**A comparative analysis of canyon and non-canyon populations of the deep-sea scavenging amphipod *Paralicella caperesca***

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*Existing population studies of deep-sea amphipods have focused species that inhabit deep-sea vent or trench environments but few cosmopolitan species have been studied. Here we provide new insight into the life history and population ecology of the pan-oceanic scavenging amphipod* Paralicella caperesca *and discuss the influence of nutrient-rich submarine canyon environments on the growth and reproduction of this species.*

*Data were collected through the dissection and measurement of 2997* P. caperesca *from 14 samples taken from abyssal plains, continental slopes, and submarine canyons in the North-East Atlantic. Sexual dimorphism was less pronounced than observed for other scavenging amphipod species but females were significantly larger and had shorter antennae than males. The size of oostegites in female* P. caperesca *varied considerably within size classes, ovaries contained a relatively large number of oocytes, and no empty ovaries were observed. These factors, in combination with absence of mature females, suggest that* P. caperesca *practices semelparity, a reproductive strategy that complements the feeding strategy this obligate necrophage.*

*Five male and seven female size-grouped cohorts were identified for* P. caperesca*. Cohorts from deep-sea submarine canyon populations showed consistently larger mean total body lengths than non-canyon cohorts. Individuals from canyon samples also expressed sexual characteristics at smaller sizes than non-canyon individuals. We hypothesise that these trends are indicative of nutrient-mediated growth, maturation, and reproduction in* P. caperesca*. The species is able grow and reproduce relatively quickly in response to increased nutrient input in canyon environments and therefore dominates scavenging amphipod assemblages.*

**Keywords:** Amphipoda, abyssal, Atlantic Ocean, deep sea, necrophage, nutrient-mediated growth, semelparity

INTRODUCTION

Scavenging amphipods are a vital component of the secondary production cycles that support numerous deep-sea ecosystems through organic carbon cycling and dispersal from large food falls (Stockton & DeLaca, 1982; Britton & Morton, 1994; Payne & Moore, 2006; Higgs et al., 2014). However, knowledge of the population ecology and life histories of this group of necrophages is limited. Recent efforts in the study of these organisms have improved our understanding of their community level ecology (e.g. Thurston, 1990; Jamieson et al., 2011; Duffy et al., 2012; Horton et al., 2013), but studies examining the factors that affect population structure and dynamics of deep-sea scavenging amphipods are underrepresented.

Population studies carried out on shallow-water amphipod species (see Sainte-Marie, 1991, for a review of pre-1990 literature; Arndt & Beuchel, 2005; Nygård et al., 2009) provide a robust framework when examining and analysing deep-sea scavenger populations. However, deep-sea amphipod population studies have primarily targeted

populations of deep-sea amphipod species from vent (Sheader et al., 2000; Sheader et al., 2004; Sheader & Van Dover, 2007), canyon (Kaïm-Malka, 2003; Kaïm-Malka, 2004; Kaïm-Malka, 2005), and trench environments (Thurston et al., 2002; Perrone et al., 2002; Blankenship et al., 2006, Eustace et al., 2013). Cosmopolitan species have been studied to a limited extent with *Eurythenes gryllus* (Lichtenstein in Mandt, 1822) (Ingram & Hessler, 1987; Thurston et al., 2002; Premke et al., 2006; d'Udekem d'Acoz & Havermans, 2015) a notable example.

have also received some attention, but few cosmopolitan species have been studied (e.g. Duffy et al., 2013).

*Paralicella caperesca* Shulenberger & Barnard, 1976 has a pan-oceanic distribution in lower latitudes. It has been recorded from wide-spread localities in the Pacific Ocean (e.g. Shulenberger & Barnard, 1976; Yayanos, 1981; Ingram & Hessler, 1983; Hendrycks & Conlan, 2003; Jamieson et al., 2011; Fujii et al., 2013) within a depth range of 1720–6200 m (Wilson et al., 1985; Hasegawa et al., 1986), from the Indian Ocean (Treude et al., 2002; Cousins et al., 2013) and the Atlantic Ocean. The species has not been recorded in the Southern Ocean south of the Antarctic Polar Front (De Broyer et al., 2004; De Broyer et al., 2007) or in the Arctic Ocean. There are many records from the North Atlantic (Barnard & Shulenberger, 1976; Thurston, 1979; Macdonald & Gilchrist, 1980; Macdonald & Gilchrist, 1982; Smith & Baldwin, 1982; Lampitt et al., 1983; Desbruyères et al., 1985; Thurston, 1990; Barnard & Karaman, 1991; Jones et al., 1998; Bühring & Christiansen, 2001; Duffy et al., 2012; Horton et al., 2013), where it has been found in baited traps deployed at 2448 m on the Mid-Atlantic ridge (Horton et al., 2013) down to 5940 m on the Iberian Abyssal Plain (Barnard & Shulenberger, 1976). Records from the Tropical and South Atlantic (Thurston, 1990; Hendrycks et al., 2010) suggest that the species has a pan-Atlantic distribution. Virtually all of the *P. caperesca* specimens reported have come from baited traps set on, or within a few metres of, the sea floor but the species can occur well up in the water column. It has been taken in mid-water trawls more than 700 m above bottom in the Atlantic (Thurston, 1990) and more than 900 m above bottom in the Pacific (Ingram & Hessler, 1983). This broad geographic and bathymetric distribution is in spite of the limited dispersive ability, obligate brooding, and direct development that is characteristic of scavenging amphipods (Lincoln, 1979).

Very little is known of the life history of *P. caperesca* beyond inferences made from mouthpart and gut morphology. The mandibular molars are conico-laminate and non-triturative, and appear to act as a ratchet mechanism pulling strips of food cut by the broad incisor processes towards the oesophagus in a manner similar to that reported for *Alicella gigantea* Chevreux, 1899 (De Broyer & Thurston, 1987). The gut is capable of huge expansion, allowing the ingestion of large amounts of food implying the species is a highly specialised scavenger (Barnard & Shulenberger, 1976; Shulenberger & Barnard, 1976; Thurston, 1979). Thurston (1979) noted that sexual dimorphism of *P. caperesca* is limited to relative antenna length and that it is unlikely that females of this species are capable of producing multiple broods. Further study will provide vital information on the life history of this species and will lead to a better understanding of specialized scavengers in the deep sea.

*Paralicella caperesca* has been identified from many traps deployed on the abyssal plains of the North Atlantic, and has also been found in particularly high numbers within submarine canyons of the Iberian Peninsula, dominating traps deployed there (Duffy et al., 2012). This allows for comparisons between populations of *P. caperesca* from submarine canyons and their counterparts from non-canyon environments. Submarine canyons form an interesting environment in which to study scavenging populations as they are often identified as hotspots of secondary production (Vetter 1995; Soliman & Rowe, 2008; De Leo et al., 2010; van Oevelen et al., 2011), which may result in an increase in the number of food falls (Higgs et al., 2014). This comparative study will allow a detailed examination of how the heightened secondary production of canyon environments is affecting growth and reproduction in scavenging amphipod species while elucidating life history traits of *P. caperesca*.

MATERIALS AND METHODS

**Sample collection**

Canyon populations of *Paralicella caperesca* were represented by specimens from seven samples, collected using baited traps, and sorted for community composition analyses (Duffy et al., 2012; Table 1). All canyon samples were collected from submarine canyons off the western Iberian Peninsula as part of the HERMES project (Figure 1; Weaver et al., 2009). Four samples were from baited trap deployments in Nazaré Canyon, two were from Setúbal Canyon, and one was from Cascais Canyon.

To allow comparison between populations in submarine canyons and those of non-canyon environments, commensurate samples were identified from the Discovery Collections (National Oceanography Centre, Southampton, UK). These non-canyon samples were chosen on the basis that they contained sufficient numbers (> 50) of *P. caperesca* and were collected at a similar depth, during the same year or at a similar time of year as the HERMES canyon samples (Table 1). Eight of these samples were from abyssal plains in the NE Atlantic (Figure 1; Porcupine, Madeira, and Cape Verde Abyssal Plains), and one was from the Porcupine Seabight. The *P. caperesca* components of canyon and non-canyon samples were analysed as outlined below.

