### Accepted Manuscript

Seasonal cycle of  ${\rm CO}_2$  from the sea ice edge to island blooms in the Scotia Sea, Southern Ocean

Elizabeth M. Jones, Dorothee C.E. Bakker, Hugh J. Venables, Nick J. Hardman-Mountford

PII: S0304-4203(15)30016-5

DOI: doi: 10.1016/j.marchem.2015.06.031

Reference: MARCHE 3285

To appear in: Marine Chemistry

Received date: 21 January 2015 Revised date: 28 June 2015 Accepted date: 30 June 2015



Please cite this article as: Jones, Elizabeth M., Bakker, Dorothee C.E., Venables, Hugh J., Hardman-Mountford, Nick J., Seasonal cycle of CO<sub>2</sub> from the sea ice edge to island blooms in the Scotia Sea, Southern Ocean, *Marine Chemistry* (2015), doi: 10.1016/j.marchem.2015.06.031

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

Seasonal cycle of CO<sub>2</sub> from the sea ice edge to island blooms in the Scotia Sea, Southern Ocean

Elizabeth M. Jones<sup>1,2</sup>\*, Dorothee C. E. Bakker<sup>1</sup>, Hugh J. Venables<sup>3</sup>, Nick J. Hardman-Mountford<sup>4</sup>

<sup>&</sup>lt;sup>1</sup>Centre for Ocean and Atmospheric Sciences, School of Environmental Sciences, University of East Anglia, Norwich NR4 7TJ, UK

<sup>&</sup>lt;sup>2</sup>Centre for Energy and Environmental Sciences, Energy and Sustainability Research Institute Groningen, University of Groningen, Nijenborgh 4, 9747 AG Groningen, The Netherlands

<sup>&</sup>lt;sup>3</sup>British Antarctic Survey, High Cross, Madingley Road, Cambridge CB3 0ET, UK

<sup>&</sup>lt;sup>4</sup>Oceans and Atmosphere Flagship, Commonwealth Scientific and Industrial Research Organisation, Floreat WA 6014, Australia

<sup>\*</sup>Corresponding author email: e.m.jones@rug.nl

#### **Abstract**

The Scotia Sea region contains some of the most productive waters of the Southern Ocean. It is also a dynamic region through the interaction of deep water masses with the atmosphere. We present a first seasonally-resolved time series of the fugacity of CO<sub>2</sub> (fCO<sub>2</sub>) from spring 2006, summer 2008, autumn 2009 and winter (potential temperature minimum) along a 1000 km transect from the pack ice to the Polar Front to quantify the effects of biology and temperature on oceanic fCO<sub>2</sub>. Substantial spring and summer decreases in sea surface fCO<sub>2</sub> occurred in phytoplankton blooms that developed in the naturally ironfertilised waters downstream (north) of South Georgia island (54-55°S, 36-38°W) and following sea ice melt (in the seasonal ice zone). The largest seasonal  $fCO_2$  amplitude ( $\Delta fCO_2$ ) of -159  $\mu$ atm was found in the South Georgia bloom. In this region, biological carbon uptake dominated the seasonal signal, reducing the winter maxima in oceanic fCO<sub>2</sub> by 257 µatm by the summer. In the Weddell-Scotia Confluence, the southern fringe of the Scotia Sea, the shift from wintertime CO<sub>2</sub>-rich conditions in ice covered waters to CO<sub>2</sub> undersaturation in the spring blooms during and upon sea ice melt created strong seasonality in oceanic  $fCO_2$ . Temperature effects on oceanic  $fCO_2$  ranged from  $\Delta fCO_2$  sst of ~55 µatm in the seasonal ice zone to almost double that downstream of South Georgia (98 µatm). The seasonal cycle of surface water fCO<sub>2</sub> in the high-nutrient low-chlorophyll region of the central Scotia Sea had the weakest biological control and lowest seasonality. Basin-wide biological processes dominated the seasonal control on oceanic fCO<sub>2</sub> (ΔfCO<sub>2 bio</sub> of 159 μatm), partially compensated (43%) by moderate temperature control  $(\Delta f CO_{2 \text{ sst}})$  of 68 µatm). The patchwork of productivity across the Scotia Sea creates regions of seasonally strong biological uptake of CO<sub>2</sub> in the Southern Ocean.

#### Key words

Oceanic carbon dioxide
Scotia Sea, Southern Ocean
Sea ice
South Georgia bloom
Biological carbon uptake

#### 1 Introduction

The Southern Ocean plays a key role in the global carbon cycle through the interaction of upper ocean processes with atmospheric carbon dioxide (CO<sub>2</sub>) and the ventilation of deep waters (Watson and Orr, 2003; Naveira Garabato et al., 2004; Takahashi et al., 2009). However, oceanic CO<sub>2</sub> data are sparse in the Southern Ocean and the magnitude and variability of seasonal and annual atmospheric CO<sub>2</sub> uptake remains relatively unknown compared to other areas of the global ocean (Metzl et al., 2006; Lenton et al., 2013; Landschützer et al., 2014). The Scotia Sea, in the Atlantic sector of the Southern Ocean, is an important component of the overturning circulation (Locarnini et al., 1993; Naveira Garabato et al., 2002; Naveira Garabato et al., 2004; Meredith et al., 2008). The small ocean basin encompasses diverse Southern Ocean ecosystems, from a seasonal ice zone to oceanic islands, where the movement of major water masses and presence of fronts of the Antarctic Circumpolar Current (ACC) generates rich biological diversity (Holm-Hansen et al., 2004; Murphy et al., 2007) and extensive biogeochemical cycling. Sea ice covers the southern Scotia Sea during winter and has a variable northward extent into the central part of the basin. The seasonal advance and retreat of sea ice coupled with frontal meandering induces connective biogeochemical pathways from southern regions (e.g., the Weddell Sea and Antarctic Peninsula), to the north (Murphy et al., 2007) contributing to the basin-wide biological diversity (Atkinson et al., 2001; Meredith et al., 2008). Limited data in the region have revealed that the patchwork of productivity creates areas of contrasting sea surface CO2 concentrations, from spring and summer CO2 super-saturation in largely un-productive waters and beneath sea ice cover (Shim et al., 2006; Jones et al., 2012) to substantial biological carbon uptake within intense seasonal blooms (Jones et al., 2010; Jones et al., 2012). The large blooms in the Scotia Sea region can contribute to sequestration of atmospheric CO<sub>2</sub> and export of carbon to the deep ocean (Schlitzer, 2002; Pollard et al., 2009; Takahashi et al., 2009; Korb et al., 2012).

The Scotia Sea is bound to the north, south and east by the Scotia Ridge, a sub-marine arc with island protrusions at the South Orkney Islands, on the South Scotia Ridge, and the island of South Georgia on the North Scotia Ridge (Fig. 1). It is a dynamic region due to the ventilation and mixing of water masses in the ACC through the interactions with rugged bathymetry and wind stress (Naveira Garabato et al., 2004). The ACC is split by a series of frontal jets, which are identified, from north to south, as the Sub-Antarctic Front (SAF), Polar Front (PF), the Southern ACC Front (SACCF) and the Southern Boundary (SB) (Orsi et al., 1995). The Polar Front passes through the western part of the Scotia Sea and leaves the

Scotia Sea for the Georgia Basin across the North Scotia Ridge (Smith et al., 2010; Venables et al., 2012). The SACCF passes through the central Scotia Sea before retroflecting around the eastern shelf of the North Scotia Ridge and into the Georgia Basin (Trathan et al., 1997; Thorpe et al., 2002). Mesoscale activity associated with the SACCF can lead to variability in hydrographic and biogeochemical parameters across the Scotia Sea (Boehme et al., 2008; Whitehouse et al., 2012), with the front located in close proximity (~50 km) to the SB (Venables et al., 2012). Circumpolar Deep Water upwells and outcrops at the surface towards the southern limit of the ACC at the SACCF and SB (Orsi et al., 1995; Pollard et al., 2002; Boehme et al., 2008). The SB is the southernmost front in the region and marks the limit of the ACC and the boundary of the Weddell-Scotia Confluence, a transition zone between the Scotia Sea to the north and Weddell Sea to the south as waters flow off the tip of the Antarctic Peninsula and from the Weddell Sea and infiltrate the ACC (Patterson and Sievers, 1980; Whitworth et al., 1994; Meredith et al., 2008; Jullion et al., 2014).

A wide range of productivity regimes occur across the Scotia Sea against a back-drop of high-nutrient low-chlorophyll (HNLC) conditions common to much of the remote Southern Ocean (Park et al., 2010; Whitehouse et al., 2012). Some of the longest lasting phytoplankton blooms in the ACC develop annually in the Georgia Basin, downstream (north) of the island of South Georgia (Atkinson et al., 2001; Korb and Whitehouse, 2004; Borrione and Schlitzer, 2013). Natural enrichment of the limiting micronutrient iron can occur from South Georgia, shelves and shallow sediments of the North Scotia Ridge (Holeton et al., 2005; Nielsdóttir et al., 2012), promoting high atmospheric CO<sub>2</sub> uptake (Jones et al., 2012). In the seasonal ice zone, blooms are frequently linked to the retreat of the ice edge (Lancelot et al., 1993; Nielsdóttir et al., 2012) and create transient and substantial CO<sub>2</sub> sinks (Jones et al., 2010). By contrast, the central Scotia Sea displays considerable spatial and temporal variability in phytoplankton biomass, where extensive blooms in one year can be followed by near-HNLC conditions in the next (Whitehouse et al., 2008; Park et al., 2010; Korb et al., 2012; Whitehouse et al., 2012). Alleviation of HNLC conditions is through iron enrichment (de Baar et al., 1995; Boyd et al., 2007) under favourable light conditions, where iron sources to surface waters in the Scotia Sea include interaction of the ACC with shallow topography, e.g., the Scotia Arc (Korb and Whitehouse, 2004; Holm-Hansen et al., 2004; Meskhidze et al., 2007; Nielsdóttir et al., 2012), melting sea ice (de Baar and de Jong, 2001; Nielsdóttir et al., 2012) and advection within the SACCF and SB from an upstream source, e.g., the Shackleton Fracture Zone and Antarctic Peninsula shelf (Ardelan et al., 2010; Zhou et al., 2010).

