

## Article (refereed) - postprint

---

Lahive, E.; Matzke, M.; Durenkamp, M.; Lawlor, A.J.; Thacker, S.A.; Pereira, M.G.; Spurgeon, D.J.; Unrine, J.M.; Svendsen, C.; Lofts, S. 2017. **Sewage sludge treated with metal nanomaterials inhibits earthworm reproduction more strongly than sludge treated with metal metals in bulk/salt forms.** *Environmental Science: Nano*, 4 (1). 78-88.  
[10.1039/C6EN00280C](https://doi.org/10.1039/C6EN00280C)

Copyright © The Royal Society of Chemistry 2017

This version available <http://nora.nerc.ac.uk/515967/>

NERC has developed NORA to enable users to access research outputs wholly or partially funded by NERC. Copyright and other rights for material on this site are retained by the rights owners. Users should read the terms and conditions of use of this material at <http://nora.nerc.ac.uk/policies.html#access>

**This document is the author's final manuscript version of the journal article following the peer review process. There may be some differences between this and the publisher's version. You are advised to consult the publisher's version if you wish to cite from this article.**

<http://www.rsc.org>

Contact CEH NORA team at  
[noraceh@ceh.ac.uk](mailto:noraceh@ceh.ac.uk)

**Sewage sludge treated with metal nanomaterials inhibits earthworm reproduction more strongly than sludge treated with metals in bulk/salt forms**

Lahive, E.<sup>a\*</sup>, Matzke, M.<sup>a</sup>, Durenkamp, M.<sup>b</sup>, Lawlor, A.J.<sup>c</sup>, Thacker, S.A.<sup>c</sup>, Pereira, M.G.<sup>c</sup>, Spurgeon, D.J.<sup>a</sup>, Unrine, J.M.<sup>d</sup>, Svendsen, C.<sup>a</sup>, Lofts, S.<sup>c</sup>

<sup>a</sup> NERC Centre for Ecology and Hydrology, Maclean Building, Benson Lane, Crowmarsh Gifford, Wallingford, OX10 8BB, United Kingdom

<sup>b</sup> Sustainable Soil and Grassland Systems, Rothamsted Research, Harpenden, Hertfordshire, AL5 2JQ, United Kingdom

<sup>c</sup> NERC Centre for Ecology and Hydrology, Lancaster Environment Centre, Lancaster, LA1 4AP, United Kingdom

<sup>d</sup> University of Kentucky, Department of Plant and Soil Sciences, 1100 S. Limestone St., Lexington, Kentucky 40546, United States.

\*Corresponding author email address: [elmhiv@ceh.ac.uk](mailto:elmhiv@ceh.ac.uk)

**Keywords:** Nanoparticles, biosolids, *Eisenia fetida*, bioavailability, toxicity, silver, zinc oxide

1 **Abstract**

2 Earthworms were exposed to soils amended with sewage sludges from a wastewater treatment plant  
3 (WWTP) treated with nanomaterials (ENMs) or metal/ionic salts. Sewage sludges were generated with  
4 either no metal added to the WWTP influent (control), ionic ZnO, AgNO<sub>3</sub> and bulk (micron sized) TiO<sub>2</sub>  
5 added (ionic metal-treated) or ZnO, Ag and TiO<sub>2</sub> ENMs added (ENM-treated). A sandy-loam soil was  
6 amended with the treated sewage sludge and aged in outdoor lysimeters for six months. Earthworms  
7 were exposed to the aged mixtures and a dilution of the mixtures (using control soil-sludge mix).  
8 Separate earthworm exposures to as-synthesized ENM and ionic metals salts (Zn/Ag singly) were  
9 carried out in the same soil. Earthworm reproduction was depressed by 90% in the high-metal ENM  
10 treatment and by 22-27% in the ionic metal and low-metal ENM soil-sludge treatments. Based on total  
11 metal concentrations in the soil-sludges the as-synthesised metal salt and ENM exposures predicted Zn  
12 was driving observed toxicity in the soil-sludge more than Ag. Earthworms from the high-metal ENM  
13 treatment accumulated significantly more Ag than other treatments whereas total Zn concentrations in  
14 the earthworms were within the range for earthworm Zn regulation for all treatments. This study  
15 suggests that current Zn limits set to provide protection against ionic metal forms may not protect soil  
16 biota where metals are input to WWTP in the ENM form.

17

18

19

20

21

22

23

24

25

## 26 **Introduction**

27 The growing use of engineered nanomaterials (ENMs) in numerous consumer products has led to an  
28 increase in their environmental inputs. ZnO, Ag and TiO<sub>2</sub> are among the most commonly used ENMs  
29 in consumer products such as cosmetics, personal care products, paints and antimicrobial treatments. A  
30 major transfer of ENMs from the point of use will be through sewer systems into wastewater treatment  
31 plants (WWTP). Within WWTP, ENMs have been shown to largely partition to sludge<sup>1,2,3</sup> the majority  
32 of which is subsequently applied to agricultural land as a fertiliser in many regions including the U.S.  
33 and the E.U..<sup>4</sup> Such disposal of sludge may result in the release of ENMs, or their transformation  
34 products, to soil ecosystems where they may cause toxicity to soil biota and/or enter food webs. As the  
35 environmental risk is yet to be fully understood, studies that simulate relevant exposure pathways  
36 relating to sludge application to land are clearly necessary to assess any potential impacts to soil biota  
37 of the incorporation of ENMs or their transformation products into sludge subsequently applied to land.

38 As-synthesised/as-manufactured nanoparticulate forms of ZnO and Ag have been shown to affect  
39 survival and life history traits of soil invertebrates, such as reproduction and growth. Ag ENMs often  
40 show effects at lower concentrations than Zn ENMs.<sup>5-9</sup> For ZnO and Ag ENMs, dissolution to their  
41 ionic forms in the soil porewater often has been related to observed toxicity,<sup>6-8</sup> although this is not  
42 always the case.<sup>9</sup> In contrast, TiO<sub>2</sub> ENMs show extremely low solubility<sup>10,11</sup> so dissolution products are  
43 unlikely to play a role in observed effects.<sup>11,12</sup> Indeed, compared to ZnO and Ag ENMs, TiO<sub>2</sub> ENMs  
44 have shown relatively low toxicity to soil organisms; **higher concentrations of TiO<sub>2</sub> ENMs are needed**  
45 **in the soil to cause mortality or reproduction effects compared to Zn or Ag.**<sup>5</sup> The bioavailability of  
46 ENMs to soil invertebrates has also been assessed by measuring the whole body metal concentrations  
47 of earthworms. In some cases tissue Ag and Zn tissue concentrations in ENM-exposed earthworms may  
48 reach concentrations which would normally result in mortality in equivalent ionic exposures but without  
49 the expected mortality effect occurring.<sup>7,8</sup> This suggests that the form in which soil invertebrates are  
50 exposed to metals (either as ENMs, ionic metal or a mixture of both) exerts important controls on metal  
51 handling and toxicity.

52 Previous soil invertebrate toxicity studies have largely considered only exposure to as-synthesised  
53 ENMs and not environmentally realistic scenarios where ENMs may have been transformed (for  
54 example in WWTP) into physicochemically distinct end products. In the WWT process Ag ENMs are  
55 completely sulfidised,<sup>1, 13-15</sup> while ZnO ENMs have been shown to become sulfidised, phosphatised or  
56 associated with FeO(OH).<sup>1</sup> In contrast, TiO<sub>2</sub> is expected to be much less likely to transform chemically,  
57 although surface properties and agglomeration/aggregation state may be altered. The only studies we  
58 are aware of investigating ENM toxicity after transformation by the WWT process found that ENM-  
59 containing sludge applied to soil inhibited nodulation in the model legume *Medicago truncatula* which  
60 could be linked with the down-regulation of genes involved in general stress responses, metal  
61 homeostasis, nodulation and nitrogen metabolism.<sup>16, 17</sup> Some adverse ecosystem responses to Ag ENMs  
62 in biosolids were found when applied to mesocosms; there were significant changes to microorganism  
63 abundance, function and community composition.<sup>18</sup> Other studies that investigated effects of ZnO  
64 ENMs in sewage sludge applied to soils found only slight effects on earthworm cocoon production and  
65 reduction in plant biomass (wheat, radish and vetch).<sup>19, 20</sup> These studies concluded that ZnO ENMs in  
66 sewage sludge pose a low environmental risk, although mostly in the latter studies as-synthesised ENMs  
67 were directly added to sludge rather than passed through the full WWT process.<sup>19, 20</sup> Thus there is still  
68 very little known about how transformed particles will behave in the environment and their subsequent  
69 bioavailability and toxicity to soil biota.