**Data collection**

For large samples (> 600 *P. caperesca* individuals) all specimens were assigned a unique identifier number and a random number generator was used to select a random subsample of 200 individuals. Selected individuals were dissected and measured under a stereo-dissecting microscope. Specimens were preserved in a variety of postures and many were damaged, making it difficult to obtain total body length measurements for all individuals. Past studies have resolved this problem using a single body-part measurement as a proxy for total body length (Chapelle, 1995; Sheader et al., 2000; Thurston et al., 2002; Sheader et al., 2004; Blankenship et al., 2006; Sheader & Van Dover, 2007; Nygård et al., 2009; Duffy et al., 2013). In this study coxal plate 4 was used as a proxy for total body length.

The total body length (distance along the dorsal margin between the anterior margin of the head and the tip of the telson) of 50 intact, randomly selected, *P. caperesca* from sample D297/15734 was measured using a digital graphics tablet and HTML-assisted Measuring System (HaMS; Duffy et al., 2013). These data were subsequently correlated to diagonal linear measures of coxal plate 4 (Figure 2; as in Chapelle, 1995), which were made using a stage graticule. Estimation of total body length for all remaining specimens was possible using the coxal plate 4 measurements as a proxy. Coxal plate 4 measurements, rather than estimated total body lengths, were used for population characterisation analyses (see below) to reduce the effect of rounding errors that would be magnified through the scaling up of measurements.

Primary and secondary sexual characteristics permitted sexing of all but the smallest individuals, which were classified as unsexed juveniles. Together with coxal plate 4, antenna 1 (length, number of articles, accessory flagellum length), antenna 2 (length, number of articles), and secondary sexual characteristics (oostegite length and presence of oostegite setae in females, presence of genital papillae in males) were also measured. Oocytes within ovaries of females at varying stages of sexual maturity were measured and counted.

**Population characterisation**

Characterisation of amphipod populations relies upon the identification of discrete cohorts, or growth stages, representing successive moults. Each of these cohorts contains individuals of similar size but not necessarily similar age, owing to growth rate being affected by a range of environmental factors. Identification of cohorts is often possible by identifying normal distributions from polymodal length frequency distribution data of total body length or a correlated proxy (Sheader et al., 2000; Thurston et al., 2002; Kaïm-Malka, 2003; Sheader et al., 2004; Kaïm-Malka, 2004; Kaïm-Malka, 2005; Premke et al., 2006; Blankenship et al., 2006; Sheader & Van Dover, 2007; Nygård et al., 2009; Duffy et al., 2013).

Oostegite measurements for females and coxal plate 4 measurements for males, females, and unsexed juveniles were plotted as probability density histograms. The identification of Gaussian components, each representing an oostegite stage or cohort, was performed initially by eye. The presence and parameters of each of these distributions were confirmed using the probability paper method (Cassie, 1954; Harding, 2009) and the ‘mixdist’ package (Macdonald & Pitcher, 1979; Macdonald & Du, 2011) in R statistical software (R Development Core Team, 2015). The goodness of fit of each of the modelled distributions was confirmed using the ANOVA function included in the ‘mixdist’ package.

**Comparison of populations**

Mann-Whitney U tests were used to compare distributions of male:female and juvenile:non-juvenile ratios between canyon and non-canyon samples. Once parameters for cohorts were accurately determined, individuals from each sample were assigned to the appropriate cohort based on 95% confidence limits (2 standard deviations from the mean). Where overlap of these limits occurred individuals were assigned to cohorts based on the relative proportions of each cohort. Following classification, population composition data were analysed using multivariate tests in PRIMER 6 (Plymouth Routines in Multivariate Ecological Research; Clarke & Gorley, 2006) statistical software. Data were root-transformed to reduce skewing and standardised to account for differing sample sizes. Following the creation of a Bray-Curtis similarity matrix populations were compared using an ANOSIM test (Clarke, 1993) with sampling area (canyon, non-canyon) as the lone factor.

RESULTS

In total, 1422 *Paralicella caperesca* from canyon samples and 1575 individuals from non-canyon samples were dissected and measured. Males, females, and juveniles were identified in all samples. No mature or ovigerous females were found. There was a positive correlation between coxal plate 4 length and total body length (L = 6.8902C + 1.0610, R2 = 0.6517; where L is the estimated total body length and C is the linear measure of coxal plate 4, Figure 2). Although untransformed coxal plate 4 measurements were used for subsequent analyses, this relationship did permit the use of coxal plate 4 measurements as an indicative proxy to estimate total body length and allow for basic morphometric description and comparison of *P. caperesca* with other scavenging amphipod species.

Estimated total body length ranged from 1.70 mm to 22.34 mm. No individuals smaller than 4.65 mm showed primary or secondary sexual characteristics and were classed as juveniles. All individuals of 8.00 mm in length or more consistently exhibited primary and secondary sexual characteristics. A number of individuals smaller than 8.00 mm, mainly from canyon samples, could be sexed.

**Sexual dimorphism**

Estimated total body length in males ranged from 4.65 mm to 18.97 mm (Median = 10.12 mm, IQ range = 8.86 mm to 10.55 mm) and in females from 4.65 mm to 22.34 mm long (Median = 10.97 mm, IQ range = 8.86 mm to 13.07 mm). There was a significant difference between the size distributions of males and females (Figure 3; Mann-Whitney U-test: n = 963, U = 82 146, p < 0.001). Males possessed significantly longer antenna 1 and antenna 2, relative to coxal plate 4 length, than females (MWU: n = 948, U = 152 385, p < 0.001; n = 950, U = 171 933, p < 0.001 respectively). The antennal differences between the sexes was most pronounced in antenna 2, with the antenna 1 to antenna 2 ratio significantly lower for males than females (MWU: n = 940, U = 45 246, p < 0.001) with males more commonly possessing second antennae of equal or greater length than their first antennae. As a function of antenna length (articles mm-1) males possessed significantly more articles per mm of antenna length than females (MWU: antenna 1, n = 753, U = 68 733, p = 0.015; antenna 2, n = 755, U = 84 405, p = 0.001).

**Sexual development**

A pair of ovaries could be identified between pereonites 3 and 7, lying either side of the gut, in females as small as 4.65 mm. Complete ovaries were recoverable from 59 individuals. Both ovaries contained approximately equal numbers of spheroid oocytes of comparable size, densely packed in a three-tiered linear arrangement. Oocyte size, calculated as the area of an ellipse, showed a weak positive correlation with coxal plate 4 length, a proxy of total body length (Figure 4a). The total number of oocytes within each whole ovary ranged from 26 to 75, with a mean of 42.4 oocytes per ovary, but there was no apparent relationship between oocyte count and estimated body size (Figure 4b).

The smallest female with visible oostegite buds was an estimated 4.65 mm in length but most females of this size had no detectable oostegites and could only be identified by ovary presence. Larger females possessed oostegites of varying size. A probability density histogram of oostegite length as a ratio of gill length (Figure 4c) identified four oostegite stages (OS1–OS4; Table 2) with minimal overlap. The presence and parameters of these stages were confirmed using the probability paper method.

No mature females with setose oostegites were found. Evidence of setal development was seen in some specimens possessing particularly large oostegites but in no case had setae evaginated. This suggests that these individuals were one moult away from sexual maturity. Testes were identifiable in the majority of males seen but were generally in poor condition making developmental classification impossible. Penile papillae in this species were small but could be identified on close examination of the underside of pereonite 7. The smallest male possessing visible penile papillae was an estimated 4.65 mm in body length.

**Population characterisation**

Size distributions differed between the canyon samples and non-canyon samples, therefore separate cohort analyses were performed for each of these environments. Individuals that could not be sexed were assigned to four identified juvenile stages (Table 3, Figures 5, 6). The parameters for the identified cohorts are comparable for the smallest individuals (J1, J2) but the separation between canyon and non-canyon cohorts becomes more pronounced at greater body length (J3, J4). Canyon cohorts have consistently larger mean total body lengths than their non-canyon counterparts.

A total of five male (Figure 5; M3–M8) and seven female (Figure 6; F3–F10) cohorts were identified (Table 3). The occurrence and parameters of these cohorts were confirmed using the probability paper method and a ‘mixdist’ model fitted to the data (ANOVA: Canyon males, d.f. = 19, χ2 = 24.08, p = 0.1930; canyon females, d.f. = 25, χ2 = 51.88, p = 0.1243; non-canyon males, d.f. = 19, χ2 = 12.47, p = 0.8646; non-canyon females, d.f. = 20, χ2 = 25.53, p = 0.2634). There was direct overlap between the largest juvenile (J3, J4) and smallest sexed (M3, M4, F3, F4) cohorts. Cohorts were numbered to reflect this.