The dynamic hydrographic and biological regimes of the Scotia Sea create a natural 'mesocosm' to investigate the seasonal cycle of oceanic CO<sub>2</sub>. Quantifying the processes that control the cycling of CO<sub>2</sub> across several typical Southern Ocean environments is fundamental to better determine the role of the polar oceans in the global carbon cycle. While the biological and biogeochemical processes highlighted in the preceding paragraph have been described, the quantification of their influence on CO<sub>2</sub> dynamics in the Scotia Sea region is limited to a few localised observations in spring and summer (Shim et al., 2006; Jones et al., 2012). This study resolves for the first time the relative contribution of biological and physical (principally temperature) controls as drivers of variability in oceanic CO<sub>2</sub> across the Scotia Sea between different seasons. We present a new time series of oceanic CO<sub>2</sub> from a ~1000 km meridional transect during spring 2006, summer 2008, autumn 2009 and winter (potential temperature minimum). Data extend from the seasonal pack ice in the Weddell-Scotia Confluence to the large phytoplankton blooms downstream (north) of South Georgia and up to the Polar Front. The oceanic context is set by considering the basin-scale spatial and temporal distribution and variability of phytoplankton blooms and sea ice cover, as determined by satellite measurements during the 2006-2009 study period. Finer resolution shipboard measurements of hydrographic variables, macronutrients and chlorophyll-a are considered to identify the physical (temperature, salinity, sea ice cover, mixed layer depth) and biological controls that drive the seasonal variability oceanic CO2 in the Scotia Sea. The data highlight the seasonal dynamics as key to understanding the carbon cycle of the polar oceans.

#### 2 Methods

### 2.1 Oceanographic sampling

Data were collected in the Scotia Sea, Atlantic sector of the Southern Ocean, during three cruises onboard RRS *James Clark Ross* in spring, summer and autumn of consecutive years as part of the British Antarctic Survey DISCOVERY-2010 program (Tarling et al. 2012). A ~1000 km meridional transect that extended from south of the South Orkney Islands to the Polar Front (Fig. 1) was occupied during austral (1) spring (JR161; 24.10.06-02.12.06), (2) summer (JR177; 31.12.07-14.02.08) and (3) autumn (JR200; 12.03.09-15.04.09). The depth of the mixed layer is defined as the depth at which the potential density exceeds that measured at 10 m by 0.05 kg m<sup>-3</sup> (Brainerd and Gregg, 1995). Mixed layer depths (mld) were determined from 2 dbar resolution depth profiles of the conductivity, temperature, depth (CTD) sensor (Seabird 911+) deployment at each hydrographic station (Venables et al., 2012). Positions of the

SB, SACCF and PF are expressed as latitudinal ranges based on absolute dynamic sea-surface height during each cruise (Venables et al., 2012). Sea surface temperature and salinity data were obtained from the underway seawater supply (Oceanlogger SeaBird Electronics SBE45 CTD) with an intake at 6.5 m depth. Salinity values are reported on the practical salinity scale. Samples for chlorophyll-a analysis (200-500 mL) were filtered through glass fibre filters (0.7 µm poresize GF/F, Fisherbrand), frozen at –20°C and then extracted into 10 mL 90% acetone for 24 hours before analysis on a fluorometer (Turner Designs TD-700). A detailed description of the chlorophyll-a method is presented in Korb and Whitehouse (2004).

#### 2.2 Sea surface CO<sub>2</sub>

### 2.2.1 Seasonal observations and CO<sub>2</sub> disequilibrium

Measurements of the fugacity of CO<sub>2</sub> (fCO<sub>2</sub>) in seawater and air were made during each cruise using a PML/ Dartcom Live CO<sub>2</sub> system (Hardman-Mountford et al., 2008), based on principles outlined in Cooper et al. (1998). The  $CO_2$  disequilibrium ( $\Delta fCO_2$ ) is the difference between the  $fCO_2$  in surface water and overlying air, where negative values represent CO<sub>2</sub> undersaturation with respect to the atmosphere. Atmospheric samples were taken from an inlet located forward at 15 m height of the RRS James Clark Ross. Mixing ratios of CO<sub>2</sub> and water were determined by infra-red detection with a LI-COR 840. The LI-COR was calibrated using secondary gas standards with nominal CO<sub>2</sub> concentrations of 250 ppm and 450 ppm in artificial air and nitrogen was the zero reference gas. All gases underwent pre- and post-cruise calibration against certified primary standards from the National Oceanic and Atmospheric Administration (NOAA) at Plymouth Marine Laboratory. Samples from the equilibrator headspace and marine air were partially dried in a condenser prior to analysis in the LI-COR. The fCO<sub>2</sub> was computed from the mixing ratios and the ship's barometric pressure and then corrected for seawater vapour pressure (Weiss and Price, 1980). Two platinum resistance thermometers located in the upper and lower part of the equilibrator determined the average warming (all measurements) of the seawater between the intake and the equilibrator as 0.6 °C ( $\sigma = 0.3$  °C; n = 3,568). Sea surface  $fCO_2$  data were corrected to sea surface temperature to account for this warming (Takahashi et al., 1993). The accuracy of the fCO<sub>2</sub> data is determined at 3 µatm from an instrument intercomparison carried out during cruise JR177 (Jones, 2010). The precision is estimated as better than 2 µatm.

Concentrations of dissolved inorganic carbon and alkalinity were measured during summer 2008 (cruise JR177) at the depth of the potential temperature minimum ( $\theta_{min}$ ) for each hydrographic station; a full

description of sample collection, analysis and data is presented in Jones et al. (2012). The potential temperature minimum represents the Winter Water (Jennings et al., 1984), the remnant of the mixed layer from the preceding winter, i.e., 2007. Parameters measured in the Winter Water during summer 2008 are taken to reflect surface conditions during winter 2007 and were used to calculate sea surface  $fCO_2$  with the CO2Sys programme (Lewis and Wallace, 1998; van Heuven et al., 2011) using thermodynamic dissociation constants for  $K_1$  and  $K_2$  by Mehrbach et al. (1973), re-fitted by Dickson and Millero (1987). The accuracy of calculated  $fCO_2$  is better than 6  $\mu$ atm (Millero, 1995). Thus, winter ( $\theta_{min}$ )  $fCO_2$  data is used to complement the spring 2006, summer 2008 and autumn 2009 sea surface  $fCO_2$  seasonal time series.

#### 2.2.2 Biological and physical CO<sub>2</sub> signals

The seasonal changes in sea surface  $fCO_2$  can be separated into biological and physical (temperature) signals (Takahashi et al., 2002). Other processes affecting oceanic  $CO_2$ , such as air-sea  $CO_2$  exchange, advection, mixing, upwelling and changes in alkalinity, e.g., from calcium carbonate precipitation (calcification) and dissolution, are not accounted for independently here and are incorporated into the 'biological' signal. To remove the effects of temperature, sea surface  $fCO_2$  data measured at observed temperature (sst<sub>obs</sub>) are normalised to the average sea surface temperature (sst<sub>ave</sub>, 1.52 °C) of spring, summer and autumn and winter ( $\theta_{min}$ ) along the transect:

$$fCO_2 at \, sst_{ave} = fCO_2 at \, sst_{obs} \times \exp\left(0.0423 \left(sst_{ave} - sst_{obs}\right)\right) \tag{1}$$

The temperature signal in sea surface  $fCO_2$  is determined from the mean  $fCO_2$  of spring, summer, autumn and winter ( $fCO_2$  ave) along the transect and the difference between sst<sub>obs</sub> and sst<sub>ave</sub>:

$$fCO_2 at sst_{obs} = fCO_{2ave} \times \exp(0.0423 \left(sst_{obs} - sst_{ave}\right))$$
(2)

This approach assumes that the observed field data cover the maxima and minima of the seasonal variations in sea surface temperature and  $fCO_2$ , which has limitations for this basin-wide study with large spatial variability. This is addressed here by using data at the temperature minimum (as a proxy for winter sea surface data) and by interpreting the seasonal changes (amplitudes) within a wider context set by satellite data. Seasonal amplitudes in oceanic  $fCO_2$  due to biological ( $\Delta fCO_2$  bio) and temperature ( $\Delta fCO_2$ 

sst) effects are obtained from the difference between the maximum and minimum values in  $fCO_2$  at  $sst_{ave}$  and  $fCO_2$  at  $sst_{obs}$ , respectively, from all spring 2006, summer 2008, autumn 2009 and winter ( $\theta_{min}$ ) data:

$$\Delta fCO_{2\ bio} = \left(fCO_2\ at\ sst_{ave}\right)_{max} - \left(fCO_2\ at\ sst_{ave}\right)_{min} \tag{3}$$

$$\Delta fCO_{2 sst} = \left(fCO_2 \ at \ sst_{obs}\right)_{max} - \left(fCO_2 \ at \ sst_{obs}\right)_{min} \tag{4}$$

The influence of the biological and temperature controls on seawater  $fCO_2$  is investigated by comparison of the difference ( $\Delta fCO_{2 \, sst} - \Delta fCO_{2 \, bio}$ ) and ratio ( $\Delta fCO_{2 \, sst} / \Delta fCO_{2 \, bio}$ ) of the two expressions. In regions of high biological activity and/ or strong seasonal variability in biological activity the difference ( $\Delta fCO_{2 \, sst} - \Delta fCO_{2 \, bio}$ ) is negative and the ratio ( $\Delta fCO_{2 \, sst} / \Delta fCO_{2 \, bio}$ ) is less than 1. In regions of low biological activity and/ or weak seasonal variability in biological activity the difference is positive and the  $\Delta fCO_2$  ratio is greater than 1 (Thomas et al., 2005).