70 Earthworms are keystone species in terrestrial environments, integral to organic matter turnover in the  
71 soil, and so are a key group for which to assess the effects of amending soils with treated sludges with  
72 ENM inputs. In this study, earthworms were exposed to soils amended with sewage sludge generated  
73 by a pilot full WWTP.<sup>1</sup> Sewage sludges were generated where either no metal was added to the WWTP  
74 influent (control); non-ENM metal salts ZnSO<sub>4</sub>, AgNO<sub>3</sub> and bulk metal (micron-sized) TiO<sub>2</sub> were added  
75 to the influent (ionic metal treatment), or ZnO, Ag and TiO<sub>2</sub> ENMs were added to the influent (ENM  
76 treatment).<sup>1, 16, 17</sup> The bioavailability and toxicity of the metals in the different soil-sludge treatments to  
77 earthworms were compared by considering the metal concentration in the soil and the total body metal  
78 concentrations in the earthworms, linking to effects on the key life history traits of growth and  
79 reproduction. Toxicity data from as-synthesised Ag/ZnO ENM and Ag/Zn ionic metal salt single

80 exposures were used to predict effects of the total Zn and Ag concentrations in the soil and the total Zn  
81 and Ag concentrations in the earthworms from the soil-sludge exposures. Given that TiO<sub>2</sub> is known to  
82 have little or no toxicity for earthworms at the concentrations in the soil-sludge treatment<sup>5,11,12</sup> and are  
83 unlikely to transform, Ti was not considered as a toxicant in this study.

84

## 85 **Materials and Methods**

### 86 *Soil-sludge mixtures for toxicity tests*

87 The sludge generation and soil-sludge mixtures are described in detail in Ma et al 2014<sup>1</sup> and Judy et al  
88 2015<sup>16</sup> respectively. In brief, a sandy loam soil (Woburn, U.K.) was amended with sewage sludge  
89 derived from spiking WWTP influent with either (1) ZnO (30 nm uncoated described previously<sup>1</sup>), Ag  
90 (50 nm stabilized with a 55 kDa average molecular weight polyvinylpyrrolidone (PVP) previously fully  
91 described<sup>21</sup>) and TiO<sub>2</sub> (27±7.5 nm, Sigma Aldrich, (Figure S1, Supporting Information)) ENMs (ENM  
92 sewage sludge), (2) ZnSO<sub>4</sub>, AgNO<sub>3</sub> and micron-sized TiO<sub>2</sub> (ionic/bulk metal sewage sludge) or with  
93 (3) no metals (control). The intended concentration for Zn in the sludge was 2800 mg Zn/kg dry mass,  
94 based on the current U.S. cumulative pollutant loading limit for Zn in soils amended with biosolids  
95 (2800 kg Zn/ha).<sup>22</sup> Ag and Ti loadings were set to give intended sludge concentrations of 100 mg Ag/kg  
96 and 2400 mg Ti/kg (dry mass), respectively, based on percentiles (98<sup>th</sup>) of concentration from the U.S.  
97 targeted national sewage sludge survey.<sup>16</sup> A total of 40 kg of dry sludge (160 kg of wet sludge at 25  
98 weight % solids) were produced from each plant<sup>1</sup> (ENM, ionic metal and control) to be used in plant  
99 studies<sup>16,17,23</sup> and in this current earthworm study. Sludges were air-dried at Rothamsted Research (UK)  
100 and mixed with the sandy loam soil in a ratio of 0.58:0.42 soil:sludge, to give a target Zn concentration  
101 of 1400 mg Zn/kg dry soil in the ionic metal treatment.<sup>16</sup> The ratio of sludge to soil was based on the  
102 current U. S. EPA cumulative pollutant loading limit for Zn and was selected following the guidelines  
103 within *Guide to the Biosolids Risk Assessments for the EPA, CFR 40 Part 503*, which results in a 1:1  
104 soil: sewage sludge ratio in the top 15 cm of soil following 10 years of application at the maximum  
105 allowable concentration of Zn in sludge.<sup>16</sup> The soil-sludge mixtures were aged in freely-draining

106 outdoor lysimeters for six months<sup>23</sup> to create a set of ‘aged’ soil-sludge mixtures. Earthworms were  
107 exposed to five aged soil-sludge treatments; three of the treatments were the 0.58:0.42 soil:sludge  
108 mixture treatments: (1) control soil-sludge (no metal addition) (2) high-metal ENM soil-sludge, (3)  
109 high-metal ionic metal soil-sludge and the two other treatments were the high-metal ENM or ionic metal  
110 soil-sludge treatments mixed with control soil-sludge in a 1:1 ratio giving a (4) low-metal ENM soil-  
111 sludge and (5) a low-metal ionic metal soil-sludge (Figure S2, Supporting Information). To confirm  
112 that the earthworm reproduction was above the minimum number stipulated by OECD guidelines (>30  
113 juveniles), a soil control (Woburn sandy loam soil) without any sludge amendment was also included  
114 as a fully replicated test treatment.

115

#### 116 *Experimental design and toxicity test procedure*

117 The soil-sludge mixtures were distributed in four replicate containers each containing 300 g dry weight  
118 of the soil-sludge mix. There were also eight soil controls, each containing 550 g dry weight giving a  
119 comparable volume of soil to the soil-sludge mixtures due to differences in the bulk densities of the test  
120 media. All soils were wet to 50% of their respective water holding capacities (Table 1) using de-ionised  
121 water and left for ten days before the organisms were introduced. *Eisenia fetida* were initially obtained  
122 from a commercial source (Blades Biological, Kent, UK) and maintained in culture soil in a controlled  
123 temperature room at  $20 \pm 1$  °C in a 12:12 hour light:dark cycle.<sup>8</sup> The toxicity test procedure followed  
124 the OECD guideline 222 (earthworm reproduction test (*Eisenia fetida/andrei*)). Groups of ten fully-  
125 clitellated earthworms (average weight 10 worms =  $3.41 \pm 0.2$  g, Mean  $\pm$  SD, n=28) were rinsed, excess  
126 moisture removed with paper towel and weighed as a batch before being added to each replicate  
127 container. Horse manure (10 g dry weight), wetted to 80% of its water holding capacity, was added to  
128 the soil-only control treatments as food.<sup>24</sup> No food was added to the soil-sludge treatments as the sludge  
129 provided a food source for the earthworms that also allowed for oral exposure.<sup>25</sup> The earthworm  
130 exposure containers were kept in a controlled temperature room at  $20 \pm 1$  °C under a 12:12 hour  
131 light:dark cycle. After 14 and 28 days of incubation, earthworm survival and batch weight were  
132 measured. Surviving adult earthworms were removed from the test containers after 28 days and three

133 earthworms from each replicate were rinsed to remove adhered soil and then kept individually on clean  
134 filter paper for 24 hours to allow them to purge their gut contents<sup>7,8</sup>. This ensured that minimal soil was  
135 left in the earthworm prior to tissue Ag and Zn analysis. The soil-sludge mixtures were the incubated  
136 for a further 28 days to allow juveniles to hatch from laid cocoons. The number of juveniles was counted  
137 as previously described.<sup>8</sup>

138 In order to compare the toxicity observed in the ENM and ionic metal soil-sludge mixtures, single  
139 compound earthworm exposures (i.e. separate exposure were set up for each compound so they were  
140 not added as mixture) to as-synthesised ZnO ENM (30 nm uncoated)<sup>7</sup> and PVP-coated Ag EMM as  
141 well as Zn ( $\text{Zn}(\text{NO}_3)_2$ ) and Ag ( $\text{AgNO}_3$ ) salts (Sigma Aldrich, UK) were set up and run using the same  
142 procedures as for the soil-sludge exposures (i.e. 28 days survival test and 56 day reproduction test). The  
143 same sandy loam (Woburn) soil was spiked with the ENMs or salt, either Zn (100, 225, 500, 1100, 2200  
144 mg Zn/kg) or Ag (9, 22.5, 56.3, 141, 352, 880, 2200 mg Ag/kg), in triplicate according to the protocol  
145 previously described.<sup>8</sup> Spiked soils were wet to 50% of the water holding capacity (Table 1) and after  
146 one week ten adult earthworms were added to each test replicate. The toxicity test set up and duration  
147 followed the same as for the soil-sludge experiments above and previously described test protocols.<sup>7,8</sup>  
148 Three surviving adult earthworms were prepared and stored for tissue Zn or Ag analysis in the same  
149 manner as for the soil-sludge treatments. It was not possible to carry out these as-synthesised exposures  
150 in the control soil-sludge due to the limited amount that could be produced by the pilot WWTP.

151

#### 152 *Soil porewater extraction*

153 Soil porewater has been identified as an uptake route for ionic metal in soils.<sup>26 27, 28</sup> To get a better  
154 measure of metal reactivity in the soil, the soil porewater was extracted by centrifugation from each  
155 replicate of the soil-sludge mixtures at the end of the exposure period (56 days), before the juveniles  
156 were counted. Two 20 g (25 g from the soil control) (dry weight equivalent) soil samples, for separate  
157 Zn and Ag analysis, were collected from each of the treatment replicates, saturated to 140 % of the  
158 water holding capacity of the soil-sludge mixture and equilibrated overnight before porewater was



159 extracted following the extraction protocol described in Whitley et al 2013 but with two amendments  
160 to the protocol.<sup>29</sup> The soil sample extracted for Ag was filtered through glass wool and ultra-filters that  
161 were pre-soaked in a 0.1 M CuSO<sub>4</sub> solution to minimise Ag ion adsorption and losses.<sup>8,30</sup> The samples  
162 were centrifuging at 4000 g for 1.5 hours (J2-HC, Beckman Coulter, California, USA) to achieve  
163 maximum porewater extraction from the soil. A total of 5 ml of the extracted porewater was placed in  
164 a 10 kD ultra-filtration device (Amicon Ultra-15 Filters, Millipore, Ireland) and centrifuged for 1.5  
165 hours at 4000 g.<sup>8</sup> The extracted porewater and ultra-filtered porewater from each replicate were analysed  
166 for Ag or Zn using ICP-OES and pH measured (Sartorius Professional Meter PP-25, Sartorius AG,  
167 Goettingen, Germany; combination pH probe, filled with 3M KCl).