Cohorts M3 and F3 were found almost exclusively in canyon samples with individuals of similar size from non-canyon environments rarely showing any sexual characteristics and therefore classified as J3 juveniles. The two cohorts containing the largest individuals (M8, F10) were not represented in canyon samples. Canyon cohorts showed consistently larger mean total body lengths than their non-canyon counterparts (Table 3; Figure 5, 6).

Linear growth factors between cohorts were comparable to those reported for *A. abyssorum* (Stebbing, 1888) (Duffy et al., 2013) and early stages of *E. gryllus* (Thurston, 1979; Ingram & Hessler, 1987). Increments between cohorts ranged from 1.10–1.44 with the largest growth increments between juvenile cohorts. Oostegite stages were distributed across female cohorts. More developed oostegites were more frequently observed in smaller individuals (F3–F6) from canyons than from non-canyon environments. The half-range of mature female body length (Sainte-Marie, 1991), a means proposed to distinguish between iteroparous and semelparous amphipod species, could not be calculated for *P. caperesca* as no mature females were available.

**Population comparison**

The male:female and juvenile:non-juvenile ratios did not differ significantly between canyon and non-canyon environments (Table 4; Mann-Whitney U-test: n = 14, U = 28.50, p = 0.620; n = 14, U = 27.50, p = 0.710 respectively). Both ratios varied widely within test groups. Multivariate analysis showed population composition based on cohort abundance varied significantly between canyon and non-canyon samples (ANOSIM: R = 0.284, p = 0.020).

DISCUSSION

**Sexual dimorphism**

Sexual dimorphism of *Paralicella caperesca*, while apparent, is less pronounced than in some other deep-sea scavenging amphipod species such as *A. abyssorum* (Duffy et al., 2013) and *E. gryllus* (Ingram & Hessler, 1987). Despite the lack of mature individuals, females were marginally larger than males in terms of estimated body length (Figure 3). This is in agreement with the pattern of inter-sexual size differences in lysianassoids demonstrated by Sainte-Marie (1991). *Paralicella caperesca* shares many characteristics with lysianassoids. As such, the observed size difference between males and females suggests that *P. caperesca* belongs to a group of species that practices non-mate-guarding pre-copulatory behaviour (Bousfield & Shih, 1994; Bousfield, 2001) as documented in lysianassoids by Conlan (1991). Males had significantly longer antennae with significantly more articles per mm of antenna length. The elongate antennae of male amphipods are believed to enhance chemosensing ability (Kaufmann, 1994) and tactile detection during mate searching (Steele, 1995).

**Sexual development**

No mature females were found amongst the 2997 specimens examined in the present study and Thurston (1979) identified only two mature females from a sample of 330 individuals taken at a depth of 4855 m in the North-East Atlantic. Ingram & Hessler (1983) did not indicate the proportion of adult female *P. caperesca* in their samples, so it is unclear as to whether the present observations represent the norm. The near-absence of mature females in baited-traps suggests that *P. caperesca* is a semelparous organism as hypothesised by Thurston (1979).

The size of oostegites in female *P. caperesca* varied considerably within instars (Table 3), indicating a wide overlap of size ranges for succeeding developmental stages. This agrees with the pattern documented by Thurston (1979) in material trapped on the Horseshoe Abyssal Plain.A re-analysis of benthopelagic material from midwater hauls taken up to at least 720 m above the Cape Verde Abyssal Plain (Thurston, 1990) showed a similar pattern (juveniles 3–8 mm, juvenile females 6–12 mm, immature females 10–17 mm, adult females 9–11 mm) with the adult females taken at least 100 m above the sea floor. This common pattern indicates that sexual development is plastic in female *P. caperesca* and that maturity is likely reached in fewer instars in canyon populations than abyssal plain populations. In contrast, there is no size overlap between non-adult and adult males implying that development is fixed or at least less plastic than in females.

Based on the contents of all complete ovaries recovered, the maximum possible brood size of *P. caperesca* ranges from 52–150 offspring with a mean estimated brood size of 84. Total oocyte counts were substantially higher than those reported by Duffy et al., (2013) for the bathyal scavenging amphipod *A. abyssorum*. Again using lysianassoid amphipods as a reference point, brood size loosely correlates with female body length but varies widely (Sainte-Marie, 1991). The estimated brood size of *P. caperesca* is substantially larger than that of most lysianassoids of similar size. The potential to produce large broods is in keeping with the hypothesis that *P. caperesca* is semelparous, investing substantial resources into one large reproductive event.

The frequency of the most developed oostegite stages (OS3, OS4) increased with increasing size-class number, however, no oostegites were setose regardless of oostegite stage and size-class. The lack of mature and ovigerous females leaves an important gap in our understanding of this species. As a result of the apparently semelparous lifestyle of *P. caperesca* it is unlikely that either of these development-classes will ever be caught using baited traps. The risks of predation at a food fall can be high (Lampitt et al., 1983; Jones et al., 1998; Janßen et al., 2000; Jamieson et al., 2009; Jamieson et al., 2011). Furthermore, the ability of *P. caperesca* to ingest huge quantities of food also results in a morphotype incompatible with brood retention (Shulenberger & Hessler, 1974; Shulenberger & Barnard, 1976; Thurston, 1979).

The hypothesis that *P. caperesca* is semelparous is further supported by the fact that no empty ovaries were identified in any females examined. A semelparous lifestyle has been identified as a successful reproductive strategy for insects in environments with stable and predictable nutrient input (Fritz et al., 1982, and references therein). If, as theorised, a semelparous strategy is practiced by *P. caperesca* this may go some way to explaining the dominance and apparent success of *Paralicella* species in submarine canyons (Duffy et al., 2012), where nutrient input is elevated and relatively consistent.

**Population characterisation and comparison**

The linear growth factors determined here for *P. caperesca* are comparable to those previously documented for this species (Thurston, 1979). These factors are lower than those of *A. abyssorum* (Duffy et al., 2013) and early instars of *E. gryllus* (Thurston, 1979; Ingram & Hessler, 1987). They are similar to those reported for *O. gerulicorbis* (Thurston, 1979, as *Orchomene*), and higher than documented growth factors for *Scopelocheirus hopei* (Costa, 1851) (Kaïm-Malka, 2003), and *Tmetonyx similis* (Sars, 1891) (Kaïm-Malka, 2005).

There are marked differences in the population composition between the canyon samples and the non-canyon samples. The estimated mean total body length for each size-class was consistently higher for canyon size-classes than it was for their counterparts from the non-canyon environments. A number of environmental factors differentiating between these two sampling areas may be responsible for these differences. Canyon sites were generally shallower than non-canyon sites owing to their position on the continental slope. Therefore, canyon sites were also in marginally warmer water (Duffy et al., 2012). While the non-canyon group included one shallower site not from the abyssal plain (D092/9756/008, 3852 m), this sample group was nevertheless skewed toward deeper sampling. Amphipod growth rates and adult body size increase at warmer temperatures (Sutcliffe et al., 1981; Highsmith & Coyle, 1991; Pöckl, 1992; Panov & McQueen, 1998; Maranhão & Marques, 2003). However, the temperature difference between sample sites in canyons and non-canyon environments was very small (< 0.1 °C) compared to the temperature ranges used in laboratory experiments, making this explanation of the size differences between the populations unlikely.

Another possible explanation for the differing body-lengths between canyon and non-canyon populations is that the two environments support separate subspecies of *P. caperesca*. Despite the many records of this species from Pacific, Atlantic, and Indian Oceans (see above), the only report of morphological variation is of minor differences between Pacific and Atlantic material (Barnard & Shulenberger, 1976). All of the material used in this study was obtained from a relatively circumscribed area in the North-East Atlantic Ocean lacking obvious physical barriers. This apparent homogeneity is in contrast with the significant morphological variation reported for *E. gryllus* (Thurston et al., 2002; Stoddart & Lowry, 2004), shown subsequently to represent a species-complex containing at least five species (Havermans et al., 2013; d'Udekem d'Acoz & Havermans, 2015). The *Eurythenes* species-complex shows a bathymetric as well as a geographic element in the genetic divergences separating the included entities (Bucklin et al., 1987, Havermans et al., 2013). All of the *P. caperesca* material used in this study came from depths exceeding 3000 m, a depth that appears critical for separating shallow and deep entities within the *Eurythenes* complex (Havermans et al, 2013). This suggests that bathymetry is not a prime cause of the differences observed.