### 2.3 Satellite chlorophyll-a and sea ice cover

Satellite ocean colour from SeaWiFS (2006-2007) and MODIS-Aqua (2006-2009) were obtained from http://oceancolor.gsfc.nasa.gov/ (July 2009) as 8 day, 9 km level 3 mapped data (Feldman and McClain, 2006a; Feldman and McClain, 2006b). Satellite derived concentrations of chlorophyll-a (chl-a) are used as an indicator of spatio-temporal variation in phytoplankton biomass. Bloom conditions are defined as having chlorophyll-a concentrations exceeding 0.75 mg chl-a m<sup>-3</sup> (Korb et al., 2012) and HNLC conditions as less than 0.2 mg m<sup>-3</sup> in the presence of high macronutrient concentrations (Whitehouse et al., 2012). The 8 day composite images were used to determine the initiation and termination of the blooms. Bloom progression could be more easily tracked using the multi-day composites rather than daily images, which were strongly affected by cloud cover. The multi-day composites maintained a sufficient temporal resolution.

Sea ice concentration data at 4 km resolution were obtained through the Operational Sea Surface Temperature and Sea Ice Analysis (OSTIA) service (http://ghrsst-pp.metoffice.com/pages/latest\_analysis/ostia.html; December 2009), part of the Group for High-Resolution Sea Surface Temperature (Stark et al., 2007). The ice edge is defined here as the terminus of consolidated sea ice along the transect, expressed as 0.5 fractional (50%) sea ice cover during each

seasonal cruise. The seasonal ice zone (SIZ) is the region between the maximum and minimum sea ice extent based on monthly fractional sea ice cover during the study period 2006-2009.

### 3 Results

### 3.1 Bloom dynamics: seasonal and year-to-year variations

The SeaWiFS and MODIS-Aqua time series (Fig. 2) shows high spatial and temporal variability in surface chlorophyll-a in the Scotia Sea (55-59.3°S, 25-55°W) and Georgia Basin (49.5-53.0°S, 25-45°W) regions throughout the 3-year study period. A general seasonal cycle can be depicted: chlorophyll-a patches appeared during spring (September, October, November), sporadic chlorophyll-a maxima occurred in the southern and central Scotia Sea in both spring and summer, followed by peaks in the Georgia Basin in summer (December, January, February), in some years persisting until autumn (March, April, May). The northern Scotia Sea regularly exhibited HNLC-type characteristics as evident from sea surface nitrate and phosphate concentrations measured during each cruise, reported in Whitehouse et al. (2012). Sea ice obscured part of the southern basin from satellite detection during the winter, while shorter daylight hours also limited data coverage (June, July, August) (hence not shown in Fig. 2).

Each phytoplankton bloom in the Scotia Sea and the Georgia Basin is ranked in terms of areal extent and longevity to give a relative 'strength' as a product of these two features (Table 1). Blooms show high inter-annual variability in extent and duration in the Scotia Sea (Fig. 3) and seasonality. In 2006/2007 a huge bloom lasted for up to 6 months and extended across the central Scotia Sea (rank product 1) (Figs. 2 and 3). Extensive blooms in the Georgia Basin occur annually, often persisting for up to 6 months from spring until autumn, peaking in intensity during the summer (rank products 2,3,4).

#### 3.2 The seasonal ice zone of the Scotia Sea

The seasonal advance and retreat of the sea ice from 2006 to 2009 can be observed from satellite derived sea ice cover to locate the position of the sea ice edge (Fig. 4). The ice typically originates in the Weddell Sea and drifts and extends northwards into the Scotia Sea during winter (Murphy et al., 1995). During February and March each year the region was consistently ice free. Inter-annual variation in the timing of the sea ice advance and retreat and maximum coverage is evident. The maximum extent of sea ice, defined as the northward limit of the sea ice cover where ice is present for the whole month, reached 57°S in September 2007; defining the northward limit of the seasonal ice zone (SIZ) for the 2006/2009 study

period. Summer-autumn 2007/2008 had a short ice free period of only 2 months (February to March) in the Scotia Sea. This was followed by relatively early seasonal sea ice advance in April 2008, making 2007/2008 a relatively 'icy' period. By contrast, sea ice had largely retreated from the Scotia Sea by December 2008, thus 2008/2009 was relatively ice free.

### 3.3 Sub-regions of the Scotia Sea

Combining remotely sensed chlorophyll-a (Fig. 2) data with sea surface hydrographic and biogeochemical data (Fig. 5), macronutrient distributions (Whitehouse et al., 2012) and frontal boundaries (Orsi et al., 1995; Venables et al., 2012), eight sub-regions were distinguished along the meridional transect for the 2006-2009 study period:

- 1. Weddell-Scotia Confluence (WSC; 62.6-59.3°S) includes the southern limit of the transect; the coldest waters and those south of the ACC, i.e., south of the SB; main part of the SIZ annually.
- 2. Southern Boundary (SB; 59.3-57.7°S) defined as the southern limit of the ACC as observed from the disappearance of Circumpolar Deep Water in potential temperature profiles during each cruise; regularly within the SIZ.
- 3. Southern ACC front (SACCF; 57.7-57.2°S) defined as the southern extent of the sub-surface 1.8°C potential temperature contour from potential temperature profiles during each cruise; regularly within the SIZ.
- 4. HNLC waters (57.2-56.0°S) defined by persistent high concentrations of nitrate and phosphate of 25-30 mmol m<sup>-3</sup> and 1.5-2.0 mmol m<sup>-3</sup> (Whitehouse et al., 2012), respectively. Chlorophyll-a concentrations are often less than 0.2 mg m<sup>-3</sup> as determined from satellite and shipboard measurements; rarely within the SIZ.
- 5. Northern Scotia Sea (nSS; 56.0-55.0°S) defined as deep (> 3000 m) waters up to the North Scotia Ridge; north of the SIZ.
- 6. North Scotia Ridge (NSR; 55.0-53.0°S) defined as shelf waters (< 3000 m) across the ridge.
- 7. Georgia Basin (GB; 53.0-50.5°S) defined as the deep (> 3000 m) basin northwest of the island of South Georgia.
- 8. Polar Front (PF; 50.5-49.5°S) defined as the northern limit of the sub-surface potential temperature minimum as observed from potential temperature profiles during each cruise; represents the northern limit of the meridional transect.

#### 3.4 Seasonal CO<sub>2</sub> disequilibrium

The degree of  $CO_2$  disequilibrium ( $\Delta fCO_2$ ) in seawater (Fig. 6) is regulated by biological production, temperature, salinity, sea ice cover and depth of the mixed layer. Temperature exerts a strong control on seawater  $fCO_2$  (Fig. 7a) through changing the solubility of the gas in seawater (Weiss and Price, 1980). A normalisation of the seasonal  $fCO_2$  time series to mean sea surface temperature has been made to remove the effects of temperature, following the empirical relationship of Takahashi et al. (2002).

Seasonal seawater  $fCO_2$  minima in spring and summer are due to photosynthetic  $CO_2$  uptake in phytoplankton blooms upon retreat of sea ice and in the naturally iron-fertilised waters downstream (north) of South Georgia island (Fig. 7c). Springtime maximum (positive) and minimum (negative)  $CO_2$  disequilibrium occurred at the southern end of the transect  $\sim 60^{\circ}$ S (Fig. 6a). High  $CO_2$  disequilibrium of 62  $\mu$ atm in cold, saline and deep mixed layers (80 m) occurred beneath sea ice (sea ice fraction 0.5-1.0) present around the South Orkney Islands (Fig. 5). In contrast,  $CO_2$  disequilibrium of -82  $\mu$ atm at  $\sim 57.5^{\circ}$ S coincided with high chlorophyll-a values of 7.9 mg m<sup>-3</sup> (Fig. 5b) in cool, less saline and shallower mixed layer (50 m) surface waters (Fig. 5c-e); symptomatic of recent sea ice melt. Springtime surface water  $fCO_2$  varied significantly with chlorophyll-a concentrations ( $r^2 = 0.79$ , m = -12.9, p < 0.001, n = 46) and salinity, an indicator of sea ice melt, ( $r^2 = 0.56$ , m = 289.1, p < 0.001, n = 781) (Fig. 7b-c). The location of the sea ice edge during spring was around 59°S (Fig. 5f). Seasonal mixed layers of 100-120 m (Fig. 5e) in the north Scotia Sea and the North Scotia Ridge had sea surface  $fCO_2$  close to atmospheric values (Fig. 5a) with very low chlorophyll-a (Fig. 5b). At the northern limit of the springtime transect,  $CO_2$  disequilibrium shifted to -50  $\mu$ atm in relatively warm surface waters of the Georgia Basin and the PF, where chlorophyll-a values were around 2 mg m<sup>-3</sup> (Fig. 5b).

