number of optical and physicochemical properties of DOM, with SAC340 being the most significant
635 predictor of protective ability. They suggested that the predictive capability of BLMs could be
636 improved by the use of SAC340 to adjust the activity factor on a water-by-water basis. In general,
637 larger, more lipophilic, more aromatic DOMs of terrigenous origin, with higher humic acid-like

638 content, and therefore higher SAC values, appear to be more protective against Cu toxicity. Despite
639 these findings, however, many BLM studies still use a global activity factor in model development
640 because this has the advantage of requiring only measurement of absolute DOC concentration for
641 application.

642

643 *Evaluation of speciation models*

644 The ability of models to predict metal speciation, for the chemical conditions relevant to BLM
645 development and application, needs to be fully evaluated. Because BLM parameterization employs
646 predicted free metal ion activity, testing should ideally be done on measurements of the free
647 activity. Speciation measurement is complex, and many methods remain under active development
648 rather than in routine use. Relatively well-established methods (e.g., ion-selective electrodes [ISEs])
649 are challenging to apply at the dissolved metal and DOC concentrations encountered in natural
650 waters because of issues such as membrane dissolution and fouling (Eriksen et al. 1999), although
651 they can be highly useful in toxicity tests, particularly acute toxicity studies (e.g., Al-Reasi et al. 2012;
652 Crémazy et al. 2016). Some have used DOC preconcentration (Ahmed et al. 2013) or continuous-flow
653 (Tait et al. 2016) systems to enable measurement at the metal:DOM ratios encountered in natural
654 waters. Competitive ligand exchange voltammetry (Xue and Sigg 1999; Cao et al. 2006) has also been
655 used, though its validity has been criticized (van Leeuwen and Town 2005; Lofts and Tipping 2011).
656 Nonetheless, continued research in this area is essential, alongside critical assessment of speciation
657 models against such data. Of particular note is a pair of landmark studies comparing the reliability
658 and performance of different trace metal speciation analytical methods and comparative model
659 performance to modeling of metal speciation (Sigg et al. 2006; Unsworth et al. 2006). Examples of
660 model predictions compared against recent measurements using ISEs (for Cu) and an ion-exchange
661 technique (for Co, Ni, Zn, and Cd) are shown in Supplemental Data, Figures S1 through S6.

662 CONSIDERATIONS FOR THE INCORPORATION OF METAL BIOAVAILABILITY MODELS INTO 663 ENVIRONMENTAL QUALITY STANDARDS

664 *Dealing with extreme waters*

665 The occurrence of extreme natural water conditions (e.g., unusual pH, hardness, DOC levels, or
666 combinations thereof) is a common reality for almost all geographical regions. These situations have
667 been recognized in working with BLMs (Van Genderen et al. 2005; Natale et al. 2007; Hoppe et al.
668 2015a, 2015b) and pose a common challenge in terms of bioavailability model development and
669 application for regulatory criteria. There are various reasons for this. First, the taxa typically used for
670 model development (i.e., organisms commonly used in all laboratory testing) may not be tolerant of
671 extreme water conditions. All of the water conditions that modify metal toxicity are themselves
672 environmental characteristics that limit habitat suitability in ways that have nothing to do with metal
673 toxicity. For example, DOM has been called an ecological driving force for aquatic ecosystems with
674 well-documented effects on the pH and primary productivity of natural waters (Steinberg et al.
675 2006). Furthermore, there is evidence that DOC can bind to the gills (Campbell et al. 1997) and alter
676 the basic physiology of ion transport in a way which can beneficially mitigate the damaging effects of
677 metals and low pH (Galvez et al. 2008; Wood et al. 2011; Duarte et al. 2016). These actions of DOC
678 are separate from their ability to reduce the bioavailability of metals by complexation. Some
679 crustaceans and snails will not thrive in culture waters of very low hardness or pH (i.e., control
680 performance will not be acceptable). Equally, they will not be present in natural waters of low
681 hardness (Lodge et al. 1987; Hooper et al. 2008; Cairns and Yan 2009). Second, the available models

682 are not generally validated for extreme water conditions, and thus, the predictions can be erroneous
683 and/or generate uncertainty. And third, the extreme water types often have specific ecological
684 assemblages with organisms of different physiological characteristics, which may or may not show
685 similar sensitivities and/or metal–bioavailability relationships (e.g., different log K values or pH
686 slopes) to organisms typically used for model development.

687 The main target for model development should be the central distribution of data, such as the 5th to
688 95th percentile of the distribution of water chemistry parameters, rather than undue focus on
689 exceptions and extreme values. The following options can be considered in deciding the derivation
690 of a new model or use of existing model for ecosystems with extreme water parameters.

691 *Extending the boundaries of existing models*

692 This includes recalibration of an existing bioavailability model with the testing of local waters and
693 organisms to extend the physicochemical boundaries of the model. A series of papers has been
694 published with methods describing how the validation boundaries of BLMs or hybrid models for Cu,
695 Ni, Zn, and Pb can be extended to also accurately predict metal toxicity under more extreme pH and
696 hardness (Van Genderen et al. 2005; Deleebeek et al. 2007; Nys et al. 2016, 2017; Van Regenmortel
697 et al. 2017).

698 *Developing new bioavailability models*

699 In general, bioavailability models should be developed and tested in media that resemble surface
700 water conditions that are within the natural limit of the test organism. Testing organisms outside
701 their usual physiological range of tolerance is inadvisable. Site-specific models should be developed
702 using toxicity testing that employs site waters and native organisms from the extreme sites.

703 *Metal mixtures*

704 Our discussions so far have treated metals as if they occur one by one in the environment. Likewise,
705 regulatory criteria are developed as if individual metals occurred in isolation, with no interactive
706 toxicities. Both are, of course, complete fiction. In the real world, metals always occur in mixtures
707 that are a function of the mineral composition of the watershed. Anthropogenic inputs will
708 invariably produce mixtures of metals, and some generalities about mixture occurrences in ambient
709 waters can be made. The most predictable metal combination is probably Cd and Zn, which seem to
710 naturally occur at close to a 1:200 mass ratio around the world (Mebane et al. 2017). Nickel and Co
711 commonly occur in association with Cu, and Pb is commonly associated with Zn; but the ratios and
712 particular combinations may be highly variable across geological domains (Salminen 2005). Empirical
713 models are not well suited to such variable scenarios, whereas mechanistic bioavailability and
714 toxicity models do provide a flexible approach to handle these combinations.

715 It has long been recognized that the single-metal framework for BLMs could logically be extended to
716 metal mixtures (Di Toro et al. 2001; Playle 2004), and much recent progress has been made in this
717 area (e.g., Farley et al. 2015; Meyer et al. 2015; Nys et al. 2018). These metal mixture modeling tools
718 may be highly useful in risk-assessment scenarios. However, because of the overwhelming diversity
719 of possible combinations, we expect that regulatory criteria to protect aquatic environments will
720 continue to be developed for individual substances for the foreseeable future. Toxic unit models
721 assume that potency-normalized concentrations of metals can be added together to predict the
722 toxicity of a metal mixture; these provide a simple approach to estimate mixture toxicity risks from
723 single-metal toxicity models (concentration addition or toxic unit models). Alternatively, predicted
724 toxic responses from single-substance toxicity models can be added (response addition or, more

725 appropriately stated, independent action models). The concentration addition approach tends to be
726 more conservative than the response addition approach; that is, concentration addition may predict
727 greater effects than observed (Van Regenmortel et al. 2017; Crémazy et al. 2018). Both approaches
728 implicitly assume that chemicals in the mixture do not physically, chemically, or biologically interact
729 and thereby overlook competition for metal binding sites on DOC and on the target biotic ligand,
730 which could make metal mixtures more or less toxic than if there were no interactions. Recent
731 studies at acutely toxic metal levels indicate that such binding interactions can occur at biotic ligands
732 (Niyogi et al. 2015; Brix et al. 2016, 2017c). However, interactions between metals for biotic ligand
733 or DOC binding sites are not predicted to be important at mixture concentrations at the low $\mu\text{g/L}$
734 levels relevant to most chronic regulatory criteria (Balistrieri and Mebane 2014). We believe that
735 mixture toxicity models are ultimately needed for the application of metal criteria, and as mentioned
736 earlier, they should be developed using a common DOM speciation platform. However, the
737 development of bioavailability-based criteria on a single-metal basis remains a reasonable approach.
738 Concerns over how to apply criteria in the ubiquitous settings with metal mixtures present should
739 not hold back the development and application of single-metal bioavailability-based criteria.

740 *Ownership and maintenance of bioavailability models*

741 Setting up the reaction equations for bioavailability models, writing code to execute them,
742 developing software to provide a functional user interface and interpretive output display,
743 documenting the construction and performance of the package, and preparing detailed
744 documentation for users is no trivial undertaking. The expectations are particularly onerous when
745 bioavailability models are used to set regulatory water criteria sufficient to protect diverse
746 communities in diverse environments with legally enforceable limits that drive costs for engineering
747 design, capital construction, operating, and monitoring. Furthermore, the use of models in public
748 policy settings requires sustaining commitments by sponsors over the long term.

749 It takes no ongoing effort to maintain regulatory criteria that are expressed as simple mathematical
750 functions of toxicity-modifying factors. For instance, some of the criteria values based on hardness
751 equations published by the US Environmental Protection Agency (1986) are still in use 3 decades on.
752 In contrast, for criteria calculated with the aid of custom software, that software needs ongoing
753 maintenance to upgrade to new operating systems, to fix bugs, and to modify the model capabilities
754 following advances in the underlying science. These maintenance needs pose a challenge to
755 regulatory authorities. They must not just provide one-time support for a model to be used in
756 criteria and then move on; an ongoing commitment to maintain the model is needed. Further, the
757 opportunistic use of model software that is not fully in the public domain to set environmental
758 regulations raises intellectual property ownership questions. Institutions supporting chemical
759 speciation models may sell licenses to partially offset their development and maintenance costs and
760 allow the developers to keep advancing their models. Regulatory authorities may hesitate to rely on
761 a software application that is not in the public domain and that cannot be guaranteed to be
762 functional indefinitely, and they may be unwilling or unable to commit to ongoing support of model
763 applications on behalf of their affected dischargers. Until these practical model support and public
764 domain issues are addressed, the pragmatic path forward is to use bioavailability models as research
765 tools to inform simpler, lower-maintenance translational tools for regulatory adoption, such as MLRs
766 and look-up tables (see section Complexity versus simplicity: Use of mechanistic and hybrid models
767 to inform development of simpler models).

768 CONSENSUS RECOMMENDATIONS FOR DEVELOPING OR APPLYING METAL BIOAVAILABILITY MODELS

769 Our workgroup reached consensus on the following points. It would be prudent for scientists and
770 environmental managers to consider these when developing or applying metal bioavailability models
771 for environmental quality standards or risk assessment.

772 1) Empirical bioavailability tools such as MLRs and look-up tables should always be informed
773 qualitatively and quantitatively by mechanistic models.

774 2) Going forward, equilibrium speciation should be considered in the design of experiments for
775 bioavailability models. We recommend a 24-h pre-equilibration period in experimental designs to
776 allay concerns of nonequilibrium test conditions. Care should be used to include speciation models
777 that encompass precipitation reactions so as to ensure that the solubility limits are respected for the
778 metal(s) of interest and that no loss of solubility occurs.

779 3) The chemical speciation model used should be tested independently of its ability to predict
780 toxicity.

781 4) Data obtained from tests conducted with organisms outside the chemistry boundaries from
782 where they live should not be used. The main target for model development should be the central
783 distribution of data, such as the 5th to 95th percentile of the distribution of water chemistry
784 parameters, rather than undue focus on exceptions and extreme values.

785 5) Some potentially important toxicity-modifying factors are currently not represented in
786 bioavailability models and have received insufficient attention in toxicity testing. Temperature is
787 probably of foremost importance; P is likely important in plant and algae models.

788 6) Plant and animal bioavailability models should not be combined because of the divergent
789 influences of pH.

790 7) pH is a unique toxicity-modifying factor, with multiple possible mechanisms. These effects are
791 currently best captured by hybrid models, which can inform improved mechanistic understanding
792 and foster better mechanistic models of pH effects. Failures of mechanistic models to explain
793 experimental data can advance our understanding of actual mechanisms.

794 8) To develop models for mixture toxicity, a common chemical speciation platform should be used.
795 This is particularly important for DOM.

796 9) To estimate bioavailability model parameters, univariate or full-factorial designs are most useful
797 because multivariate designs may miss responses for variables with limited effect. In univariate test
798 designs, nonsimultaneous testing can introduce confounding variability. In real laboratories in the
799 real world, this may be unavoidable, but repeating treatments between studies is important.

800 10) For best practice during chronic tests, combined waterborne and dietary matched exposures
801 should be performed. These should be based on natural live diets equilibrated with the associated
802 waterborne metal concentration. However, the absence of such designs should not be a criterion for
803 rejecting currently available chronic data.

804 11) There is a need to develop mechanistically based bioavailability models for behavioral toxicity
805 and to consider these for future regulatory application. Such models should be built from the ground
806 up using behavioral endpoints, rather than by adjusting the sensitivity parameter in existing BLMs.

807

808 12)

809 If bioavailability models are to be used in environmental regulation, ongoing support and availability
810 for use of the models in the public domain are essential. Until this can be guaranteed, simpler,
811 lower-maintenance translational tools based on bioavailability models, such as MLRs and look-up
812 tables, may be preferable.