Supported by the differences seen between populations of *A. abyssorum* in areas of different surface productivity (Duffy et al., 2013), it appears that the most probable reason for larger total body length in canyon size-classes is a result of the increased nutrient input to canyon environments. While the availability of large food-falls to deep canyon sites is yet unknown, increased nutrient availability and heightened secondary production have been documented (Vetter, 1995; Soliman & Rowe, 2008; De Leo et al., 2010; van Oevelen et al., 2011). The high abundances of scavenging amphipods found in submarine canyons (Duffy et al., 2012) lends further support to the theory that nutrient availability in an environment has significant effects on scavenging amphipods at both the community and population levels.

There are several possible explanations for the earlier onset of reproductive development in submarine canyon populations. As with growth rate, earlier reproductive development has also been attributed to temperature variations in other amphipod species (Sheader, 1983; Highsmith & Coyle, 1991), however, as previously discussed, water temperature in the sampled canyons was only marginally higher than at non-canyon sites (< 0.1 °C; Duffy et al., 2012). Once again, the difference in nutrient availability between the two areas is an important consideration. A link between nutrient availability and sexual development in *P. caperesca* has been previously hypothesised by Thurston (1979). With an abundant food supply, individuals in canyons are able to apply resources to both growth and reproductive output allowing the onset of earlier reproductive development (Sutcliffe et al., 1981). Earlier maturation and oostegite development with nutrient rich conditions has been recorded in the scavenging amphipod *T. similis* (Kaïm-Malka, 2004). This developmental plasticity in response to food supply has also been observed in other amphipod species from very different habitats, such as the estuarine *Gammarus duebeni* Liljeborg, 1852 (Sheader, 1983) and the vent-dwelling *Bouvierella curtirama* Bellan-Santini & Thurston, 1996 (Sheader et al., 2004). In the food-limited abyssal plains resources are more constrained and therefore input into reproductive development is delayed until further growth has been achieved.

**Ecological implications**

*Paralicella caperesca* has the characteristics of a semelparous organism capable of one reproductive event in a lifetime and, as demonstrated by its abundance and ubiquity (Thurston, 1979; Lampitt et al., 1983; Thurston, 1990; Barnard & Karaman, 1991; Jones et al., 1998; Duffy et al., 2012; Horton et al., 2013; Duffy et al., 2013), it is very successful in abyssal habitats. In his comparison of two scavenging fish species, *Coryphaenoides acrolepis* (Bean, 1884) and *C. armatus* (Hector, 1875), Drazen (2002) hypothesised that the latter, an abyssal species, is semelparous owing to the nutrient regime of the abyssal zone. While food supplies in the abyss are low, they are also generally stable. This means survivorship of both juveniles and adults is relatively constant, making semelparity a successful and energetically conservative reproductive strategy. The same appears to be true for *P. caperesca*.

The small, sexually mature, *P. caperesca* females identified in material reported by Thurston (1979, 1990) together with variability in sexual development across size-classes reported in this study suggest that maturation, and hence reproduction, of *P. caperesca* relies on an environmental trigger rather than being purely age or size dependent. There is strong evidence that this trigger is primarily nutrient availability. As an obligate scavenger capable of gorging itself (Shulenberger & Barnard, 1976; Thurston, 1979), *P. caperesca* can capitalise on any high nutrient inputs into the abyss, such as large food falls, maturing and reproducing when these events occur. The expected increase in large food falls (Higgs et al., 2014) driven by increased sediment input to the submarine canyon system and heightened productivity (Vetter, 1995; Soliman & Rowe, 2008; De Leo et al., 2010; van Oevelen et al., 2011) and the semelparity of *P. caperesca* allow this species to dominate canyon communities (Duffy et al., 2012).

The semelparous lifestyle of *P. caperesca* provides an interesting contrast to *A. abyssorum*, another deep-sea scavenging amphipod that has been studied in comparative detail (Duffy et al., 2013), which is more often associated with the bathyal zone (Horton et al., 2013). *Abyssorchomene abyssorum* expresses the characteristics of an iteroparous organism capable of producing multiple broods of similar size to other iteroparous amphipod species (Sainte-Marie, 1991; Duffy et al., 2013). It is a less specialised scavenger than *P. caperesca* and believed to be a facultative rather than obligate scavenger (Shulenberger & Barnard, 1976; Thurston, 1979). This may go some way toward explaining the differing reproductive strategies of these two species. Investing in a single, large, reproductive output in response to a food fall allows *P. caperesca* to maximise its success under the feast-famine regime experienced by obligate scavengers in abyssal environments and capitalise when the only nutrient resource it can consume becomes available. As *A. abyssorum* cannot gorge to the same extent as a specialist necrophage (Shulenberger & Barnard, 1976; Dahl, 1979; Thurston, 1979), a more generalist feeding strategy where smaller amounts are consumed at more regular intervals is likely. This strategy lends itself to the multiple smaller reproductive outputs of iteroparity. These differing reproductive and scavenging strategies have allowed these two species to become highly successful and abundant within their respective environments (Duffy et al., 2012; Horton et al., 2013).

*Abyssorchomene abyssorum* and *P. caperesca* have bathymetric distributions that overlap to a limited degree (Thurston, 1990; Horton et al., 2013). Hydrostatic pressure and water temperature are often identified as factors controlling scavenging amphipod distributions and community composition (Thurston, 1990; Thurston et al., 2002) but trophic conditions have also been implicated as a contributory factor (Premke et al., 2006; Horton et al., 2013). Our improved understanding of the contrasting feeding and reproductive strategies of scavenging amphipod species supports this theory. Obligate necrophages, such as *P. caperesca*, prevail in abyssal habitats, where background nutrients are poor and large inputs are likely to be patchy and often infrequent, through facultative breeding (i.e. based on food availability). If food and sufficient energy are available individuals will gorge, mature rapidly, breed, brood, and die. Where episodic nutrient-rich inputs, such as large food falls, occur more frequently, species capable of earlier maturation and a large reproductive output when trophic conditions are favourable are able to dominate communities (Duffy et al., 2012). In regions where the trophic conditions are more stable and nutrients are less limited, generalists following an iteroparous lifestyle, with multiple small reproductive events, are at an advantage. *A. abyssorum* has adopted this lifestyle (Duffy et al., 2013) which may facilitate community dominance in shallower deep-sea habitats such as mid-ocean ridges (Horton et al., 2013). Deep-sea scavenging fishes of the genus *Coryphaenoides* show similar patterns in reproductive strategy differences between abyssal and bathyal habitats (Drazen, 2002).

This study presents strong evidence for semelparity and nutrient-mediated growth, maturation, and reproduction in *Paralicella caperesca*. Comparing canyon and non-canyon populations of this species highlights how this strategy affects population dynamics, with canyon cohorts having consistently larger body size and expressing sexual characteristics at a smaller size. These findings provide important insights into the ecology of deep-sea scavenging amphipods. Semelparity complements the obligate necrophagy practised by *P. caperesca* and has been pivotal to the success of this species in both abyssal and deep-sea submarine canyon habitats.

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REFERENCES

**Arndt C. E. and Beuchel F.** (2005). Life history and population dynamics of the Arctic sympagic amphipods *Onisimus nanseni* Sars and *O. glacialis* Sars (Gammaridea: Lysianassidae). *Polar Biology* 29, 239–248. doi:10.1007/s00300-005-0045-x

**Barnard J. L. and Karaman G. S.** (1991). The families and genera of marine gammaridean Amphipoda (except marine gammaroidea). Part 2. *Records of the Australian Museum, Supplement* 13, 419–866. doi:10.3853/j.0812-7387.13.1991.367

**Barnard J. L. and Shulenberger E.** (1976). Clarification of the abyssal amphipod, *Paralicella tenuipes* Chevreux. *Crustaceana* 31, 267–274. doi:10.1163/156854076x00053

**Bellan-Santini D. and Thurston M. H.** (1996). Amphipoda of the hydrothermal vents along the mid-Atlantic Ridge. *Journal of Natural History* 30, 685–702. doi:10.1080/00222939600770381

**Blankenship L. E., Yayanos A. A., Cadien D. B. and Levin L. A.** (2006). Vertical zonation patterns of scavenging amphipods from the Hadal zone of the Tonga and Kermadec Trenches. *Deep Sea Research Part I: Oceanographic Research Papers* 53, 48–61. doi:10.1016/j.dsr.2005.09.006

**Bousfield E. L.** (2001). An updated commentary on phyletic classification of the amphipod Crustacea and its application to the North American fauna. *Amphipacifica* 3, 49–119

**Bousfield E. L. and Shih C. T.** (1994). The phyletic classification of amphipod crustaceans: problems in resolution. *Amphipacifica* 1, 76–134.