The summertime distribution of  $fCO_2$  was dominated by intense undersaturation in the Weddell-Scotia Confluence, northern Scotia Sea regions (nSS and NSR) and Georgia Basin (Fig. 5a, 6b). A swift transition from  $fCO_2$  saturation to substantial undersaturation of -100 µatm in the Weddell-Scotia Confluence (Fig. 5a) coincided with the transformation from cold, salty waters beneath sea ice to less saline, stratified surface waters with chlorophyll-a values of 3.5 mg m<sup>-3</sup> (Fig. 5b-d) upon extensive sea ice melt (Fig. 5f). These trends are reflected in significant summertime sea surface  $fCO_2$  correlation with chlorophyll-a concentrations ( $r^2 = 0.38$ , m = -13.8, p < 0.001, n = 180) and salinity ( $r^2 = 0.24$ , m = 117.2, p < 0.001, n = 714) for all data (Fig. 7b-c). The summertime ice edge was located around 62°S (Fig. 5f).

A summer minimum in  $fCO_2$  undersaturation of -116 µatm was measured alongside very high concentrations of chlorophyll-a over 10 mg m<sup>-3</sup> in the relatively warm, stratified waters of the Georgia Basin (Fig. 5). By contrast, the central Scotia Sea (SACCF and HNLC) was moderately supersaturated as  $CO_2$  disequilibrium approached 20 µatm with deeper mixed layers of about 80 m in the presence of low chlorophyll-a (Figs. 5a-b and 5e).

Surface waters had a  $fCO_2$  range between 310  $\mu$ atm and 400  $\mu$ atm for the entire transect during autumn (Figs. 5 and 7). Disequilibrium of  $CO_2$  changed from oversaturation (20  $\mu$ atm) in relatively warm and saline waters of the central Scotia Sea, sub-regions SACCF and HNLC (Fig. 7a-b) to undersaturation (-50  $\mu$ atm) in the Georgia Basin, where waters were fresher and chlorophyll-a values were high at 5 mg m<sup>-3</sup>. Such surface water patterns created a significant and moderately strong relationship between autumnal  $fCO_2$  and salinity ( $r^2 = 0.67$ , m = 111.36, p < 0.001, n = 684). A second area of  $fCO_2$  undersaturation (-40  $\mu$ atm) and smaller chlorophyll-a peak (2.5 mg m<sup>-3</sup>) occurred near the South Orkney Islands, contributing to a weak but significant trend between  $fCO_2$  and chlorophyll-a ( $r^2 = 0.12$ , m = -8.9, p < 0.001, n = 131) during the autumn (Fig. 7b-c).

Oceanic  $fCO_2$  in Winter Water ( $\theta_{min}$  data) indicate that sea surface  $fCO_2$  during the winter is mostly supersaturated with respect to atmospheric  $CO_2$  (Fig. 5a). High values between 380-450 µatm coincide with high salinities (> 34.2) and low temperatures (<  $-1.35^{\circ}C$ ) beneath almost total ice cover in the Weddell-Scotia Confluence (Figs. 5 and 7). Winter sea surface  $fCO_2$  maxima (440-455 µatm) occurred in deep mixed layers (120-130 m) in the Georgia Basin (Fig. 5). No winter data are available for the Polar Front region.

The strongest seasonal biological signals ( $\Delta f CO_{2 \text{ bio}}$ ) of 257  $\mu$ atm and 196  $\mu$ atm occurred in the Georgia Basin and Weddell-Scotia Confluence, respectively (Fig. 8a). This reflects the oceanic  $f CO_2$  changes due to seasonal photosynthetic uptake of  $CO_2$  in these regions. Temperature control ( $\Delta f CO_2$  sst) had comparatively little variation along the meridional transect with a slightly greater effect in the north compared to the south (Fig. 8a). Strong seasonal warming driving strong thermodynamic changes in oceanic  $CO_2$  near the Polar Front has been previously observed along a south-north transect at 52°W in the Scotia Sea (Shim et al., 2006). The southern sub-regions lie within the seasonal ice zone where partial sea ice cover is still present during spring and summer (Fig. 4) and this reduces the time for sea surface warming. The northern sub-regions experience greater seasonal warming (Fig. 5d), where the maximum

temperature control of 98  $\mu$ atm occurred in the Georgia Basin (Fig. 8a). The HNLC waters in the central Scotia Sea displayed the lowest biological control of 97  $\mu$ atm and weakest net seasonal variability of the whole region with a  $\Delta f$ CO<sub>2</sub> ratio ( $\Delta f$ CO<sub>2</sub> sst /  $\Delta f$ CO<sub>2</sub> bio) of 0.7 (Fig. 8b).

- 4 Discussion
- 4.1 Strong CO<sub>2</sub> dynamics in the seasonal ice zone

Large seasonal fCO<sub>2</sub> amplitude is observed in the Weddell-Scotia Confluence through a synergy of weak temperature control (ΔfCO<sub>2 sst</sub>), typical of the southern Scotia Sea, and strong biological CO<sub>2</sub> uptake (ΔfCO<sub>2 bio</sub>) during the spring and summer sea ice thaw. The dominant seasonal biological control (ΔfCO<sub>2</sub> bio) of 196 µatm in the meltwaters of the Weddell-Scotia Confluence generated the substantial summertime CO<sub>2</sub> disequilibrium of around -120 µatm, possibly with a minor contribution from calcium carbonate dissolution in the sea ice meltwater, respectively (Jones et al., 2010). For the whole SIZ (WSC, SB, SACCF), the distribution of sea surface fCO<sub>2</sub> displayed synchronicity to the seasonal transition from ice covered to ice free waters. The wintertime fCO<sub>2</sub> supersaturation in very cold, salty waters of the Weddell-Scotia Confluence is indicative of entrainment of CO<sub>2</sub> beneath sea ice. Throughout winter and early spring, remineralisation of organic matter, respiration, vertical mixing and brine rejection enable supersaturation of CO<sub>2</sub> beneath the pack ice (Nomura et al., 2006; Rysgaard et al., 2007). Ice covered waters of the Weddell-Scotia Confluence in spring were a potential source of CO<sub>2</sub> to the atmosphere as indicated by large, positive CO<sub>2</sub> disequilibrium. Breaking up of the sea ice near the South Orkney Islands could enable immediate release of the winter CO<sub>2</sub> 'reserve' prior to springtime photosynthetic activity and biological CO<sub>2</sub> drawdown that occurs during and after sea ice melt (Bakker et al., 2008; Jones et al., 2010).

The shift from CO<sub>2</sub> saturation to undersaturation in close proximity aligns with the retreating seasonal ice edge, where phytoplankton blooms developed and strong CO<sub>2</sub> disequilibrium appeared to 'track' the southward retreating ice edge. Thawing sea ice led to stronger stratification and reduced surface ice cover, thus exposing algal cells to increased light levels alongside potential additional iron inputs. Iron might originate from melting sea ice (Sedwick and DiTullio, 1997; Nielsdóttir et al., 2012), shallow sediments from local sources such as the South Orkney Islands (Nielsdóttir et al., 2012), or upstream sources, e.g., the shelf of the Antarctic Peninsula (Hewes et al., 2008; Ardelan et al., 2009; Dulaiova et al., 2009). Iron (re)-supply to the central Scotia Sea could also result from upwelling of deep waters following water

mass-topography interactions, e.g., the Antarctic Peninsula; melting sea ice and icebergs and subsequent transport eastwards within the SACCF (Sokolov and Rintoul, 2007; Dulaiova et al., 2009; Ardelan et al., 2010; Zhou et al., 2010; Venables et al., 2012). The large transit time from an upstream source to bloom location enables biological assimilation of the iron and physical dispersion, accounting for the temporal relief from iron stress and relatively short-lived blooms in the central Scotia Sea region (Korb et al., 2012; Whitehouse et al., 2012). Consumption of the reserve of macronutrients was evident within the diatom-cryptophyte blooms that were observed in spring 2006 (Korb et al., 2012; Whitehouse et al., 2012) in the shallow surface waters, symptomatic of recent sea ice melt. The emergence of cryptophytes in the Weddel-Scotia Confluence and at the SB by the summer was indicative of a species shift from an ice edge diatom bloom (Jacques and Panouse, 1991; Buma et al., 1992; Korb et al., 2012).