168

#### 169 *Chemical analysis*

170 Approximately 0.75 g of air-dried soil and soil-sludge mixtures or 0.5 g of freeze-dried whole  
171 earthworm were refluxed with a 3:1 mixture of hydrochloric and nitric acids (Merck, 'Aristar' grade)  
172 at 140°C for 2.5 h. After digestion the solutions were allowed to cool and then filtered using Whatman  
173 number 540 (12.5 cm diameter) filter papers that were pre-soaked with a 0.1 M CuSO<sub>4</sub> solution (Sigma-  
174 Aldrich, 'purum' grade). Digests were made up to a final volume of 50 ml with 0.5% v/v nitric acid and  
175 stored at 4°C prior to analysis for either Ag or Al and Zn. A 1 ml aliquot of porewater was digested  
176 with a 3:1 mixture of hydrochloric and nitric acids (Merck, 'Aristar') using closed Teflon vessels in a  
177 microwave digestion system (CEM Corporation, MARSXpress). The digests were heated to 180°C over  
178 a period of 30 minutes and then held at this temperature for a further 30 minutes. Digests were allowed  
179 to cool and then made up to a final volume of 50 ml with 1% v/v hydrochloric acid. The soil, porewater  
180 and earthworm digests were analysed by inductively coupled plasma mass spectrometry (ICPMS) using  
181 a Perkin Elmer Nexion 300D ICPMS instrument. The details of the procedures for checking the  
182 efficiency of the digestions, digest dilutions and instrument calibration are in the Supporting  
183 Information. For the primary element of interest, Ag, the ICPMS instrument detection limit was 0.14  
184 µg/l (mean blank + 3σ reagent blank, n=10) and the instrument method had a precision of 1.4 % (CoV,  
185 at 5 µg/l, n = 10).

186 Total metal concentrations in the earthworms were corrected, if necessary, for metal due to soil residues  
187 remaining in the gut following depuration. This was done using the total Al concentrations in the soils  
188 and earthworms. Aluminium was used to correct as it is naturally present at readily detectable  
189 concentrations in the soil and largely present in non-bioavailable forms, thus the concentrations in the  
190 worms could be attributed to residual soil present in the gut rather than to uptake into the tissues.

191 The expression used for correction was

$$192 \quad \{M\}_{\text{worm,corr}} = \{M\}_{\text{worm}} - m * \{Al\}_{\text{worm}} \quad (1)$$

193 where  $\{M\}_{\text{worm}}$  and  $\{Al\}_{\text{worm}}$  are the measured metal and Al concentrations in the worm and  $\{M\}_{\text{worm,corr}}$   
194 is the corrected tissue concentration. The term  $m$  is the slope of the linear regression of the measured  
195 worm metal against the measured worm Al. Separate regressions were done for body burden  
196 concentrations of worms exposed to each soil-sludge mixture. Significant relationships (regression  
197  $p < 0.05$ ) were found for Al and Ag or Zn concentrations in worms exposed to either the ionic metal or  
198 ENM-treated sludges, so corrections were applied to the total Ag and Zn concentration in the  
199 earthworms from these exposures.

200

### 201 *Data analysis*

202 Survival, weight change and reproduction were first checked for normal variance structure using the  
203 Anderson-Darling normality test and log transformed if required. Comparisons of survival,  
204 reproduction, and weight change, total Ag and Zn concentrations in the earthworms, total and ultra-  
205 filtered metal concentrations in the porewaters across all the treatments were carried out in Minitab 16  
206 using analysis of variance (ANOVA). Where significant differences were found, the Tukey test was  
207 used to identify the pattern of significant differences among treatments. Total and ultra-filtered  
208 concentrations for each treatment were also compared using an unstacked ANOVA.

209 To estimate response parameters for the as-synthesised ENMs and metal salts earthworm exposures,  
210 data for reproduction (juvenile production rate) was used to fit a three-parameter log-logistic model

211 (Equation 2) to obtain estimates for the EC<sub>50</sub> values based on total metal in the soil and total metal  
212 concentrations in the earthworms. Models were fitted in the form:

$$213 \quad y = y_{max}/(1+(c/EC_{50})^b) \quad (2)$$

214 Where  $y_{max}$  is the upper asymptote, EC<sub>50</sub> is the concentration (soil/body) resulting in a 50% effect on  
215 the measured endpoint (EC<sub>50</sub>) and  $b$  the slope parameter. Model fits to derive parameters with associated  
216 standard errors were completed using SigmaPlot. EC<sub>25</sub> and EC<sub>90</sub> values were also estimated from the  
217 dose response curves.

218

## 219 **Results**

### 220 *Test validation*

221 The earthworms in the sandy loam soil control produced more than the minimum 30  
222 juveniles/individuals ( $39 \pm 10$  juveniles; Mean  $\pm$  SD; n=6) thus validating the test procedure.<sup>24</sup> The  
223 earthworms in the control soil-sludge treatment both gained more weight,  $29 \pm 11\%$  weigh increase  
224 compared to a  $4.5 \pm 4.9\%$  weight loss in the soil control and produced 2.5 times more juveniles ( $97 \pm$   
225  $14.6$  juveniles, n=4) than the earthworms in the soil control over the test (Table 2). This improved  
226 performance of the sludge–exposed earthworms is likely to be related to the superior quality of food in  
227 the organic–rich sewage sludge compared to the soil control. Hence to identify adverse effects of the  
228 sludge treated with ENM or ioinic metal all comparisons were made to the control sludge treatment  
229 throughout the study and not the soil control.

230 For the as-synthesised Ag and Zn ENM and ionic metal exposures in the sandy loam soil, concentration–  
231 response relationships were obtained for all exposures. It was possible to calculate EC<sub>25</sub>, EC<sub>50</sub> and EC<sub>90</sub>  
232 values based on the total Ag or Zn concentration in the soil and the total Ag and Zn concentration in  
233 the earthworms in all cases (Table S1, Supporting Information).

234

### 235 *Soil metal concentrations and earthworm responses*

236 The metal concentrations in the ENM and ionic metal soil-sludge mixes are shown in Table 1. The Zn  
237 concentrations were close to the target value of 1400 mg/kg, being on average 114% and 97% of the  
238 target in the high-metal ionic and ENM treatments respectively. Recovery of Ag was also close to the  
239 intended Ag concentrations, being 111% and 94% of the target in the ionic and ENM mixtures  
240 respectively (Table 1).

241 Earthworm survival and reproduction were clearly decreased more in the high-metal ENM soil-sludge  
242 treatment compared to all other treatments (Figure 1, Table 2). Earthworm survival was reduced by  
243 25% and reproduction was significantly reduced by 90% compared to the control soil-sludge treatment  
244 (ANOVA:  $F = 110.25$ ,  $p < 0.001$ ) (Table 2). In comparison the ionic metal soil-sludge treatments and  
245 the low-metal ENM treatment reduced reproduction, although not significantly, by 25-30 % compared  
246 to the control soil-sludge (ANOVA:  $F = 2.55$ ,  $p = 0.12$ ) (Figure 1) and there was 100% survival (Table  
247 2). Earthworm weight change did not vary significantly across any of the soil-sludge treatments  
248 (ANOVA:  $F = 2.07$ ,  $p = 0.135$ ) (Table 2). The Zn concentrations in each of the soil-sludge treatments  
249 were above the  $EC_{25}$  and  $EC_{90}$  effect concentrations for the ionic metal and low metal ENM as-  
250 synthesised exposures, respectively (Figure 1a, Table S1). Only the  $EC_{90}$  value for the as-synthesised  
251 Zn ENM (1926 mg Zn/kg) was above the Zn concentration in the high metal soil-sludge treatment (1690  
252 mgZn/kg). In the case of Ag, all the soil-sludge treatments with the exception of the high metal ENM  
253 treatment had higher Ag soil concentrations than the  $EC_{25}$  or  $EC_{90}$  effect concentrations in the ionic  
254 metal as-synthesised concentration-response curves (Figure 1b). The Ag soil concentration high metal  
255 ENM treatment (94 mg Ag/kg) was most similar to the ionic metal as-synthesised  $EC_{90}$  value (74 mg  
256 Ag/kg and indeed fell along as-synthesised ionic metal DRC.