**Britton J. C. and Morton B.** (1994). Marine carrion and scavengers. *Oceanography and Marine Biology: an Annual Review* 32, 369–434.

**Bucklin A., Wilson R.R. and Smith K.L.** (1987). Genetic differentiation of seamount and basin populations of the deep-sea amphipod *Eurythenes gryllus*. *Deep Sea Research Part A Oceanographic Research Papers* 34, 1795–1810. doi: 10.1016/0198-0149(87)90054-9

**Bühring S. I. and Christiansen B.** (2001). Lipids in selected abyssal benthopelagic animals: links to the epipelagic zone? *Progress in Oceanography* 50, 369–382. doi:10.1016/s0079-6611(01)00061-1

**Cassie R. M.** (1954). Some uses of probability paper in the analysis of size frequency distributions. *Australian Journal of Marine and Freshwater Research* 5, 513–522. doi:10.1071/MF9540513

**Chapelle G.** (1995). Estimating size of amphipods in life cycle studies: what to measure and what for? *Polskie Archiwum Hydrobiologii* 42, 295–302.

**Chevreux E.** (1899). Sur quelques intèressantes espèces d'amphipodes provenant de la dernière campagne du yacht Princesse Alice. *Bulletin De La Societe Zoologique De France* 24, 152–158.

**Clarke K. R.** (1993). Non‐parametric multivariate analyses of changes in community structure. *Australian Journal of Ecology* 18, 117–143. doi:10.1111/j.1442-9993.1993.tb00438.x

**Clarke K. R. and Gorley R. N.** (2006). *PRIMER v6: User Manual/Tutorial*. Plymouth: PRIMER-E.

**Conlan K. E.** (1991). Precopulatory mating behavior and sexual dimorphism in the amphipod Crustacea. *Hydrobiologia* 223, 255–282. doi:10.1007/bf00047644

**Costa A.** (1851). Catalogo de crostacei del regno di Napoli. *Anfipodi Gammaridi. Fauna Regno Napoli* 7.

**Cousins N. J., Horton T., Wigham B. D. and Bagley P. M.** (2013). Abyssal scavenging demersal fauna at two areas of contrasting productivity on the Subantarctic Crozet Plateau, southern Indian Ocean. *African Journal of Marine Science*, 35, 299–306. doi:10.2989/1814232X.2013.802747

**d'Udekem d'Acoz C. and Havermans C.** (2015). Contribution to the systematics of the genus *Eurythenes* S.I. Smith in Scudder, 1882: an integrative study (Crustacea: Amphipoda: Lysianassoidea: Eurytheneidae). *Zootaxa* 3971, 1–80. doi:10.11646/zootaxa.3971.1.1

**Dahl E.** (1979). Deep-sea carrion feeding amphipods: evolutionary patterns in niche adaptation. *Oikos* 33, 167. doi:10.2307/3543994

**De Broyer C., Lowry J. K., Jażdżewski K. and Robert H.** (2007). Catalogue of the Gammaridean and Corophiidean Amphipoda (Crustacea) of the Southern Ocean with distribution and ecological data. In C. De Broyer, *Census of Antarctic Marine Life Synopsis of the Amphipoda of the Southern Ocean*.

**De Broyer C., Nyssen F. and Dauby P.** (2004). The crustacean scavenger guild in Antarctic shelf, bathyal and abyssal communities. *Deep Sea Research Part II: Topical Studies in Oceanography* 51, 1733–1752. doi:10.1016/j.dsr2.2004.06.032

**De Broyer C. and Thurston M. H.** (1987). New Atlantic material and redescription of the type specimens of the giant abyssal amphipod *Alicella gigantea* Chevreux (Crustacea). *Zoologica Scripta* 16, 335–350.

**De Leo F. C., Smith C. R., Rowden A. A., Bowden D. A. and Clark M. R.** (2010). Submarine canyons: hotspots of benthic biomass and productivity in the deep sea. *Proceedings of the Royal Society B: Biological Sciences* 277, 2783–2792. doi:10.1098/rspb.2010.0462

**Desbruyères D., Geistdoerfer P., Ingram C. L., Khripounoff A. and Lagardère J. P.** (1985). Répartition des populations de l’épibenthos carnivore. In L. Laubier & C. Monniot, *Peuplements profonds du golfe de Gascogne* (pp. 233–252). Paris: IFREMER.

**Drazen J. C.** (2002). Energy budgets and feeding rates of *Coryphaenoides acrolepis* and *C. armatus*. *Marine Biology*, 140, 677–686. doi:10.1007/s00227-001-0747-8

**Duffy G. A., Horton T. and Billett D. S. M.** (2012). Deep-sea scavenging amphipod assemblages from the submarine canyons of the Western Iberian Peninsula. *Biogeosciences*, 9, 4861–4869. doi:10.5194/bg-9-4861-2012

**Duffy G. A., Horton T., Sheader M. and Thurston M. H.** (2013). Population structure of *Abyssorchomene abyssorum* (Stebbing, 1888) (Amphipoda: Lysianassoidea), a scavenging amphipod from the Mid-Atlantic Ridge in the vicinity of the Charlie-Gibbs Fracture Zone. *Deep Sea Research Part II: Topical Studies in Oceanography* 98, 360–369. doi:10.1016/j.dsr2.2013.02.004

**Eustace R. M., Kilgallen N. M., Lacey N. C. and Jamieson A. J.** (2013). Population structure of the hadal amphipod *Hirondellea gigas* (Amphipoda: Lysianassoidea) from the Izu-Bonin Trench. *Journal of Crustacean Biology* 33, 793–801. doi:10.1163/1937240X-00002193

**Fritz R. S., Stamp N. E. and Halverson T. G.** (1982). Iteroparity and semelparity in insects. *The American Naturalist* 120, 264. doi:10.1086/283987

**Fujii T., Kilgallen, N. M. Rowden, A. A. and Jamieson A. J.** (2013). Deep-sea amphipod community structure across abyssal to hadal depths in the Peru-Chile and Kermadec trenches. *Marine Ecology Progress Series* 492, 125–138. doi:10.3354/meps10489

**Harding J. P.** (2009). The use of probability paper for the graphical analysis of polymodal frequency distributions. *Journal of the Marine Biological Association of the UK* 28, 141. doi:10.1017/S0025315400055259

**Hasegawa M., Kurohiji Y., Takayanagi S., Sawadaishi S. and Yao M.** (1986). Collection of fish and Amphipoda from abyssal sea-floor at 30 °N – 147 °E using traps tied to 10000 m wire of research vessel. *Bulletin of Tokai Regional Fisheries Research Laboratory* 119, 65–75.