Bloom development in the Weddell-Scotia Confluence and southern Scotia Sea is affected by the start and rate of sea ice retreat (Comiso et al., 1993; Korb et al., 2005; Murphy et al., 2007). During winterspring 2006, sea ice reached its maximum northward extent at 57°S in August. Sea ice retreat began in September, exposing a large part of the SIZ to increased light levels and a meltwater stabilised water column potentially fertilised by iron and sea ice algae. Subsequently, a large bloom had developed by October 2006, which was identified as the strongest Scotia Sea bloom during the 3 year study period, accompanied by the negative CO<sub>2</sub> disequilibrium in the region of the SACCF and SB (Fig. 5). The intense bloom of spring-summer season 2006/7 was the strongest bloom of the entire region from 2006-2009 (Figs. 2 and 3) where the 'plume-like' structure of the bloom largely followed the course of the SACCF. High primary productivity drove the CO<sub>2</sub> undersaturation in the springtime diatom-dominated bloom (Korb et al., 2012), which then shifted to CO<sub>2</sub> saturation in a decaying bloom. The years 2007 and 2008 represented high and low wintertime sea ice extent during the study period, respectively, both with patchy and short-lived blooms in the SIZ after ice melt. This demonstrates the influence of sea ice on the biological activity of the Scotia Sea and contributed to large variations in bloom features in spring and summer. Large interannual variability in phytoplankton blooms in the Scotia Sea is imprinted on the seasonality observed during the spring, summer and autumn transects, i.e., bloom areal coverage ranged from 41,000 to 463,000 km<sup>2</sup> during the spring-summer season in different years across the Scotia Sea (Table 1). This is reflected in the year-to-year variability in marine productivity during the growing season (Constable et al., 2003; Smith and Comiso, 2008; Park et al., 2010; Korb et al., 2012). When present, blooms are regularly co-located to the broad position of the SACCF, which is visible to some extent during most spring seasons as a band extending west-east across the Scotia Sea (Fig. 2). The

location-range of the SACCF from all three cruises was close to the northern most extent of the SIZ at 57°S and about 700 km downstream of the Antarctic Peninsula (Venables et al., 2012). Mesoscale activity associated with the SACCF was evident from variability in silicic acid during autumn (Venables et al., 2012; Whitehouse et al., 2012). Sea ice retreat occurred relatively late in October 2006 but by November 2006 had reached comparable extent to the other years. Following such rapid ice retreat, optimal light conditions and iron supply from an upstream source (perhaps in an episodic manner related to mesoscale activity) presented favourable conditions for sporadic blooms, such as that observed in spring-summer 2006/7. This is supported by the shift from productive diatom blooms with chlorophyll-a values greater than 7 mg m<sup>-3</sup> and subsequent springtime CO<sub>2</sub> disequilibrium minimum of -80 µatm, to unproductive dinoflagellate communities (Korb et al., 2012) and fCO<sub>2</sub> saturated, warmer waters in the summer and autumn. Years of reduced sea ice cover (e.g., 2008) are exposed to more wintertime wind-driven mixing and possible enhanced outgassing of CO<sub>2</sub>. Greater seasonal warming is likely to occur on the open sea surface and hence a larger temperature control is expected. Conversely, in icy years (e.g., 2007) where the seasonal temperature control is dampened through sea ice cover, comparatively greater ice edge perimeter could enhance bloom formation events upon sea ice melt, under favourable iron and light conditions, tipping the balance to larger biological control.

The magnitude of seasonal biological ( $\Delta f CO_{2 \text{ bio}}$ ) and temperature ( $\Delta f CO_{2 \text{ sst}}$ ) effects is further depicted by considering the difference between and respective ratio of  $\Delta f CO_{2 \text{ sst}}$  and  $\Delta f CO_{2 \text{ bio}}$ , revealing that seasonal changes in oceanic  $f CO_2$  in the Weddell-Scotia Confluence are substantial and highly variable (Fig. 8b). Despite large year to year variability, the blooms that develop in the wake of sea ice retreat exert a strong biological control on oceanic  $CO_2$  in the Weddell-Scotia Confluence to generate one of the largest seasonal amplitudes in  $CO_2$  disequilibrium in the Scotia Sea region.

### 4.2 Transient CO<sub>2</sub> uptake in the central Scotia Sea: sporadic blooms and HNLC waters

The central Scotia Sea displayed typical HNLC conditions, with the exception of a spring bloom in 2006, and had the lowest seasonal change in sea surface  $fCO_2$  ( $\Delta fCO_2$  sst-bio of -29  $\mu$ atm). This region displayed the weakest biological control on oceanic  $CO_2$  of 97  $\mu$ atm, compensated by moderate temperature control of 68  $\mu$ atm. Sea surface  $fCO_2$  remained close to the atmospheric value in summer and autumn and reached supersaturation during the winter. Chlorophyll-a concentrations were less than 0.2 mg m<sup>-3</sup> with a resident dinoflagellate population (Korb et al., 2012) in summer and autumn. However, the large springtime phytoplankton bloom in 2006 that extended into the HNLC sub-region generated  $fCO_2$  undersaturation.

Thus, the region displays overall weak seasonality in oceanic CO<sub>2</sub> with predominantly HNLC conditions. However, the presence of sporadic seasonal blooms influences the seasonal signal where biological carbon uptake becomes a strong driver of oceanic CO<sub>2</sub>, largely compensated by moderate temperature control.

### 4.3 Northern Scotia Sea: episodic CO<sub>2</sub> uptake superimposed on systematic CO<sub>2</sub> seasonal cycles

The northern Scotia Sea (nSS, NSR) exhibits a systematic seasonal cycle in  $CO_2$  disequilibrium from (winter/spring) supersaturation to (summer) undersaturation to (autumn/winter) saturation. This is driven by large biological control ( $\Delta f CO_2$  bio) of 146-151  $\mu$ atm, partially compensated by temperature effects ( $\Delta f CO_2$  sst) of 70-75  $\mu$ atm. Maximum sea surface  $f CO_2$  in deep mixed layers from winter to spring is reduced to a summertime  $f CO_2$  minimum as biological production (chlorophyll-a) increases during the growing season.

Blooms in this region are variable, often building up biomass (chlorophyll-a) during spring, sometimes reaching a peak in summer and occasionally persisting until the autumn. Such fluctuations could be a result of seasonal iron depletion and/ or re-supply, including inputs from interactions with the North Scotia Ridge and episodic iron enrichment events, i.e., Patagonian dust deposition (Erickson III et al., 2003). In the established diatom bloom during summer 2008, the nSS-NSR sub-regions revealed distinct depletions in nitrate and phosphate (Korb et al., 2012; Whitehouse et al., 2012) as chlorophyll-a values reached 1.2 mg m<sup>-3</sup>. This was accompanied by the observed seasonal shift in CO<sub>2</sub> disequilibrium from springtime (super)saturation into summertime undersaturation close to the North Scotia Ridge. By autumn, sea surface warming within a decaying diatom bloom and emergence of heterotrophic dinoflagellates (Korb et al., 2012) enabled sea surface fCO<sub>2</sub> to increase towards near saturation, demonstrating the temperature control in this region. The frequent blooms that develop by episodic events are superimposed on to seasonal warming, leading to little seasonality and overall net biological control in the northern Scotia Sea and across the North Scotia Ridge.

### 4.4 Substantial seasonal CO<sub>2</sub> uptake in the South Georgia bloom

Despite the highest wintertime sea surface  $fCO_2$  values and strongest temperature control of the six subregions studied, substantial and sustained biological carbon uptake in the South Georgia bloom yielded very low sea surface  $fCO_2$  from spring to summer to autumn, resulting in dominant biological control on oceanic  $CO_2$  across the Georgia Basin (Fig. 8).

Wintertime maximum sea surface fCO<sub>2</sub> of up to 455 µatm in the Georgia Basin is the likely result of deep mixing, enabling CO<sub>2</sub>-rich waters to permeate the winter mixed layer (Winter Water) (Jones et al., 2012). At this time, the sea surface reached fCO<sub>2</sub> supersaturation and was a potential strong source of CO<sub>2</sub> to the atmosphere. By the onset of spring, sea surface fCO<sub>2</sub> was reduced to near-saturation levels accompanied by moderate chlorophyll-a concentrations in the Georgia Basin. At this time the phytoplankton community was possibly light limited (heterotrophic dinoflagellates were dominant) and iron concentrations were low (Korb et al., 2012; Nielsdóttir et al., 2012). By summer, shallowing mixed layers and optimal light conditions were accompanied by peaks in chlorophyll-a concentrations of 11 mg m<sup>-3</sup>, water column chlorophyll-a and rates of primary production (Korb et al., 2012). There was a shift in the phytoplankton community towards large diatoms. Waters were characterised by high surface iron concentrations exceeding 5 nM (Nielsdóttir et al., 2012) and a lack of phytoplankton response to iron addition experiments (Hinz et al., 2012); all indicative of an iron replete environment (Korb and Whitehouse, 2004; Korb et al., 2008). The sea surface reached a summertime minimum in CO<sub>2</sub> disequilibrium of -116 µatm that counteracted seasonal effects of sea surface warming. Substantial biological  $CO_2$  uptake across the Georgia Basin created one of the strongest summertime sinks of 12.9  $\pm$ 11.7 mmol m<sup>-2</sup> day<sup>-1</sup> for atmospheric CO<sub>2</sub> and highest seasonal inorganic carbon depletion of  $4.6 \pm 0.8$ mol m<sup>-2</sup> in naturally iron fertilised ice-free waters of the Southern Ocean (Jones et al., 2012).

By the autumn, dinoflagellates had increased (Korb et al., 2012) and chlorophyll-a values still remained high. Respiration acted to slightly increase sea surface  $fCO_2$ . The northern part of the transect had the largest seasonal increase in sea surface temperature. The data showed a latitudinal increase in temperature control across the Scotia Sea region to a maximum  $\Delta fCO_2$  sst of 98 µatm in the Georgia Basin. Considering the difference and ratio in seasonal  $\Delta fCO_2$  of biological and temperature effects, the phytoplankton blooms downstream (north) of South Georgia are strong, annual features that lead to dominant seasonal biological control of oceanic  $CO_2$  of 257 µatm.

The annual occurrence of vast phytoplankton blooms downstream, to the north, of South Georgia is observed during the whole study period. High sea surface chlorophyll-a and productivity in this region have been previously documented (Korb and Whitehouse 2004; Korb et al., 2008; Borrione and Schlitzer, 2013). Based on bloom area and duration, the longest lived, widespread blooms in the Georgia Basin occurred during the summer seasons and represented some of the strongest blooms in the wider Scotia Sea region from 2006-2009 (Table 1). These blooms often start during early spring, persisting until the

autumn and regularly spread across the whole Georgia Basin. The areal limits of the bloom are strongly correlated to the contours of sea surface dynamic height (Venables et al., 2012), which can be seen by enhanced chlorophyll-a plumes within the jets of the SACCF and the PF to the south and north of the basin, respectively (Figs. 1 and 2).