813 *Supplemental Data*

814 The Supplemental Data are available on the [at DOI: 10.1002/etc.4560](https://doi.org/10.1002/etc.4560).

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825

826

827 *References*

- 828 Adams W, Blust R, Dwyer R, Mount D, Nordheim E, Rodriguez PH, Spry D. 2020. Bioavailability
829 assessment of metals in freshwater environments: A historical review. *Environ Toxicol Chem* 39: 48–
830 59.
- 831 Ahmed IAM, Hamilton-Taylor J, Lofts S, Meeussen JCL, Lin C, Zhang H, Davison W. 2013. Testing
832 copper-speciation predictions in freshwaters over a wide range of metal–organic matter ratios.
833 *Environ Sci Technol* 47: 1487– 1495.
- 834 Alabaster JS, Lloyd R. 1980. *Water Quality Criteria for Freshwater Fish*. Butterworth, Boston, MA,
835 USA.
- 836 Al-Reasi HA, Smith SD, Wood CM. 2012. Evaluating the ameliorative effect of natural dissolved
837 organic matter (DOM) quality on copper toxicity to *Daphnia magna*: Improving the BLM.
838 *Ecotoxicology* 21: 524– 537.
- 839 Alsop D, Ng TYT, Chowdhury MJ, Wood CM. 2016. Interactions of waterborne and dietborne Pb in
840 rainbow trout, *Oncorhynchus mykiss*: Bioaccumulation, physiological responses, and chronic toxicity.
841 *Aquat Toxicol* 177: 343– 354.
- 842 Alsop DH, Wood CM. 2000. Kinetic analysis of zinc accumulation in the gills of juvenile rainbow trout:
843 Effects of zinc acclimation and implications for biotic ligand modeling. *Environ Toxicol Chem* 19:
844 1911– 1918.
- 845 Antunes PMC, Kreager NJ. 2014. Lead toxicity to *Lemna minor* predicted using a metal speciation
846 chemistry approach. *Environ Toxicol Chem* 33: 2225– 2233.
- 847 Balistrieri LS, Mebane CA. 2014. Predicting the toxicity of metal mixtures. *Sci Total Environ* 466–467:
848 788– 799.
- 849 Balistrieri LS, Mebane CA, Schmidt TS, Keller WB. 2015. Expanding metal mixture toxicity models to
850 natural stream and lake invertebrate communities. *Environ Toxicol Chem* 34: 761– 776.
- 851 Balistrieri LS, Nimick DA, Mebane CA. 2012. Assessing time-integrated dissolved concentrations and
852 predicting toxicity of metals during diel cycling in streams. *Sci Total Environ* 425: 155– 168.
- 853 HL Bergman, EJ Dorward-King, eds. 1997. *Reassessment of Metals Criteria for Aquatic Life*
854 *Protection: Priorities for Research and Implementation*. SETAC Pellston Workshop on Reassessment
855 of Metals Criteria for Aquatic Life Protection. SETAC, Pensacola, FL, USA.
- 856 Besser JM, Brumbaugh WG, Brunson EL, Ingersoll CG. 2005. Acute and chronic toxicity of lead in
857 water and diet to the amphipod *Hyalella azteca*. *Environ Toxicol Chem* 24: 1807– 1815.
- 858 Bianchini A, Wood CM. 2002. Physiological effects of chronic silver exposure in *Daphnia magna*.
859 *Comp Biochem Physiol C Toxicol Pharmacol* 133: 137– 145.
- 860 Bianchini A, Wood CM. 2003. Mechanism of acute silver toxicity in *Daphnia magna*. *Environ Toxicol*
861 *Chem* 22: 1361– 1367.
- 862 Bielmyer GK, Grosell M, Brix KV. 2006. Toxicity of silver, zinc, copper, and nickel to the copepod
863 *Acartia tonsa* exposed via a phytoplankton diet. *Environ Sci Technol* 40: 2063– 2068.
- 864 Bielmyer GK, Grosell M, Paquin PR, Mathews R, Wu KB, Santore RC, Brix KV. 2007. Validation study
865 of the acute biotic ligand model for silver. *Environ Toxicol Chem* 26: 2241– 2246.

866 Birceanu O, Chowdhury MJ, Gillis PL, McGeer JC, Wood CM, Wilkie MP. 2008. Modes of metal
867 toxicity and impaired branchial ionoregulation in rainbow trout exposed to mixtures of Pb and Cd in
868 soft water. *Aquat Toxicol* 89: 222– 231.

869 Boullemant A, Lavoie M, Fortin C, Campbell PGC. 2009. Uptake of hydrophobic metal complexes by
870 three freshwater algae: Unexpected influence of pH. *Environ Sci Technol* 43: 3308– 3314.

871 Brinkman SF, Woodling JD. 2014. Acclimation and deacclimation of brown trout (*Salmo trutta*) to
872 zinc and copper singly and in combination with cadmium or copper. *Arch Environ Contam Toxicol* 67:
873 214– 223.

874 Brix KV, DeForest DK, Tear LM, Grosell M, Adams WJ. 2017a. Use of multiple linear regression
875 models for setting water quality criteria for copper: A complementary approach to the biotic ligand
876 model. *Environ Sci Technol* 51: 5182– 5192.

877 Brix KV, DeForest DK, Tear L, Peijnenburg W, Peters A, Traudt E, Erikson R. 2020. Development of
878 empirical bioavailability models for metals. *Environ Toxicol Chem* 39: 85– 100.

879 Brix KV, Schlegel CE, Garman ER. 2017b. The mechanisms of nickel toxicity in aquatic environments:
880 An adverse outcome pathway analysis. *Environ Toxicol Chem* 36: 1128– 1137.

881 Brix KV, Tellis MS, Crémazy A, Wood CM. 2016. Characterization of the effects of binary metal
882 mixtures on short-term uptake of Ag, Cu, and Ni by rainbow trout (*Oncorhynchus mykiss*). *Aquat*
883 *Toxicol* 180: 236– 246.

884 Brix KV, Tellis MS, Crémazy A, Wood CM. 2017c. Characterization of the effects of binary metal
885 mixtures on short-term uptake of Cd, Pb, and Zn by rainbow trout (*Oncorhynchus mykiss*). *Aquat*
886 *Toxicol* 193: 217– 227.

887 Bryan SE, Tipping E, Hamilton-Taylor J. 2002. Comparison of measured and modelled copper binding
888 by natural organic matter in freshwaters. *Comp Biochem Physiol C Toxicol Pharmacol* 133: 37– 49.

889 Cain DJ, Croteau M-N, Fuller CC, Ringwood AH. 2016. Dietary uptake of Cu sorbed to hydrous iron
890 oxide is linked to cellular toxicity and feeding inhibition in a benthic grazer. *Environ Sci Technol* 50:
891 1552– 1560.

892 Cairns A, Yan N. 2009. A review of the influence of low ambient calcium concentrations on
893 freshwater daphniids, gammarids, and crayfish. *Environ Rev* 17: 67– 79.

894 CASCampbell PGC, Twiss MR, Wilkinson KJ. 1997. Accumulation of natural organic matter on the
895 surfaces of living cells: Implications for the interaction of toxic solutes with aquatic biota. *Can J Fish*
896 *Aquat Sci* 54: 2543– 2554.

897 Canadian Council of Ministers of the Environment. 2007. Canadian Water Quality Guidelines for the
898 Protection of Aquatic Life (Summary Table, Ver 7.1). Winnipeg, MB, Canada. [cited 2019 November
899 10]. Available
900 from: https://www.ccme.ca/en/resources/canadian_environmental_quality_guidelines/index.html

901 Cao J, Xue H, Sigg L. 2006. Effects of pH and Ca competition on complexation of cadmium by fulvic
902 acids and by natural organic ligands from a river and a lake. *Aquat Geochem* 12: 375– 387.

903 Chapman GA. 1985. Acclimation as a factor influencing metal criteria. In RC Bahner, DJ Hansen, eds,
904 *Aquatic Toxicology and Hazard Assessment: Eighth Symposium (STP 891-EB)*, Vol STP 891. ASTM
905 International, Philadelphia, PA, USA, pp 119– 136.

- 906 Clearwater SJ, Farag AM, Meyer JS. 2002. Bioavailability and toxicity of dietborne copper and zinc to
907 fish. *Comp Biochem Physiol C Toxicol Pharmacol* 132: 269– 313.
- 908 Clifford M, McGeer JC. 2009. Development of a biotic ligand model for the acute toxicity of zinc to
909 *Daphnia pulex* in soft waters. *Aquat Toxicol* 91: 26– 32.
- 910 Clifford M, McGeer JC. 2010. Development of a biotic ligand model to predict the acute toxicity of
911 cadmium to *Daphnia pulex*. *Aquat Toxicol* 98: 1– 7.
- 912 Crémazy A, Brix KV, Wood CM. 2018. Chronic toxicity of binary mixtures of six metals (Ag, Cd, Cu, Ni,
913 Pb, and Zn) to the great pond snail *Lymnaea stagnalis*. *Environ Sci Technol* 52: 5979– 5988.
- 914 Crémazy A, Wood CM, Ng TYT, Smith DS, Chowdhury MJ. 2017. Experimentally derived acute and
915 chronic copper biotic ligand models for rainbow trout. *Aquat Toxicol* 192: 224– 240.
- 916 Crémazy A, Wood CM, Smith DS, Ferreira MS, Johannsson OE, Giacomini M, Val AL. 2016.
917 Investigating copper toxicity in the tropical fish cardinal tetra (*Paracheirodon axelrodi*) in natural
918 Amazonian waters: Measurements, modeling, and reality. *Aquat Toxicol* 180: 353– 363.
- 919 DeForest DK, Brix KV, Tear LM, Adams WJ. 2018. Multiple linear regression models for predicting
920 chronic aluminum toxicity to freshwater aquatic organisms and developing water quality guidelines.
921 *Environ Toxicol Chem* 37: 80– 90.
- 922 DeForest DK, Gensemer RW, Van Genderen EJ, Gorsuch JW. 2011. Protectiveness of water quality
923 criteria for copper in western United States waters relative to predicted olfactory responses in
924 juvenile Pacific salmon. *Integr Environ Assess Manag* 7: 336– 347.
- 925 DeForest DK, Meyer JS. 2015. Critical review: Toxicity of dietborne metals to aquatic organisms. *Crit*
926 *Rev Environ Sci Technol* 45: 1176– 1241.
- 927 DeForest DK, Santore RC, Ryan AC, Church BG, Chowdhury MJ, Brix KV. 2017. Development of biotic
928 ligand model–based freshwater aquatic life criteria for lead following US Environmental Protection
929 Agency guidelines. *Environ Toxicol Chem* 36: 2965– 2973.
- 930 DeForest DK, Van Genderen EJ. 2012. Application of U.S. EPA guidelines in a bioavailability-based
931 assessment of ambient water quality criteria for zinc in freshwater. *Environ Toxicol Chem* 31: 1264–
932 1272.
- 933 Deleebeek NME, De Schamphelaere KAC, Janssen CR. 2007. A bioavailability model predicting the
934 toxicity of nickel to rainbow trout (*Oncorhynchus mykiss*) and fathead minnow (*Pimephales*
935 *promelas*) in synthetic and natural waters. *Ecotoxicol Environ Saf* 67: 1– 13.
- 936 Deleebeek NME, De Schamphelaere KAC, Janssen CR. 2008. A novel method for predicting chronic
937 nickel bioavailability and toxicity to *Daphnia magna* in artificial and natural waters. *Environ Toxicol*
938 *Chem* 27: 2097– 2107.
- 939 Deleebeek NME, De Schamphelaere KAC, Janssen CR. 2009. Effects of Mg²⁺ and H⁺ on the toxicity
940 of Ni²⁺ to the unicellular green alga *Pseudokirchneriella subcapitata*: Model development and
941 validation with surface waters. *Sci Total Environ* 407: 1901– 1914.
- 942 De Schamphelaere KAC. 2018. Chronic copper gBAM for fish: Investigating possibilities and
943 limitations of a generalised bioavailability model (gBAM) for predicting chronic copper toxicity to
944 freshwater fish. Ghent University, Laboratory for Environmental Toxicology and Aquatic Ecology.

945 Gent, Belgium. [cited 2019 November 10]. Available from:
946 <https://biblio.ugent.be/publication/8562866>

947 De Schamphelaere KAC, Forrez I, Dierckens K, Sorgeloos P, Janssen CR. 2007. Chronic toxicity of
948 dietary copper to *Daphnia magna*. *Aquat Toxicol* 81: 409– 418.

949 De Schamphelaere KAC, Heijerick DG, Janssen CR. 2002. Refinement and field validation of a biotic
950 ligand model predicting acute copper toxicity to *Daphnia magna*. *Comp Biochem Physiol C Toxicol*
951 *Pharmacol* 133: 243– 258.

952 De Schamphelaere KAC, Janssen CR. 2002. A biotic ligand model predicting acute copper toxicity for
953 *Daphnia magna*: The effects of calcium, magnesium, sodium, potassium, and pH. *Environ Sci Technol*
954 36: 48– 54.

955 De Schamphelaere KAC, Janssen CR. 2004a. Development and field validation of a biotic ligand
956 model predicting chronic copper toxicity to *Daphnia magna*. *Environ Toxicol Chem* 23: 1365– 1375.

957 De Schamphelaere KAC, Janssen CR. 2004b. Effects of dissolved organic carbon concentration and
958 source, pH, and water hardness on chronic toxicity of copper to *Daphnia magna*. *Environ Toxicol*
959 *Chem* 23: 1115– 1122.

960 De Schamphelaere KAC, Janssen CR. 2006. Bioavailability models for predicting copper toxicity to
961 freshwater green microalgae as a function of water chemistry. *Environ Sci Technol* 40: 4514– 4522.

962 De Schamphelaere KAC, Lofts S, Janssen CR. 2005a. Bioavailability models for predicting acute and
963 chronic toxicity of zinc to algae, daphnids, and fish in natural surface waters. *Environ Toxicol Chem*
964 24: 1190– 1197.

965 De Schamphelaere KAC, Stauber JL, Wilde KL, Markich SJ, Brown PL, Franklin NM, Creighton NM,
966 Janssen CR. 2005b. Toward a biotic ligand model for freshwater green algae: Surface-bound and
967 internal copper are better predictors of toxicity than free Cu²⁺-ion activity when pH is varied. *Environ*
968 *Sci Technol* 39: 2067– 2072.