**Havermans C., Sonet G., d’Udekem d’Acoz C., Nagy Z. T., Martin P., Brix S., Riehl T., Agrawal H. and Held C.** (2013). Genetic and morphological divergences in the cosmopolitan deep-sea amphipod *Eurythenes gryllus* reveal a diverse abyss and a bipolar species. *PLoS ONE* 8, e74218–15. doi:10.1371/journal.pone.0074218

**Hendrycks E. A. and Conlan, K. E.** (2003). New and unusual abyssal gammaridean Amphipoda from the north-east Pacific. *Journal of Natural History* 37, 2303–2368. doi:10.1080/00222930210138926

**Hendrycks E. A., De Broyer C. and Havermans C.** (2010) *Preliminary notes on baited trap amphipods from the DIVA-3 cruise*. Paper presented at XIVth International Colloquium on Amphipoda. Sevilla, Spain. http://hdl.handle.net/2078/121215

**Higgs N. D., Gates A. R. and Jones D. O. B.** (2014). Fish food in the deep sea: revisiting the role of large food-falls. *PLoS ONE* 9, e96016. doi:10.1371/journal.pone.0096016.s006

**Highsmith R. C. and Coyle K. O.** (1991). Amphipod life histories: community structure, impact of temperature on decoupled growth and maturation rates, productivity, and P: B ratios. *American Zoologist* 31, 861–873. doi:10.1093/icb/31.6.861

**Horton T., Thurston M. H. and Duffy G. A.** (2013). Community composition of scavenging amphipods at bathyal depths on the Mid-Atlantic Ridge. *Deep Sea Research Part II: Topical Studies in Oceanography* 98, 352–359. doi:10.1016/j.dsr2.2013.01.032

**Ingram C. L. and Hessler R. R.** (1983). Distribution and behavior of scavenging amphipods from the central North Pacific. *Deep Sea Research Part A Oceanographic Research Papers* 30, 683–706. doi:10.1016/0198-0149(83)90017-1

**Ingram C. L. and Hessler R. R.** (1987). Population biology of the deep-sea amphipod *Eurythenes gryllus*: inferences from instar analyses. *Deep Sea Research Part A Oceanographic Research Papers* 34, 1889–1910. doi:10.1016/0198-0149(87)90090-2

**Jamieson A. J., Fujii T., Solan M., Matsumoto A. K., Bagle, P. M. and Priede I. G.** (2009). First findings of decapod crustacea in the hadal zone. *Deep Sea Research Part I: Oceanographic Research Papers* 56, 641–647. doi:10.1016/j.dsr.2008.11.003

**Jamieson A. J., Kilgallen N. M., Rowden A. A., Fujii T., Horton T., Lörz A. N., Kitazawad K. and Priede I. G.** (2011). Bait-attending fauna of the Kermadec Trench, SW Pacific Ocean Evidence for an ecotone across the abyssal–hadal transition zone. *Deep Sea Research Part I: Oceanographic Research Papers* 58, 49–62. doi:10.1016/j.dsr.2010.11.003

**Janßen F., Treude T. and Witte U.** (2000). Scavenger assemblages under differing trophic conditions: a case study in the deep Arabian Sea. *Deep Sea Research Part II: Topical Studies in Oceanography* 47, 2999–3026. doi:10.1016/s0967-0645(00)00056-4

**Jones E. G., Collins M. A., Bagley P. M., Addison S. and Priede I. G.** (1998). The fate of cetacean carcasses in the deep sea: observations on consumption rates and succession of scavenging species in the abyssal north-east Atlantic Ocean 265, 1119–1127. doi:10.1098/rspb.1998.0407

**Kaïm-Malka R. A.** (2003). Biology and life cycle of *Scopelocheirus hopei* (A. Costa, 1851), a scavenging amphipod from the continental slope of the Mediterranean. *Journal of Natural History* 37, 2547–2578. doi:10.1080/00222930210155693

**Kaïm-Malka R. A.** (2004). Oostegite development during the sexual maturation of females of *Tmetonyx similis* (G. O. Sars, 1891) (Amphipoda, Lysianassidae). *Journal of Natural History* 38, 2403–2424. doi:10.1080/00222930310001647398

**Kaïm-Malka R. A.** (2005). Biology and life cycle of *Tmetonyx similis* (G. O. Sars, 1891) (Amphipoda, Lysianassidae), a scavenging amphipod from the continental slope of the Mediterranean. *Journal of Natural History* 39, 3163–3186. doi:10.1080/00222930500240502

**Kaufmann R. S.** (1994). Structure and function of chemoreceptors in scavenging lysianassoid amphipods. *Journal of Crustacean Biology* 14, 54. doi:10.2307/1549055

**Lampitt R. S., Merrett N. R. and Thurston M. H.** (1983). Inter-relations of necrophagous amphipods, a fish predator, and tidal currents in the deep sea. *Marine Biology* 74, 73–78. doi:10.1007/BF00394277

**Lichtenstein H.** (1822). *Observationes in historiam naturalem et anatomiam comparatam*. In M. W. Mandt, *itinere Groenlandico factae* (pp. 31–37).

**Liljeborg V.** (1852). Norges Crustacéer. *Öfversigt af Kongliga Vetenskaps-Akademiens Förhandlingar­­­* 8, 19–25.

**Lincoln, R. J.** (1979). *British Marine Amphipoda: Gammaridea.* London: British Museum (Natural History).

**Macdonald A. G. and Gilchrist I.** (1980). Effects of hydraulic decompression and compression on deep-sea amphipods. *Comparative Biochemistry and Physiology a-Physiology* 67, 149–153. doi:10.1016/0300-9629(80)90420-X

**Macdonald A. G. and Gilchrist I.** (1982). The pressure tolerance of deep sea amphipods collected at their ambient high pressure. *Comparative Biochemistry and Physiology Part A: Physiology* 71, 349–352. doi:10.1016/0300-9629(82)90415-7

**Macdonald P. D. M. and Du J.** (2011). *mixdist: Finite Mixture Distribution Models. R package version 0.5-4. http://CRAN.Rproject.org/package=mixdist*.

**Macdonald P. D. M. and Pitcher T. J.** (1979). Age-Groups from size-frequency data: A versatile and efficient method of analyzing distribution mixtures. *Journal of the Fisheries Research Board of Canada* 36, 987–1001. doi:10.1139/f79-137

**Maranhão P. and Marques J. C.** (2003). The influence of temperature and salinity on the duration of embryonic development, fecundity and growth of the amphipod *Echinogammarus marinus* Leach (Gammaridae). *Acta Oecologica* 24, 5–13. doi:10.1016/S1146-609X(02)00003-6

**Nygård H., Vihtakari M. and Berge J.** (2009). Life history of *Onisimus caricus* (Amphipoda: Lysianassoidea) in a high Arctic fjord. *Aquatic Biology* 5, 63–74. doi:10.3354/ab00142

**Panov V. E. and McQueen D. J.** (1998). Effects of temperature on individual growth rate and body size of a freshwater amphipod. *Canadian Journal of Zoology* 76, 1107–1116. doi:10.1139/cjz-76-6-1107

**Payne L. X. and Moore J. W.** (2006). Mobile scavengers create hotspots of freshwater productivity. *Oikos* 115, 69–80

**Perrone F. M., Dell'Anno, A., Danovaro, R., Della Croce N. and Thurston M. H.** (2002). Population biology of *Hirondellea* sp. nov. (Amphipoda : Gammaridea : Lysianassoidea) from the Atacama Trench (south-east Pacific Ocean). *Journal of the Marine Biological Association of the UK* 82, 419–425. doi:10.1017/S0025315402005672

**Pöckl M.** (1992). Effects of temperature, age and body size on moulting and growth in the freshwater amphipods *Gammarus fossarum* and *G. roeseli*. *Freshwater Biology*, 27, 211–225. doi:10.1111/j.1365-2427.1992.tb00534.x

**Premke K., Klages M. and Arntz W. E.** (2006). Aggregations of Arctic deep-sea scavengers at large food falls: temporal distribution, consumption rates and population structure. *Marine Ecology Progress Series* 325, 121–135.

**R Development Core Team**. (2015). *R: A Language and Environment for Statistical Computing*. Vienna: R Foundation for Statistical Computing.

**Sainte-Marie B.** (1991). A review of the reproductive bionomics of aquatic gammaridean amphipods: variation of life history traits with latitude, depth, salinity and superfamily. *Hydrobiologia* 223, 189–227. doi:10.1007/978-94-011-3542-9\_19

**Sars G. O.** (1891). *An account of the Crustacea of Norway with short descriptions and figures of all the species. Volume 1, Amphipoda, Parts 4-9* (pp. 69–212). Copenhagen: A Cammermeyer.

**Sheader M.** (1983). The reproductive biology and ecology of *Gammarus duebeni* (Crustacea: Amphipoda) in southern England. *Journal of the Marine Biological Association of the UK* 63, 517. doi:10.1017/s0025315400070855

**Sheader M. and Van Dover C. L.** (2007). Temporal and spatial variation in the reproductive ecology of the vent-endemic amphipod *Ventiella sulfuris* in the eastern Pacific. *Marine Ecology Progress Series* 331, 181–194.