The maximum areal extent of the South Georgia bloom in summer 2007/8 was determined as 325,000 km² (rank 2) and was the strongest South Georgia bloom from the 2006-2009 study period (Table 1). The presence of weakly and strongly silicified diatoms in sediment traps from 2000 m depth in the Georgia Basin suggest that there is a high particulate flux of organic carbon to depth (Korb et al., 2010; Korb et al., 2012; Whitehouse et al., 2012) and that the South Georgia bloom is another naturally iron fertilised hotspot of carbon export to the deep Southern Ocean. The blooms of summer seasons 2005/6, 2006/7, 2008/9 are of similar duration and extent (Fig. 3) and therefore it can be inferred that, based on observations during a 'typical' summer, the South Georgia bloom creates considerable atmospheric CO<sub>2</sub> uptake and concomitant sequestration of organic carbon to depth and exerts a strong biological control on the annual cycle of oceanic CO<sub>2</sub>.

### 4.5 Biological processes drive seasonal signals in oceanic CO<sub>2</sub> across the Scotia Sea

The new seasonally-resolved time series in oceanic  $CO_2$  has allowed, for the first time, an assessment of the relative contribution of biological and temperature effects that drive the variability in oceanic  $CO_2$  across the Scotia Sea region from spring to summer, autumn and winter. The seasonal basin-wide biological effect on oceanic  $CO_2$ , determined as the mean of  $\Delta f CO_2$  bio for all sub-regions, is 159  $\mu$ atm. The mean temperature effect ( $\Delta f CO_2$  sst) of 68  $\mu$ atm partly (43%) compensates the biological control. This demonstrates the dominance of seasonal biological processes on oceanic  $CO_2$  from the seasonal ice zone to the Polar Front. The relationship developed by Takahashi et al. (2002) to distinguish temperature and biological signals in sea surface  $f CO_2$  (2) assigns all non-thermodynamic controls to biological processes. Subsequently, air-sea  $CO_2$  exchange, advection, mixing, upwelling, calcification and carbonate dissolution are incorporated into the seasonal 'biological' signal. Uncertainties in the derived temperature and biological signals exist if the seasonal sea surface  $f CO_2$  maxima and minima occur at finer temporal scales or at other locations than the seasonal meridional transects, which could be possible for the Scotia Sea, which has large spatial variability (as demonstrated by satellite imagery). However, the combination of (a) quasi-continuous sea surface  $f CO_2$ , temperature and salinity data; (b) frequent chlorophyll-a (Korb et al., 2012) and macronutrient (Whitehouse et al., 2012) measurements; (c) regular hydrographic stations

from spring (October-November) 2006 to summer (January-February) to autumn (April-May) 2009 and the inclusion of winter (August-September) 2007 data from summer potential temperature minimum during this study provides sufficient spatio-temporal coverage to elucidate the main physical and biological controls on CO<sub>2</sub> disequilibrium in the Scotia Sea.

#### 5 Conclusion

A first seasonally-resolved time series of oceanic CO<sub>2</sub> quantifies the effects of biological processes and temperature as drivers of the seasonal CO<sub>2</sub> cycle in the Scotia Sea. Spring, summer, autumn and winter fCO<sub>2</sub> data along a 1000 km meridional transect show that biological effects dominate temperature effects to control the seasonal signal of sea surface  $fCO_2$ . The naturally iron fertilised phytoplankton blooms that develop downstream (north) of the island of South Georgia each year promote substantial CO<sub>2</sub> uptake from spring to autumn. Despite wintertime CO<sub>2</sub> supersaturation and compensating temperature controls, the South Georgia blooms generate the largest seasonal fCO<sub>2</sub> amplitude for the entire Scotia Sea region. The seasonal ice zone was characterised by a large seasonal biological control and a weak temperature control. Strong seasonality in the presence of sea ice and phytoplankton blooms displayed a synergy with oceanic CO<sub>2</sub> in spring and summer. Ice-covered, CO<sub>2</sub>-rich waters were swiftly transformed to regions of large CO<sub>2</sub> undersaturation as biological carbon uptake rapidly reduced the winter CO<sub>2</sub> reserve following sea ice melt. Oceanic CO<sub>2</sub> varied significantly with chlorophyll-a concentrations and sea ice cover, as strong CO<sub>2</sub> disequilibrium tracked the southward retreating ice edge. The weakest seasonal biological signal occurred in high-nutrient low-chlorophyll and CO<sub>2</sub>-saturated waters of the central Scotia Sea. At the basin scale, biological processes exceeded temperature effects from the seasonal ice zone up to the Polar Front. The highly productive areas of the Scotia Sea region generate seasonally strong biological uptake of atmospheric CO<sub>2</sub> in the Southern Ocean.

#### Acknowledgements

The authors extend great thanks to the British Antarctic Survey and the DISCOVERY-2010 program, the Captain, officers, crew and scientists onboard RRS *James Clark Ross* during cruises JR161, JR177 and JR200. Special thanks to Ian Brown and Gerald Moore, Plymouth Marine Laboratory, and the British Antarctic Survey for continual support of the CO<sub>2</sub> instrument and to Andy Watson and Mario Hoppema for insightful discussions and comments that helped in preparing the manuscript. Supporting data, statistical and practical advice was gratefully received from Rebecca Korb, Min Gordon, Michiel de Roo,

Geraint Tarling, Angus Atkinson and Sophie Fielding. NASA, the SeaWiFS/MODIS programme and OSTIA at GHRSST are acknowledged for satellite data. Financial support was given by a NERC PhD studentship to EMJ (NE/F14/G6/115), under the auspices of the Centre for observation of Air-Sea Interactions and fluXes (CASIX) and the Antarctic Funding Initiative (CGS8/28).

#### References

- Ardelan, M.V., Holm-Hansen, O., Hewes, C.D., Reiss, C.S., Silva, N.S., Dulaiova, H., Steinnes, E., Sakshaug, E., 2010. Natural iron enrichment around the Antarctic Peninsula in the Southern Ocean. Biogeosciences 7, 11-25. doi:10.5194/bg-7-11-2010.
- Atkinson, A., Whitehouse, M.J., Priddle, J., Cripps, G.C., Ward, P., Brandon, M.A., 2001. South Georgia, Antarctica: a productive, cold water, pelagic ecosystem. Mar. Ecol. Prog. Ser. 216, 279-308.
- Bakker, D.C.E., Hoppema, M., Schroder, M., Geibert, W., de Baar, H.J.W., 2008. A rapid transition from ice covered CO<sub>2</sub>-rich waters to a biologically mediated CO<sub>2</sub> sink in the eastern Weddell Gyre. Biogeosciences 5, 1373-1386.
- Blain, S., Quéguiner, B., Armand, L., Belviso, S., Bombled, B., Bopp, L., Bowie, A., Brunet, C., Brussaard, C., Carlotti, F., Christaki, U., Corbière, A., Durand, I., Ebersbach, F., Fuda, J.-L., Garcia, N., Gerringa, L., Griffiths, B., Guigue, C., Guillerm, C., Jacquet, S., Jeandel, C., Lann, P., Lefèvre, D., Lo Monaco, C., Malits, A., Mosseri, J., Obernosterer, I., Park, Y.-H., Picheral, M., Pondaven, P., Remenyi, T., Sandroni, V., Sarthou, G., Savoye, N., Scouarnec, L., Souhart, M., Thuiller, D., Timmermans, K., Trull, T., Uitz, J., van Beek, P., Veldhuis, M., Vincent, D., Viollier, E., Vong, L., Wagener, T., 2007. Effect of natural iron fertilization on carbon sequestration in the Southern Ocean. Nature 446, 1070-1075. doi:10.1038/nature05700.
- Boehme, L., Meredith, M.P., Thorpe, S.E., Biuw, M., Fedak, M., 2008. Antarctic Circumpolar Current frontal system in the South Atlantic: monitoring using merged Argo and animal-borne sensor data.

  J. Geophys. Res. 113 (C09012). doi:10.1029/2007JC004647.
- Borrione, I., Schlitzer, R., 2013. Distribution and recurrence of phytoplankton blooms around South Georgia, Southern Ocean, Biogeosciences 10, 217-231. doi: 10.5194/bg-10-217-2013.
- Boyd, P.W., Jickells, T., Law, C.S., Blain, S., Boyle, E.A., Buesseler, K.O., Coale, K.H., Cullen, J.J., de Baar, H.J.W., Follows, M., Harvey, M., Lancelot, C., Levasseur, M., Owens, N.P.J., Pollard, R.T., Rivkin, R., Sarmiento, J., Schoemann, V., Smetacek, V., Takeda, S., Tsuda, A., Turner, S., Watson, A.J., 2007. Mesoscale Iron Enrichment Experiments 1993-2005: Synthesis and Future Directions. Science 315, 612-617. doi: 10.1126/science.1131669.
- Brainerd, K.E., Gregg, M.C., 1995. Surface mixed and mixing layer depths. Deep-Sea Res. I 42, 1521-1543. doi:10.1016/0967-0637(95)00068-H.