257

#### 258 *Total metal concentrations in the earthworms*

259 Earthworms exposed in the control soil-sludge had significantly higher total Ag concentrations ( $0.881$   
260  $\pm 0.129$   $\mu\text{g Ag/g}$ ), than those from the soil control ( $0.036 \pm 0.011$   $\mu\text{g Ag/g}$ ), although both had  
261 significantly lower total Ag concentrations than in all other treatments (Figure 2b, Table 2). Total Ag

262 concentrations in earthworms from the ENM and ionic metal soil-sludge treatments were only  
263 compared to those from the control soil-sludge. The total Zn concentrations in the earthworms across  
264 all the soil-sludge treatments ranged from  $86.9 \pm 26.4 \mu\text{g Zn/g}$  to  $122 \pm 11.8 \mu\text{g Zn/g}$  (Figure 2a, Table  
265 2). Exposure of earthworms to the ionic metal and ENM soil-sludge treatments did not result in  
266 significantly higher total Zn concentrations in earthworms compared to the control soil-sludge (Figure  
267 2a). Total Zn concentrations in earthworms from the soil-sludge treatments were all below effect  
268 concentrations ( $\text{EC}_{25}/\text{EC}_{90}$ ) shown in the concentration-response curves from the as-synthesised ionic  
269 metal and ENM Zn exposures (Figure 2a). There was a poor correlation between total Zn concentrations  
270 in earthworms and the observed effect on reproduction across the soil-sludge treatments ( $r^2=0.007$ ).  
271 This suggests that Zn exposure in all the mixtures was within the physiological tolerance range of the  
272 earthworms for Zn ( $100\text{-}200 \mu\text{g Zn/g}$ )<sup>31</sup> although soil concentrations were above what would usually  
273 be tolerated in as-synthesised Zn exposures.

274 Earthworms exposed in the high-metal ENM soil-sludge treatment had significantly higher total Ag  
275 concentrations than earthworms from all of the other treatments, with the exception of the low-metal  
276 ionic metal soil-sludge treatment (Figure 2b, Table 2). There was a strong relationship between the total  
277 Ag concentrations in earthworms from the soil-sludge treatments and the observed effects on  
278 reproduction ( $r^2=0.864$ ). Earthworm from the soil-sludge treatments had total Ag concentrations that  
279 were also less than the effect concentrations ( $\text{EC}_{25}/\text{EC}_{90}$ ) seen in the concentration-response curves from  
280 the as-synthesised ionic metal and ENM Ag exposures (Figure 2b, Table S1). However the earthworms  
281 from the high-metal ENM soil-sludge treatment accumulated significantly more Ag than those in other  
282 treatments ( $9 \text{ mg Ag/kg}$ ) which was most similar to the  $\text{EC}_{90}$  for total Ag concentrations in earthworms  
283 ( $10.6 \text{ mg Ag/kg}$ ) from the ionic metal as-synthesised exposure (Figure 2b).

284

#### 285 *Porewater metal concentrations*

286 Zn concentrations in the soil porewater were dependent on the total soil Zn concentrations; porewater  
287 in the high-metal treatments had greater Zn concentrations than in the low-metal treatments (Figure 3a).

288 The porewater Zn concentrations were significantly higher in the ionic metal soil-sludge treatments  
289 compared to the ENM soil-sludge treatments (ANOVA:  $F = 144.58$ ,  $p < 0.01$ ). Ultra-filtered porewater  
290 Zn concentrations did not differ significantly from the total porewater concentrations in any of the  
291 treatments (ANOVA:  $F = 0.22$ ,  $p > 0.05$ ) (Figure 3a). Soil porewater Ag concentrations were  
292 significantly higher in the high-metal ENM and the two ionic metal soil-sludge treatments than in the  
293 control soil-sludge and low-metal ENM treatments (ANOVA:  $F = 17.09$ ,  $p < 0.001$ ) (Figure 3b).  
294 Ultrafiltration significantly reduced the porewater Ag concentrations, for both ENM and ionic metal  
295 sludge treatments and Ag concentrations in the ultra-filtered porewaters did not differ across the soil-  
296 sludge treatments ( $F = 1.35$ ;  $p > 0.05$ ) (Figure 3b).

297

## 298 **Discussion**

299 The application of sewage sludge to soils represents a realistic pathway for nanomaterials, or their  
300 transformation products, to enter terrestrial ecosystems. In order to understand, and ultimately regulate,  
301 the use and input of nanomaterials into the environment it is necessary to assess the risks resulting from  
302 land application of sludge produced from WWTP receiving inputs of nanomaterials, in scenarios that  
303 are realistic and representative of the final exposure for soil organisms. Ag and Zn nanomaterials were  
304 transformed by the wastewater treatment process into forms that were more thermodynamically stable  
305 under WWTP conditions, becoming largely or almost completely sulphidised or phosphatised.<sup>1, 2</sup>  
306 Crucially, ionic forms of Zn and Ag were also transformed, to a similar extent, producing essentially  
307 identical solid-phase speciation (coordination environment and oxidation state) in both the ENM-  
308 and ionic metal-treated sludges.<sup>1, 16</sup> There is evidence that Ag nanomaterial sulphidation reduces  
309 toxicity in controlled laboratory studies<sup>32, 33</sup> with similar passivation of Zn toxicity expected.<sup>34</sup>  
310 Durenkamp et al 2016 found that very little metal was leached during the six month aging process (in  
311 total over six month - 5 ug Zn/g and 2 ug Ag/g) and that there was no difference in speciation between  
312 the ENM and ionic metal forms of Ag and Zn.<sup>1, 23</sup> However they did find that the inorganic N form did  
313 change; at the beginning of the aging process (i.e. fresh sludge) the majority was present in the form of  
314  $\text{NH}_4^+$  (a toxic form for earthworms) whereas  $\text{NO}_3^-$  dominated (up to 90%) at the end, but was the same

315 in the ionic metal and ENM treatments.<sup>23</sup> Earthworms in all soil-sludge treatments (control, ionic metal  
316 and ENM) gained similar weight over the duration of the exposure. However in this study, clear and  
317 significant differences were observed between the effects on earthworm reproduction when exposed in  
318 soils mixed with sludges derived from WWTP lines treated with either ENM or ionic metal forms.  
319 Although all earthworms gained weight in the three sludge treatments (control, ionic metal and ENM)  
320 over the duration of the exposure the ENM treated sludge depressed earthworm reproduction four times  
321 more than the same sludge treated with ionic metals. These results suggest the hypothesis that sludges  
322 showing similar solid-phase speciation of Zn and Ag should result in similar toxicity, regardless of the  
323 form of the spike, is incorrect. A similar conclusion was reached by Judy et al 2015 for effects on the  
324 legume *Medicago truncatula*.<sup>16</sup> Judy et al 2015 used the same aged soil-sludge mixture as in this study  
325 and showed that the solid-phase speciation did not differ between ENM and ionic/bulk metal treatments.  
326 The solid-phase speciation was determined using X-ray absorption spectroscopy (XAS), which  
327 measures the oxidation state and local coordination environment of metals. It is possible that although  
328 the metals had similar coordination environments, the mineral particles may have had different sizes,  
329 morphologies, crystal structure or other nano-scale attributes that differed between treatments that are  
330 not measured by XAS which is an Angstrom-scale characterisation.<sup>16</sup> However given the greater  
331 toxicity of the ENM treatment, the U.S. regulations for ionic metals in sludge may not protect soil biota  
332 in the case of sludge derived from WWTP primarily receiving inputs of Zn and/or Ag in the form of  
333 ENMs.

334 The sludges contained Zn, Ag and Ti added to the WWTP inflow in either the ENM or ionic metal form  
335 and previous work had shown that similar solid-phase speciation of Zn and Ag was found in the ENM  
336 and the ionic metal sludges.<sup>1, 16</sup> Consequently the ionic metal effect data (EC<sub>50</sub>) for Zn and Ag were  
337 used to model the responses in both the ionic metal and ENM soil-sludge treatments. Thus, the toxic  
338 effects observed in the ENM soil-sludge treatments were compared to predicted effects (EC<sub>25</sub> or EC<sub>90</sub>)  
339 calculated from both the ionic metal and the ENM as-synthesised effect data. The Ti concentrations in  
340 the soil-sludge treatments were between 1180 and 2467 mg Ti/kg<sup>16</sup> about 10 times lower than exposure  
341 concentration (10000 mg Ti/kg) where only slight effects of TiO<sub>2</sub> ENMs were found on reproduction

342 reported in toxicity studies.<sup>11</sup> Hence in this study we assume that effects due to Ti were negligible and  
343 so were not included in the assessment of toxicity. The total Zn and Ag metal concentrations in the  
344 ENM and ionic metal sludge treatments were effectively the same. The total Zn and Ag soil  
345 concentrations were above the observed effect concentrations (EC<sub>25</sub> or EC<sub>90</sub>) in the as-synthesised ionic  
346 metals soil exposures, particularly for Zn, but the predicted toxicity was not realised in the low-metal  
347 ENM or the ionic metal soil-sludge treatments. This could be expected when the exposure medium is  
348 considered; the sludge treatments had much higher organic matter content than the sandy loamy soil  
349 alone in which the as-synthesised metal exposures were conducted. It is widely established that ionic  
350 metals can show lower toxicity in soil with high organic matter.<sup>35, 36</sup> Another consideration is the aging  
351 of metals in soil which is known to greatly influence their toxicity to organisms in soils. In the case of  
352 ionic metals the aging in soils has been well described typically showing metals to become less toxic to  
353 organisms as they become more associated and bound to the solid phase in soils.<sup>37, 38 39</sup> Hence, a  
354 leaching-aging factor of 3 has been recommended to be applied to laboratory data in order to account  
355 for aging in the field and leaching of salts both of which will lower Zn toxicity.<sup>37</sup> In this study the Zn  
356 concentration in the ENM soil-sludge treatment which caused a 90% effect (1690 mg Zn/kg) was about  
357 three times greater than the ionic metal Zn EC<sub>90</sub> (605 mg Zn/kg). This means the safety factor applied  
358 to ionic metal response data may not be fully protective for Zn and certainly not Ag in cases where the  
359 metal is in the form of an aged or transformed ENM. Indeed Diez et al. 2015 showed that Ag ENM  
360 toxicity to earthworms increased over a one year time period (EC<sub>50</sub> reduced from 1420 to 34mg Ag/kg)  
361 compared to a decrease in Ag ionic metal toxicity (EC<sub>50</sub> increased from 49 to 104 mg Ag/kg)<sup>8</sup> which  
362 emphasizes the limitations of short-term exposures to as-synthesised ENMs in predicting ultimate  
363 toxicity. Overall in this study the ENM as-synthesised exposures showed low ENM toxicity compared  
364 to ionic metals and did not predict the level of effect observed in the high-metal ENM treatment better  
365 than ionic metal exposures.