**Sheader M., Van Dover C. L. and Shank T. M.** (2000). Structure and function of *Halice hesmonectes* (Amphipoda: Pardaliscidae) swarms from hydrothermal vents in the eastern Pacific. *Marine Biology* 136, 901–911. doi:10.1007/s002270000300

**Sheader M., Van Dover C. L. and Thurston M. H.** (2004). Reproductive ecology of *Bouvierella curtirama* (Amphipoda: Eusiridae) from chemically distinct vents in the Lucky Strike vent field, Mid-Atlantic Ridge. *Marine Biology* 144, 503–514. doi:10.1007/s00227-003-1211-8

**Shulenberger E. and Barnard J. L.** (1976). Amphipods from an abyssal trap set in the North Pacific Gyre. *Crustaceana* 31, 241–258. doi:10.1163/156854076x00035

**Shulenberger E. and Hessler R. R.** (1974). Scavenging abyssal benthic amphipods trapped under oligotrophic central North Pacific Gyre waters. *Marine Biology* 28, 185–187. doi:10.1007/bf00387296

**Smith K. L. and Baldwin R. J.** (1982). Scavenging deep-sea amphipods - effects of food odor on oxygen-consumption and a proposed metabolic strategy. *Marine Biology* 68, 287–298. doi:10.1007/BF00409595

**Soliman Y. S. and Rowe G. T.** (2008). Secondary production of *Ampelisca mississippiana* Soliman and Wicksten 2007 (Amphipoda, Crustacea) in the head of the Mississippi Canyon, northern Gulf of Mexico. *Deep Sea Research Part II: Topical Studies in Oceanography* 55, 2692–2698. doi:10.1016/j.dsr2.2008.07.019

**Stebbing T. R. R.** (1888). Report on the Amphipoda collected by H.M.S. *Challenger* during the years 1873-76. *Report on the Scientific Results of the Voyage of H.M.S. Challenger During the Years During the Years 1873–76, Zoology* 29, 1737.

**Steele D. H.** (1995). Sexual dimorphism in mature gammaridean amphipods. *Polskie Archiwum Hydrobiologii* 4, 303–317.

**Stockton W. L. and DeLaca T. E.** (1982). Food falls in the deep sea: occurrence, quality, and significance. *Deep Sea Research Part A Oceanographic Research Papers* 29, 157–169. doi:10.1016/0198-0149(82)90106-6

**Stoddart H. E. and Lowry J. K.** (2004). The deep-sea lysianassoid genus *Eurythenes* (Crustacea, Amphipoda, Eurytheneidae n. fam.). *Zoosystema* 26, 425–468.

**Sutcliffe D. W., Carrick T. R. and Willoughby L. G.** (1981). Effects of diet, body size, age and temperature on growth-rates in the amphipod *Gammarus pulex*. *Freshwater Biology* 11, 183–214. doi:10.1111/j.1365-2427.1981.tb01252.x

**Thurston M. H.** (1979). Scavenging abyssal amphipods from the North-East Atlantic Ocean. *Marine Biology* 51, 55–68. doi:10.1007/BF00389031

**Thurston M. H.** (1990). Abyssal Necrophagous Amphipods (Crustacea, Amphipoda) in the Northeast and Tropical Atlantic-Ocean. *Progress in Oceanography* 24, 257–274. doi:10.1016/0079-6611(90)90036-2

**Thurston M. H., Petrillo M. and Della Croce N.** (2002). Population structure of the necrophagous amphipod *Eurythenes gryllus* (Amphipoda: Gammaridea) from the Atacama Trench (south-east Pacific Ocean). *Journal of the Marine Biological Association of the UK* 8, 205–211. doi:10.1017/s0025315402005374

**Treude T., Janssen F., Queisser W. and Witte U.** (2002). Metabolism and decompression tolerance of scavenging lysianassoid deep-sea amphipods. *Deep Sea Research Part I: Oceanographic Research Papers* 49, 1281–1289.

**van Oevelen D., Soetaert K., García R., de Stigter H. C., Cunha M. R., Pusceddu A. and Danovaro, R.** (2011). Canyon conditions impact carbon flows in food webs of three sections of the Nazaré canyon. *Deep Sea Research Part II: Topical Studies in Oceanography* 58, 2461–2476. doi:10.1016/j.dsr2.2011.04.009

**Vetter E. W.** (1995). Detritus-based patches of high secondary production in the nearshore benthos. *Marine Ecology Progress Series* 120, 251–262. doi:10.3354/meps120251

**Weaver P. and Gunn V.** (2009). HERMES: Hotspot Ecosystem Research on the Margins of European Seas. *Oceanography* 22, 12–15. doi:10.5670/oceanog.2009.01

**Wilson R. R., Smith K. L. and Rosenblatt R. H.** (1985). Megafauna associated with bathyal seamounts in the central North Pacific-Ocean. *Deep Sea Research Part A Oceanographic Research Papers* 32, 1243–1254. doi:10.1016/0198-0149(85)90007-X

**Yayanos A. A.** (1981). Reversible inactivation of deep-sea amphipods (*Paralicella Capresca*) by a decompression from 601 bars to atmospheric-pressure. *Comparative Biochemistry and Physiology Part A: Physiology* 69, 563–565. doi:10.1016/0300-9629(81)93020-6

FIGURE LEGENDS

**Fig. 1.** Bathymetric chart of study areas coloured by water depth. Porcupine Abyssal Plain / Seabight and Iberian Margin shown in detail. Circles indicate non-canyon sampling sites and triangles indicate canyon sampling sites. See table 1 for detailed sample collection information.

**Fig. 2.** Representation of coxal plate 4 of *Paralicella caperesca* (redrawn from Shulenberger & Barnard, 1976). Arrowed line indicates the diagonal linear measurement (as in Chapelle, 1995) used in this study as a proxy for total body length.

**Fig. 3.** Comparison between juvenile, male, and female *Paralicella caperesca* from all sampling sites. Estimated total body length, using coxal plate 4 as a proxy, for males, juveniles, and females (A). Length of antennae as a function of coxal plate 4 length for antenna 1 and antenna 2 in male and female *P. caperesca* (B). Median and interquartile ranges shown, closed circles indicate individuals with estimated body length outside the of the interquartile range.

**Fig. 4.** Reproductive characteristics of female *Paralicella caperesca* individuals examined in this study. Oocyte size (A) and number of oocytes in each complete ovary (B) relative to the coxal plate 4 length, a proxy for body length, of each female where oocytes could be identified or complete ovaries were recovered. Oocyte size, calculated as the area of an ellipse, was weakly correlated to coxal plate 4 length (R2 = 0.3946). Number of oocytes showed no correlation with coxal plate 4 length. Probability density histogram for oostegite:gill ratios for all female *P. caperesca* possessing oostegites from combined canyon and non-canyon samples (C). Normal distributions, identified using ‘mixdist’ package of R statistical software, indicated. Thin lines represent the distribution of each oostegite stage, thick line represents entire population.

**Fig. 5.** Probability density histograms of coxal plate 4, a proxy for total body length, measures for all unsexed juvenile (unshaded) and male (shaded) *Paralicella caperesca* from canyon (A) and non-canyon (B) samples. Normal distributions, identified using ‘mixdist’ package of R statistical software, indicated. Thin lines represent distribution of each cohort, thick line represents entire population.

**Fig. 6.** Probability density histograms of coxal plate 4, a proxy for total body length, measures for all unsexed juvenile (unshaded) and female (shaded) *Paralicella caperesca* from canyon (A) and non-canyon (B) samples. Normal distributions, identified using ‘mixdist’ package of R statistical software, indicated. Thin lines represent distribution of each cohort, thick line represents entire population.

TABLES

**Table 1.** Baited trap deployments used *Paralicella caperesca* used for this analysis. Shaded samples are from canyon sites using material from Duffy et al., (2012) with canyon name indicated in area column. Non-shaded samples are from non-canyon sites. Area abbreviations: PSB, Porcupine Seabight; CVP, Cape Verde Abyssal Plain; PAP, Porcupine Abyssal Plain; MAP, Madeira Abyssal Plain.

|  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- |
| **Station** | **Area** | **Location** | | **Depth** | **Date deployed** | **Bottom time** |
|  |  | **Latitude** | **Longitude** | **(m)** | **(dd/mm/yyyy)** | **(hh:mm)** |
| CD179/56817 | Setúbal | 38.1528 | -09.6000 | 3194 | 25/04/06 | 31:14 |
| CD179/56855 | Nazaré | 39.5027 | -09.9050 | 3499 | 11/05/06 | 23:02 |
| D297/15734 | Nazaré | 39.4963 | -09.9648 | 3600 | 01/08/05 | 21:28 |
| CD179/56837 | Cascais | 38.3662 | -09.8834 | 4230 | 02/05/06 | 30:17 |
| D297/15741 | Nazaré | 39.5825 | -10.2750 | 4286 | 04/08/05 | 24:22 |
| CD179/56847 | Nazaré | 39.5917 | -10.3167 | 4403 | 07/05/06 | 24:34 |
| CD179/56839 | Setúbal | 38.1095 | -09.9697 | 4445 | 04/05/06 | 24:12 |
| D092/9756/008 | PSB | 49.8933 | -13.9117 | 3852 | 13/04/79 | 04:15 |
| D204/12600/044 | CVP | 21.0867 | -31.1100 | 4540 | 09/10/93 | 10:45 |
| C085/52216/005 | PAP | 48.8337 | -16.5070 | 4842 | 23/06/85 | 11:05 |
| D229/13200/031 | PAP | 48.8132 | -16.3905 | 4842 | 11/07/97 | 05:22 |
| JC071/034 | PAP | 48.9642 | 16.5020 | 4846 | 05/05/12 | 40:00 |
| JC071/020 | PAP | 49.0048 | -16.4497 | 4847 | 04/05/12 | 17:00 |
| D194/12174/020 | MAP | 31.1267 | -21.1667 | 4941 | 18/08/90 | 15:17 |

**Table 2.** Counts of oostegite stage, identified by oostegite:gill ratios (OGR), for each female cohort of *Paralicella caperesca* from canyon and non-canyon sites.

|  |  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
|  | **Oostegite stage** | **Mean OGR** | **F3** | **F4** | **F5** | **F6** | **F7** | **F8** | **F9** | **F10** |
| **Canyon** | OS0 | - | 3 | 0 | 1 | 1 | 0 | 0 | 0 | 0 |
|  | OS1 | 0.16 | 26 | 14 | 14 | 0 | 0 | 0 | 0 | 0 |
|  | OS2 | 0.40 | 6 | 12 | 25 | 11 | 1 | 0 | 0 | 0 |
|  | OS3 | 0.64 | 1 | 4 | 76 | 112 | 4 | 1 | 1 | 0 |
|  | OS4 | 0.88 | 1 | 0 | 3 | 7 | 1 | 0 | 0 | 0 |
| **Non-canyon** | OS0 | - | 2 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
|  | OS1 | 0.16 | 6 | 57 | 21 | 8 | 7 | 0 | 1 | 0 |
|  | OS2 | 0.40 | 2 | 18 | 30 | 29 | 9 | 2 | 2 | 0 |
|  | OS3 | 0.64 | 0 | 0 | 3 | 51 | 12 | 14 | 26 | 1 |
|  | OS4 | 0.88 | 0 | 0 | 0 | 2 | 3 | 1 | 0 | 0 |

**Table 3.** Mean coxal plate 4 measurement and estimated total body length (TBL) for all cohorts of *Paralicella caperesca* identified in this study from canyon and non-canyon sites. Linear growth factor between successive cohorts in parentheses.

|  |  |  |  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
|  | **Juv. cohorts** | **C4 mean** | **TBL** |  | **Male cohorts** | **C4 mean** | **TBL** |  | **Female cohorts** | **C4 mean** | **TBL** |  |
| **Canyon** |  |  |  |  |  |  |  |  |  |  |  |  |
| J1 | 0.40 | 3.79 |  |  |  |  |  |  |  |  |  |
|  |  |  | (1.44) |  |  |  |  |  |  |  |  |
| J2 | 0.64 | 5.45 |  |  |  |  |  |  |  |  |  |
|  |  |  | (1.27) |  |  |  |  |  |  |  |  |
| J3 | 0.85 | 6.90 |  | M3 | 0.85 | 6.90 |  | F3 | 0.86 | 6.95 |  |
|  |  |  | (1.22) |  |  |  | (1.22) |  |  |  | (1.26) |
|  | J4 | 1.07 | 8.41 |  | M4 | 1.07 | 8.41 |  | F4 | 1.11 | 8.73 |  |
|  |  |  |  |  |  |  |  | (1.22) |  |  |  | (1.19) |
|  |  |  |  |  | M5 | 1.33 | 10.25 |  | F5 | 1.36 | 10.41 |  |
|  |  |  |  |  |  |  |  | (1.33) |  |  |  | (1.17) |
|  |  |  |  |  | M6 | 1.83 | 13.64 |  | F6 | 1.61 | 12.16 |  |
|  |  |  |  |  |  |  |  | (1.09) |  |  |  | (1.24) |
|  |  |  |  |  | M7 | 2.00 | 14.84 |  | F7 | 2.04 | 15.12 |  |
|  |  |  |  |  |  |  |  |  |  |  |  | (1.18) |
|  |  |  |  |  | M8 | - |  |  | F8 | 2.43 | 17.78 |  |
|  |  |  |  |  |  |  |  |  |  |  |  | (1.08) |
|  |  |  |  |  |  |  |  |  | F9 | 2.63 | 19.16 |  |
|  |  |  |  |  |  |  |  |  |  |  |  |  |
|  |  |  |  |  |  |  |  |  | F10 | - |  |  |
| **Non-canyon** |  |  |  |  |  |  |  |  |  |  |  |  |
| J1 | 0.44 | 4.07 |  |  |  |  |  |  |  |  |  |
|  |  |  | (1.32) |  |  |  |  |  |  |  |  |
| J2 | 0.62 | 5.36 |  |  |  |  |  |  |  |  |  |
|  |  |  | (1.20) |  |  |  |  |  |  |  |  |
| J3 | 0.78 | 6.43 |  | M3 | - |  |  | F3 | - |  |  |
|  |  |  | (1.21) |  |  |  |  |  |  |  |  |
| J4 | 0.98 | 7.80 |  | M4 | 1.07 | 8.44 |  | F4 | 1.03 | 8.19 |  |
|  |  |  |  |  |  |  | (1.17) |  |  |  | (1.22) |
|  |  |  |  |  | M5 | 1.28 | 9.90 |  | F5 | 1.30 | 10.02 |  |
|  |  |  |  |  |  |  |  | (1.29) |  |  |  | (1.15) |
|  |  |  |  |  | M6 | 1.70 | 12.77 |  | F6 | 1.52 | 11.53 |  |
|  |  |  |  |  |  |  |  | (1.12) |  |  |  | (1.20) |
|  |  |  |  |  | M7 | 1.91 | 14.24 |  | F7 | 1.85 | 13.81 |  |
|  |  |  |  |  |  |  |  | (1.12) |  |  |  | (1.08) |
|  |  |  |  |  | M8 | 2.15 | 15.90 |  | F8 | 2.02 | 14.98 |  |
|  |  |  |  |  |  |  |  |  |  |  |  | (1.12) |
|  |  |  |  |  |  |  |  |  | F9 | 2.27 | 16.72 |  |
|  |  |  |  |  |  |  |  |  |  |  |  | (1.19) |
|  |  |  |  |  |  |  |  |  | F10 | 2.73 | 19.84 |  |

**Table 4.** Total counts of males, females, and juveniles for all samples. Population composition ratios used for univariate comparison shown.

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| **Station** | **Total Males** | **Total Females** | **Juveniles** | **M:F ratio** | **Mature:Juv. Ratio** |
| CD179/56817 | 22 | 13 | 28 | 1.69 | 1.25 |
| CD179/56855 | 27 | 18 | 35 | 1.50 | 1.29 |
| D297/15734 | 81 | 4 | 81 | 20.25 | 1.05 |
| CD179/56837 | 80 | 33 | 400 | 2.42 | 0.28 |
| D297/15741 | 8 | 12 | 180 | 0.67 | 0.11 |
| CD179/56847 | 38 | 19 | 143 | 2.00 | 0.40 |
| CD179/56839 | 80 | 33 | 400 | 2.42 | 0.28 |
| D092/9756/008 | 71 | 46 | 35 | 1.54 | 3.34 |
| D204/12600/044 | 21 | 14 | 127 | 1.50 | 0.28 |
| C085/52216/005 | 55 | 17 | 258 | 3.24 | 0.28 |
| D229/13200/031 | 20 | 23 | 41 | 0.87 | 1.05 |
| JC071/034 | 46 | 64 | 88 | 0.72 | 1.25 |
| JC071/020 | 33 | 58 | 109 | 0.57 | 0.83 |
| D194/12174/020 | 21 | 13 | 16 | 1.62 | 2.13 |