- Buma, A.G.J., Gieskes, W.W.C., Thomsen, H.A., 1992. Abundance of cryptophyceae and chlorophyll b containing organisms in the Weddell-Scotia Confluence area in the spring of 1988. Polar Biology 12, 43-52. doi:10.1007/BF00239964.
- Comiso, J.C., McClain, C.R., Sullivan, C.W., Ryan, J.P., Leonard, C.L., 1993. Coastal zone color scanner pigment concentrations in the Southern-Ocean and relationships to geophysical surface-features. J. Geophys. Res. Oceans 98, 2419-2451. doi:10.1029/92JC02505.
- Constable, A.J., Nicol, S., Strutton, P.G., 2003. Southern Ocean productivity in relation to spatial and temporal variation in the physical environment. J. Geophys. Res. 108 (C4), 8079. doi:10.1029/2001JC001270.
- Cooper, D.J., Watson, A.J., Ling, R.D., 1998. Variation of pCO<sub>2</sub> along a North Atlantic shipping route (U.K. to the Caribbean): A year of automated observations. Marine Chem. 60, 147-164. doi:10.1016/S0304-4203(97)00082-0.
- de Baar, H.J.W., de Jong, J. T.M., 2001. Distribution, sources and sinks of iron in seawater, in: Turner, D., Hunter, K. (Eds.), The Biogeochemistry of Iron in Seawater. Wiley, New York, pp. 123-253.
- de Baar, H.J.W., Jong, J.T.M., Bakker, D.C.E., Löscher, B.M., Veth, C., Bathmann, U., 1995. Importance of iron for plankton blooms and carbon dioxide drawdown in the Southern Ocean. Nature 373, 412-415. doi:10.1038/373412a0.
- Dulaiova, H., Ardelan, M.V., Henderson, P.B., Charette, M.A., 2009. Shelf-derived iron inputs drive biological productivity in the southern Drake Passage. Global Biogeochem. Cy. 23, doi:10.1029/2008GB003406.
- Erickson III, D.J., Hernandez, J.L., Ginoux, P., Gregg, W.W., McClain, C., 2003. Atmospheric iron delivery and surface ocean biological activity in the Southern Ocean and Patagonian region. Geophys. Res. Lett. 30, doi:10.1029/2003GL017241.
- Feldman, G.C., McClain, C.R., 2006a. Ocean Color Web, MODIS Reprocessing 1.1, NASA Goddard Space Flight Center. N. Kuring and S. W. Bailey.
- Feldman, G.C., McClain, C.R., 2006b. Ocean Color Web, SeaWiFS Reprocessing 5.1, NASA Goddard Space Flight Center. N. Kuring and S. W. Bailey.
- GEBCO, 2001. General Bathymetric Chart of the Oceans Digital Atlas. British Oceanographic Data Centre, Liverpool, U.K.
- Hardman-Mountford, N.J., Moore, G., Bakker, D.C.E., Watson, A.J., Schuster, U., Barciela, R., Hines, A., Moncoiffé, G., Brown, J., Dye, S., Blackford, J., Somerfield, P.J., Holt, J., Hydes, D.J.,

- Aitken, J., 2008. An operational monitoring system to provide indicators of CO<sub>2</sub>-related variables in the ocean. ICES J. Marine Sci. 65, 1498-1503. doi: 10.1093/icesjms/fsn110.
- Hewes, C.D., Reiss, C.S., Kahru, M., Mitchell, B.G., Holm-Hansen, O., 2008. Control of phytoplankton biomass by dilution and mixed layer depth in the western Weddell-Scotia Confluence. Mar. Ecol. Prog. Ser. 366, 15-29. doi:10.3354/meps07515.
- Hinz, D.J., Nielsdóttir, M.C., Korb, R.E., Whitehouse, M.J., Poulton, A.J., Moore, C.M., Achterberg, E.P., Bibby, T.S., 2012. Responses of microplankton community structure to iron addition in the Scotia Sea. Deep-Sea Res. II, 59-60, 36-46. doi:10.1016/j.dsr2.2011.08.006.
- Holeton, C.L., Nédélec, F., Sanders, R., Brown, L., Moore, C.M., Stevens, D.P., Heywood, K.J., Statham, P.J., Lucas, C.H., 2005. Physiological state of phytoplankton communities in the southwest Atlantic sector of the Southern Ocean, as measured by fast repetition rate fluorometry. Polar Biology 29, 44-52. doi:10.1007/s00300-005-0028-y.
- Holm-Hansen, O., Naganobu, M., Kawaguchi, S., Kameda, T., Krasovski, I., Tchernyshkov, P., Priddle, J., Korb, R., Brandon, M., Demer, D., Hewitt, R.P., Kahru, M., Hewes, C.D., 2004. Factors influencing the distribution, biomass, and productivity of phytoplankton in the Scotia Sea and adjoining waters. Deep-Sea Res. II 51, 1333-1350. doi: doi:10.1016/j.dsr2.2004.06.015..
- Jacques, G., Panouse, M., 1991. Biomass and composition of size fractionated phytoplankton in the Weddell-Scotia Confluence area. Polar Biology 11, 315-328. doi:10.1007/BF00239024.
- Jones, E.M., Bakker, D.C.E., Venables, H.J., Watson, A.J., 2012. Dynamic seasonal cycling of inorganic carbon downstream of South Georgia, Southern Ocean. Deep-Sea Res. II 59-60, 25-35. doi:10.1016/j.dsr2.2011.08.001.
- Jones, E.M., Bakker, D.C.E., Venables, H.J., Whitehouse, M.J., Korb, R.E., Watson, A.J., 2010. Rapid changes in surface water carbonate chemistry during Antarctic sea ice melt. Tellus 62B, 621-635. doi:10.1111/j.1600-0889.2010.00496.x.
- Jones, E.M., 2010. The Marine Carbon Cycle of the Scotia Sea, Southern Ocean. Ph.D. Thesis. University of East Anglia. UEA e-prints.
- Jullion, L., Naveira Garabato, A.C., Bacon, S., Meredith, M.P., Brown, P.J., Torres-Valdés, S., Speer, K.G., Holland, P.R., Dong, J., Bakker, D.C.E., Hoppema, M., Loose, B., Venables, H.J., Jenkins, W.J., Messias, M.-J., Fahrbach, E., 2014. The contribution of the Weddell Gyre to the lower limb of the Global Overturning Circulation. J. Geophys. Res. Oceans 119, 3357-3377. doi: 10.1002/2013JC009725.

- Korb, R.E., Whitehouse, M.J., 2004. Contrasting primary production regimes around South Georgia, Southern Ocean: large blooms versus high nutrient, low chlorophyll waters. Deep-Sea Res. I 51, 721-738. doi:10.1016/j.dsr.2004.02.006.
- Korb, R.E., Whitehouse, M. J., Thorpe, S. E., Gordon, M., 2005. Primary production across the Scotia Sea in relation to the physico-chemical environment. J. Mar. Syst. 57, 231-249. doi:10.1016/j.jmarsys.2005.04.009.
- Korb, R.E., Whitehouse, M.J., Atkinson, A., Thorpe, S.E., 2008. Magnitude and maintenance of the phytoplankton bloom at South Georgia: a naturally iron replete environment. Mar. Ecol. Progr. Ser. 368, 75-91. doi:10.3354/meps07525.
- Korb, R.E., Whitehouse, M.J., Gordon, M., Ward, P., Poulton, A.J., 2010. Summer microplankton community structure across the Scotia Sea: implications for biological carbon export. Biogeosciences 7, 343-356. doi:10.5194/bg-7-343-2010.
- Korb, R.E., Whitehouse, M.J., Ward, P., Gordon, M., Venables, H.J., 2012. Regional and seasonal differences in microplankton biomass, productivity and structure across the Scotia Sea and implications to the export of biogenic carbon. Deep-Sea Res. II, 59-60, 67-77. doi:10.1016/j.dsr2.2011.06.006.
- Landschützer, P., Gruber, N., Bakker, D.C.E., Schuster, U., 2014. Recent variability of the global ocean carbon sink. Global Biogeo. Cycles, 28, 927-949. doi: 10.1002/2014GB004853.
- Lancelot, C., Mathot, S., Veth, C., de Baar, H.J.W., 1993. Factors controlling phytoplankton ice edge blooms in the Marginal Ice Zone of the northwestern Weddell Sea during sea ice retreat 1988 field observations and mathematical modelling. Polar Biology 13, 377-387. doi:10.1007/BF01681979.
- Lenton, A., Tilbrook, B., Law, R.M., Bakker, D.C.E., Doney, S.C., Gruber, N., Ishii, M., Hoppema, M., Lovenduski, N.S., Matear, R.J., McNeil, B.I., Metzl, N., Mikaloff Fletcher, S.E., Monteiro, P.M.S., Rödenbeck, C., Sweeney, C., Takahashi, T., 2013. Sea–air CO<sub>2</sub> fluxes in the Southern Ocean for the period 1990–2009. Biogeosciences, 10, 4037-4054. doi:10.5194/bgd-10-285-2013.
- Locarnini, R.A., Whitworth, T., Nowlin, W.D., 1993. The importance of the Scotia Sea on the outflow of Weddell Sea Deep Water. J. Marine Res. 51, 135-153. doi:http://dx.doi.org/10.1357/0022240933223846.
- Meredith, M.P., Naveira Garabato, A.C., Gordon, A.L., Johnson, G.C., 2008. Evolution of the Deep and Bottom Waters of the Scotia Sea, Southern Ocean, during 1995-2005. J. Climate 21, 3327-3342. doi:http://dx.doi.org/10.1175/2007JCLI2238.1.