969 De Schamphelaere KAC, Vasconcelos FM, Heijerick DG, Tack FMG, Delbeke K, Allen HE, Janssen CR.
970 2003. Development and field validation of a predictive copper toxicity model for the green alga
971 *Pseudokirchneriella subcapitata*. *Environ Toxicol Chem* 22: 2454– 2465.

972 De Schamphelaere KAC, Vasconcelos FM, Tack FMG, Allen HE, Janssen CR. 2004. Effect of dissolved
973 organic matter source on acute copper toxicity to *Daphnia magna*. *Environ Toxicol Chem* 23: 1248–
974 1255.

975 Dew WA, Wood CM, Pyle GG. 2012. Effects of continuous copper exposure and calcium on the
976 olfactory response of fathead minnows. *Environ Sci Technol* 46: 9019– 9026.

977 Di Toro DM, Allen HE, Bergman HL, Meyer JS, Paquin PR, Santore RC. 2001. Biotic ligand model of the
978 acute toxicity of metals. 1. Technical basis. *Environ Toxicol Chem* 20: 2383– 2396.

979 Duarte RM, Smith DS, Val AL, Wood CM. 2016. Dissolved organic carbon from the upper Rio Negro
980 protects zebrafish (*Danio rerio*) against ionoregulatory disturbances caused by low pH exposure. *Sci*
981 *Rep* 6:20377.

982 Dwane GC, Tipping E. 1998. Testing a humic speciation model by titration of copper-amended
983 natural waters. *Environ Int* 24: 609– 616.

- 984 Erickson RJ, Benoit DA, Mattson VR, Nelson HP, Leonard EN. 1996. The effects of water chemistry on
985 the toxicity of copper to fathead minnows. *Environ Toxicol Chem* 15: 181– 193.
- 986 Erickson RJ, Brooke LT, Kahl MD, Vende Venter F, Harting SL, Markee TP, Spehar RL. 1998. Effects of
987 laboratory test conditions on the toxicity of silver to aquatic organisms. *Environ Toxicol Chem* 17:
988 572– 578.
- 989 Eriksen RS, Mackey DJ, Alexander P, Marco RD, Wang XD. 1999. Continuous flow methods for
990 evaluating the response of a copper ion selective electrode to total and free copper in seawater. *J*
991 *Environ Monit* 1: 483– 487.
- 992 European Environment Agency. 2018. Nutrients in freshwater in Europe. [cited 2019 November 10].
993 Available from: [https://www.eea.europa.eu/data-and-maps/indicators/nutrients-in-](https://www.eea.europa.eu/data-and-maps/indicators/nutrients-in-freshwater/nutrients-in-freshwater-assessment-published-6)
994 [freshwater/nutrients-in-freshwater-assessment-published-6](https://www.eea.europa.eu/data-and-maps/indicators/nutrients-in-freshwater/nutrients-in-freshwater-assessment-published-6)
- 995 Farley KJ, Meyer JS. 2015. Metal mixtures model evaluation project: 3. Lessons learned and steps
996 forward. *Environ Toxicol Chem* 34: 821– 832.
- 997 Farley KJ, Meyer JS, Balistrieri LS, De Schamphelaere KAC, Iwasaki Y, Janssen CR, Kamo M, Lofts S,
998 Mebane CA, Naito W, Ryan AC, Santore RC, Tipping E. 2015. Metal mixture modeling evaluation
999 project: 2. Comparison of four modeling approaches. *Environ Toxicol Chem* 34: 741– 753.
- 1000 Franklin NM, Glover CN, Nicol JA, Wood CM. 2005. Calcium/cadmium interactions at uptake surfaces
1001 in rainbow trout: Waterborne versus dietary routes of exposure. *Environ Toxicol Chem* 24: 2954–
1002 2964.
- 1003 Galvez F, Donini A, Playle RC, Smith DS, O'Donnell MJ, Wood CM. 2008. A matter of potential
1004 concern: Natural organic matter alters the electrical properties of fish gills. *Environ Sci Technol* 42:
1005 9385– 9390.
- 1006 Gao C, De Schamphelaere KAC, Smolders E. 2016. Zinc toxicity to the alga *Pseudokirchneriella*
1007 *subcapitata* decreases under phosphate limiting growth conditions. *Aquat Toxicol* 173: 74– 82.
- 1008 Garman ER, Meyer JS, Bergeron CM, Blewett TA, Clements WH, Elias MC, Farley KJ, Gissi F, Ryan AC
1009 2020. Validation of bioavailability-based toxicity models for metals. *Environ Toxicol Chem* 39: 101–
1010 117.
- 1011 Glover CN, Playle RC, Wood CM. 2005. Heterogeneity of natural organic matter amelioration of silver
1012 toxicity to *Daphnia magna*: Effect of source and equilibration time. *Environ Toxicol Chem* 24: 2934–
1013 2940.
- 1014 Golding LA, Borgmann U, Dixon DG. 2013. Cadmium bioavailability to *Hyalella azteca* from a
1015 periphyton diet compared to an artificial diet and application of a biokinetic model. *Aquat Toxicol*
1016 126: 291– 298.
- 1017 Green WW, Mirza RS, Wood CM, Pyle GG. 2010. Copper binding dynamics and olfactory impairment
1018 in fathead minnows (*Pimephales promelas*). *Environ Sci Technol* 44: 1431– 1437.
- 1019 Gustafsson JP. 2001. Modeling the acid–base properties and metal complexation of humic
1020 substances with the Stockholm humic model. *J Colloid Interface Sci* 244: 102– 112.
- 1021 Hansen JA, Lipton J, Welsh PG. 2002. Relative sensitivity of bull trout (*Salvelinus confluentus*) and
1022 rainbow trout (*Oncorhynchus mykiss*) to acute copper toxicity. *Environ Toxicol Chem* 21: 633– 639.

- 1023 Heijerick DG, De Schamphelaere KAC, Janssen CR. 2002. Predicting acute zinc toxicity for *Daphnia*
1024 *magna* as a function of key water chemistry characteristics: Development and validation of a biotic
1025 ligand model. *Environ Toxicol Chem* 21: 1309– 1315.
- 1026 Heijerick DG, De Schamphelaere KAC, Van Sprang PA, Janssen CR. 2005. Development of a chronic
1027 zinc biotic ligand model for *Daphnia magna*. *Ecotoxicol Environ Saf* 62: 1– 10.
- 1028 Heugens EHW, Jager T, Creyghton R, Kraak MHS, Hendriks AJ, Van Straalen NM, Admiraal W. 2003.
1029 Temperature-dependent effects of cadmium on *Daphnia magna*: Accumulation versus sensitivity.
1030 *Environ Sci Technol* 37: 2145– 2151.
- 1031 Hodson PV, Sprague JB. 1975. Temperature-induced changes in acute toxicity of zinc to Atlantic
1032 salmon (*Salmo salar*). *Journal of the Fisheries Research Board of Canada* 33: 1– 10.
- 1033 Holm-Jensen I. 1948. Osmotic regulation in *Daphnia magna* under physiological conditions and in the
1034 presence of heavy metals. *Det Kongelige Danske Videnskabernes Selskab Biologiske Meddelelser*,
1035 Vol 20. Copenhage, Denmark.
- 1036 Hook SE, Fisher NS. 2001. Reproductive toxicity of metals in calanoid copepods. *Mar Biol* 138: 1131–
1037 1140.
- 1038 Hook SE, Fisher NS. 2002. Relating the reproductive toxicity of five ingested metals in calanoid
1039 copepods with sulfur affinity. *Mar Environ Res* 53: 161– 174.
- 1040 Hooper HL, Connon R, Callaghan A, Fryer G, Yarwood-Buchanan S, Biggs J, Maund SJ, Hutchinson TH,
1041 Sibly RM. 2008. The ecological niche of *Daphnia magna* characterized using population growth rate.
1042 *Ecology* 89: 1015– 1022.
- 1043 Hoppe S, Garmo ØA, Leppanen MT, Borg H, Ndungu K. 2015a. Soft and sour: The challenge of setting
1044 environmental quality standards for bioavailable metal concentration in Fennoscandinavian
1045 freshwaters. *Environ Sci Policy* 54: 210– 217.
- 1046 Hoppe S, Gustafsson JP, Borg H, Breitholtz M. 2015b. Evaluation of current copper bioavailability
1047 tools for soft freshwaters in Sweden. *Ecotoxicol Environ Saf* 114: 143– 149.
- 1048 Irving EC, Baird DJ, Culp JM. 2003. Ecotoxicological responses of the mayfly *Baetis tricaudatus* to
1049 dietary and waterborne cadmium: Implications for toxicity testing. *Environ Toxicol Chem* 22: 1058–
1050 1064.
- 1051 Janes N, Playle RC. 1995. Modeling silver binding to gills of rainbow trout (*Oncorhynchus mykiss*).
1052 *Environ Toxicol Chem* 14: 1847– 1858.
- 1053 Jones JRE. 1938. The relative toxicity of salts of lead, zinc and copper to the stickleback (*Gasterosteus*
1054 *aculeatus* L.) and the effect of calcium on the toxicity of lead and zinc salts. *J Exp Biol* 15: 394– 407.
- 1055 Kayhanian M, Stransky C, Bay S, Lau SL, Stenstrom MK. 2008. Toxicity of urban highway runoff with
1056 respect to storm duration. *Sci Total Environ* 389: 386– 406.
- 1057 Kim SD, Ma H, Allen HE, Cha DK. 1999. Influence of dissolved organic matter on the toxicity of copper
1058 to *Ceriodaphnia dubia*: Effect of complexation kinetics. *Environ Toxicol Chem* 18: 2433– 2437.
- 1059 Kinniburgh DG, Milne CJ, Benedetti MF, Pinheiro JP, Filius J, Koopal LK, Van Riemsdijk WH. 1996.
1060 Metal ion binding by humic acid: Application of the NICA-Donnan model. *Environ Sci Technol* 30:
1061 1687– 1698.

- 1062 Kinraide TB. 2006. Plasma membrane surface potential (ψ_{PM}) as a determinant of ion bioavailability:
1063 A critical analysis of new and published toxicological studies and a simplified method for the
1064 computation of plant ψ_{PM} . *Environ Toxicol Chem* 25: 3188– 3198.
- 1065 Kolts JM, Boese CJ, Meyer JS. 2009. Effects of dietborne copper and silver on reproduction by
1066 *Ceriodaphnia dubia*. *Environ Toxicol Chem* 28: 71– 85.
- 1067 Lavoie M, Le Faucheur S, Boullemant A, Fortin C, Campbell PGC. 2012. The influence of pH on algal
1068 cell membrane permeability and its implications for the uptake of lipophilic metal complexes. *J*
1069 *Phycol* 48: 293– 302.
- 1070 Lloyd R, Herbert DWM. 1962. The effect of the environment on the toxicity of poisons to fish.
1071 *Journal of the Institution of Public Health Engineers* 61: 132– 145.
- 1072 Lodge DM, Brown KM, Klosiewski SP, Stein RA, Covich AP, Leathers BK, Brönmark C. 1987.
1073 Distribution of freshwater snails: Spatial scale and the relative importance of physicochemical and
1074 biotic factors. *Am Malacol Bull* 5: 73– 84.
- 1075 Web of Science® Lofts S, Tipping E. 2011. Assessing WHAM/model VII against field measurements of
1076 free metal ion concentrations: Model performance and the role of uncertainty in parameters and
1077 inputs. *Environ Chem* 8: 501– 516.
- 1078 Louis Y, Garnier C, Lenoble V, Mounier S, Cukrov N, Omanović D, Pižeta I. 2009. Kinetic and
1079 equilibrium studies of copper–dissolved organic matter complexation in water column of the
1080 stratified Krka River estuary (Croatia). *Mar Chem* 114: 110– 119.
- 1081 Ma H, Kim SD, Cha DK, Allen HE. 1999. Effects of kinetics of complexation by humic acid on toxicity of
1082 copper to *Ceriodaphnia dubia*. *Environ Toxicol Chem* 18: 828– 837.
- 1083 Macdonald A, Silk L, Schwartz ML, Playle RC. 2002. A lead–gill binding model to predict acute lead
1084 toxicity to rainbow trout (*Oncorhynchus mykiss*). *Comp Biochem Physiol C Toxicol Pharmacol* 133:
1085 227– 242.
- 1086 MacRae RK, Smith DE, Swoboda-Colberg N, Meyer JS, Bergman HL. 1999. Copper binding affinity of
1087 rainbow trout (*Oncorhynchus mykiss*) and brook trout (*Salvelinus fontinalis*) gills: Implications for
1088 assessing bioavailable metal. *Environ Toxicol Chem* 18: 1180– 1189.
- 1089 McDonald DG, Wood CM. 1993. Branchial mechanisms of acclimation to metals in freshwater fish. In
1090 JC Rankin, FB Jensen, eds, *Fish Ecophysiology*, Vol 9—Chapman & Hall Fish and Fisheries Series.
1091 Springer, Dordrecht, the Netherlands, pp 297– 321.
- 1092 McGeer JC, Playle RC, Wood CM, Galvez F. 2000. A physiologically based biotic ligand model for
1093 predicting the acute toxicity of waterborne silver to rainbow trout in freshwaters. *Environ Sci*
1094 *Technol* 34: 4199– 4207.
- 1095 McIntyre JK, Baldwin DH, Meador JP, Scholz NL. 2008. Chemosensory deprivation in juvenile coho
1096 salmon exposed to dissolved copper under varying water chemistry conditions. *Environ Sci Technol*
1097 42: 1352– 1358.
- 1098 Mebane CA, Hennessy DP, Dillon FS. 2010. Incubating rainbow trout in soft water increased their
1099 later sensitivity to cadmium and zinc. *Water Air Soil Pollut* 205: 245– 250.
- 1100 Mebane CA, Schmidt TS, Balistriero LS. 2017. Larval aquatic insect responses to cadmium and zinc in
1101 experimental streams. *Environ Toxicol Chem* 36: 749– 762.

- 1102 Meyer JS, Adams WJ. 2010. Relationship between biotic ligand model–based water quality criteria
1103 and avoidance and olfactory responses to copper by fish. *Environ Toxicol Chem* 29: 2096– 2103.
- 1104 Meyer JS, Adams WJ, Brix KV, Luoma SN, Mount DR, Stubblefield WA, Wood CM, eds. 2005. Toxicity
1105 of Dietborne Metals to Aquatic Organisms. Society of Environmental Toxicology and Chemistry,
1106 Pensacola, FL, USA.
- 1107 Meyer JS, DeForest DK. 2018. Protectiveness of copper water quality criteria against impairment of
1108 behavior and chemo/mechanosensory responses: An update. *Environ Toxicol Chem* 37: 1260– 1279.
- 1109 Meyer JS, Farley KJ, Garman ER. 2015. Metal mixtures modeling evaluation: 1. Background. *Environ*
1110 *Toxicol Chem* 34: 726– 740.
- 1111 Meyer JS, Santore RC, Bobbitt JP, Debrey LD, Boese CJ, Paquin PR, Allen HE, Bergman HL, Di Toro
1112 DM. 1999. Binding of nickel and copper to fish gills predicts toxicity when water hardness varies, but
1113 free-ion activity does not. *Environ Sci Technol* 33: 913– 916.
- 1114 Meylan S, Behra R, Sigg L. 2003. Accumulation of copper and zinc in periphyton in response to
1115 dynamic variations of metal speciation in freshwater. *Environ Sci Technol* 37: 5204– 5212.
- 1116 Mirza RS, Green WW, Connor S, Weeks ACW, Wood CM, Pyle GG. 2009. Do you smell what I smell?
1117 Olfactory impairment in wild yellow perch from metal-contaminated waters. *Ecotoxicol Environ Saf*
1118 72: 677– 683.
- 1119 Morel FMM. 1983. *Principles of Aquatic Chemistry*. Wiley Interscience, New York, NY, USA.
- 1120 Morgan TP, Wood CM. 2004. A relationship between gill silver accumulation and acute silver toxicity
1121 in the freshwater rainbow trout: Support for the acute silver biotic ligand model. *Environ Toxicol*
1122 *Chem* 23: 1261– 1267.
- 1123 Naddy RB, Rehner AB, McNerney GR, Gorsuch JW, Kramer JR, Wood CM, Paquin PR, Stubblefield
1124 WA. 2007. Comparison of short-term chronic and chronic silver toxicity to fathead minnows in
1125 unamended and sodium chloride–amended waters. *Environ Toxicol Chem* 26: 1922– 1930.
- 1126 Naddy RB, Stubblefield WA, Bell RA, Wu KB, Santore RC, Paquin PR. 2018. Influence of varying water
1127 quality parameters on the acute toxicity of silver to the freshwater cladoceran, *Ceriodaphnia dubia*.
1128 *Bull Environ Contam Toxicol* 100: 69– 75.
- 1129 Natale OE, Gómez CE, Leis MV. 2007. Application of the biotic ligand model for regulatory purposes
1130 to selected rivers in Argentina with extreme water-quality characteristics. *Integr Environ Assess*
1131 *Manag* 3: 517– 528.
- 1132 Ng TY-T, Chowdhury MJ, Wood CM. 2010. Can the biotic ligand model predict Cu toxicity across a
1133 range of pHs in softwater-acclimated rainbow trout? *Environ Sci Technol* 44: 6263– 6268.
- 1134 Nimick DA, Gammon JR, Parker SR. 2011. Diel biogeochemical processes and their effect on the
1135 aqueous chemistry of streams: A review. *Chem Geol* 283: 3– 17.
- 1136 Niyogi S, Kent R, Wood CM. 2008. Effects of water chemistry variables on gill binding and acute
1137 toxicity of cadmium in rainbow trout (*Oncorhynchus mykiss*): A biotic ligand model (BLM) approach.
1138 *Comp Biochem Physiol C Toxicol Pharmacol* 148: 305– 314.

- 1139 Niyogi S, Nadella SR, Wood CM. 2015. Interactive effects of waterborne metals in binary mixtures on
1140 short-term gill-metal binding and ion uptake in rainbow trout (*Oncorhynchus mykiss*). *Aquat Toxicol*
1141 165: 109– 119.
- 1142 Niyogi S, Wood CM. 2003. Effects of chronic waterborne and dietary metal exposures on gill metal-
1143 binding: Implications for the biotic ligand model. *Hum Ecol Risk Assess* 9: 813– 846.
- 1144 Niyogi S, Wood CM. 2004. Biotic ligand model, a flexible tool for developing site-specific water
1145 quality guidelines for metals. *Environ Sci Technol* 38: 6177– 6192.
- 1146 Nys C, Janssen CR, De Schamphelaere KAC. 2013. Pb-diet: An investigation of the potential toxicity of
1147 dietary Pb to *Ceriodaphnia dubia*. Report prepared for International Lead Zinc Research
1148 Organization, Durham, NC, USA. [cited 2019 November 10]. Available from: [https://www.ila-
1149 lead.org/UserFiles/File/UGent-Pb_Diet_final_report_July9th2013.pdf](https://www.ila-lead.org/UserFiles/File/UGent-Pb_Diet_final_report_July9th2013.pdf)
- 1150 Nys, C, Janssen, CR, Van Sprang, P, and De Schamphelaere, KAC. 2016. The effect of pH on chronic
1151 aquatic nickel toxicity is dependent on the pH itself: Extending the chronic nickel bioavailability
1152 models. *Environ Toxicol Chem* 35: 1097– 1106.
- 1153 Nys C, Janssen CR, Mager EM, Esbaugh AJ, Brix KV, Grosell M, Stubblefield WA, Holtze K, De
1154 Schamphelaere KAC. 2014. Development and validation of a biotic ligand model for predicting
1155 chronic toxicity of lead to *Ceriodaphnia dubia*. *Environ Toxicol Chem* 33: 394– 403.
- 1156 Nys C, Van Regenmortel T, Janssen CR, Oorts K, Smolders E, De Schamphelaere KAC. 2018. A
1157 framework for ecological risk assessment of metal mixtures in aquatic systems. *Environ Toxicol*
1158 *Chem* 37: 623– 642.
- 1159 Nys C, Versieren L, Cordery KI, Blust R, Smolders E, De Schamphelaere KAC. 2017. Systematic
1160 evaluation of chronic metal-mixture toxicity to three species and implications for risk assessment.
1161 *Environ Sci Technol* 51: 4615– 4623.
- 1162 Organisation for Economic Co-operation and Development. 2011. Test No. 201: Freshwater alga and
1163 cyanobacteria, growth inhibition test. OECD Guidelines for the Testing of Chemicals. Paris, France.
- 1164 Pagenkopf GK. 1983. Gill surface interaction model for trace-metal toxicity to fishes: Role of
1165 complexation, pH, and water hardness. *Environ Sci Technol* 17: 342– 347.
- 1166 Paquin PR, Di Toro DM. 2008. Silver biotic ligand model (BLM): Refinement of an acute BLM for
1167 silver. 99-ECO-1-2T. Water Environment Research Foundation, Alexandria, VA, USA.
- 1168 Paquin PR, Gorsuch JW, Apte S, Batley GE, Bowles KC, Campbell PGC, Delos CG, Di Toro DM, Dwyer
1169 FJ, Galvez F, Gensemer RW, Goss GC, Hogstrand C, Janssen CR, McGeer JC, Naddy RB, Playle RC,
1170 Santore RC, Schneider U, Stubblefield WA, Wood CM, Wu KB. 2002. The biotic ligand model: A
1171 historical overview. *Comp Biochem Physiol C Toxicol Pharmacol* 133: 3– 35.
- 1172 Paquin PR, Santore RC, Wu KB, Kavvas CD, Di Toro DM. 2000. The biotic ligand model: A model of
1173 the acute toxicity of metals to aquatic life. *Environ Sci Policy* 3(Suppl. 1): 175– 182.
- 1174 Pereira CMS, Deruytter D, Blust R, De Schamphelaere KAC. 2017. Effect of temperature on chronic
1175 toxicity of copper, zinc, and nickel to *Daphnia magna*. *Environ Toxicol Chem* 36: 1909– 1916.
- 1176 Peters A, Lofts S, Merrington G, Brown B, Stubblefield W, Harlow K. 2011. Development of biotic
1177 ligand models for chronic manganese toxicity to fish, invertebrates, and algae. *Environ Toxicol Chem*
1178 30: 2407– 2415.

- 1179 Peters A, Schlekot CE, Merrington G. 2016. Does the scientific underpinning of regulatory tools to
1180 estimate bioavailability of nickel in freshwaters matter? The European-wide environmental quality
1181 standard for nickel. *Environ Toxicol Chem* 35: 2397–2404.
- 1182 Playle RC. 2004. Using multiple metal–gill binding models and the toxic unit concept to help
1183 reconcile multiple-metal toxicity results. *Aquat Toxicol* 67: 359–370.
- 1184 Playle RC, Dixon DG, Burnison BK. 1993a. Copper and cadmium binding to fish gills: Estimates of
1185 metal–gill stability constants and modelling of metal accumulation. *Can J Fish Aquat Sci* 50: 2678–
1186 2687.
- 1187 Playle RC, Dixon DG, Burnison BK. 1993b. Copper and cadmium binding to fish gills: Modification by
1188 dissolved organic carbon and synthetic ligands. *Can J Fish Aquat Sci* 50: 2667–2677.
- 1189 Playle RC, Wood CM. 1989. Water chemistry changes in the gill micro-environment of rainbow trout:
1190 Experimental observations and theory. *J Comp Physiol B Biochem Syst Environ Physiol* 159: 527–
1191 537.
- 1192 CASPyle GG, Wood CM. 2007. Predicting non-scents: Rationale for a chemosensory-based biotic
1193 ligand model. *Australian Journal of Ecotoxicology* 13: 47–51.
- 1194 CASRao DGVP, Khan MAQ. 2000. Zebra mussels: Enhancement of copper toxicity by high
1195 temperature and its relationship with respiration and metabolism. *Water Environ Res* 72: 175–178.
- 1196 Rodriguez PH, Arbildua J, Villavicencio G, Mejías R, Urrestarazu P, Jiménez M. 2012. Copper acute
1197 and chronic toxicity to *D. magna*: Sensitivity at three different hardness at pH 6.3 (MES buffered) in
1198 the presence of 2 mg/L DOC. Chilean Mining and Metallurgy Research Center, Laboratory of
1199 Ecotoxicology and Chemistry of Trace Metals, Centro de Investigación Minera y Metalúrgica,
1200 Santiago, Chile.
- 1201 Rogers JT, Patel M, Gilmour KM, Wood CM. 2005. Mechanisms behind Pb-induced disruption of Na⁺
1202 and Cl[–] balance in rainbow trout (*Oncorhynchus mykiss*). *Am J Physiol Regul Integr Comp Physiol*
1203 289: R463–R472.
- 1204 Rogers JT, Wood CM. 2004. Characterization of branchial lead–calcium interaction in the freshwater
1205 rainbow trout *Oncorhynchus mykiss*. *J Exp Biol* 207: 813–825.
- 1206 Roy I, Hare L. 1999. Relative importance of water and food as cadmium sources to the predatory
1207 insect *Sialis velata* (Megaloptera). *Can J Fish Aquat Sci* 56: 1143–1149.
- 1208 R Salminen, ed. 2005. *Geochemical Atlas of Europe. Part 1: Background Information, Methodology
1209 and Maps.* Geological Survey of Finland, Espoo, Finland. [cited 2019 November 10]. Available from:
1210 <http://www.gtk.fi/publ/foregsatlas/>
- 1211 Santore RC, Paquin PR, Di Toro DM, Allen HE, Meyer JS. 2001. Biotic ligand model of the acute
1212 toxicity of metals. 2. Application to acute copper toxicity in freshwater fish and *Daphnia*. *Environ
1213 Toxicol Chem* 20: 2397–2402.
- 1214 Santore RC, Ryan AC, Kroglund F, Teien HC, Rodriguez PH, Stubblefield WA, Cardwell AS, Adams WJ,
1215 Nordheim E. 2018. Development and application of a biotic ligand model for predicting the toxicity
1216 of dissolved and precipitated aluminum. *Environ Toxicol Chem* 37: 70–79.

- 1217 Schlekot CE, Van Genderen EJ, De Schamphelaere KAC, Antunes PMC, Rogevich EC, Stubblefield WA.
1218 2010. Cross-species extrapolation of chronic nickel biotic ligand models. *Sci Total Environ* 408: 6148–
1219 6157.
- 1220 Sigg L, Black F, Buffle J, Cao J, Cleven R, Davison W, Galceran J, Gunkel P, Kalis E, Kistler D, Martin M,
1221 Noël S, Nur Y, Odzak N, Puy J, van Riemsdijk W, Temminghoff E, Tercier-Waeber M-L, Toepperwien S,
1222 Town RM, Unsworth E, Warnken KW, Weng L, Xue H, Zhang H. 2006. Comparison of analytical
1223 techniques for dynamic trace metal speciation in natural freshwaters. *Environ Sci Technol* 40: 1934–
1224 1941.
- 1225 Steinberg CEW, Kamara S, Prokhotskaya VY, Manusadžianas L, Karasyova TA, Timofeyev MA, Jie Z,
1226 Paul A, Meinelt T, Farjalla VF, Matsuo AYO, Burnison BK, Menzel R. 2006. Dissolved humic
1227 substances—ecological driving forces from the individual to the ecosystem level? *Freshw Biol* 51:
1228 1189– 1210.
- 1229 Stephan CE, Mount DI, Hansen DJ, Gentile JH, Chapman GA, Brungs WA. 1985. Guidelines for
1230 deriving numerical national water quality criteria for the protection of aquatic organisms and their
1231 uses. EPA 822-R-85-100. US Environmental Protection Agency, Duluth, MN; Narragansett, RI; and
1232 Corvallis, OR.
- 1233 Stephenson M, Turner MA. 1993. A field study of cadmium dynamics in periphyton and in *Hyalella*
1234 *azteca* (Crustacea: Amphipoda). *Water Air Soil Pollut* 68: 341– 361.
- 1235 Stockdale A, Tipping E, Lofts S, Ormerod SJ, Clements WH, Blust R. 2010. Toxicity of proton–metal
1236 mixtures in the field: Linking stream macroinvertebrate species diversity to chemical speciation and
1237 bioavailability. *Aquat Toxicol* 100: 112– 119.
- 1238 Stumm W, Morgan JJ. 1996. *Aquatic Chemistry: Chemical Equilibria and Rates in Natural Waters*, 3rd
1239 ed. Wiley Interscience, New York, NY, USA.
- 1240 Tait TN, Rabson LM, Diamond RL, Cooper CA, McGeer JC, Smith DS. 2016. Determination of cupric
1241 ion concentrations in marine waters: An improved procedure and comparison with other speciation
1242 methods. *Environ Chem* 13: 140– 148.
- 1243 Timmermans KR, Spijkerman E, Tonkes M, Govers H. 1992. Cadmium and zinc uptake by two species
1244 of aquatic invertebrate predators from dietary and aqueous sources. *Can J Fish Aquat Sci* 49: 655–
1245 662.
- 1246 Tipping E. 1994. WHAM—A chemical equilibrium model and computer code for waters, sediments,
1247 and soils incorporating a discrete site/electrostatic model of ion-binding by humic substances.
1248 *Comput Geosci* 20: 973– 1023.
- 1249 Tipping E. 1998. Humic Ion-Binding Model VI: An improved description of the interactions of protons
1250 and metal ions with humic substances. *Aquat Geochem* 4: 3– 48.
- 1251 Tipping E, Hurley MA. 1992. A unifying model of cation binding by humic substances. *Geochim*
1252 *Cosmochim Acta* 56: 3627– 3641.
- 1253 Tipping E, Lofts S. 2013. Metal mixture toxicity to aquatic biota in laboratory experiments;
1254 application of the WHAM-FTOX model. *Aquat Toxicol* 142–143: 114– 122.
- 1255 Tipping E, Lofts S. 2014. Testing WHAM-FTOX with laboratory toxicity data for mixtures of metals
1256 (Cu, Zn, Cd, Ag, Pb). *Environ Toxicol Chem* 34: 788– 798.

- 1257 Web of Science®Tipping E, Lofts S, Sonke JE. 2011. Humic Ion-Binding Model VII: A revised
1258 parameterisation of cation-binding by humic substances. *Environ Chem* 8: 225– 235.
- 1259 Tipping E, Stockdale A, Lofts S. 2019. Systematic analysis of freshwater metal toxicity with WHAM-
1260 FTOX. *Aquat Toxicol* 212: 128– 137.
- 1261 Todd AS, Brinkman SF, Wolf RE, Lamothe PJ, Smith KS, Ranville JF. 2009. Use of an enriched stable-
1262 isotope approach to determine the gill-zinc binding properties of juvenile rainbow trout
1263 (*Oncorhynchus mykiss*) during acute zinc exposures in hard and soft waters. *Environ Toxicol Chem*
1264 28: 1233– 1243.
- 1265 Tomczyk N, Parr TB, Gray E, Iburg J, Capps K. 2018. Trophic strategies influence metal
1266 bioaccumulation in detritus-based, aquatic food webs. *Environ Sci Technol* 52: 11886– 11894.
- 1267 Unsworth ER, Warnken KW, Zhang H, Davison W, Black F, Buffle J, Cao Y, Cleven RFMJ, Galceran J,
1268 Gunkel P, Kalis E, Kistler D, van Leeuwen HP, Martin M, Noël S, Nur Y, Odzak N, Puy J, van Riemsdijk
1269 W, Sigg L, Temminghoff E, Tercier-Waeber M-L, Toepperwien S, Town RM, Weng L, Xue H. 2006.
1270 Model predictions of metal speciation in freshwaters compared to measurements by in situ
1271 techniques. *Environ Sci Technol* 40: 1942– 1949.
- 1272 US Environmental Protection Agency. 1986. Quality criteria for water. EPA 440/5-86-001.
1273 Washington, DC.
- 1274 US Environmental Protection Agency. 2002. Short-term methods for estimating the chronic toxicity
1275 of effluents and receiving waters to freshwater organisms, 4th ed. EPA-821-R-02-013. Cincinnati, OH.
- 1276 US Environmental Protection Agency. 2007. Aquatic life ambient freshwater quality criteria—
1277 Copper. EPA-822-R-07-001. Washington, DC.
- 1278 US Environmental Protection Agency. 2017. Draft aquatic life ambient water quality criteria for
1279 aluminum. EPA-822-P-17-001. Washington, DC.
- 1280 Vandenberg JA, Ryan MC, Nuell DD, Chu A. 2005. Field evaluation of mixing length and attenuation
1281 of nutrients and fecal coliform in a wastewater effluent plume. *Environ Monit Assess* 107: 45– 57.
- 1282 Van Genderen EJ, Ryan AC, Tomasso JR, Klaine SJ. 2005. Evaluation of acute copper toxicity to larval
1283 fathead minnows (*Pimephales promelas*) in soft surface waters. *Environ Toxicol Chem* 24: 408– 414.
- 1284 Van Genderen EJ, Stauber JL, Delos C, Eignor D, Gensemer RW, McGeer J, Merrington G, Whitehouse
1285 P. 2020. Best practices and derivation and application of thresholds for metals using bioavailability-
1286 based approaches. *Environ Toxicol Chem* 39: 118– 130.
- 1287 van Leeuwen HP, Town RM. 2005. Kinetic limitations in measuring stabilities of metal complexes by
1288 competitive ligand exchange-adsorptive stripping voltammetry (CLE-AdSV). *Environ Sci Technol* 39:
1289 7217– 7225.
- 1290 Van Regenmortel T, Janssen CR, De Schamphelaere KAC. 2015. Comparison of the capacity of two
1291 biotic ligand models to predict chronic copper toxicity to two *Daphnia magna* clones and formulation
1292 of a generalized bioavailability model. *Environ Toxicol Chem* 34: 1597– 1608.
- 1293 Van Regenmortel T, Nys C, Janssen CR, Lofts S, De Schamphelaere KAC. 2017. Comparison of four
1294 methods for bioavailability-based risk assessment of mixtures of Cu, Zn and Ni in freshwater. *Environ*
1295 *Toxicol Chem* 36: 2123– 2138.

- 1296 Van Sprang PA, Nys C, Blust RJP, Chowdhury J, Gustafsson JP, Janssen CJ, De Schamphelaere KAC.
1297 2016. The derivation of effects threshold concentrations of lead for European freshwater
1298 ecosystems. *Environ Toxicol Chem* 35: 1310– 1320.
- 1299 Van Sprang PA, Vangheluwe ML, Van Hyfte A, Heijerick DG, Vandenbroele M, Verdonck FAM,
1300 Delbeke K, Dwyer RL, Adams WJ. 2008. Effects to freshwater organisms. In European Union Risk
1301 Assessment Report: Voluntary risk assessment of copper, copper II sulphate pentahydrate,
1302 copper(I)oxide, copper(I)oxide, dicopper chloride trihydroxide. European Copper Institute, Brussels,
1303 Belgium. p 194. [cited 2019 November 10]. Available from: [http://echa.europa.eu/copper-voluntary-](http://echa.europa.eu/copper-voluntary-risk-assessment-reports)
1304 [risk-assessment-reports](http://echa.europa.eu/copper-voluntary-risk-assessment-reports)
- 1305 Van Sprang PA, Verdonck FAM, Van Assche F, Regoli L, De Schamphelaere KAC. 2009. Environmental
1306 risk assessment of zinc in European freshwaters: A critical appraisal. *Sci Total Environ* 407: 5373–
1307 5391.
- 1308 Veltman K, Huijbregts MAJ, Hendriks AJ. 2010. Integration of biotic ligand models (BLM) and
1309 bioaccumulation kinetics into a mechanistic framework for metal uptake in aquatic organisms.
1310 *Environ Sci Technol* 44: 5022– 5028.
- 1311 Verschoor AJ, Vijver MG, Vink JPM. 2017. Refinement and cross-validation of nickel bioavailability in
1312 PNEC-Pro, a regulatory tool for site-specific risk assessment of metals in surface water. *Environ*
1313 *Toxicol Chem* 36: 2367– 2376.
- 1314 Vukov O, Smith DS, McGeer JC. 2016. Acute dysprosium toxicity to *Daphnia pulex* and *Hyalella*
1315 *azteca* and development of the biotic ligand approach. *Aquat Toxicol* 170: 142– 151.
- 1316 Wang N, Mebane CA, Kunz JL, Ingersoll CG, Brumbaugh WG, Santore RC, Gorsuch JW, Arnold WR.
1317 2011. Influence of DOC on toxicity of copper to a unionid mussel (*Villosa iris*) and a cladoceran
1318 (*Ceriodaphnia dubia*) in acute and chronic water exposures. *Environ Toxicol Chem* 30: 2115– 2125.
- 1319 Welsh PG, Lipton J, Mebane CA, Marr JCA. 2008. Influence of flow-through and renewal exposures
1320 on the toxicity of copper to rainbow trout. *Ecotoxicol Environ Saf* 69: 199– 208.
- 1321 Wood CM, Al-Reasi HA, Smith DS. 2011. The two faces of DOC. *Aquat Toxicol* 105(Suppl.): 3– 8.
- 1322 CM Wood, AP Farrell, CJ Brauner, eds. 2012a. Homeostasis and Toxicology of Essential Metals, Vol
1323 31A—Fish Physiology. Academic, London, UK.
- 1324 CM Wood, AP Farrell, CJ Brauner, eds. 2012b. Homeostasis and Toxicology of Non-Essential Metals,
1325 Vol 31A—Fish Physiology. Academic, London, UK.
- 1326 Xie L, Funk DH, Buchwalter DB. 2010. Trophic transfer of Cd from natural periphyton to the grazing
1327 mayfly *Centroptilum triangulifer* in a life cycle test. *Environ Pollut* 158: 272– 277.
- 1328 Xue H, Sigg L. 1999. Comparison of the complexation of Cu and Cd by humic or fulvic acids and by
1329 ligands observed in lake waters. *Aquat Geochem* 5: 313– 335.
- 1330 Zhao C-M, Campbell PGC, Wilkinson KJ. 2016. When are metal complexes bioavailable? *Environ*
1331 *Chem* 13: 425– 433.
- 1332 Zitko P, Carson WV, Carson WG. 1973. Prediction of incipient lethal levels of copper to juvenile
1333 Atlantic salmon in the presence of humic acid by cupric electrode. *Bull Environ Contam Toxicol* 10:
1334 265– 271.

1335 Table 1. Summary of studies in which physiological mechanisms (e.g., gill enzyme inhibition or body ion loss) and/or the actual metal burden at the biotic
 1336 ligand (e.g., gill metal concentration) have been measured

Metal	Organism	Physiological endpoints	Toxicity endpoints	Sources
Ag	Daphnia magna	1-h whole-body accumulation, gill enzyme inhibition	Mortality not measured	(Bianchini and Wood 2003)
Ag	Rainbow trout	2- to 3 h accumulations, gill enzyme inhibition	Accumulation and 96-h mortality data related across different studies	(Janes and Playle 1995; McGeer et al. 2000)
Ag	Rainbow trout	3- and 24-h gill accumulation	Mortality at 96 h	(Morgan and Wood 2004)
Al	Atlantic salmon	140-h accumulation	Mortality at 140 h	(Santore et al. 2018)
Cd	Rainbow trout	3-h gill accumulation	Mortality at 96 h	(Niyogi et al. 2008)
Cd, Cu	Fathead minnow	2- to 3-h gill accumulations	Mortality at 96 h	(Playle et al. 1993a, 1993b)
Cd, Cu, Pb, Zn	Rainbow trout	0.75- to 24-h gill accumulation	Accumulation and 96-h mortality data related across different studies	(Balistrieri and Mebane 2014)
Cd, Pb	Rainbow trout	3- and 24-h gill accumulation; Ca ²⁺ and Na ⁺ influx	Mortality at 96 h	(Birceanu et al. 2008)
Cu	Rainbow trout	24-h gill accumulations	Mortality at 120 h	(MacRae et al. 1999)
Cu	Rainbow trout	24-h gill accumulations	Mortality at 96 h and 30 d	(Ng et al. 2010)
Cu	Rainbow trout	24-h gill accumulations from previous work	Mortality at 96 h and 30 d	(Crémazy et al. 2017)
Ni, Cu	Fathead minnow	2- to 3-h gill accumulation	Mortality at 96 h	(Meyer et al. 1999)
Pb	Rainbow trout	3-h gill accumulation	Time to mortality of a single concentration in different waters	(Macdonald et al. 2002)
Pb	Rainbow trout	0- to 96-h Pb accumulation, enzyme inhibition, ion flux rates	Mortality not measured	(Rogers et al. 2005; Rogers and Wood 2004)
Zn	Rainbow trout	0.5- to 72-h gill accumulations	Mortality at 96 h	(Alsop and Wood 2000)
Zn	Rainbow trout	0.75- and 3-h gill accumulations	Mortality at 96 h	(Todd et al. 2009)

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1339 Table 2. Selected examples of available freshwater bioavailability-based modes of metal toxicity

Metal	Model type or name	Reference	Organisms used in development	Organisms presumed applicable to	Necessary inputs	Example applications	Notes
Al	Acute and chronic BLM	(Santore et al. 2018)	Various fish, invertebrates and green algae	All freshwater aquatic life	"BLM"	Model supported a simplified MLR model which became federal water quality guidance for United States (US Environmental Protection Agency 2017)	Toxicity is considered a function of both dissolved and precipitated Al, caused by either ionoregulatory or respiratory disturbance depending on pH
Cd	Acute BLM	(Niyogi et al. 2008)	Rainbow trout		"BLM"		Classic BLM with binding coefficients derived from measured gill accumulation
Cd	Acute BLM	(Clifford and McGeer 2010)	<i>Daphnia pulex</i>		BLM		Binding coefficients calculated from toxicity values from a wide variety of water types
Cu	Acute BLM	(Di Toro et al. 2001; Santore et al. 2001; US Environmental Protection Agency 2007)	Fish, <i>Daphnia</i>	All freshwater animals	"BLM"	Acute model extrapolated to chronic criterion by ACR; adopted as federal water quality guidance in United States;	Me-BL log K values from fish gill accumulation tests, toxicity data used to adjust CA values for <i>Daphnia</i> and others

						some adoption by states	
Cu	Generalized bioavailability model (gBAM)	(De Schampelaere et al. 2003; Van Sprang et al. 2008)	Green algae	All primary producers	pH ("BLM" set needed for FIA calculation)	ERA for REACH, EQS (Europe)	pH only factor affecting algal growth when expressed as FIA; full "BLM" chemistry needed for FIA
Cu	Chronic BLM	(De Schampelaere and Janssen 2004a)	<i>Daphnia magna</i>	All invertebrates	"BLM"	ERA for REACH, EQS (Europe)	Only H and Na considered to compete with Cu; CuOH ⁺ and CuCO ₃ also bind to BL and significantly contribute to toxicity
Cu	Chronic gBAM	(Van Regenmortel et al. 2015)	<i>Daphnia magna</i>	All invertebrates	"BLM"		Toxicity of FIA predicted as a function of pH, Na, Ca, Mg
Cu	Chronic BLM	(Crémazy et al. 2017)	Rainbow trout	Fish	"BLM"		Toxicity decreased (higher effect concentrations) with increasing pH, but pattern reversed at high pH (>8); even in long-term exposures, Cu was an acute toxicant
Ni	Hybrid BLM	(Deleebeeck et al. 2007)	Rainbow trout, fathead minnow	Fish	"BLM"		Nonlinear pH response could not be modeled as a single-site BLM;

							model predicted toxicity well, but the exact mechanisms by which Ca, Mg, and pH modify Ni toxicity were undetermined
Ni	Hybrid BLM	(Deleebeeck et al. 2008)	<i>Daphnia magna</i>	All invertebrates	"BLM"		Stronger effect of pH on chronic toxicity than acute; similar issues and resolution with nonlinear pH response as their fish model
Ni	Hybrid BLM	(Deleebeeck et al. 2009)	Green algae	All primary producers	"BLM"		Predictions from classic BLM limited to a narrow pH range; incorporating a nonlinear pH function expanded prediction ranges
Ni	Chronic BLMs	(Schlekat et al. 2010)	Fish, daphnids, duckweed, snails, rotifers, insects, <i>Chironomus dilutus</i>	All freshwater aquatic life	"BLM"	ERA for REACH, EQS (Europe)	Tested existing BLMs; BLMs developed with cladocerans outperformed BLMs developed for more taxonomically similar taxa
Pb	Chronic BLM	(Nys et al. 2014)	<i>Ceriodaphnia dubia</i>	Freshwater invertebrates	"BLM"	ERA for REACH, EQS (Europe)	pH had a strong influence on toxicity but not Ca; H ⁺ (as

							log K_{BL-H}) was the only competitive constant needed
Pb	gBAM	(Van Sprang et al. 2016)	Algae and fish	All aquatic organisms	"BLM"	ERA for REACH, EQS (Europe)	Toxicity of FIA for algae is a function of pH and DOC only; for <i>Ceriodaphnia</i> , pH; and for fish, pH and Ca
Pb	Acute and chronic BLMs	(DeForest et al. 2017)	Daphnids, mayflies, fathead minnow, rotifer, snails	All aquatic animals	"BLM"		Various animal toxicity data sets were fit by adjusting critical accumulation values with a single set of unidentate log K values; includes $PbOH^+$ toxicity
Zn	Chronic BLM	(Heijerick et al. 2005)	<i>Daphnia magna</i>	<i>Daphnia magna</i>	"BLM"	RA for REACH, EQS (Europe)	Toxicity of FIA increased (lower effect concentrations) with increasing pH
Zn	gBAM	(De Schamphelaere et al. 2005a)	Green algae	All primary producers	pH ("BLM" set needed for FIA calculation)	RA for REACH, EQS (Europe)	Toxicity of FIA and dissolved Zn increased (higher effect concentrations) with increasing pH; pH was the only factor affecting Zn suppression of algal growth when expressed as FIA;

							full "BLM" chemistry needed for FIA
Zn	Chronic BLM	(De Schampelaere et al. 2005a)	<i>Daphnia magna</i> and rainbow trout	All invertebrates and fish	"BLM"	ERA for REACH, EQS (Europe)	Similar model structure and log K values between <i>Daphnia</i> and trout
Zn	Acute BLM	(Clifford and McGeer 2009)	<i>Daphnia pulex</i>		BLM		Binding coefficients calculated from toxicity values from a wide variety of water types
Zn	Acute and chronic BLMs	(DeForest and Van Genderen 2012)	Various aquatic animals and rotifers, <i>Brachionus calyciflorus</i>	All aquatic animals	"BLM"	ERA for REACH, EQS (Europe)	Multiple BLM log K values from previous models were averaged to produce a "unified BLM which performed well predicting toxicity to a phylogenetically diverse group of species, including daphnids, rotifers, snails, and fish"
Al, Cu, Pb, Zn	"F-Tox" humic acid model	(Stockdale et al. 2010)	Field surveys of stream aquatic insect communities	Macroinvertebrate communities	"BLM"		Humic acid used as proxy for bioaccumulation on insect biotic ligands; fitted potency factors were used to relate modeled accumulation of

							metals to species richness
24 metals	“F-Tox” humic acid model	(Tipping and Lofts 2013, 2014; Tipping et al. 2019)	Fish, cladocerans, lettuce	Various aquatic species	“BLM”		F-Tox humic acid model expanded to predict effects in classic single-species toxicity tests; fitted potency factors were used to relate modeled accumulation of metals to various endpoints
Al, Cd, Cu, Ni, Pb, Zn	“Tox” (generalization of the F-Tox humic acid model)	(Balistreri and Mebane 2014; Balistreri et al. 2015; Mebane et al. 2017)	Trout in single-species toxicity tests; stream and lake aquatic invertebrate communities	Natural aquatic communities	“BLM”		Similar to F-Tox but generalized to use various bioaccumulation models, including but not limited to humic acid
Cd, Cu, Zn	2-pKa bidentate model with Tox addition	(Farley and Meyer 2015; Mebane et al. 2017)	Trout, cladocerans, stream insect communities	Freshwater animals	“BLM”		“Streamlined” mixture models used bidentate, single-site accumulation models with Tox addition to predict effects; bidentate structure increased flexibility for handling nonlinear pH responses

Ag, Cd, Cu, Zn	BLM and biodynamic hybrid model	(Veltman et al. 2010)	Various freshwater fish	Freshwater fish	"BLM" chemistry; dietary and gill uptake, assimilation, and elimination rates; organism size		The covalent index, reflecting metal affinity for proteins, was used to estimate metal adsorption efficiencies
Pb	Surface complexation	(Antunes and Kreager 2014)	Duckweed (<i>Lemna minor</i>)	Vascular plants	"BLM"		Used the oxide surface complexation model within WHAM VII
Cationic metals	Plasma membrane	(Kinraide 2006)	Wheat (<i>Triticum aestivum</i>)	Vascular and single-cell plants	I "BLM set"		Most successful when calculated with free ion activities. Concept is broadly applicable to aquatic organisms

1340 "BLM set" of model inputs are temperature, pH, dissolved organic carbon, Ca, Mg, Na, K, Cl⁻, SO₄, and alkalinity or dissolved inorganic carbon.

1341 ACR = acute to chronic effects ratio; BLM = biotic ligand model; CA = critical accumulation values on the biotic ligand associated with a level of effect;

1342 EQS = environmental quality standard; ERA = environmental risk assessment; FIA = free ion activity; log *K* = stability constant for cations to biotic ligands;

1343 MLR = multiple linear regression; RA = response addition; REACH = Registration, Evaluation, Authorisation and Restriction of Chemicals; WHAM

1344 VII = Windermere humic aqueous model VII.

1345 **Figures**

1346 Figure 1. An example of a biotic ligand model calibrated to measure rainbow trout gill Ag
1347 accumulation data, over a variety of water chemistry conditions. In most cases, the patterns of
1348 measured and calculated accumulations matched well (redrawn from Janes and Playle 1995).

1349 Figure 2. The biotic ligand model (BLM) used to predict gill accumulation in Figure 1 also predicts
1350 toxicity well. Median lethal concentrations of Ag experimentally obtained with *Ceriodaphnia dubia*
1351 from laboratory (open symbols) or natural (closed symbols) water exposures compare well with
1352 calculated BLM predictions. Solid diagonal is line of perfect agreement; dashed lines denote a factor
1353 of ± 2 deviations from the 1:1 line. (Bielmyer et al. 2007; Paquin and Di Toro 2008; Naddy et al.
1354 2018). LC50 = median lethal concentration.

1355 Figure 3. Effect of pH on dissolved zinc (upper left) and copper toxicity (upper right) to *Daphnia*
1356 *magna* and *Pseudokirchneriella*. The figure for zinc shows the originally reported 72-h median effect
1357 concentration (EC50) for algae biomass (Heijerick et al. 2002) and the 21-d reproductive EC50
1358 (Heijerick et al. 2005). The figure for copper shows simulated toxicity data for the same endpoints,
1359 using multiple linear regression models fitted to data from a multivariate test design, as reported for
1360 algae (De Schamphelaere et al. 2003) and *D. magna* (De Schamphelaere and Janssen 2004b). The 2
1361 upper panels show a clearly distinct effect trend of pH on dissolved metal toxicity and form clear
1362 examples that strongly suggest that merging algae and animal bioavailability models into a single
1363 model is not appropriate. The lower left panel shows that, when expressed on a free ion activity
1364 basis (data also from Heijerick et al. 2002, 2005), the direction of the effect of pH on zinc toxicity is
1365 the same for algae and *Daphnia*, but the magnitude of the effect is clearly stronger for algae than for
1366 daphnids. The strong effect on free zinc ion toxicity for algae dominates over the speciation effect of
1367 pH, overall resulting in increased toxicity at higher pH for algae. In contrast, at higher pH, the
1368 speciation effect dominates for *Daphnia*, explaining the decreasing toxicity toward higher pH levels.
1369 A similar reasoning applies to copper (not shown). Data in the lower left panel have been used to
1370 construct and apply separate bioavailability models for algae and invertebrates in normalizing
1371 toxicity data for criteria and predicted-no-effect concentration derivation in Europe. The lower right
1372 panel shows possible regulatory implications of the different bioavailability relationships with pH
1373 (data taken from Supplementary 5 in Van Sprang et al. [2009]). The sensitivity ratio of algae versus
1374 invertebrates (geometric mean of normalized no-observed-effect concentration of all
1375 invertebrates/algae) is simulated as a function of pH for 2250 water samples. This panel shows that,
1376 although on average algae show similar sensitivity as invertebrates at low pH (~6.5), algae become
1377 by far more sensitive with increasing pH, up to approximately 10-fold and more at pH 8 and above.
1378 Algae also become increasingly more sensitive than invertebrates at higher pH for copper and lead
1379 (not shown). NOEC = no-observed-effect concentration.

1380 Figure 4. Simulated relationships between toxicity of the free metal ion and pH, presented on a log-
1381 scale versus pH (upper panel) or on a linear scale versus H^+ ion activity, according to various
1382 assumed mechanisms, that is, the classic biotic ligand model (BLM) with a unidentate binding site
1383 (blue), a BLM with a bidentate binding site (orange), contribution of the hydroxide complex to
1384 toxicity (gray), and assuming that humic acid is a good multiple-site surrogate for the biotic ligand.
1385 This figure shows that, even if the emerging relationships are not perfectly linear (except for the
1386 classic BLM in the lower panel and the bidentate model in the upper panel), reasonably good linear
1387 fits can be obtained. EC_x = x% effect concentration; WHAM = Windermere humic aqueous model.

1388 Figure 5. An example of nonunique solutions possible when deriving biotic ligand models by fitting
1389 toxicity test data without regard to actual critical accumulations (LA50). The parameters for model A

1390 are grounded in experimental findings. Model B fits the toxicity data just as well, even though the
1391 large reciprocal changes to the log K and critical accumulation fractions make the latter
1392 physiologically implausible (see online Supplemental Data SI-2 in Farley et al. 2015). The solid
1393 diagonal line is the line of 1:1 agreement; the dotted and dashed diagonal lines are a factor of ± 2
1394 deviations from the 1:1 line. BL = biotic ligand; LA50 = short-term accumulation at the biotic ligand
1395 that is predictive of 50% mortality at a later time; LC50 = median lethal concentration.

1396 Figure 6. (A) Modeled 30-d 20% lethal concentrations (LC20s) as a function of dissolved organic
1397 carbon (DOC; at pH 7) for rainbow trout (more sensitive) and a 10 times less sensitive hypothetical
1398 fish species, presented on linear and log scale, and (B) 30-d LC20s as a function of DOC for rainbow
1399 trout at pH 6 and 8, presented on linear and log scale. EC_x = x% effect concentration.

1400 Figure 7. Biotic ligand models (BLMs) assume equilibrium conditions, which may not be true with
1401 metal–dissolved organic carbon (DOC) binding that can take up to 24 h to reach equilibrium in some
1402 tests. The data shown here for Cu and DOC sampled in a stream during a rainstorm show that
1403 concentrations can change rapidly and not necessarily in synchrony. Copper is expected to have
1404 greater bioavailability and toxicity in nonequilibrium conditions than would be predicted by
1405 equilibrium-based BLMs. Data from Balistrieri et al. (2012).

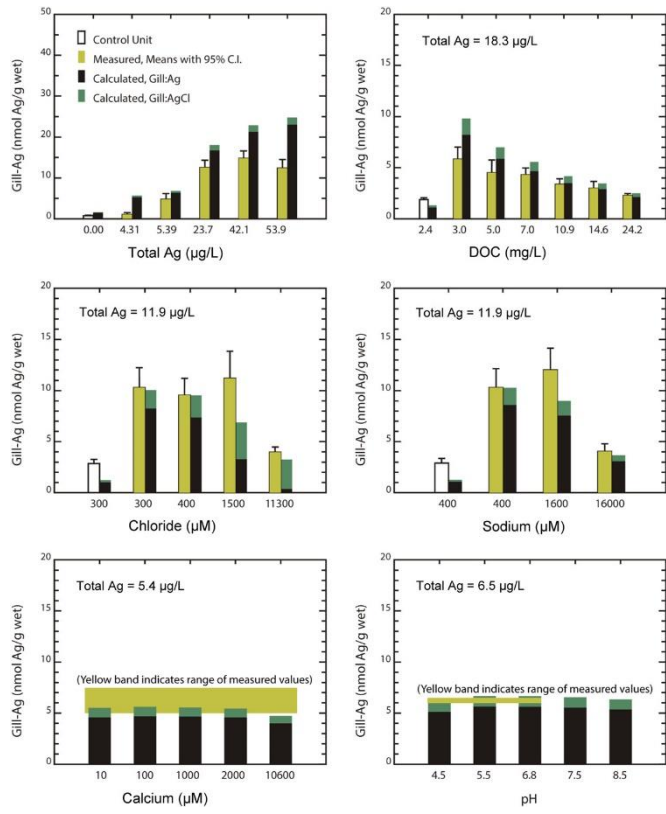
1406 Figure 8. Comparisons of biotic ligand model (BLM)–predicted and measured fathead minnow
1407 median lethal concentrations (LC50s) in flow-through and static test designs. (A) Repeated tests of
1408 same-age fish from the same broodstock in constant exposure water (unamended Lake Superior
1409 reference water) varied by a factor of approximately ± 2 , which is the origin of the “factor-of-2” rule
1410 of thumb for evaluating BLM performance. (B) Measured and BLM-predicted LC50s from tests
1411 amending Lake Superior water with added major ions or dissolved organic carbon. Both comparisons
1412 show that Cu tended to be more toxic in flow-through than static test designs, which is likely at least
1413 partially related to nonequilibrium conditions in the flow-through tests. Original data from Erickson
1414 et al. (1996) plotted after Santore et al. (2001) but using the US Environmental Protection Agency's
1415 (2007) updated fathead minnow species mean critical accumulation value of 2.97 nmol/g wet
1416 weight. The solid diagonal line is the line of 1:1 agreement; the dotted and dashed diagonal lines are
1417 a factor of ± 2 deviations from the 1:1 line.

1418 Figure 9. Kinetic patterns of free Cu concentrations over the course of a toxicity test in the presence
1419 of 5 mg/L dissolved organic carbon in 4 different hypothetical experimental conditions. In each
1420 simulation, the dissolved Cu concentration is 10 $\mu\text{g/L}$. In each panel, the free Cu concentration at
1421 equilibrium is shown as a horizontal dashed blue line, and the simulated free Cu in the experiment as
1422 a function of time is shown as a red line. For a static exposure with no pre-equilibration (A), the free
1423 Cu (Cu^{2+}) at the start of the test is elevated but decreases over time until it is near equilibrium at
1424 approximately 30 h. For a static exposure with a 24-h pre-equilibration period (B), the free Cu is
1425 close to equilibrium for the entire test duration. For a static test with daily renewals (C), the free Cu
1426 is elevated at the beginning of each renewal and then decreases but never reaches equilibrium. In a
1427 flow-through test (D) with a 1-h residence time, the free Cu is constant but far from equilibrium.

1428 Figure 10. Laboratory toxicity testing of metals with increased dissolved organic matter (DOM)
1429 should consider metal–DOM equilibrium time. Silver was more toxic to *Daphnia magna* (lower 24-h
1430 LC50s) in tests initiated after 3-h metal–DOM contact time than in tests initiated after 24-h contact
1431 time (Glover et al. 2005). DOC = dissolved organic carbon; LC50 = median lethal concentration.

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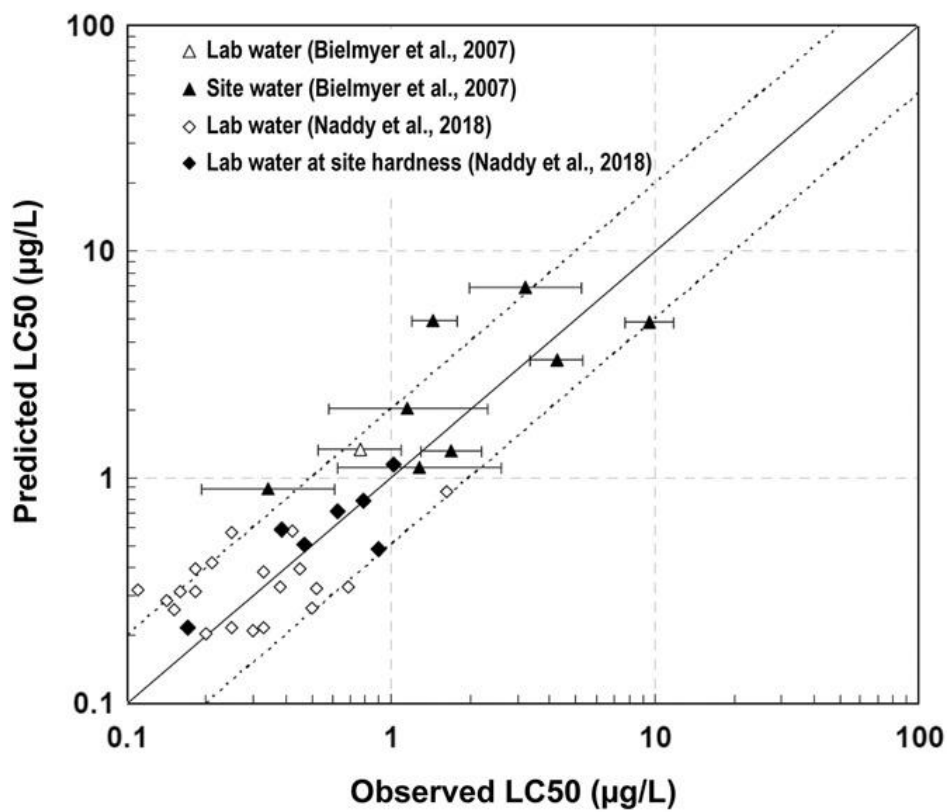
1433 Figure 1.



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1436 Figure 2.

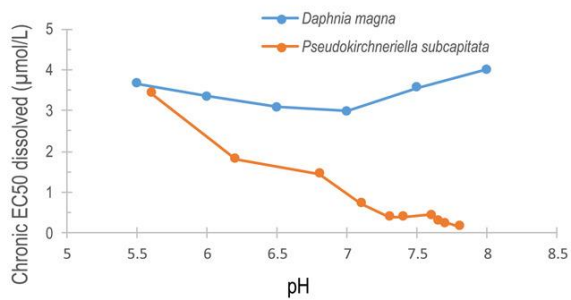


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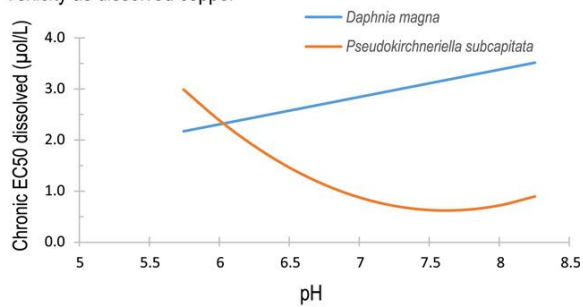
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1439 Figure 3.

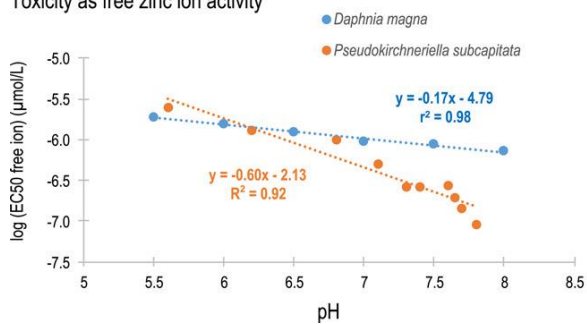
Toxicity as dissolved zinc



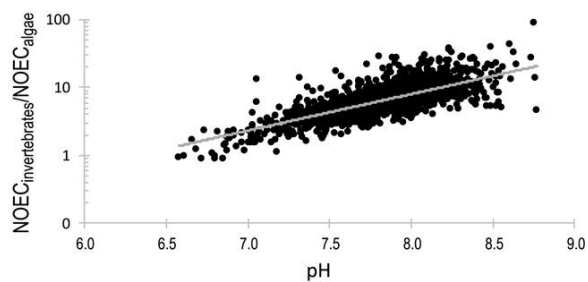
Toxicity as dissolved copper



Toxicity as free zinc ion activity



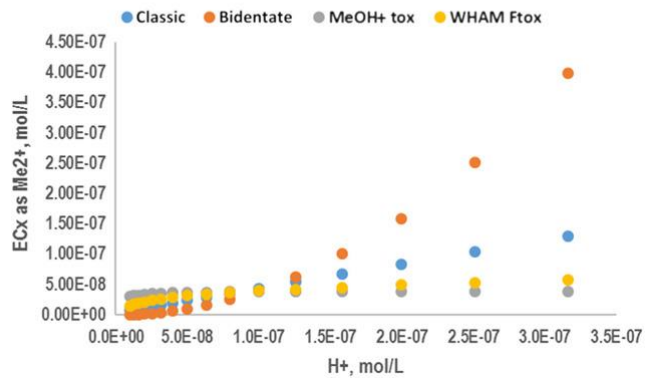
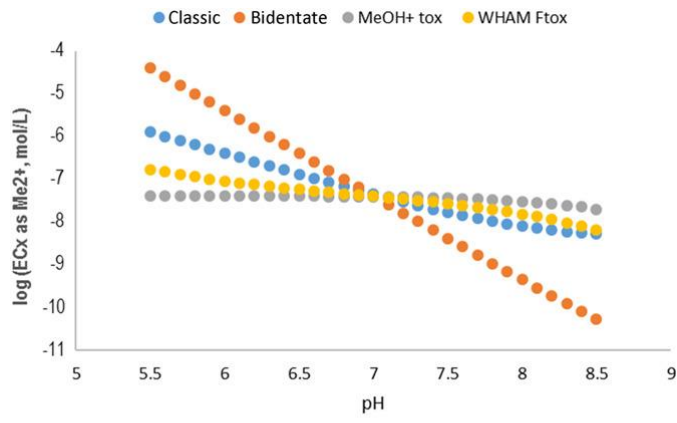
Simulated zinc sensitivity ratio in field waters



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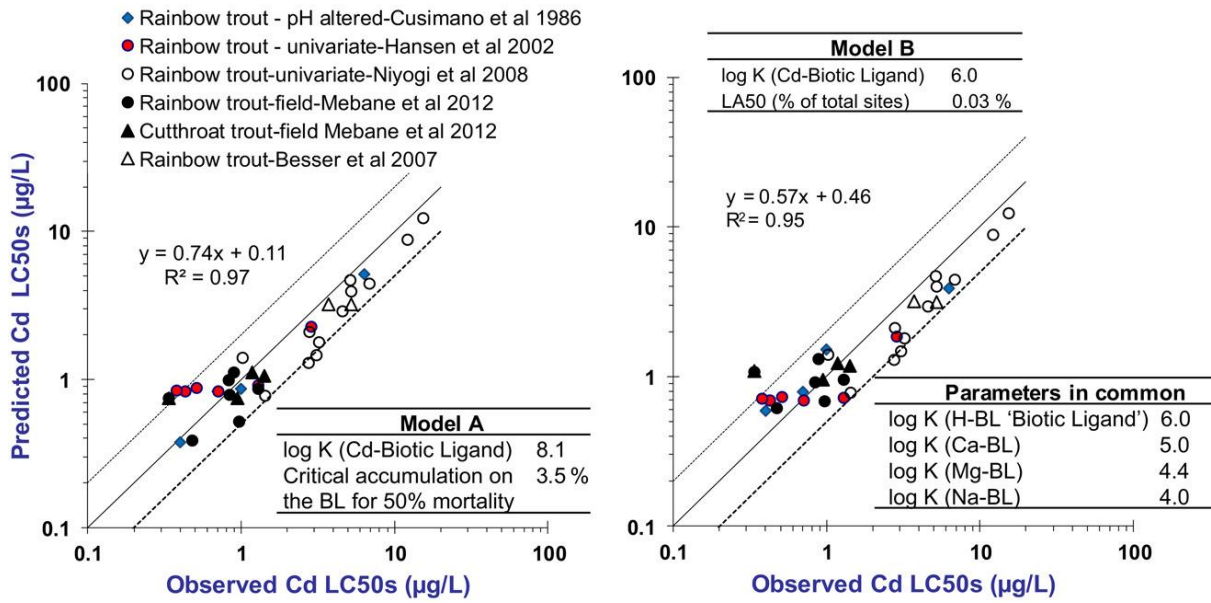
1442 Figure 4.



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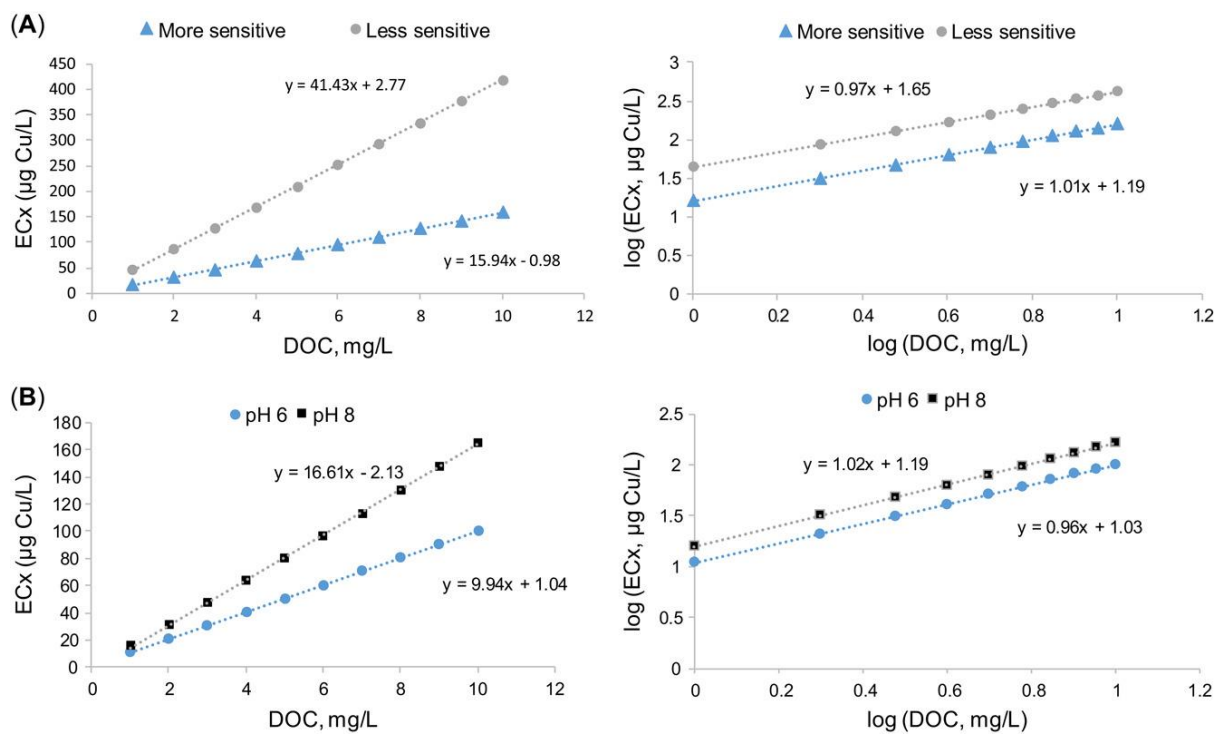
1445 Figure 5.



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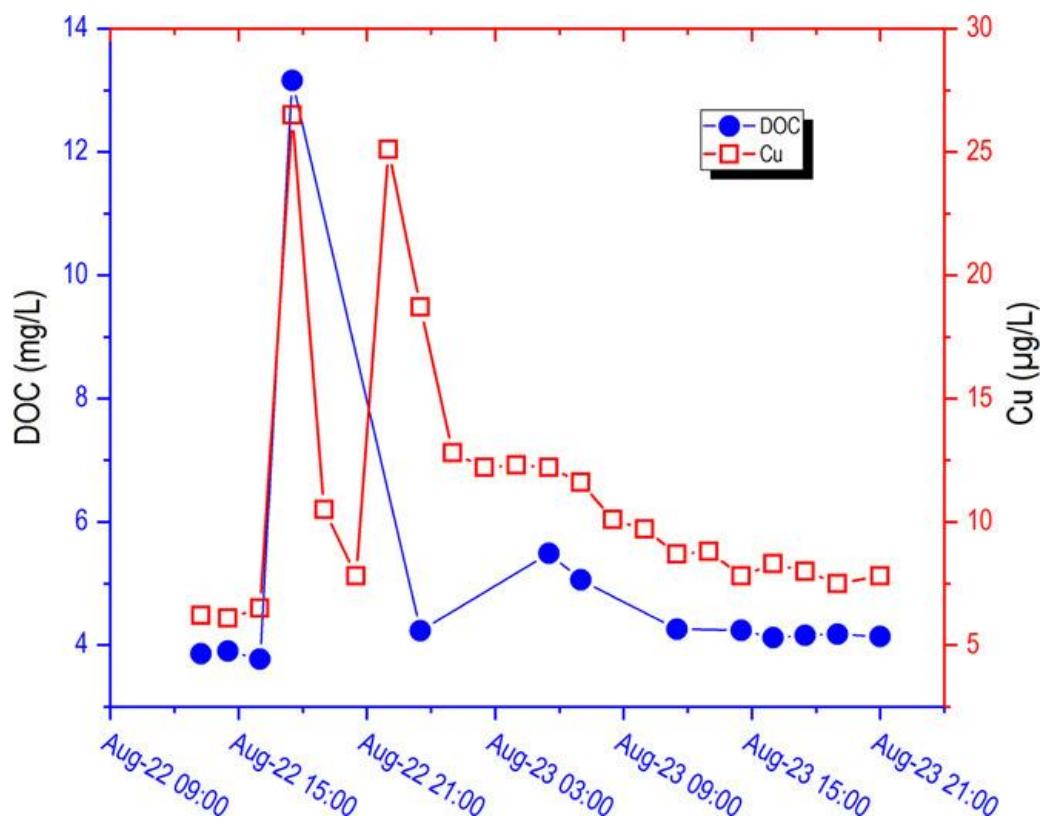
1448 Figure 6.



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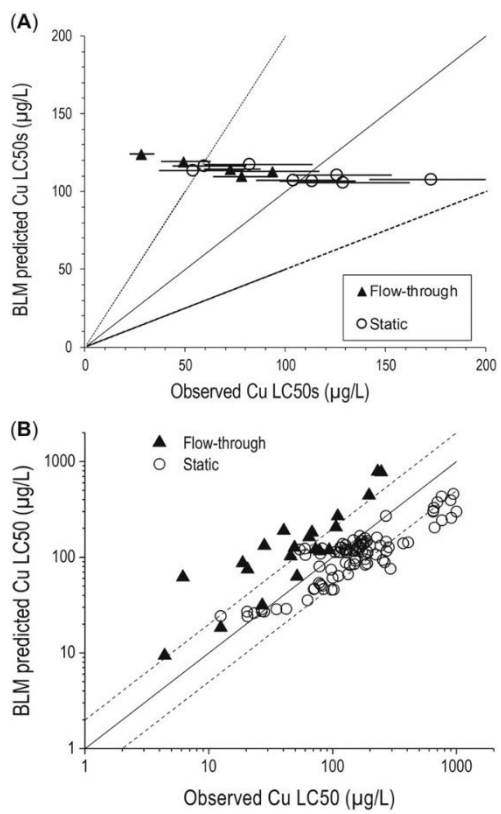
1451 Figure 7.



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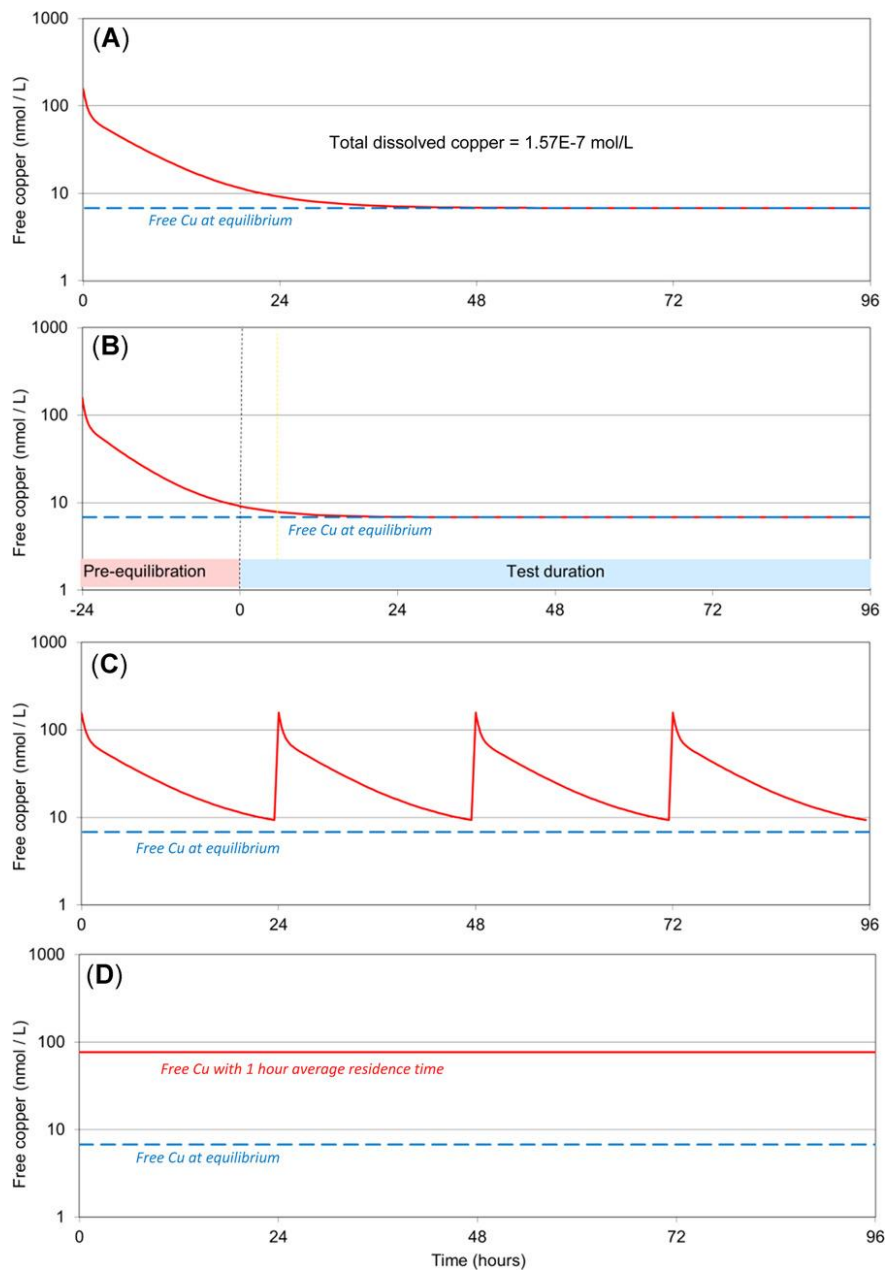
1454 Figure 8.



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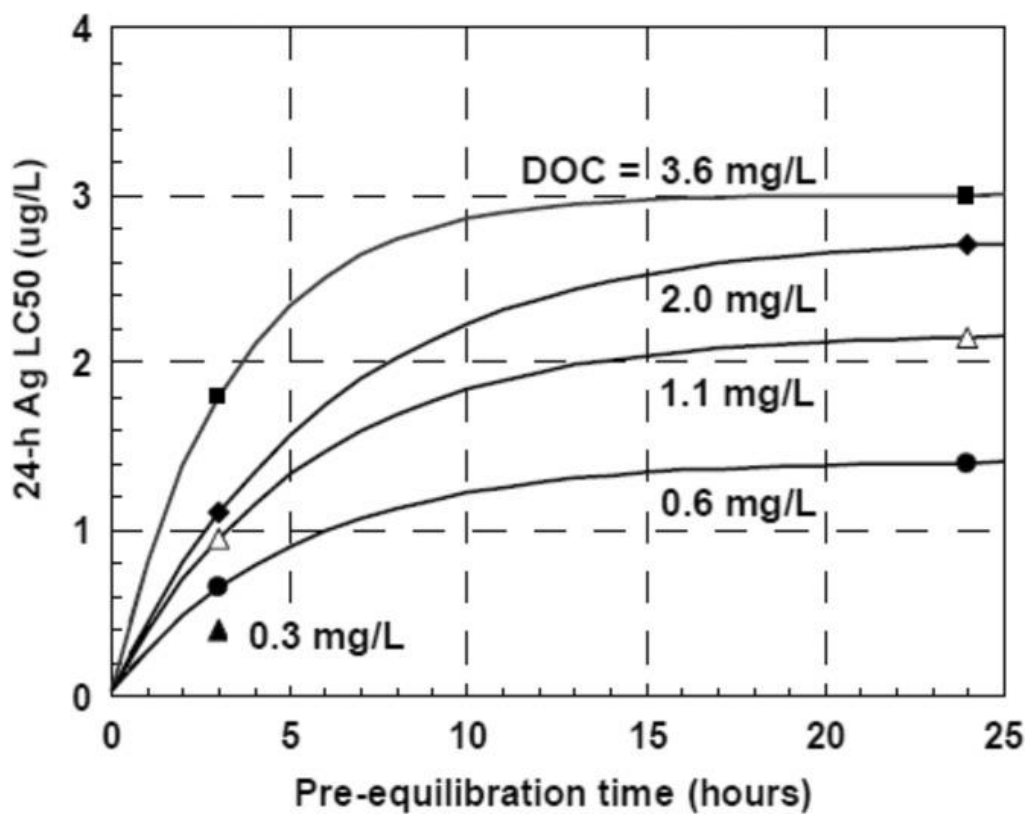
1457 Figure 9.



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1460 Figure 10.



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