366 Porewater measurements of metal in toxicity exposures may be used to explain variability in the  
367 solubility and thus the chemical reactivity of the metals across treatments. For ionic metals, increased  
368 solubility suggests greater bioavailability, though caution needs to be applied when considering



369 porewater metal concentrations across soil types, due to the additional influence of variables such as  
370 the porewater pH and organic matter on metal availability.<sup>40, 41</sup> However, the low variability of pH and  
371 dissolved organic carbon across the different sludge treatments suggests that the variability in the  
372 porewater metal concentrations could be usefully used as a surrogate for metal reactivity and hence the  
373 bioavailability of ionic forms. Accordingly, if the observed uptake and toxicity were due to uptake of  
374 ionic Ag and/or Zn, it would be expected that the porewater concentrations of at least one metal would  
375 be higher in the ENM treatments compared to the ionic treatments. A small number of studies that have  
376 investigated the aging processes of ENMs in soils have shown the progression of metal toxicity to be  
377 different from that of ionic metals and that over time ENMs will undergo dissolution into the porewater  
378 which has been linked with greater toxicity.<sup>6, 8, 42</sup> However, porewater concentrations of both Zn and  
379 Ag were consistently higher in the ionic treatments. Therefore, conventional patterns of ionic metal  
380 bioavailability cannot explain the observed effects and accumulation.

381 Organism body concentrations, in principle, provide the closest direct link to exposure since they  
382 integrate bioavailability and effects. Ag concentrations in the earthworm tissues varied significantly  
383 across the treatments; earthworms exposed to the ENM sludge accumulated more Ag than the ionic  
384 metal treatments. In the high-metal ENM treatment there was also significantly greater accumulation  
385 of Zn in the ENM treatment than the ionic treatment. A similar pattern was observed by Judy et al  
386 2015., in *M. truncatula* where shoot concentrations of all three metals were higher in the ENM treatment  
387 than ionic/bulk, although only statistically significant for Zn.<sup>16</sup> Total Zn concentrations in the  
388 earthworms were within the physiological limits for earthworm Zn regulation,<sup>31</sup> and showed no  
389 relationship to total soil concentrations or to effects. However, the possibility of effects due to Zn cannot  
390 be precluded, as the earthworms may become stressed as a result of the energy requirements to maintain  
391 a physiologically stable body concentrations in the face of a Zn stress.<sup>43</sup> Total Ag concentrations in  
392 earthworms did show a strong relationship with effect, across both the ENM and ionic metal treatments.  
393 The effects of the soil-sludge treatments were more similar to the ionic metal as-synthesised response  
394 curve compared to the ENM as-synthesised response. However as the effects in the high metal ENM  
395 treatment were observed at slightly lower total Ag concentrations in earthworms than those expected

396 from the as-synthesised ionic metal or ENM exposures it would suggest that either both Ag and Zn  
397 contribute to the effects or that the transformations of the metals in the WWTP system increase their  
398 toxic potency relative to the as-synthesised forms. For example, it is possible Ag<sub>2</sub>S particles are being  
399 taken up and entering different locations in cells and then undergoing dissolution locally. There is  
400 evidence for the apparent changing toxicity of Ag ENMs in soils to earthworms; Diez et al. 2015 found  
401 that the EC<sub>50</sub> (as total Ag concentration in the earthworms) for Ag initially spiked into a soil in the ENM  
402 form decreased from 64 µg Ag/g total Ag concentration in earthworms on initial toxicity testing to 7  
403 µg Ag/g after incubation of Ag in the soil for a year.<sup>8</sup> This trend was interpreted as being due to  
404 differential uptake of Ag ENMs and ionic Ag, coupled with gradual dissolution of Ag ENMs to ionic  
405 Ag over the incubation period. Thus, it is not possible to draw definite conclusions regarding the relative  
406 role of Zn and Ag in exerting toxic effects in the sludge treatments, since their toxic potencies may be  
407 dependent upon their chemical speciation. Furthermore, the differences in toxic effect observed across  
408 the ionic and ENM treatments suggest that the toxic potencies of the forms in the final sludges have  
409 been influenced by the nature of the starting metal form (i.e. ionic metal or ENM), despite the  
410 observation that the bulk phase speciation was similar in both sludge treatments. Indeed given that  
411 toxicity was unexpectedly highest in the high-metal ENM treatments, more research is required into the  
412 physicochemical form and distribution of the metals in the sludges to draw more definitive links with  
413 the observed toxicity.

414 This study was designed to represent the worst case scenario for ENM contamination associated with  
415 sludge application to soils. At present the maximum allowable concentrations are only set for Zn and  
416 these are to provide protection against the toxicity of metal salt forms. Although the metal salt exposures  
417 over-predicted toxicity for most of the sludge treatments it more closely predicted the ENM toxicity  
418 following transformation and aging. This study clearly shows as-synthesised ENM exposure studies do  
419 not accurately predict the toxicity of ENMs in environmentally realistic scenarios (aged and  
420 transformed after WWTP). Studies which show ENMs to be more toxic than the ionic metal are rare  
421 but there is a growing body of evidence that aging ENMs<sup>8</sup> and/or exposure in more environmentally  
422 realistic forms such as sludge treated with ENMs<sup>16-18</sup> can result in greater toxicity than when treated

423 with the ionic/bulk metal forms. Although previous studies have demonstrated that sulfidation and  
 424 phosphatation of Ag and ZnO nanomaterials greatly reduces their toxicity,<sup>14, 34 44</sup> ENMs can be more  
 425 toxic than ionic metals after undergoing similar transformations. When both materials were aged the  
 426 ENM metal forms were more toxic than the metal salt form suggesting that current Zn limits may not  
 427 protect soil biota if the majority of metals enter the WWTPs from which these sludges are produced in  
 428 the ENM form.

429

430

### 431 **Tables**

432 Table 1: Total soil Ag, Zn and Ti concentrations, pH values and dissolved organic carbon concentration  
 433 in porewaters and the water holding capacity for each of the soil-sludge mixtures and the sandy loam  
 434 control soil (Woburn).<sup>†</sup>

Treatment	Total Ag	Total Zn	Total Ti	Porewater	Porewater	Water
	(mg Ag/kg dry mass)	(mg Zn/kg dry mass)	(mg Ti/kg dry mass) <sup>‡</sup>	pH	dissolved organic carbon (µg/ml)	holding capacity (ml/100 g)
Control	2.84 ± 0.35	321.5 ± 7.19	1180 ± 32.7	7.10 ± 0.03	283 ± 24.7	94
High metal ionic metal	111 ± 7.75	1600 ± 52.3	2365 ± 61.8	7.02 ± 0.04	304 ± 11.5	92
High metal ENM <sup>a</sup>	94.3 ± 4.77	1360 ± 64.8	2467 ± 181	7.06 ± 0.06	299 ± 6.83	92
Low metal ionic metal	71.2 ± 2.21	985 ± 34	n.d.	7.30 ± 0.06	272 ± 1.5	93
Low metal ENM	51.6 ± 2.42	853 ± 35.7	n.d.	7.09 ± 0.08	314 ± 15	93
Soil control	0.09 ± 0.02	39.2 ± 0.71	n.d.	7.31 ± 0.08	119 ± 11.1	32

435 <sup>†</sup>Each value represents mean ± one standard deviation <sup>‡</sup>Data from Judy et al 2015.<sup>16</sup> n.d. means that  
 436 the measurements were not determined. <sup>a</sup>ENM = engineered nanomaterials

437

438

439

440

441

442 Table 2: The survival, percentage weight change, reproduction and total Zn and Ag concentrations in  
443 earthworms for the soil-sludge treatments and the sandy loam control soil (Woburn).<sup>†</sup>

Treatment	% Survival	% Weight change	Reproduction (Juveniles per worm per week)	Total Zn concentration in earthworms (µg Zn/g)	Total Ag concentration in earthworms (µg Ag/g)
Control	97.5 ± 5 <sup>a</sup>	27.9 ± 11.5 <sup>a</sup>	2.45 ± 0.318 <sup>a</sup>	116 ± 14.9 <sup>a</sup>	0.881 ± 0.129 <sup>b</sup>
High metal ionic metal	100 <sup>a</sup>	58.1 ± 5.02 <sup>a</sup>	1.90 ± 0.617 <sup>a</sup>	86.9 ± 26.4 <sup>b</sup>	3.28 ± 1.86 <sup>c</sup>
High metal ENM <sup>a</sup>	75 ± 2.65 <sup>a</sup>	41.1 ± 13.1 <sup>a</sup>	0.236 ± 0.277 <sup>b</sup>	113 ± 18.3 <sup>a</sup>	8.99 ± 2.75 <sup>d</sup>
Low metal ionic metal	97.5 ± 5 <sup>a</sup>	48.1 ± 29.4 <sup>a</sup>	1.71 ± 0.367 <sup>a</sup>	122 ± 11.8 <sup>a</sup>	5.16 ± 0.925 <sup>cd</sup>
Low metal ENM	100 <sup>a</sup>	57.4 ± 18.1 <sup>a</sup>	1.688 ± 0.483 <sup>a</sup>	118 ± 17.5 <sup>a</sup>	4.62 ± 1.55 <sup>bc</sup>
Soil control	100 <sup>a</sup>	-4.46 ± 4.92 <sup>b</sup>	0.969 ± 0.267 <sup>a</sup>	90.3 ± 7.42 <sup>b</sup>	0.036 ± 0.011 <sup>a</sup>

444 <sup>†</sup>Each value represents mean ± one standard deviation; Survival, weight change and reproduction had  
445 n=4, Zn, Ag concentration: n=12. Means with the same superscript letters are not significantly  
446 different (*p*>0.05). <sup>a</sup>ENM = engineered nanomaterials

447

448

449

450

451

452 **Figure captions**

453 **Figure 1:** The normalised reproduction response (normalised to reproduction in the control soil-sludge)  
454 with increasing soil (a) Zn or (b) Ag concentrations. The data points are response data from the five  
455 soil-sludge treatments. Solid line = ENM concentration-response curve, dashed line = ionic metal  
456 concentration-response curve for Zn or Ag in sandy loam control soil. The grey shaded areas around  
457 the response curves represent the 95% confidence intervals around the curves. The black star represents

458 the EU limit (86 / 278 /EEC) (max. 300 mg/kg) and the white star the US limit<sup>22</sup> (1400 mg/kg) for Zn  
459 in soil from sludge application to land.

460

461 **Figure 2:** The normalised reproduction response (normalised to reproduction in the control soil-sludge)  
462 with increasing total (a) Zn or (b) Ag concentrations in earthworms. The data points are response data  
463 from the five soil-sludge mixtures. Error bars are the standard deviations of total metal concentrations  
464 in earthworms from three replicate earthworms in each treatment replicate. The model fits are from the  
465 as-synthesised ENM and bulk metal Zn and Ag exposure data. Solid line = ENM concentration-  
466 response curve, dashed line = ionic metal concentration-response curve for Zn or Ag sandy loam control  
467 soil. The grey shaded areas around the response curves represent the 95% confidence intervals around  
468 the curves. Vertical grey lines = limits for Zn regulation by earthworms.<sup>31</sup>

469

470 **Figure 3:** The total and ultra-filtered porewater concentrations of (a) Zn and (b) Ag in the soil-sludge  
471 mixtures extracted from the soils at the end of the toxicity exposure. Different letters denote significant  
472 differences between the total and ultra-filtered metal concentrations in the soil-sludge treatments.  
473 Asterisks next to the letters indicate where the total metal concentration in the porewater was  
474 significantly different from the ultra-filtered porewater concentration for the same soil-sludge treatment.  
475 Error bars are standard deviations.

476

477

478

479

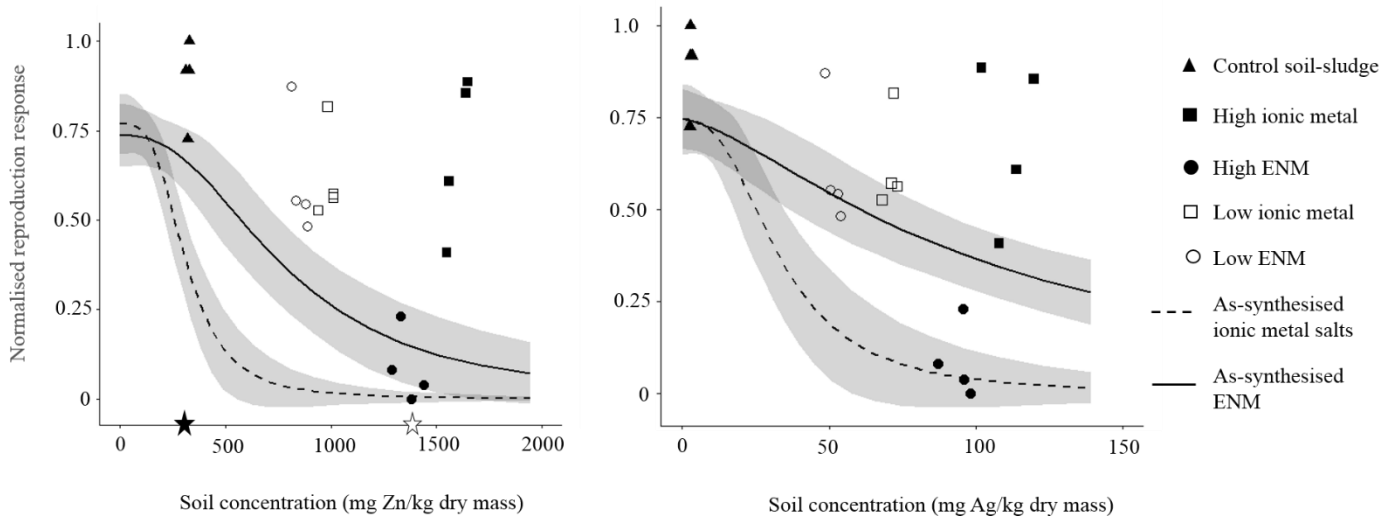
480

481

482

483

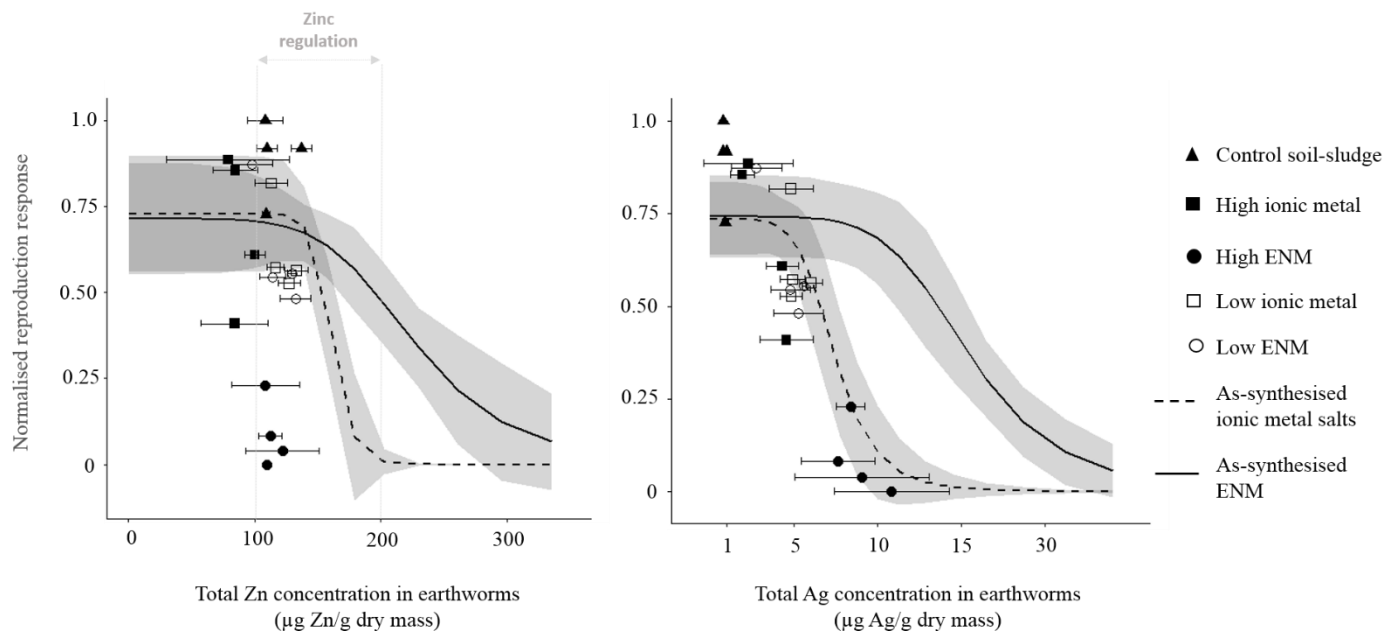
484 **Figures**



485

486 **Figure 1**

487

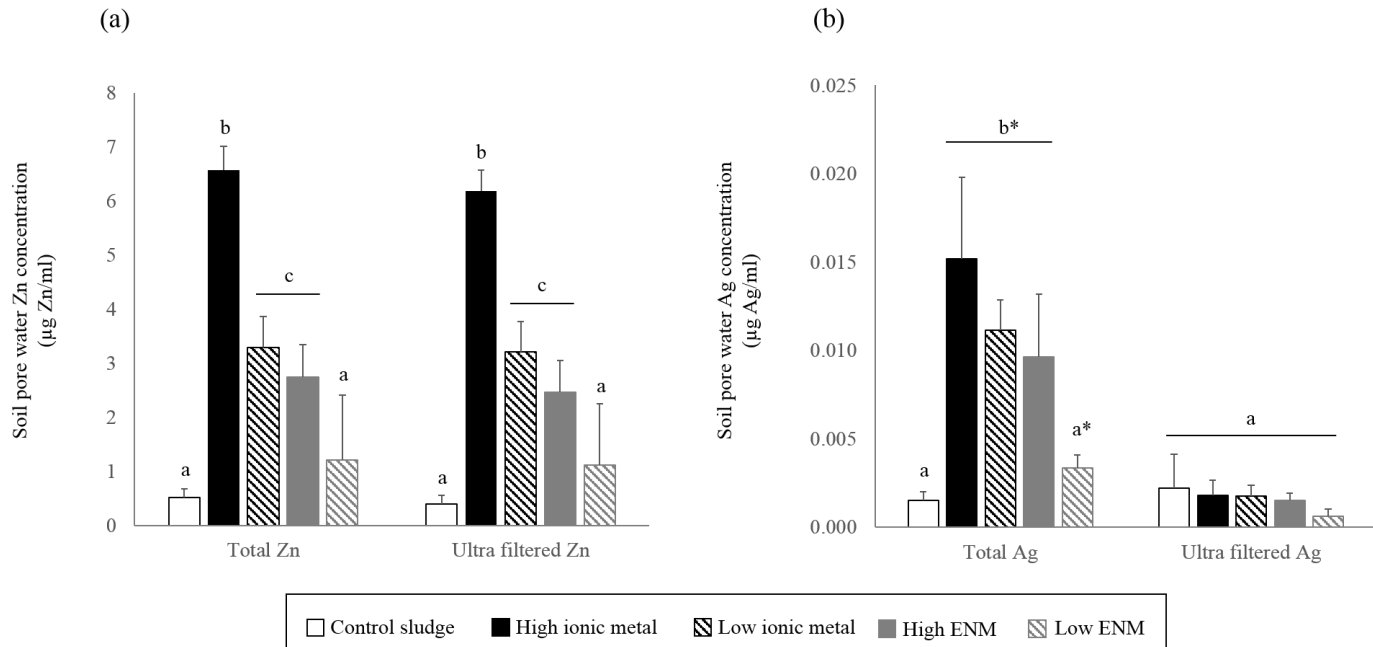


488

489

490 **Figure 2**

491



492

493 **Figure 3**

494

495 **Acknowledgements**

496 The authors wish to thank S. Marinakos at Duke University, Department of Civil and Environmental  
497 Engineering for her work in preparing particles. We also wish to acknowledge A. Romero and A.A.  
498 Horton for their assistance in the laboratory toxicity test work and A. Robinson for his assistance in  
499 creating the figures for the text. This work was funded by the Natural Environment Research Council  
500 Transatlantic Initiative for Nanotechnology and the Environment (TINE) grant NE/H013679/1, and  
501 NanoFASE EU Horizon 2020 research and innovation programme under grant agreement No.646002.  
502 M.D. was also supported by the UK Biotechnology and Biological Sciences Research Council  
503 (BBS/E/C/00005094). M.M. was supported by the Marie-Curie FP7-PEOPLE-2011-IEF,  
504 Micronanotox (PIEF-GA-2011-303140).

505

506  
507  
508  
509  
510  
511  
512  
513  
514  
515  
516  
517  
518  
519  
520  
521  
522  
523  
524  
525  
526  
527  
528  
529  
530  
531  
532  
533  
534  
535  
536  
537  
538  
539  
540  
541  
542  
543  
544  
545  
546  
547  
548  
549  
550  
551  
552  
553  
554

## References

1. Ma, R.; Levard, C.; Judy, J. D.; Unrine, J. M.; Durenkamp, M.; Martin, B.; Jefferson, B.; Lowry, G. V., Fate of zinc oxide and silver nanoparticles in a pilot wastewater treatment plant and in processed biosolids. *Environ. Sci. Technol.* **2014**, *48* (1), 104-112.
2. Kim, B.; Park, C.-S.; Murayama, M.; Hochella, M. F., Discovery and characterization of silver sulfide nanoparticles in final sewage sludge products. *Environ. Sci. Technol.* **2010**, *44* (19), 7509-7514.
3. Kaegi, R.; Voegelin, A.; Sinnet, B.; Zuleeg, S.; Hagedorfer, H.; Burkhardt, M.; Siegrist, H., Behavior of metallic silver nanoparticles in a pilot wastewater treatment plant. *Environ. Sci. Technol.* **2011**, *45* (9), 3902-3908.
4. Kelessidis, A.; Stasinakis, A. S., Comparative study of the methods used for treatment and final disposal of sewage sludge in European countries. *Waste Manage.* **2012**, *32* (6), 1186-1195.
5. Tourinho, P. S.; van Gestel, C. A. M.; Lofts, S.; Svendsen, C.; Soares, A. M. V. M.; Loureiro, S., Metal-based nanoparticles in soil: Fate, behavior, and effects on soil invertebrates. *Environ. Toxicol. Chem.* **2012**, *31* (8), 1679-1692.
6. Kool, P. L.; Ortiz, M. D.; van Gestel, C. A. M., Chronic toxicity of ZnO nanoparticles, non-nano ZnO and ZnCl<sub>2</sub> to *Folsomia candida* (Collembola) in relation to bioavailability in soil. *Environ. Pollut.* **2011**, *159* (10), 2713-2719.
7. Heggelund, L. R.; Diez-Ortiz, M.; Lofts, S.; Lahive, E.; Jurkschat, K.; Wojnarowicz, J.; Cedergreen, N.; Spurgeon, D.; Svendsen, C., Soil pH effects on the comparative toxicity of dissolved zinc, non-nano and nano ZnO to the earthworm *Eisenia fetida*. *Nanotoxicology* **2013**, *8* (5), 559-572.
8. Diez-Ortiz, M.; Lahive, E.; George, S.; Ter Schure, A.; Van Gestel, C. A. M.; Jurkschat, K.; Svendsen, C.; Spurgeon, D. J., Short-term soil bioassays may not reveal the full toxicity potential for nanomaterials; bioavailability and toxicity of silver ions (AgNO<sub>3</sub>) and silver nanoparticles to earthworm *Eisenia fetida* in long-term aged soils. *Environ. Pollut.* **2015**, *203*, 191-198.
9. Manzo, S.; Rocco, A.; Carotenuto, R.; De Luca Picione, F.; Miglietta, M.; Rametta, G.; Di Francia, G., Investigation of ZnO nanoparticles' ecotoxicological effects towards different soil organisms. *Environ. Sci. and Pollut. Res.* **2011**, *18* (5), 756-763.
10. Wang, H.; Wick, R. L.; Xing, B., Toxicity of nanoparticulate and bulk ZnO, Al<sub>2</sub>O<sub>3</sub> and TiO<sub>2</sub> to the nematode *Caenorhabditis elegans*. *Environ. Pollut.* **2009**, *157* (4), 1171-1177.
11. McShane, H.; Sarrazin, M.; Whalen, J. K.; Hendershot, W. H.; Sunahara, G. I., Reproductive and behavioral responses of earthworms exposed to nano-sized titanium dioxide in soil. *Environ. Toxicol. Chem.* **2012**, *31* (1), 184-193.
12. Simonin, M.; Guyonnet, J. P.; Martins, J. M. F.; Ginot, M.; Richaume, A., Influence of soil properties on the toxicity of TiO<sub>2</sub> nanoparticles on carbon mineralization and bacterial abundance. *J. Hazard. Mater.* **2015**, *283*, 529-535.



- 555 13. Lombi, E.; Donner, E.; Taheri, S.; Tavakkoli, E.; Jämting, Å. K.; McClure, S.; Naidu, R.;  
556 Miller, B. W.; Scheckel, K. G.; Vasilev, K., Transformation of four silver/silver chloride nanoparticles  
557 during anaerobic treatment of wastewater and post-processing of sewage sludge. *Environ. Pollut.* **2013**,  
558 *176*, 193-197.
- 559 14. Levard, C.; Hotze, E. M.; Lowry, G. V.; Brown, G. E., Environmental transformations of silver  
560 nanoparticles: Impact on stability and toxicity. *Environ. Sci. Technol.* **2012**, *46* (13), 6900-6914.  
561
- 562 15. Pradas del Real, A. E.; Castillo-Michel, H.; Kaegi, R.; Sinnet, B.; Magnin, V.; Findling, N.;  
563 Villanova, J.; Carrière, M.; Santaella, C.; Fernández-Martínez, A.; Levard, C.; Sarret, G., Fate of Ag-  
564 NPs in sewage sludge after application on agricultural soils. *Environ. Sci. Technol.* **2016**, *50* (4), 1759-  
565 1768.  
566
- 567 16. Judy, J. D.; McNear, D. H.; Chen, C.; Lewis, R. W.; Tsyusko, O. V.; Bertsch, P. M.; Rao, W.;  
568 Stegemeier, J.; Lowry, G. V.; McGrath, S. P.; Durenkamp, M.; Unrine, J. M., Nanomaterials in  
569 biosolids inhibit nodulation, shift microbial community composition, and result in increased metal  
570 uptake relative to bulk/dissolved metals. *Environ. Sci. Technol.* **2015**, *49* (14), 8751-8758.  
571
- 572 17. Chen, C.; Unrine, J. M.; Judy, J. D.; Lewis, R. W.; Guo, J.; McNear, D. H.; Tsyusko, O. V.,  
573 Toxicogenomic responses of the model legume *Medicago truncatula* to aged biosolids containing a  
574 mixture of nanomaterials (TiO<sub>2</sub>, Ag, and ZnO) from a pilot wastewater treatment plant. *Environ. Sci.*  
575 *Technol.* **2015**, *49* (14), 8759-8768.  
576
- 577 18. Colman, B. P.; Arnaout, C. L.; Anciaux, S.; Gunsch, C. K.; Hochella, M. F., Jr.; Kim, B.;  
578 Lowry, G. V.; McGill, B. M.; Reinsch, B. C.; Richardson, C. J.; Unrine, J. M.; Wright, J. P.; Yin, L.;  
579 Bernhardt, E. S., Low concentrations of silver nanoparticles in biosolids cause adverse ecosystem  
580 responses under realistic field scenario. *PLoS ONE* **2013**, *8* (2), e57189.  
581
- 582 19. Fernández, M. D.; Alonso-Blázquez, M. N.; García-Gómez, C.; Babin, M., Evaluation of zinc  
583 oxide nanoparticle toxicity in sludge products applied to agricultural soil using multispecies soil  
584 systems. *Sci. Total Environ.* **2014**, 497-498, 688-696.  
585
- 586 20. García-Gómez, C.; Fernández, M.; Babin, M., Ecotoxicological evaluation of sewage sludge  
587 contaminated with zinc oxide nanoparticles. *Arch. of Environ. Con. Tox. I* **2014**, *67* (4), 494-506.  
588
- 589 21. Cheng, Y.; Yin, L.; Lin, S.; Wiesner, M.; Bernhardt, E.; Liu, J., Toxicity reduction of polymer-  
590 stabilized silver nanoparticles by sunlight. *J. Phys. Chem. C* **2011**, *115* (11), 4425-4432.  
591
- 592 22. EPA, A Guide to the Biosolids Risk Assessments for the EPA Part 503 Rule. In Agency, E. P.,  
593 Ed. Washington DC, 1995.  
594
- 595 23. Durenkamp, M.; Pawlett, M.; Ritz, K.; Harris, J. A.; Neal, A. L.; McGrath, S. P., Nanoparticles  
596 within WWTP sludges have minimal impact on leachate quality and soil microbial community structure  
597 and function. *Environ. Pollut.* **2016**, *211*, 399-405.  
598
- 599 24. OECD, *Test No. 222: Earthworm Reproduction Test (Eisenia fetida/Eisenia andrei)*. OECD  
600 Publishing. **2004**  
601
- 602 25. Sinha, R. K.; Herat, S.; Bharambe, G.; Brahmabhatt, A., Vermistabilization of sewage sludge  
603 (biosolids) by earthworms: converting a potential biohazard destined for landfill disposal into a  
604 pathogen-free, nutritive and safe biofertilizer for farms. *Waste Manage Res* **2010**, *28* (10), 872-881.  
605
- 606 26. Vijver, M. G.; Vink, J. P. M.; Miermans, C. J. H.; van Gestel, C. A. M., Oral sealing using glue:  
607 a new method to distinguish between intestinal and dermal uptake of metals in earthworms. *Soil Biol*  
608 *Biochem* **2003**, *35* (1), 125-132.

- 609 27. Sauv , S.; Hendershot, W.; Allen, H. E., Solid-solution partitioning of metals in contaminated  
610 soils: Dependence on pH, total metal burden, and organic matter. *Environ. Sci. Technol.* **2000**, *34* (7),  
611 1125-1131.  
612
- 613 28. Thakali, S.; Allen, H. E.; Di Toro, D. M.; Ponizovsky, A. A.; Rooney, C. P.; Zhao, F.-J.;  
614 McGrath, S. P.; Criel, P.; Van Eeckhout, H.; Janssen, C. R.; Oorts, K.; Smolders, E., Terrestrial biotic  
615 ligand model. 2. Application to Ni and Cu toxicities to plants, invertebrates, and microbes in soil.  
616 *Environ. Sci. Technol.* **2006**, *40* (22), 7094-7100.  
617
- 618 29. Whitley, A. R.; Levard, C.; Oostveen, E.; Bertsch, P. M.; Matocha, C. J.; Kammer, F. v. d.;  
619 Unrine, J. M., Behavior of Ag nanoparticles in soil: Effects of particle surface coating, aging and sewage  
620 sludge amendment. *Environ. Pollut.* **2013**, *182*, 141-149.  
621
- 622 30. Cornelis, G.; Kirby, J. K.; Beak, D.; Chittleborough, D.; McLaughlin, M. J., A method for  
623 determination of retention of silver and cerium oxide manufactured nanoparticles in soils. *Environ.*  
624 *Chem.* **2010**, *7*, (3), 298-308.  
625
- 626 31. Lock, K.; Janssen, C. R., Zinc and cadmium body burdens in terrestrial oligochaetes: Use and  
627 significance in environmental risk assessment. *Environ. Toxicol. Chem.* **2001**, *20*, (9), 2067-2072.  
628
- 629 32. Reinsch, B. C.; Levard, C.; Li, Z.; Ma, R.; Wise, A.; Gregory, K. B.; Brown, G. E.; Lowry, G.  
630 V., Sulfidation of silver nanoparticles decreases *Escherichia coli* growth inhibition *Environ. Sci.*  
631 *Technol.* **2012**, *46*, (13), 6992-7000.  
632
- 633 33. Doolette, C. L.; McLaughlin, M. J.; Kirby, J. K.; Navarro, D. A., Bioavailability of silver and  
634 silver sulfide nanoparticles to lettuce (*Lactuca sativa*): Effect of agricultural amendments on plant  
635 uptake. *J. Hazard. Mater.* **2015**, *300*, 788-795.  
636
- 637 34. Rathnayake, S. Transformations, bioavailability and toxicity of manufactured ZnO  
638 nanomaterials in wastewater. University of Kentucky, 2013.  
639
- 640 35. Spurgeon, D. J.; Hopkin, S. P., Effects of variations of the organic matter content and pH of  
641 soils on the availability and toxicity of zinc to the earthworm *Eisenia fetida*. *Pedobiologia* **1996**, *40*,  
642 80-96.  
643
- 644 36. Crommentuijn, T.; Doornekamp, A.; Van Gestel, C. A. M., Bioavailability and ecological  
645 effects of cadmium on *Folsomia candida* (Willem) in an artificial soil substrate as influenced by pH  
646 and organic matter. *Appl. Soil Ecol.* **1997**, *5* (3), 261-271.  
647
- 648 37. Smolders, E.; Oorts, K.; Van Sprang, P.; Schoeters, I.; Janssen, C. R.; McGrath, S. P.;  
649 McLaughlin, M. J., Toxicity of trace metals in soil as affected by soil type and aging after  
650 contamination: Using calibrated bioavailability models to set ecological soil standards. *Environ.*  
651 *Toxicol. Chem.* **2009**, *28* (8), 1633-1642.  
652
- 653 38. Smit, C. E.; Van Gestel, C. A. M., Effects of soil type, prepercolation, and ageing on  
654 bioaccumulation and toxicity of zinc for the springtail *Folsomia candida*. *Environ. Toxicol. Chem.*  
655 **1998**, *17* (6), 1132-1141.  
656
- 657 39. Lock, K.; Janssen, C. R., Influence of ageing on zinc bioavailability in soils. *Environ. Pollut.*  
658 **2003**, *126* (3), 371-374.  
659
- 660 40. Lofts, S.; Spurgeon, D. J.; Svendsen, C.; Tipping, E., Deriving soil critical limits for Cu, Zn,  
661 Cd, and Pb: A method based on free ion concentrations. *Environ. Sci. Technol.* **2004**, *38* (13), 3623-  
662 3631.  
663

- 664 41. Spurgeon, D. J.; Lofts, S.; Hankard, P. K.; Toal, M.; McLellan, D.; Fishwick, S.; Svendsen, C.,  
665 Effect of pH on metal speciation and resulting metal uptake and toxicity for earthworms. *Environ.*  
666 *Toxicol. Chem.* **2006**, 25 (3), 788-796.  
667
- 668 42. Coutris, C.; Joner, E. J.; Oughton, D. H., Aging and soil organic matter content affect the fate  
669 of silver nanoparticles in soil. *Sci.Total Environ.* **2012**, 420, 327-333.  
670
- 671 43. van Gestel, C. A. M.; Dirven-van Breemen, E. M.; Baerselman, R., Proceedings of the 2nd  
672 European Conference on Ecotoxicology Accumulation and elimination of cadmium, chromium and  
673 zinc and effects on growth and reproduction in *Eisenia andrei* (Oligochaeta, Annelida). *Sci.Total*  
674 *Environ.* **1993**, 134, 585-597.  
675
- 676 44. Starnes, D. L.; Unrine, J. M.; Starnes, C. P.; Collin, B. E.; Oostveen, E. K.; Ma, R.; Lowry, G.  
677 V.; Bertsch, P. M.; Tsyusko, O. V., Impact of sulfidation on the bioavailability and toxicity of silver  
678 nanoparticles to *Caenorhabditis elegans*. *Environ. Pollut.* **2015**, 196, 239-246.

679

680

681

682

683