- Meredith, M.P., Watkins, J.L., Murphy, E.J., Cunningham, N.J., Wood, A.G., Korb, R.E., Whitehouse, M.J., Thorpe, S.E., Vivier, F., 2003. An anticyclonic circulation above the Northwest Georgia Rise. Geophys. Res. Lett. 30(20), 2061, 3327-3342. doi:10.1029/2003GL018039.
- Meskhidze, N., Nenes, A., Chameides, W.L., Luo, C., Mahowald, N., 2007. Atlantic Southern Ocean productivity: fertilization from above or below? Global Biogeochem. Cycl. 21, GB2006. doi:10.1029/2006GB002711.
- Metzl, N., Brunet, C., Jabaud-Jan, A., Poisson, A., Schauer, B., 2006. Summer and winter air—sea CO<sub>2</sub> fluxes in the Southern Ocean. Deep-Sea Res. I 53, 1548-1563. doi:10.1016/j.dsr.2006.07.006.
- Murphy, E.J., Watkins, J., Trathan, P.N., Reid, K., Meredith, M.P., Thorpe, S.E., Johnson, N.M., Clarke,
  A., Tarling, G., Collins, M.A., Forcada, J., Shreece, R.S., Atkinson, A., Korb, R.E., Whitehouse,
  M.J., Ward, P., Rodhouse, P.G., Enderlein, P., Hirst, A.G., Martin, A.R., Hill, S.L., Staniland, I.J.,
  Pond, D.W., Briggs, D.R., Cunningham, N.J., Fleming, A.H., 2007. Spatial and temporal
  operation of the Scotia Sea ecosystem: a review of large-scale links in a krill centred food web.
  Philos. Trans. R. Soc. B 362, 113-148. doi:10.1098/rstb.2006.1957.
- Murphy, E.J., Clarke, A., Symon, C., Priddle, J., 1995. Temporal variation in Antarctic sea-ice: analysis of a long- term fast-ice record rom the South-Orkney Islands. Deep-Sea Res. I 42, 1045-1062. doi:10.1016/0967-0637(95)00057-D.
- Naveira Garabato, A.C., Heywood, K.J., Stevens, D.P., 2002. Modification and pathways of Southern Ocean Deep Waters in the Scotia Sea. Deep-Sea Res. I 49, 681-705. doi:10.1016/S0967-0637(01)00071-1.
- Naveira Garabato, A.C., Polzin, K.L., King, B.A., Heywood, K.J., Visbeck, M., 2004. Widespread intense turbulent mixing in the Southern Ocean. Science 303, 210-213. doi:10.1126/science.1090929.
- Nielsdóttir, M.C., Bibby, T.S., Moore, C.M., Hinz, D.J., Sanders, R., Whitehouse, M.J., Korb, R.E., Achterberg, E.P., 2012. Seasonal and spatial dynamics of iron availability in the Scotia Sea. Marine Chemistry. 130-131, 62-72. doi:10.1016/j.marchem.2011.12.004.
- Nomura, D., Inoue, H.Y., Yoyota T., 2006. The effect of sea-ice growth on air-sea  $CO_2$  flux in a tank experiment. Tellus 58B, 418-426. doi:10.1111/j.1600-0889.2006.00204.x.
- Orsi, A.H., Whitworth, T., Nowlin, W.D., 1995. On the meridional extent and fronts of the Antarctic Circumpolar Current. Deep-Sea Res. I 42, 641-673. doi:10.1016/0967-0637(95)00021-W.
- Park, J., Im-Sang, O., Hyun-Cheol, K., Sinjae, Y., 2010. Variability of SeaWiFs chlorophyll-a in the southwest Atlantic sector of the Southern Ocean: strong topographic effects and weak seasonality. Deep-Sea Res. I 57, 604-620. doi:10.1016/j.dsr.2010.01.004.

- Patterson, S.L., Sievers, H.A., 1980. The Weddell-Scotia Confluence. J. Phys. Oceanogr. 10, 1584-1610. doi: http://dx.doi.org/10.1175/1520-0485(1980)010<1584:TWSC>2.0.CO;2.
- Pollard, R.T., Salter, I., Sanders, R.J., Lucas, M.I., Moore, C.M., Mills, R.A., Statham, P.J., Allen, J.T., Baker, A.R., Bakker, D.C.E., Charette, M.A., Fielding, S., Fones, G.R., French, M., Hickman, A.E., Holland, R.J., Hughes, J.A., Jickells, T.D., Lampitt, R.S., Morris, E.E., Nedelec, F.H., Nielsdottír, M., Planquette, H., Popova, E.E., Poulton, A.J.,Read,J.F.,Seeyave,S.,Smith,T., Stinchcombe, M., Taylor, S., Thomalla, S., Venables, H.J., Williamson, R., Zubkov, M.V., 2009. Southern Ocean deep-water carbon export enhanced by natural iron fertilization. Nature 457, 577-581. doi:10.1038/nature07716.
- Pollard, R.T., Lucas, M.I., Read, J.F., 2002. Physical controls on biogeochemical zonation in the Southern Ocean. Deep-Sea Res. II 49, 3289-3305. doi:10.1016/S0967-0645(02)00084-X.
- Rysgaard, S., Glud, R.N., Sejr, M.K., Bendtsen, J., Christensen, P.B., 2007. Inorganic carbon transport during sea ice growth and decay: A carbon pump in polar seas. J. Geophys. Res. 112. doi: 10.1029/2006JC003572.
- Schlitzer, R., 2002. Carbon export fluxes in the Southern Ocean: results from inverse modelling and comparison with satellite based estimates. Deep-Sea Res. II 49, 1623-1644. doi:10.1016/S0967-0645(02)00004-8.
- Sedwick, P.N., DiTullio, G.R., 1997. Regulation of algal blooms in Antarctic shelf waters by the release of iron from melting sea ice. Geophys. Res. Lett. 24, 2515-2518. doi:10.1029/97GL02596.
- Shim, J., Kang, Y.C., Kim, D., Choi, S.-H., 2006. Distribution of net community production and surface pCO<sub>2</sub> in the Scotia Sea, Antarctica, during austral spring 2001. Mar. Chem. 101, 68-84. doi:10.1016/j.marchem.2005.12.007.
- Smith, I.J., Stevens, D.P., Heywood, K.J., Meredith, M.P., 2010. The flow of the Antarctic Circumpolar Current over the North Scotia Ridge. Deep-Sea Res. I 57, 14-28. doi:10.1016/j.dsr.2009.10.010.
- Smith, W.O., Comiso, J.C., 2008. Influence of sea ice on primary productivity in the Southern Ocean: A satellite perspective. J. Geophys. Res. 113, doi:10.1029/2007JC004251.
- Sokolov, S., Rintoul, S.R., 2007. On the relationship between fronts of the Antarctic Circumpolar Current and surface chlorophyll concentrations in the Southern Ocean. J. Geophys. Res. 112, doi:10.1029/2006JC004072.
- Stark, J.D., Donlon, C.J., Martin, M.J., McCulloch, M.E., 2007. OSTIA: An operational, high resolution, real time, global sea surface temperature analysis system. In: Marine Challenges: coastline to deep sea. Conference proceedings. Oceans '07 IEEE Aberdeen.

- Tarling, G.A., Ward, P., Atkinson, A., Collins, M., Murphy, E.J., 2012. Editorial: Discovery 2010: understanding the Scotia Sea food web. Deep-Sea Research II 59-60, 1-13.
- Takahashi, T., Olafsson, J., Goddard, J.G., Chipman, D.W., Sutherland, S.C., 1993. Seasonal variation of CO<sub>2</sub> and nutrients in the high latitude surface oceans a comparative study. Global Biogeochem. Cyc. 7, 843-878. doi:10.1029/93GB02263.
- Takahashi, T., Sutherland, S.C., Sweeney, C., Poisson, A., Metzl, N., Tilbrook, B., Bates, N., Wanninkhof, R., Feely, R.A., Sabine, C.L., Olafsson, J., Nojiri, Y., 2002. Global sea-air CO<sub>2</sub> flux based on climatological surface ocean pCO<sub>2</sub>, and seasonal biological and temperature effects. Deep-Sea Res. II 49, 1601-1622. doi:10.1016/S0967-0645(02)00003-6.
- Takahashi, T., Sutherland, S.C., Wanninkhof, R., Sweeney, C., Feely, R.A., Chipman, D.W., Hales, B., Friederich, G., Chavez, F., Sabine, C.L., Watson, A.J., Bakker, D.C.E., Schuster, U., Metzl, Yoshikawa-Inoue, H., Ishii, M., Midorikawa, T., Nojiri, Y., Körtzinger, Steinhoff, T., Hoppema, M., Olafsson, J., Arnarson, T.S., Tilbrook, B., Johannessen, T., Olsen, A., Bellerby, R., Wong, C.S., Delille, B., Bates, N.R., de Baar, H.J.W., 2009. Climatological mean and decadal change in surface ocean pCO<sub>2</sub>, and net sea-air CO<sub>2</sub> flux over the global oceans. Deep-Sea Res. II 56, 554-577. doi:10.1016/j.dsr2.2008.12.009.
- Thomas, H, Bozec, Y., Elkalay, K., de Baarm H.J.W., Borges, A.V., Schiettecatte, L.-S., 2005. Controls of the surface water partial pressure of CO<sub>2</sub> in the North Sea. Biogeosciences, 2, 323-334.
- Thorpe, S.E., Heywood, K.J., Brandon, M.A., Stevens, D.P., 2002. Variability of the Southern Antarctic Circumpolar Current Front north of South Georgia. J. Mar. Sys., 37, 87-105. doi:10.1016/S0924-7963(02)00197-5.
- Trathan, P.N., Brandon, M.A., Murphy, E.J., Thorpe, S.E., 1997. Characterisation of the Antarctic Polar Frontal Zone to the north of South Georgia in summer 1984. J. Geophys. Res. 102, 10483-10497.
- van Heuven, S., Pierrot, D., Rae, J.W.B., