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## **Acclimation and adaptation components of the temperature dependence of plant photosynthesis at the global scale**

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## Summary

- The temperature response of photosynthesis is one of the key factors determining predicted responses to warming in global vegetation models (GVMs). The response may vary geographically, due to genetic adaptation to climate, and temporally, due to acclimation to changes in ambient temperature. Our goal was to develop a robust quantitative global model representing acclimation and adaptation of photosynthetic temperature responses.
- We quantified and modelled key mechanisms responsible for photosynthetic temperature acclimation and adaptation using a global dataset of photosynthetic CO<sub>2</sub> response curves including data from 141 C<sub>3</sub> species from tropical rainforest to Arctic tundra. We separated temperature acclimation and adaptation processes by considering seasonal and common-garden datasets, respectively.
- The observed global variation in the temperature optimum of photosynthesis was primarily explained by biochemical limitations to photosynthesis, rather than stomatal conductance or respiration. We found acclimation to growth temperature

to be a stronger driver of this variation, than adaptation to temperature at climate of origin.

- We developed a summary model to represent photosynthetic temperature responses and showed that it predicted the observed global variation in optimal temperatures with high accuracy. This novel algorithm should enable improved prediction of the function of global ecosystems in a warming climate.

**Key words:** Global vegetation models (GVM), climate of origin, growth temperature,  $V_{\text{cmax}}$ ,  $J_{\text{max}}$ , maximum carboxylation capacity, maximum electron transport rate,  $AC_i$  curves

## Introduction

The capacity of species to cope with increasing growth temperature is one of the key determinants in range shifts and local extinction of species because their distribution and range limits closely follow temperature isolines (Battisti *et al.*, 2005). Evidence suggests that many species are adapted to their thermal environment of origin (Berry & Björkman, 1980) but also exhibit the capacity to adjust to temporal variations in the temperature of their environment (Rehfeldt *et al.*, 2001; Valladares *et al.*, 2014). However, the mechanisms that determine these responses are not well understood, making it challenging to predict the fate of plants in a changing climate.

Global vegetation models (GVMs) are one of the principal tools used to predict future terrestrial vegetation carbon balance (Rogers *et al.*, 2017a; Mercado *et al.*, 2018). The temperature response of leaf-scale net photosynthesis (referred to as  $A_n$ -T response hereafter) is one of the key processes in these models. The effect of warming on modelled photosynthesis depends on the  $A_n$ -T response function used in the model, and in particular, the optimum temperature of photosynthesis ( $T_{\text{opt}A}$ ) (Booth *et al.*, 2012). Decades of empirical studies have shown that the  $A_n$ -T responses of plants vary geographically, suggesting genetic adaptation of species to their climate of origin (Fryer & Ledig, 1972; Slatyer, 1977; Slatyer, 1978; Berry & Björkman, 1980; Gunderson *et al.*, 2009). Considerable evidence also shows that plants have the capacity to adjust the  $A_n$ -T response following temporal changes in ambient temperature, a response known as thermal acclimation (Way & Sage, 2008; Hall *et al.*, 2013; Way & Yamori, 2014; Yamaguchi *et al.*, 2016; Way *et al.*, 2017). In a recent review, Yamori *et al.* (2014) reported inherent differences in the  $A_n$ -T response and its acclimation capacity among photosynthetic pathways ( $C_3$ ,  $C_4$  and CAM) and

functional types (annual vs perennial, deciduous vs evergreen) that often differ in their climatic distributions. However, the current representations of  $A_n$ -T response in GVMs do not capture this empirical knowledge well (Smith & Dukes, 2013; Lombardozzi *et al.*, 2015; Smith *et al.*, 2016; Mercado *et al.*, 2018). Most GVMs use either a single  $A_n$ -T response function for all species or represent broad geographical variation in the  $A_n$ -T response by using plant functional type(s) (PFTs)-specific functions without considering thermal acclimation. Robust representation of adaptation and acclimation of  $A_n$ -T response in GVMs is challenging as we lack a quantitative assessment of acclimation and adaptation of photosynthetic temperature responses on a global scale (Stinziano *et al.*, 2017).

Many GVMs incorporate the biochemical model of  $C_3$  photosynthesis (Farquhar *et al.*, 1980; Rogers *et al.*, 2017a ; referred to as FvCB hereafter). Therefore it is both tractable and valuable to encapsulate the mechanisms of photosynthetic temperature adaptation and acclimation in terms of parameters of the Farquhar model (Hikosaka *et al.*, 1999; Dreyer *et al.*, 2001; Medlyn *et al.*, 2002b; Dillaway & Kruger, 2010). The model has two key parameters, for which the temperature response is particularly important; the maximum rate of ribulose-1,5-bisphosphate carboxylase-oxygenase (Rubisco) activity ( $V_{cmax}$ ) and the maximum potential electron transport rate ( $J_{max}$ ) (Farquhar *et al.*, 1980). GVMs use two basic functional forms to characterize the instantaneous temperature response of the key FvCB model parameters, namely the standard and peaked Arrhenius functions (Medlyn *et al.*, 2002a). Most empirical studies of the instantaneous temperature response of  $V_{cmax}$  and  $J_{max}$  have used the peaked Arrhenius model, which has four key parameters; the basal rate of either  $V_{cmax}$  or  $J_{max}$  at a standard temperature of 25°C ( $V_{cmax25}$  or  $J_{max25}$ ), the activation energy ( $E_a$ ), the de-activation energy ( $H_d$ ), and the entropy term ( $\Delta S$ ). The peaked Arrhenius model can also be used to calculate the optimum temperatures of  $V_{cmax}$  ( $Topt_V$ ) and  $J_{max}$  ( $Topt_J$ ). These parameters have now been documented for a wide range of species from different biomes and PFTs (Onoda *et al.*, 2005; Rogers *et al.*, 2017b; Slot & Winter, 2017). Evidence suggests that the Arrhenius model parameters vary significantly across plant taxa but also that these parameters have the capacity to acclimate to the growth temperature (Crous *et al.*, 2013; Crous *et al.*, 2018).

Several meta-analytic studies have attempted to characterise species variation in the model parameters. Medlyn *et al.* (2002a) compared the temperature response of key FvCB model parameters across different species but reported a poor relationship overall between the optimum temperature for photosynthesis and the temperature of the growing environment.

They reported lower  $Topt_V$  and  $Topt_J$  for plants grown in boreal compared to temperate climates, but it was unclear whether this difference was due to inherent genetic differences among the boreal and temperate species, or acclimation to prevailing growth temperature. In an analysis of 23 species, (Hikosaka *et al.*, 2006) identified two important mechanisms of photosynthetic temperature acclimation, namely  $E_a$  of  $V_{cmax}$  ( $Ea_V$ ) and  $J_{max}$  ( $Ea_J$ ) and the ratio of  $J_{max} : V_{cmax}$  ( $JV_r$ ). The most comprehensive synthesis to date of the biochemically-based plant photosynthetic temperature response is that of Kattge and Knorr (2007), who compared the instantaneous temperature response of  $V_{cmax}$  and  $J_{max}$  across 36 species. This study found a lack of thermal acclimation of  $Ea_V$  and  $Ea_J$  but reported significant acclimation relationships for  $JV_r$  and  $\Delta S$  of  $V_{cmax}$  ( $\Delta S_V$ ) and  $J_{max}$  ( $\Delta S_J$ ). Importantly, Kattge and Knorr (2007) synthesised these relationships into a simple and generalizable form that enabled direct implementation into GVMs, thus providing a means to quantify the effect of thermal acclimation of photosynthesis on terrestrial carbon cycle predictions (Chen & Zhuang, 2013; Lombardozzi *et al.*, 2015; Smith *et al.*, 2016) as well as on biophysical consequences in future climates (Smith *et al.*, 2017).

Despite the success of the Kattge and Knorr (2007) algorithms, the functions have several limitations. Firstly, the parameterization process did not consider potential inter-specific differences in photosynthetic temperature response; all changes were attributed to differences in growth temperature. Hence, the response incorporates elements of both temperature adaptation and acclimation without resolving the extent of the contribution of the two processes. Given that acclimation can occur over days and adaptation takes many generations, the importance of resolving the relative contribution of the two processes is critical. Recently, Mercado *et al.*, (2018) showed that assuming the relationships represent both adaptation and acclimation, or adaptation only, leads to significantly different conclusions about the trajectory of future terrestrial carbon storage under warming. Their results further highlight the importance of separating photosynthetic thermal adaptation and acclimation when simulating current and future carbon storage. However, to date, few studies have separated species differences in temperature adaptation from temperature acclimation processes (Lin *et al.*, 2013).

Secondly, the data used to derive the Kattge and Knorr (2007) functions came mainly from northern temperate and boreal trees and lacked globally important PFTs such as tropical forests and Arctic tundra. As a result, the growth temperature range only varied from 11 to 29°C (Kattge and Knorr 2007), which is substantially narrower than growth temperatures

simulated in GVMs. Therefore, the analysis of Kattge and Knorr (2007) could be improved with a broader global dataset directly addressing the relative roles of temperature acclimation and adaptation.

Thirdly, the ability of the acclimation functions to capture the observed differences in temperature optima of light saturated net photosynthesis ( $T_{opt_A}$ ) has not been directly tested. It is not clear whether making adjustments to  $T_{opt_V}$  and  $T_{opt_J}$  improves the ability of models to capture changes in  $T_{opt_A}$ ; some studies have reported similar  $T_{opt_A}$  values even with significantly different  $T_{opt_J}$  among species (Vårhammar *et al.*, 2015). Moreover, the photosynthetic temperature response is controlled not only by the photosynthetic biochemistry, but also by stomatal and respiratory processes. Sensitivity analysis suggests that all three component processes are equally important in determining the  $T_{opt_A}$  at leaf scale (Lin *et al.*, 2012) as well as at canopy scale (Tan *et al.*, 2017) but none of the previous review studies addressed how the latter two components affected  $T_{opt_A}$ .

Given the need for robust representation of photosynthetic temperature acclimation and adaptation in GVMs, and its importance in predicting future global carbon budget (Lombardozzi *et al.*, 2015, Smith *et al.*, 2016, Mercado *et al.*, 2018) and climate (Smith *et al.*, 2017), we quantified and modelled the mechanisms that underlie the observed differences in  $T_{opt_A}$  among species and growth temperatures. We hypothesized that  $T_{opt_A}$  would be strongly driven by adaptation to the climate of origin, while temperature acclimation would further modify the temperature optimum in response to seasonal changes in temperature of the growth environment. To test these hypotheses, we compiled a global database of photosynthetic CO<sub>2</sub> response curves measured at multiple leaf temperatures to simultaneously resolve the temperature optima of  $A_{net}$ ,  $V_{cmax}$  and  $J_{max}$ . The data comprised a total of 141 species from tropical rainforests to Arctic tundra. Included in this database were datasets: (i) from common-garden studies, which were used to quantify effects of adaptation alone on  $T_{opt_A}$ ; and (ii) comprising time course studies that measured plants under contrasting prevailing ambient temperatures, which are used to quantify effects of temperature acclimation alone. We combined the identified effects of climate adaptation and temperature acclimation to derive a general global model of temperature responses that is then tested against (iii) a third, independent, biogeographic dataset measured on mature plants growing in their native environments across the globe.

## Materials and methods

### *Data sources*

We compiled a global database of datasets consisting of leaf photosynthetic CO<sub>2</sub> response measurements (referred to as ACi curves hereafter) measured at multiple leaf temperatures and saturating irradiance levels. The database covers 141 species from 38 experiments conducted around the world (Fig. S1, Table S1). Site latitude ranged from 42°48' S to 71°16' N and mean annual growing season temperature (long-term average temperature of months where mean monthly temperature is above 0°C) ranged from 3 to 30°C.

The method of data collection was consistent across all datasets. In most datasets, measurements were started at ambient CO<sub>2</sub> levels (360-400 ppm; depending on the year of data collection) and changed stepwise through a series of subambient (40-400 ppm) to superambient saturating CO<sub>2</sub> concentrations (400-2000 ppm). The same measurement protocol was repeated on the same leaf at different leaf temperatures. Measurements were made at saturating irradiance (Table S1) using a portable photosynthesis system with standard leaf chambers, in most cases the Licor 6400 (Licor Biosciences, Lincoln, NE, USA) although some measurements were made with the Walz-CMS system (Walz, Effeltrich, Germany). We visually inspected every ACi curve in the dataset for possible outliers and erroneous data points (i.e. negative intercellular CO<sub>2</sub> concentrations). We used criteria based on De Kauwe *et al.* (2016) to screen individual ACi curves for the analysis performed in this paper. Curves were excluded from the analysis if the fitted function (see below) had a  $r^2 < 0.99$  (however, if the number of replicates available for a given occasion was limited, the threshold  $r^2$  was reduced to 0.90; ~9% of the total ACi curves included in the analysis). After screening, the dataset contained a total of 3498 ACi curves measured at leaf temperatures ranging from 1 to 50°C.

### *Estimating temperature optimum for leaf net photosynthesis ( $T_{optA}$ )*

Ambient leaf net photosynthesis ( $A_{net}$ ) at each temperature was obtained from either the initial direct measurements at ambient CO<sub>2</sub> concentrations or extracted from the ACi curves. For curves where the first point was not measured at ambient CO<sub>2</sub> level, we extracted the  $A_{net}$  value at the measured sample CO<sub>2</sub> concentration falling between 300 and 400 ppm. We estimated the temperature optimum for  $A_{net}$ ,  $T_{optA}$ , by fitting a widely used model of instantaneous photosynthetic temperature response (Gunderson *et al.*, 2009; Crous *et al.*,

2013; Sendall *et al.*, 2015; Vårhammar *et al.*, 2015) (Eqn 1) to the net photosynthesis measurements. The model is a quadratic equation, expressed as:

$$A_{net} = A_{opt} - b(T - T_{opt_A})^2 \quad \text{Eqn 1}$$

where  $A_{net}$  is the net photosynthetic rate ( $\mu\text{mol m}^{-2} \text{s}^{-1}$ ) at a given leaf temperature,  $T$  ( $^{\circ}\text{C}$ ),  $T_{opt_A}$  is the temperature optimum for photosynthesis ( $^{\circ}\text{C}$ )  $A_{opt}$  is the net photosynthetic rate at  $T_{opt_A}$ , and the parameter  $b$  (unitless) describes the degree of curvature of the relationship.

### *Parameterising biochemical component processes of photosynthesis*

We used the FvCB model to characterize photosynthetic biochemical component processes. The model represents leaf net photosynthesis rate as the minimum of three rates; the Rubisco carboxylation limited photosynthetic rate ( $W_c$ ), the RuBP-regeneration limited photosynthetic rate ( $W_j$ ), and the triose phosphate utilization limited rate ( $W_p$ ). The widely used formulation and parameterization of the FvCB model is of the form (Eqn 2-6).

$$A_{net} = \min(W_c, W_j, W_p) \left(1 - \frac{\Gamma^*}{C_i}\right) - R_L \quad \text{Eqn 2}$$

$$W_c = V_{cmax} \frac{C_i}{C_i + K_c \left(1 + \frac{O_i}{K_o}\right)} \quad \text{Eqn 3}$$

$$W_j = \frac{J}{4} \frac{C_i}{(C_i + 2\Gamma^*)} \quad \text{Eqn 4}$$

$$W_p = 3 TPU \quad \text{Eqn 5}$$

where  $V_{cmax}$  is the maximum rate of ribulose-1,5-bisphosphate carboxylase-oxygenase (Rubisco) activity,  $C_i$  and  $O_i$  ( $\mu\text{mol mol}^{-1}$ ) are intercellular  $\text{CO}_2$  and  $\text{O}_2$  concentrations respectively,  $K_c$  and  $K_o$  ( $\mu\text{mol mol}^{-1}$ ) are Michaelis–Menten coefficients of Rubisco activity for  $\text{CO}_2$  and  $\text{O}_2$  respectively,  $\Gamma^*$  ( $\mu\text{mol mol}^{-1}$ ) is the  $\text{CO}_2$  compensation point in the absence of photorespiration,  $TPU$  ( $\mu\text{mol m}^{-2} \text{s}^{-1}$ ) is the rate of triose phosphate export from the chloroplast,  $R_L$  ( $\mu\text{mol m}^{-2} \text{s}^{-1}$ ) is the non-photorespiratory  $\text{CO}_2$  evolution in the light, and  $J$  ( $\mu\text{mol m}^{-2} \text{s}^{-1}$ ) is the rate of electron transport at a given light level.  $J$  is related to incident photosynthetically active photon flux density ( $Q$ ,  $\mu\text{mol m}^{-2} \text{s}^{-1}$ ) by

$$\theta J^2 - (\alpha Q + J_{max})J + \alpha Q J_{max} = 0 \quad \text{Eqn 6}$$

where  $J_{\max}$  ( $\mu\text{mol m}^{-2} \text{s}^{-1}$ ) is the potential rate of electron transport,  $\alpha$  ( $\mu\text{mol mol}^{-1}$ ) is the quantum yield of electron transport, and  $\theta$  (dimensionless) is the curvature of the light response curve (Farquhar *et al.*, 1980; Medlyn *et al.*, 2002a; Medlyn *et al.*, 2002b; Kattge & Knorr, 2007; Sharkey *et al.*, 2007).

We parameterized Eqns 3 – 6 using the *fitacis* function within the *plantecophys* package (Duursma, 2015) in R version 3.3.2 (R Development Core Team, 2012). We assumed the Bernacchi *et al.* (2001) kinetic constants for the temperature response of  $K_c$ ,  $K_o$  and  $\Gamma^*$  as given in Medlyn *et al.* (2002a). We used measurement  $Q$  in Eqn 6 whenever available (see Table S1); otherwise we assumed a fixed value of  $1800 \mu\text{mol m}^{-2} \text{s}^{-1}$ . We assumed constant values of  $\alpha$  ( $0.24 \mu\text{mol mol}^{-1}$ ) and  $\theta$  (0.85; unitless) for all datasets (Medlyn *et al.*, 2007); these parameter values have a relatively minor effect on the magnitude of estimated  $J_{\max}$  (Medlyn *et al.*, 2002a). The estimated parameters,  $V_{\text{cmax}}$  and  $J_{\max}$ , are apparent values as we assumed infinite mesophyll conductance ( $g_m$ ). The significance of  $g_m$  for  $V_{\text{cmax}}$  and  $J_{\max}$  estimates and their temperature response has been discussed elsewhere (Crous *et al.*, 2013; Bahar *et al.*, 2018). Here, there are insufficient data to quantify  $g_m$  and hence it would have been inappropriate to include in our analysis (see Rogers *et al.*, 2017a).

We tested two ACi curve fitting routines; one with and one without TPU limitation (Eqn 5). Accounting for TPU limitation in the FvCB model did not affect the estimated photosynthetic capacities, apparent  $V_{\text{cmax}}$  and  $J_{\max}$  (Fig. S2) suggesting that at ambient  $\text{CO}_2$  levels, net photosynthesis was rarely limited by TPU (results not shown). Hence, we focused on the temperature responses of apparent  $V_{\text{cmax}}$  and  $J_{\max}$  as the principal biochemical components affecting the  $T_{\text{optA}}$ .

The temperature responses of  $V_{\text{cmax}}$  and  $J_{\max}$  were fitted using the peaked Arrhenius function:

$$k_{T_k} = k_{25} \exp \left[ \frac{E_a(T_k - 298.15)}{(298.15 R T_k)} \right] \frac{1 + \exp \left( \frac{298.15 \Delta S - H_d}{298.15 R} \right)}{1 + \exp \left( \frac{T_k \Delta S + H_d}{T_k R} \right)} \quad \text{Eqn 7}$$

where  $k_{T_k}$  is the process rate (i.e.  $V_{\text{cmax}}$  or  $J_{\max}$ ;  $\mu\text{mol m}^{-2} \text{s}^{-1}$ ) at a given temperature,  $T_k$  (K),  $k_{25}$  is the process rate at 25 °C,  $R$  is the universal gas constant ( $8.314 \text{ J mol}^{-1} \text{ K}^{-1}$ ), and  $E_a$  ( $\text{kJ mol}^{-1}$ ) is the activation energy term that describes the exponential increase in enzyme activity with the increase in temperature,  $H_d$  ( $\text{kJ mol}^{-1}$ ) is the deactivation energy term that describes, e.g. decline in enzyme activity at higher temperature due to denaturation of enzymes, and  $\Delta S$  ( $\text{J mol}^{-1} \text{ K}^{-1}$ ) is the entropy term which characterize the changes in reaction rate caused by

substrate concentration (Johnson *et al.*, 1942). To avoid over-parameterization, we assumed a fixed value of 200000 J mol<sup>-1</sup> for  $H_d$  in Eqn 7 for all species (Dreyer *et al.*, 2001; Medlyn *et al.*, 2002a).

The optimum temperature for  $k_{Tk}$  is given by:

$$T_{opt} = \frac{H_d}{\Delta S - R \ln\left(\frac{E_a}{H_d - E_a}\right)} \quad \text{Eqn 8}$$

### *Assessing the contribution of stomatal and respiratory processes*

The optimum temperature for photosynthesis is determined by stomatal and respiratory processes as well as biochemical processes (Medlyn *et al.*, 2002a; Lin *et al.*, 2012). Stomatal conductance values are potentially affected by the measurement protocol used in ACi curve measurements which rarely replicates the ambient conditions. Therefore, to assess the relative contribution of stomatal processes to  $Topt_A$ , we calculated the net photosynthesis rate at a fixed  $C_i$  of 275  $\mu\text{mol mol}^{-1}$  from each ACi curve, interpolating the curve using the FvCB model with parameters fitted to that curve. A fixed  $C_i$  of 275  $\mu\text{mol mol}^{-1}$  was chosen as it roughly corresponds to 70% of ambient  $[\text{CO}_2]$ . When the photosynthetic rate is scaled to a common  $C_i$ , it eliminates the effect of variation in stomatal conductance on photosynthesis, isolating the temperature effects on photosynthetic biochemistry. Similar to net photosynthesis, the temperature optimum for photosynthesis at a fixed  $C_i$  ( $Topt_{A275}$ ) was estimated for each species by fitting Eqn 1. We compared  $Topt_{A275}$  with  $Topt_A$  to estimate the effect of variation in stomatal conductance on the temperature optimum for photosynthesis.

We fitted standard Arrhenius function (Eqn 9) to  $R_L$  values obtained from ACi curves to assess the effect of respiratory component processes on  $Topt_A$ . We estimated two parameters  $R_{L25}$  ( $R_L$  at 25°C) and activation energy of  $R_L$  ( $E_a$ ). Similar to  $J_{max}$  and  $V_{cmax}$ , linear regression was used to test for temperature adaptation and acclimation of  $R_L$ .

$$R_L = R_{L25} \exp\left(\frac{E_a(T_k - 298.15)}{298.15 R T_k}\right) \quad \text{Eqn 9}$$

where,  $R_{L25}$  is the rate of respiration in light at 25°C

## *Test for local adaptation and seasonal temperature acclimation of $T_{optA}$*

We divided the database into three subsets: (i) mature plants growing in their native environments; (ii) common-garden datasets; and (iii) datasets with seasonal photosynthetic measurements. We used a subset of the data collected in mature plants (i) to identify the patterns in photosynthetic temperature responses of plants in native environments and for model evaluation. Temperature responses in this subset include the effects of both adaptation to the native environment, and acclimation to the prevailing temperature. We used the common garden (ii) and seasonal measurements (iii) subsets to estimate the relative contributions of adaptation and acclimation, respectively, in determining the observed trends with temperature for plants in native environments

For plants growing in native environments, we derived relationships between photosynthetic parameters and the prevailing temperature of the growing environment defined as the mean air temperature for the 30 days prior to gas exchange measurements (Kattge & Knorr, 2007) ( $T_{growth}$ ), to identify the temporal trends in photosynthetic temperature responses. We derived  $T_{growth}$  using on-site measured real time daily air temperature for most of the datasets, but for three datasets (Hinoki cypress, Japan; Mongolian oak, Japan; and Scots pine, Finland; Table S1), we extracted  $T_{growth}$  values from the original publications as on-site temperature measurements were not available. We used a general linear model to parameterise the observed responses in mature plants dataset (Eqn 10)

$$f(T_{growth}) = a + bT_{growth} \quad \text{Eqn 10}$$

where  $a$  and  $b$  are the intercept and slope respectively.

Seasonal datasets provide the opportunity to test the acclimation capacity of different species to temporal changes in the ambient temperature of the growing environment. Here, we correlated photosynthetic parameters with growth temperature,  $T_{growth}$ , defined as the mean air temperature for the 30 days prior to gas exchange measurements. Similar to the mature plants dataset, we derived  $T_{growth}$  using on-site measured daily air temperature for most of the datasets. For datasets where real-time meteorological data were not available, we extracted  $T_{growth}$  values from the original publications.

Common gardens provide an opportunity to test for adaptation, as species with different climates of origin are grown at a common growth temperature. The common garden datasets included field trials and experiments in controlled environmental conditions which

included two or more species or provenances with contrasting climates of origin. We located the seed source of each species or provenance (latitude and longitude) using published information (Table S1). We used 30" resolution WorldClim climatology data (WorldClim 1.4;(Hijmans *et al.*, 2005)) to estimate long-term average (1960-1990) air temperature at seed source. With reference to the species selection criteria used in several common garden studies (Lin *et al.*, 2013; Vårhammar *et al.*, 2015), we defined mean maximum air temperature of the warmest month at species' seed source as the species' home temperature ( $T_{home}$ ) and derived relationships between photosynthetic parameters and  $T_{home}$  to test for adaptation of species'  $A_n$ -T response to climate of origin. We repeated the same analysis with two other forms of species' home temperature, 1. mean growing season air temperature and 2. mean temperature of the warmest quarter, to test whether our results were altered depending on the definition of climate of origin.

For both common garden and seasonal subsets, we used linear regression against  $T_{home}$  and  $T_{growth}$  (Eqns 11, 12) to test for temperature adaptation and acclimation, respectively, of  $Topt_A$ ,  $Topt_{A_{275}}$ , the photosynthetic biochemical parameters ( $V_{cmax}$  and  $J_{max}$ ), and their temperature response parameters (see Eqns 7 and 8). To test the effect of different biochemical parameters on temperature optimum for photosynthesis, we used linear regression between  $Topt_{A_{275}}$  and temperature response parameters of  $V_{cmax}$  and  $J_{max}$ .

#### *Representing acclimation and adaptation in vegetation models*

We derived functions to represent photosynthetic temperature acclimation and adaptation in GVMs. If a given parameter showed only acclimation to growth temperature, the function used was:

$$f(T_{growth}) = A_{ac} + \alpha_{ac}T_{growth} \quad \text{Eqn 11}$$

where,  $A_{ac}$  is the parameter value when  $T_{growth}=0$  and  $\alpha_{ac}$  is the acclimation coefficient ( $^{\circ}\text{C}^{-1}$ )

If a parameter showed only adaptation to climate of origin, the function was:

$$f(T_{home}) = A_{ad} + \alpha_{ad}T_{home} \quad \text{Eqn 12}$$

We combined Eqns 11 and 12 to represent both acclimation and adaptation, defined as

$$f(T_{home}, T_{growth}) = A_{ad} + \alpha_{ad}T_{home} + \delta_{ac}(T_{growth} - T_{home}) \quad \text{Eqn 13}$$

here,  $\delta_{ac}$  is the acclimation coefficient corresponding to a unit deviation in  $T_{growth}$  from the species'  $T_{home}$  ( $^{\circ}\text{C}^{-1}$ ). We parameterised Eqn 11 and 12 independently using data from seasonal photosynthetic response studies (Eqn 11) and common garden experiments (Eqn 12). Eqn 13 was parameterised using combined seasonal and common garden datasets. We implemented the modified functions into the FvCB model (see Duursma, 2015) to simulate photosynthetic temperature response curves at a constant  $C_i$  of  $275 \mu\text{mol mol}^{-1}$  and tested how well the leaf scale photosynthesis model captured the observed temperature optimum of photosynthesis in the mature plants dataset. This provided an independent comparison as the mature plants dataset was not used to parameterise the temperature acclimation and adaptation functions (Eqn 11-13).

### *Statistical analysis*

Parameters of Eqn 1, 7-9 were estimated in a non-linear mixed model framework (Zuur *et al.*, 2009) using the *nlme* function within the *nlme* package in R version 3.3.2 (R Development Core Team, 2012). Replicate trees and/or leaves of the same species were included as random effects in model. However, when datasets contained measurements of multiple species (e.g. Brazilian rainforests, Australian rainforests and Australian semi-arid woodland datasets, Table S1), individual species were considered as a random variable in the model. Similarly, Eqns 11-13 were parameterized in a linear mixed model framework using the inverse of the standard error (SE) of each parameter of Eqn 1, 7-9 as the weighting scale to account for parameter uncertainty (Zuur *et al.*, 2009; Lin *et al.*, 2015). We tested whether the model parameters (Eqn 11-13) significantly differed among datasets (and/or species) by fitting linear mixed models with and without random slopes and intercepts for each dataset (and or species). These models were then compared using a likelihood ratio test (Zuur *et al.*, 2009) to determine whether the acclimation and adaptation coefficients differed among species. We used standard model validation tools (normal quantile plots and residual plots) to test the underlying assumptions in linear mixed models and used marginal and conditional  $r^2$  values to evaluate the goodness of fit (Nakagawa & Schielzeth, 2013). The complete database used for this analysis is available as a public data product (Kumarathunge *et al.*, 2018) and code used for the entire analysis is publicly available through <https://bitbucket.org/Kumarathunge/photom>

## Results

### *Temperature optimum for net photosynthesis at saturating irradiance ( $T_{opt_A}$ )*

The temperature optimum for leaf level net photosynthesis at saturating irradiance ( $T_{opt_A}$ ) of mature plants in their natural habitats was strongly correlated with the temperature of the growth environment ( $T_{growth}$ ; mean air temperature of preceding 30 days) (Fig. 1a, Table 1). Values of  $T_{opt_A}$  ranged from 16.3 to 32.4 °C, where the minimum and maximum values were observed for Arctic vegetation and tropical evergreen trees, respectively. The rate of increase in  $T_{opt_A}$  was  $0.62 \pm 0.07$  °C per °C increase in  $T_{growth}$ .

In the seasonal dataset (Fig. 1b), we found strong evidence for acclimation of  $T_{opt_A}$  to the prevailing growth temperature.  $T_{opt_A}$  showed a significant increasing trend with  $T_{growth}$ . The mean rate of increase in  $T_{opt_A}$  was  $0.34 \pm 0.05$  °C per unit increase in  $T_{growth}$  (Table 1). In contrast, no trend was observed with climate of origin in common garden studies (Table 1). Here, we tested for a relationship between  $T_{opt_A}$  and the  $T_{home}$  (1960-1990 mean maximum air temperature of the warmest month at species' seed source) and we did not find any significant relationship for  $T_{opt_A}$  with  $T_{home}$ . (Fig. 1c, Table 1). The results were similar for the two alternative definitions of the climate of origin (Table S2). The lack of a significant relationship with the species' home temperature in the common garden datasets suggests that the variation in  $T_{opt_A}$  of mature plants across ecosystems (Fig. 1a) is more strongly driven by acclimation to growth temperatures (Fig. 1b) than by local adaptation to climate of origin (Fig. 1c).

### *Temperature optimum for photosynthesis at a common $C_i$ ( $T_{opt_{A275}}$ )*

Similar to  $T_{opt_A}$ ,  $T_{opt_{A275}}$  showed a strong correlation with  $T_{growth}$  in mature plants across ecosystems (Fig. 1d, Table 1). We found no significant differences in either intercept or slope of the linear regression between  $T_{opt_A}$  and  $T_{opt_{A275}}$  vs  $T_{growth}$  (Table 1), in both the mature (Fig 1a, d) and seasonal (Fig 1b, e) datasets, strongly suggesting that the observed variation in  $T_{opt_A}$  among ecosystems is not due to variation in the stomatal limitation of  $T_{opt_A}$ . This result also suggests that the observed seasonal pattern of  $T_{opt_A}$  (Fig. 1b) was not driven by stomatal processes but rather by the effects of photosynthetic biochemical processes. Similar to  $T_{opt_A}$ , species in common garden studies did not show significant trends for  $T_{opt_{A275}}$  with  $T_{home}$  (Fig. 1f).

## Temperature dependence of biochemical capacities, $J_{max}$ & $V_{cmax}$

Similar to  $Topt_A$ , we found a strong increase in both  $Topt_V$  and  $Topt_J$  with  $T_{growth}$  in the mature plants dataset (Fig. 2a, d). The slopes of the linear regression with  $T_{growth}$  were similar for  $Topt_V$  and  $Topt_J$  ( $0.71 \pm 0.20$  and  $0.63 \pm 0.15^\circ\text{C}^\circ\text{C}^{-1}$  respectively). These sensitivities are similar in magnitude to the sensitivity of  $Topt_A$  and  $Topt_{A275}$  to  $T_{growth}$  in the mature plants dataset. For  $V_{cmax}$ , the trend in  $Topt$  was caused by an increase ( $p \approx 0.06$ ) in  $Ea_V$  with increasing  $T_{growth}$ , and a strong decline in  $\Delta S_V$  (Fig. 2b, c). For  $J_{max}$ , however, there was no change in  $Ea_J$ , only a decline in  $\Delta S_J$  with increasing  $T_{growth}$  (Fig. 2e, f).

We decomposed the observed trends across biomes shown in Fig. 2 by looking at seasonal datasets (Fig. 3) and common garden studies (Fig. 4) independently to identify the effect of seasonal acclimation and local adaptation of photosynthetic biochemical component processes. We found a strong increase in  $Topt_V$  and  $Topt_J$  with  $T_{growth}$  (Fig. 3a, d). The rate of increase in  $Topt_J$  per unit increase in  $T_{growth}$  was slightly higher than the  $Topt_V$  (Table 1) but the difference was not significant. Further, these sensitivities were found to be similar to the sensitivity of both  $Topt_A$  and  $Topt_{A275}$  to  $T_{growth}$ . Similar to the mature plants dataset, we found a significant positive trend for  $Ea_V$  and a decreasing trend ( $p \approx 0.08$ ) for  $\Delta S_V$  with increasing  $T_{growth}$ . (Fig. 3b, c). For  $J_{max}$ , however, there was no change in  $Ea_J$ , only a strong decline in  $\Delta S_J$  with increasing  $T_{growth}$ . (Fig. 3e, f).

We found no evidence to support adaptation of  $Topt_V$ ,  $Ea_V$  and  $\Delta S_V$  to climate of origin as there were no significant trends observed with temperature at species' seed source (i.e.  $T_{home}$ ) in the common garden dataset (Fig. 4a, b, c). These observations were consistent with the lack of significant trends for  $Topt_A$  in the common garden dataset. However,  $Topt_J$  and  $\Delta S_J$  showed significant trends with  $T_{home}$  (Fig. 4d, e, f; Table 1), suggesting adaptation of both parameters to climate of origin. The results were similar for the two alternative definitions of the climate of origin (Table S2).

## The balance between $J_{max}$ and $V_{cmax}$

We found no detectable correlation between  $T_{growth}$  and the basal rate of  $V_{cmax}$  at a standard temperature  $25^\circ\text{C}$  for mature plants in their natural habitats, but the basal rate of  $J_{max}$  showed a strong decrease (Fig. 5a, b). The ratio of  $J_{max} \cdot V_{cmax}$  at  $25^\circ\text{C}$  ( $JV_r$ ) showed a significant decrease with increasing  $T_{growth}$  (Fig. 5c, Table 1). We excluded the Scots pine,

Finland dataset when fitting linear regression as the  $JV_r$  value significantly departed from the general trend, and was therefore identified as an outlier (black circle in Fig. 5c).

Basal rates of  $V_{cmax}$  and  $J_{max}$  did not show significant trends with  $T_{growth}$ , but  $JV_r$  responded negatively to  $T_{growth}$  in the seasonal dataset (Fig. 5d: f). We found no evidence to support adaptation of basal rates of  $V_{cmax}$  and  $J_{max}$  to climate of origin; no parameters showed any significant trend with  $T_{home}$  in the common garden dataset (Fig. 5g, h, Table 1). However, there was evidence of adaptation of  $JV_r$  to climate of origin, as  $JV_r$  showed a significant decrease with  $T_{home}$  in the common garden dataset (Fig. 5i, Table 1).

#### *Assessing the role of day respiration*

We found no detectable trends (Fig. S3, Table 1) for either  $R_{L25}$  or  $E_a$  of mature plants in native environments. Similar results were found for common garden studies and no seasonal trends were observed for either  $R_{L25}$  or  $E_a$  in the seasonal dataset. However, the data showed a slight negative trend for  $R_{L25}:V_{cmax25}$  ratio with increasing  $T_{growth}$  (of mature plants in native environments) and  $T_{growth}$  (of seasonal datasets) (Fig. S4). Also we observed negative  $E_a$  values in all three datasets (Fig. S4).

#### *Model to represent acclimation and adaptation in vegetation models*

Our results provide evidence that changes in the temperature response of photosynthesis among datasets are principally driven by acclimation of photosynthetic biochemistry to growth temperature. Both  $Ea_V$  and  $JV_r$  showed strong acclimation to growth temperature with significant (albeit weak) acclimation of  $\Delta S_V$ . We found little evidence to support local adaptation of photosynthetic biochemistry to climate of origin. Only  $JV_r$  and  $\Delta S_J$  showed statistically significant, but weak signals of local adaptation. We further tested whether variation in  $Ea_V$  and  $JV_r$  can explain the seasonal acclimation of temperature optimum of photosynthesis observed in the seasonal dataset using linear regression analysis ( $JV_r$  and  $Ea_V$  vs  $Topt_{A275}$ ). We found a strong negative trend for the relationship between  $JV_r$  and  $Topt_{A275}$  (Fig. 6a).  $Topt_{A275}$  increased by  $\sim 6^\circ\text{C}$  for a unit decrease in  $JV_r$ . Also, we found significant trend between  $Ea_V$  and  $Topt_{A275}$ ;  $Topt_{A275}$  increased by  $\sim 0.2^\circ\text{C}$  for a unit increase in  $Ea_V$  (Fig. 6b). Therefore, the observed trends in  $Topt_A$  of mature plants in native habitats (Fig. 1a) can be explained by the effect of growth temperature on  $Ea_V$ ,  $\Delta S_V$ ,  $JV_r$  and the

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effects of both growth temperature and climate of origin on  $\Delta S_J$  and  $JV_r$ . Hence, photosynthetic temperature acclimation and adaptation can be implemented in GVMs using these parameters. Therefore, we modified the baseline peaked Arrhenius functions (Eqn 8) to represent i) temporal variability of  $Ea_V$  and  $\Delta S_V$  using Eqn 12, ii) geographical and temporal variation of  $JV_r$  ratio at 25°C and  $\Delta S_J$  using Eqn 13. The full final model is given in Table 2.

We found that the new temperature response functions were able to predict the temperature optima of photosynthesis observed in field-grown mature plants with a high degree of accuracy ( $r^2=0.80$ ). The slope ( $1.09\pm 0.15$ ) and intercept ( $-2.20\pm 4.10$ ) of the linear regression between the predicted and observed  $Topt_A$  were not significantly different from unity and zero respectively (Fig. 7a, Table S3). Our new model outperformed the Kattge & Knorr (2007) algorithms, which tend to underpredict  $Topt_A$  (Fig. 7b, Table S3). Further, the use of PFT-specific values of  $V_{cmax}$ , together with a standard unacclimated photosynthetic temperature responses (Leuning, 2002), was not able to predict the observed variability in  $Topt_A$  as it predicts a  $Topt_A \approx 25^\circ\text{C}$  for all datasets (Fig 7a). Note that the mature plant dataset was not included in fitting Eqn 11-13, so that the predicted  $Topt_{A275}$  in Fig. 7a was independent of the data used to derived the model parameters.

## Discussion

We developed new mathematical functions to represent the photosynthetic temperature response in vegetation models to account for both acclimation to growth temperature and adaptation to climate of origin using a global database that contains more than 140 species. We found acclimation to growth temperature to be the principal driver of the photosynthetic temperature response, and observed only a few modest effects of adaptation to temperature at the climate of origin. The observed variation of temperature optimum for leaf net photosynthesis was primarily explained by the photosynthetic biochemical component processes rather than stomatal or respiratory processes. The new temperature response functions presented here capture the observed  $Topt_A$  across biomes with higher degree of accuracy than previously proposed algorithms.

## *Adaptation of $T_{opt_A}$ to climate of origin*

Despite a significant range in long term mean temperature at species' seed sources, we found no predictable relationship for  $T_{opt_A}$  with climate of origin when species were grown in common gardens. Therefore, our results do not support the hypothesis  $T_{opt_A}$  is adapted to species' climate of origin (hypothesis 1). Our results contrast with previous studies which found that  $T_{opt_A}$  is related to species climate of origin (Fryer & Ledig, 1972; Slatyer, 1977; Slatyer, 1978; Robakowski *et al.*, 2012), but there are a number of studies which compare the temperature response of photosynthesis and report a lack of local adaptation of  $T_{opt_A}$  (Ledig & Korbobo, 1983; Gunderson *et al.*, 2000). We propose two hypotheses to explain the lack of local adaptation of  $T_{opt_A}$ ; i) there is a lack of specialization in photosynthetic biochemistry in relation to climate of origin and ii) the capacity of species to adjust their  $T_{opt_A}$  to temporal variations in local thermal environment could mask ecotypic thermal adaptation of  $T_{opt_A}$  (Robakowski *et al.*, 2012).

With respect to hypothesis (i), Rubisco activity is one of the key photosynthetic biochemical determinants and one of the most temperature responsive physiological process (Galmés *et al.*, 2015). Several lines of evidence suggest that Rubisco catalytic properties, including the relative specificity for  $\text{CO}_2/\text{O}_2$  ( $S_{c/o}$ ), the Michaelis–Menten constants for  $\text{CO}_2$  ( $K_c$ ) and  $\text{O}_2$  ( $K_o$ ), and the maximum turnover of carboxylation ( $k_c$ ), differ among species that have evolved under different thermal environments (Andersson & Backlund, 2008; Galmés *et al.*, 2014). However, it is not clear whether these differential responses are due to genetic adaptation of Rubisco kinetics to climate of origin or to the temporal effects of growth temperature. Galmés *et al.* (2015) argued that closely related species could be less adapted to their current thermal environment due to past strategies that limit adaptation of Rubisco to new thermal regimes (Lambers *et al.*, 2008). This hypothesis was further supported by Savir *et al.* (2010) who suggested point mutations may not cause a significant improvement in Rubisco activity due to its close optimality in the net photosynthetic rate (Tcherkez *et al.*, 2006). As a result, the adaptive evolution of Rubisco to novel thermal environments may be rare, as adaptation to a local environment will be working against the selective pressure to cope with seasonal and annual temperature variations and would reduce species fitness, and expansion into new niches with different thermal environments. Other than the parameters  $\Delta S_J$  and  $JV_r$ , our results do not show evidence for thermal adaptation of photosynthetic biochemical parameters. Thus we suggest that the lack of local adaptation of  $T_{opt_A}$  may be

partially explained by the lack of specialization in photosynthetic biochemistry, particularly Rubisco kinetic properties to species climate of origin.

Regarding ii), we suggest that the capacity of Rubisco kinetic properties to adjust to temporal variations in growth temperature could potentially mask the species' pre-adaptive responses to their original thermal environments. Here, we show strong evidence for the acclimation of  $T_{opt_A}$  to species  $T_{growth}$  which is primarily due to the variations in photosynthetic biochemical component processes  $JV_r$ ,  $Ea_V$ ,  $\Delta S_V$  and  $\Delta S_J$  in relation to the seasonal temperature dynamics. Potential mechanisms by which the kinetic properties of Rubisco could be altered in response to changes in temperature include structural changes in the Rubisco enzyme itself (Huner & Macdowall, 1979; Huner, 1985; Yamori *et al.*, 2006); changes in the concentration of other photosynthetic enzymes such as Rubisco activase (Yamori *et al.*, 2005, Yamori *et al.*, 2011); expression of cold/heat stable isozymes (Yamori *et al.*, 2006); and by alterations in membrane fluidity (Falcone *et al.*, 2004). A number of previous studies have demonstrated short-term acclimation of Rubisco kinetics to growth temperature (Medlyn *et al.*, 2002b; Yamori *et al.*, 2006; Kattge & Knorr, 2007; Lin *et al.*, 2013; Yamaguchi *et al.*, 2016; Smith & Dukes, 2017; Crous *et al.*, 2018) although the sensitivities of the responses varied. In addition, studies that have compared the acclimation capacity of multiple species in common growth temperatures have shown similar direction and magnitude of short-term temperature acclimation of  $T_{opt_A}$  (Berry & Björkman, 1980; Sendall *et al.*, 2015) and Rubisco kinetics (Lin *et al.*, 2013; Smith & Dukes, 2017) across species irrespective of their climate of origin. Therefore, we argue that the capacity of species to adjust their photosynthetic biochemistry to temporal variations in growth temperature provides a fitness advantage over that of local climatic adaptation of  $T_{opt_A}$  and its related mechanisms, by enabling species to optimize carbon balance in their current habitat (Hikosaka *et al.*, 2006).

The lack of a temperature adaptation response in this study contrasts with the results of a previous meta-analysis which found both evolutionary changes and an acclimation effect on  $T_{opt_A}$  (Yamori *et al.*, 2014). Our common garden studies compared closely related species (or provenances of the same species) in most cases. The most climatically divergent sets of species included in this study were those of Vårhammar *et al.* (2015) (lowland and montane tropical species) and Dillaway and Kruger (2010) (North American boreal and temperate deciduous species; see Table S1). In contrast, Yamori *et al.* (2014) compared temperature responses of C<sub>3</sub>, C<sub>4</sub> and CAM plants and found evidence of evolutionary shifts among these

functional groups. Other common garden studies with taxonomically diverse species have also provided evidence for evolutionary changes in  $T_{opt_A}$  in relation to climate of origin (Cunningham & Read, 2002; Reich *et al.*, 2015).

### *Acclimation of $T_{opt_A}$ to growth temperature*

Our observations of seasonal photosynthetic temperature response datasets suggest that the seasonal plasticity in  $T_{opt_A}$  is principally driven by (i) the adjustment of the temperature response of the Rubisco limited photosynthetic rate and (ii) the balance between Rubisco limited and electron transport limited photosynthetic rates. These two mechanisms control the seasonal shifts in  $T_{opt_A}$  as follows. First, at biologically relevant leaf temperatures, the light saturated net photosynthetic rate is mostly limited by Rubisco activity (Rogers & Humphries, 2000; De Kauwe *et al.*, 2016; Yamaguchi *et al.*, 2016). An increase in  $E_{a_V}$  along with a decrease in  $\Delta S_V$  increases the Rubisco-limited photosynthetic rate with temperature, and thus affects the shape of the photosynthetic temperature response. The rate of increase in  $E_{a_V}$  with  $T_{growth}$  in this study ( $1.14 \text{ kJ mol}^{-1} \text{ }^\circ\text{C}^{-1}$ ) aligns closely with previous reports (Hikosaka *et al.*, 2006:  $1.01 \text{ kJ mol}^{-1} \text{ }^\circ\text{C}^{-1}$ ). A number of potential causes have been suggested for variations in  $E_{a_V}$  across species, including mesophyll conductance to  $\text{CO}_2$  diffusion (Bernacchi *et al.*, 2002; Warren *et al.*, 2007; Walker *et al.*, 2013; von Caemmerer & Evans, 2015), kinetic properties of Rubisco (Yamori *et al.*, 2006), distribution of leaf nitrogen among photosynthetic proteins (Yin *et al.*, 2018) and the influence of other enzymes that affect the *in vivo* activity of Rubisco (Onoda *et al.*, 2005). Further, the Rubisco activation status could also be a significant factor contributing to the observed trends in  $E_{a_V}$  with  $T_{growth}$  as evidence suggested that, plants have the capacity to maintain high Rubisco activation status through an increase in Rubisco activase concentration and expression of heat stable Rubisco activase isoforms (Crafts-Brandner & Salvucci, 2000; Sage *et al.*, 2008; Yamori *et al.*, 2014). However, not all authors find a change in  $E_{a_V}$  with growth temperature. Kattge and Knorr (2007) did not find any temperature acclimation in  $E_{a_V}$ . They argued that the choice of a standard, rather than peaked, Arrhenius model to fit the temperature response for  $V_{cmax}$  without considering the deactivation energy would be a possible reason for the observed acclimation responses of  $E_{a_V}$  in previous studies (e.g. Hikosaka *et al.* 2006). However, here we used the peaked Arrhenius model, and thus the acclimation of  $E_{a_V}$  that we observed is not an artifact of model choice.

The second important mechanism for acclimation was a change in the magnitude of  $JV_r$ , as has also been observed by (Kattge & Knorr, 2007; Crous *et al.*, 2013; Lin *et al.*, 2013; Crous *et al.*, 2018). The ratio determines the transition between the two limiting steps,  $W_c$  and  $W_j$ . As the temperature responses of  $W_c$  and  $W_j$  are different from each other with different optimum temperatures ( $T_{opt}$  of  $W_c < T_{opt}$  of  $W_j$ ),  $T_{optA}$  is potentially determined by the limiting step (von Caemmerer & Farquhar, 1981; Hikosaka, 1997). At higher  $JV_r$ , the photosynthetic rate is mostly limited by RuBP carboxylation, therefore,  $T_{optA}$  tends to be a lower value and vice versa.

The acclimation capacity of  $\Delta S_V$  observed in this study ( $-0.38 \text{ J mol}^{-1} \text{ K}^{-1}$ ) was lower compared to the  $-1.07 \text{ J mol}^{-1} \text{ K}^{-1} \text{ }^\circ\text{C}^{-1}$  reported in (Kattge & Knorr, 2007). The higher sensitivity observed in Kattge and Knorr (2007) would potentially be explained by the lack of variation in  $Ea_V$ . Both  $Ea_V$  and  $\Delta S_V$  are correlated: a high sensitivity in  $Ea_V$  to  $T_{growth}$  would potentially cause  $\Delta S_V$  to be less sensitive and vice versa.

We observed changes in  $JV_r$  with temperature in all three datasets (Fig. 5), but only the mature plant dataset showed a change in either of the two terms contributing to this ratio. In this dataset, the reduction in  $JV_r$  is driven by a reduction in  $J_{max25}$ , whereas in the other two datasets, there is no overall effect on either  $V_{cmax25}$  or  $J_{max25}$ . Some previous studies have observed changes in  $V_{cmax25}$  with growth temperature in more limited datasets (Way & Oren, 2010; Lin *et al.*, 2013; Ali *et al.*, 2015; Scafaro *et al.*, 2017; Crous *et al.*, 2018; Smith & Dukes, 2018), but here we did not find any consistent pattern in  $V_{cmax25}$ . It appears that  $JV_r$  responded strongly and consistently to growth temperature, but whether this is achieved by increasing  $V_{cmax}$ , decreasing  $J_{max}$ , or both, is highly variable. We speculate that the global pattern in  $J_{max}$  observed in Figure 5b may be a response to increasing light availability in the tropics, following the co-limitation hypothesis, as proposed by Dong *et al.* (2017), rather than a response to growth temperature.

#### *Improved temperature response functions for photosynthetic capacity*

We demonstrate acclimation to growth temperature to be the principal driver, and only a few modest effects of adaptation, in photosynthetic temperature responses at global scale. Our results highlight the limitation of using a fixed set of parameters to determine  $T_{optA}$ , and challenge the use of PFT-specific  $V_{cmax25}$  and  $J_{max25}$  with a fixed set of temperature response parameters without accounting for temperature acclimation and adaptation (Leuning, 2002) in global vegetation models (Harper *et al.*, 2016; Rogers *et al.*, 2017a). We also demonstrate that the current representation of photosynthetic temperature acclimation (Kattge & Knorr,

2007) that has been implemented in some vegetation models (Smith & Dukes, 2013; Lombardozzi *et al.*, 2015; Smith *et al.*, 2016), was not able to predict the observed patterns in  $T_{optA}$  across biomes.

We proposed new algorithms for temperature response that are based on a broad range of data, account for both geographical and temporal variability in photosynthetic biochemical component processes, and are able to capture observed variation of  $T_{optA}$  across biomes with a high degree of accuracy. The temperature response functions that we propose have a broad temperature domain ( $\sim 3 - 37$  °C) which should enable their use in GVMs without outer domain uncertainties (Stinziano *et al.*, 2017)), a limitation of the algorithms proposed previously (Katte & Knorr, 2007) that are widely implemented in GVMs (BETHY, CLM4.5, Orchidee). Due to these advantages, our new photosynthetic temperature algorithms provide an improved representation of geographical and temporal variability in  $T_{optA}$  and should ultimately improve the accuracy of predicted future C cycle in GVMs.

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## Author contribution statement

Project conceived by BEM. Analyses designed and carried out by DPK with guidance from BEM, JED, MGT, and contributions from MGDK. Manuscript writing led by DPK, BEM and JED. All other co-authors (MJA, MB, FJC, KRC, MAC, LAC, JQC, KYC, DND, ED, DSE, OG, QH, KH, AMJ, JWGK, ELK, LMM, YO, PBR, AR, MS, NGS, LT, DTT, HFT, EST, JU, AV, GW, JMW, DAW) contributed data, ideas, and edited the manuscript.

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## Supporting Information

**Fig. S1** Distribution of the dataset used in this study

**Fig. S2** Relationship between apparent  $V_{\text{cmax}}$  and  $J_{\text{max}}$  values derived using two  $AC_i$  curve fitting routines; with and without accounting for TPU limitation

**Fig. S3** Temperature response parameters of photosynthetic respiratory component processes

**Fig. S4**  $R_{L25} : V_{\text{cmax}25}$  ratio at a standard leaf temperature 25°C

**Table S1** List of data sources

**Table S2** Results of the linear mixed effect models fitted for common garden dataset to test for adaptation of photosynthetic temperature response parameters to species' climate of origin

**Table S3** Results of the linear regression analysis between observed and modelled temperature optimum for photosynthesis at a fixed  $C_i$  of 275  $\mu\text{mol mol}^{-1}$  using model parameterizations given in Table 2 in the main text and Kattge & Knorr (2007) algorithm.

**Fig. 1** Temperature optimum for (a–c) leaf net photosynthesis ( $Topt_A$ ) and (d–f) net photosynthesis at an intercellular  $\text{CO}_2$  concentration of 275  $\mu\text{mol mol}^{-1}$  ( $Topt_{A275}$ ) of mature plants growing in their native environments (a, d), species in field (grown at ambient growth temperatures) measured at least in two or more seasons (b, e) and species or provenances from contrasting climates of origin grown in common growth temperatures (common gardens or controlled environments; c, f).  $T_{\text{growth}}$  is the mean air temperature of preceding 30 d.  $T_{\text{home}}$  is the long-term (1960–1990) mean maximum temperature of the warmest month at species' seed origin. Different colours in (a, b) depict plant functional types: orange, tropical evergreen angiosperms (EA-Tr); light blue, arctic tundra; red, temperate deciduous angiosperms (DA-Te); blue, temperate evergreen angiosperms (EA-Te); green, boreal evergreen gymnosperms (EG-Br); purple, temperate evergreen gymnosperms (EG-Te); in (c–f) different datasets. The thick black lines are: (a, d) least-squares linear regression fits; (b, c, e, f) linear mixed-effect model fits with random intercepts for each dataset. The thin lines in

respective colours are the fitted random intercept models for individual datasets. Error bars represent  $\pm 1$  SE.

**Fig. 2** Biochemical temperature response parameters for the mature plants dataset in relation to mean air temperature of preceding 30 d ( $T_{growth}$ ). Different colours represent plant functional types as in Fig. 1(a, d). Solid and dotted lines in each panel are the least-squares linear regression fits (this study; coefficients and  $r^2$  values given in Table 1) and the linear models proposed by Kattge & Knorr (2007), respectively. Error bars represent  $\pm 1$  SE. Legend follows Fig. 1(a, d).

**Fig. 3** Biochemical temperature response parameters for the Seasonal dataset in relation to mean air temperature of preceding 30 d ( $T_{growth}$ ). Data were measured on field-grown plants (including whole-tree chamber experiments) in two or more seasons. Solid and dotted lines in each panel are the linear mixed-effect model fits (this study; coefficients and  $r^2$  values are given in Table 1) and the linear models proposed by Kattge & Knorr (2007), respectively. Error bars represent  $\pm 1$  SE. Legend follows Fig. 1(b, e).

**Fig. 4** Biochemical temperature response parameters for the Common garden dataset in relation to the long-term (1960–1990) mean maximum temperature of the warmest month at species' seed origin ( $T_{home}$ ). Data were measured in species or provenances from contrasting climates of origin grown at common growth temperatures (common gardens and controlled environments). Solid lines in each panel are the linear mixed-effect model fits (this study; coefficients and  $r^2$  values are given in Table 1). Error bars represent  $\pm 1$  SE. Legend follows Fig. 1(c, f).

**Fig. 5**  $V_{cmax}$ ,  $J_{max}$  and  $J_{max} \cdot V_{cmax}$  ratio ( $JV_r$ ) at a standard leaf temperature ( $25^\circ\text{C}$ ) of (a–c) mature plants growing in their native environments; (d–f) field-grown plants measured in two or more seasons; and (g–i) species or provenances from contrasting climates of origin grown in common growth temperatures (common gardens or controlled environments).  $T_{growth}$  is the mean air temperature of preceding 30 d.  $T_{home}$  is the long-term (1960–1990) mean maximum temperature of the warmest month at species' seed origin, respectively. Solid lines in each panel are the least-squares linear regression fits (b, c), linear mixed-effect model fits with random intercepts for each dataset (f, i). One outlier is circled in (c) (see text). Error bars represent  $\pm 1$  SE. Legend follows Fig. 1.

**Fig. 6** Relationship between  $JV_r$  and temperature optimum for photosynthesis at a fixed intercellular  $\text{CO}_2$  concentration of  $275 \mu\text{mol mol}^{-1}$  ( $T_{opt_{A275}}$ ) (a) and relationship between  $E_{aV}$  and  $T_{opt_{A275}}$  (b). Data were measured on field-grown plants (including whole-tree chamber experiments) in two or more seasons. Lines in each panel are the linear mixed effect regression model fits (a,  $T_{opt_{A275}} = 35.78 - 5.93 \times JV_r$ ,  $R^2 = 0.36$ ; b,  $T_{opt_{A275}} = 13.11 + 0.20 \times E_{aV_{cmax}}$ ,  $R^2 = 0.49$ ). Error bars represent  $\pm 1$  SE.

**Fig. 7** Observed and modelled temperature optimum for photosynthesis at a fixed  $C_i$  of  $275 \mu\text{mol mol}^{-1}$  using model parameterizations given in Table 2. (a) With acclimation and adaptation functions developed in this study ( $y = 1.09x - 2.20$ ,  $r^2=0.80$ ), (b) Kattge & Knorr (2007) acclimation function ( $y = 1.58x - 13.82$ ,  $r^2=0.83$ ). The crossed circle in the  $x$ -axis of (a) depicts the predicted  $T_{opt_{A275}}$  with a fixed set of parameters without acclimation and adaptation (Leuning, 2002). Colours reflect Plant Functional Types as in Fig 1. Thin lines, 1 : 1 relationship; thick lines, least-squares regression fit. In (a), the intercepts and the slope of the linear regression were not significantly different from zero and unity respectively (Supporting Information Table S3). Error bars represent  $\pm 1$  SE.

**Table 1** Results of the linear regression analysis of the parameters of Eqn 1, 8 and 9.

For common garden and seasonal datasets, linear mixed models were fit accounting for between datasets variations of a given parameter (see the Materials and Methods section for details). For mature plants in native environments, parameter values were derived by fitting simple linear regression models (Eqn 10). Values in parentheses are standard errors of estimates. Bold values are the significant parameters at  $\alpha = 0.05$ .

Mature plants in native environment (Eqn 10)					Seasonal dataset (Eqn 11)					Common garden dataset (Eqn 12)				
Parameter	<i>a</i>	<i>b</i>	<i>r</i> <sup>2</sup>	<i>P</i> -value	<i>A</i> <sub>ac</sub>	$\alpha$ <sub>ac</sub>	<i>r</i> <sup>2</sup> (Marginal)	<i>r</i> <sup>2</sup> (Conditional)	<i>P</i> - value	<i>A</i> <sub>ad</sub>	$\alpha$ <sub>ad</sub>	<i>r</i> <sup>2</sup> (Marginal)	<i>r</i> <sup>2</sup> (Conditional)	<i>P</i> -value
<i>Topt</i> <sub>A</sub>	<b>12.5 (1.4)</b>	<b>0.62 (0.1)</b>	<b>0.80</b>	<b>&lt;0.001</b>	<b>18.2 (1.1)</b>	<b>0.34 (0.05)</b>	<b>0.27</b>	<b>0.87</b>	<b>&lt;0.001</b>	24.8 (2.1)	0.07 (0.1)	0.01	0.71	0.309
<i>Topt</i> <sub>A<sub>275</sub></sub>	<b>14.9 (1.5)</b>	<b>0.63 (0.1)</b>	<b>0.84</b>	<b>&lt;0.001</b>	<b>20.5 (1.2)</b>	<b>0.24 (0.05)</b>	<b>0.16</b>	<b>0.85</b>	<b>&lt;0.001</b>	26.8 (2.3)	0.07 (0.1)	0.03	0.30	0.400
<i>Biochemical parameters</i>														
<i>V</i> <sub>cmax25</sub>	85.3 (16.7)	-1.84 (0.8)	0.19	0.404	58.2 (12.0)	0.50 (0.4)	0.01	0.94	0.252	33.4 (28.0)	1.62 (0.9)	0.07	0.91	0.096
<i>J</i> <sub>max25</sub>	<b>194.7 (24.1)</b>	<b>-5.13 (1.2)</b>	<b>0.53</b>	<b>&lt;0.001</b>	141.3(18.8)	-1.35 (0.7)	0.03	0.95	0.053	92.7 (47.2)	1.63 (1.6)	0.02	0.95	0.312
<i>Ea</i> <sub>v</sub>	<b>48.7 (7.8)</b>	<b>0.82 (0.4)</b>	<b>0.14</b>	<b>0.067</b>	39.7 (6.2)	<b>1.14 (0.3)</b>	<b>0.32</b>	<b>0.91</b>	<b>&lt;0.001</b>	79.4 (13.1)	-0.37 (0.5)	0.14	0.14	0.450
<i>Ea</i> <sub>f</sub>	43.5 (9.8)	-0.19 (0.5)	0.05	0.7143	27.2 (5.0)	0.26 (0.3)	0.04	0.82	0.325	51.5 (8.7)	-0.38 (0.3)	0.20	0.20	0.247
$\Delta S$ <sub>v</sub>	<b>662.0 (8.7)</b>	<b>-1.31 (0.5)</b>	<b>0.30</b>	<b>0.011</b>	645.1 (4.6)	-0.38 (0.2)	0.09	0.82	0.089	647.9 (9.5)	-0.36 (0.3)	0.08	0.66	0.302
$\Delta S$ <sub>f</sub>	<b>667.3 (7.8)</b>	<b>-1.34 (0.4)</b>	<b>0.36</b>	<b>0.005</b>	<b>653.9 (4.6)</b>	<b>-0.85 (0.2)</b>	<b>0.22</b>	<b>0.94</b>	<b>&lt;0.001</b>	<b>662.3 (7.5)</b>	<b>-0.99 (0.3)</b>	<b>0.49</b>	<b>0.84</b>	<b>&lt;0.001</b>
<i>Topt</i> <sub>v</sub>	<b>24.3 (3.8)</b>	<b>0.71 (0.2)</b>	<b>0.40</b>	<b>0.002</b>	<b>30.3 (1.9)</b>	<b>0.36 (0.1)</b>	<b>0.23</b>	<b>0.77</b>	<b>&lt;0.001</b>	34.3 (3.3)	0.12 (0.1)	0.05	0.36	0.335
<i>Topt</i> <sub>f</sub>	<b>19.9 (2.9)</b>	<b>0.63 (0.2)</b>	<b>0.52</b>	<b>&lt;0.001</b>	<b>27.6 (1.8)</b>	<b>0.31 (0.1)</b>	<b>0.13</b>	<b>0.91</b>	<b>&lt;0.001</b>	<b>24.8 (3.4)</b>	<b>0.42 (0.1)</b>	<b>0.42</b>	<b>0.60</b>	<b>&lt;0.001</b>
<i>JV</i> <sub>r</sub>	<b>2.9 (0.2)</b>	<b>-0.06 (0.01)</b>	<b>0.66</b>	<b>&lt;0.001</b>	<b>2.3 (0.2)</b>	<b>-0.03 (0.01)</b>	<b>0.07</b>	<b>0.17</b>	<b>&lt;0.001</b>	<b>2.5 (0.3)</b>	<b>-0.03 (0.01)</b>	<b>0.13</b>	<b>0.64</b>	<b>0.005</b>
<i>Respiratory parameters</i>														
<i>R</i> <sub>L25</sub>	2.8 (0.5)	-0.09 (0.03)	0.38	0.0037	1.54 (0.42)	-0.01 (0.02)	0.01	0.25	0.502	1.16 (0.45)	0.01 (0.01)	0.01	0.61	0.583
<i>Ea</i>	-20.7 (14.3)	1.18 (0.78)	0.07	0.1508	-9.17 (11.49)	0.42 (0.61)	0.02	0.83	0.485	-4.25 (43.38)	0.12 (1.57)	0.01	0.93	0.937
<i>R</i> <sub>L25</sub> : <i>V</i> <sub>cmax25</sub>	0.036 (0.01)	-0.001 (0.0003)	0.22	0.033	<b>0.03 (0.01)</b>	<b>(0.0003)</b>	<b>0.04</b>	<b>0.60</b>	<b>0.043</b>	0.03 (0.01)	-0.0005 (0.0004)	0.06	0.53	0.149

**Table 2** Parameters of the temperature acclimation and adaptation functions developed in this study.

$T_{home}$ , long-term (1960–1990) mean maximum temperature of the warmest month;  $T_{growth}$ , mean air temperature of preceding 30 d. Plant functional types: DA-Te, temperate deciduous angiosperms; EA-Te, temperate evergreen angiosperms; EG-Te, temperate evergreen gymnosperms; EG-Br, boreal evergreen gymnosperms; EA-Tr, tropical evergreen angiosperms; Arctic tundra, Arctic spp.

Parameter	Model representation	Value	Units
$V_{max25}$	PFT specific	DA-Te	39.0
		EA-Te	82.9
		EG-Te	42.8
		EG-Br	80.4
		EA-Tr	39.4
		Arctic tundra	78.3
$J_{max25}$	Acclimation + Adaptation	$V_{max25} \times JV_r$	$\mu\text{mol m}^{-2}\text{s}^{-1}$
$JV_r$	Acclimation + Adaptation	$2.56 - 0.0375T_{home} - 0.0202(T_{growth} - T_{home})$	unitless
$Ea_V$	Acclimation	$42.6 + 1.14T_{growth}$	$\text{kJ mol}^{-1}$
$Ea_J$	Global mean	40.71	$\text{kJ mol}^{-1}$
$\Delta S_V$	Acclimation	$645.13 - 0.38T_{growth}$	$\text{J mol}^{-1} \text{K}^{-1}$
$\Delta S_J$	Acclimation + Adaptation	$658.77 - 0.84T_{home} - 0.52(T_{growth} - T_{home})$	$\text{J mol}^{-1} \text{K}^{-1}$













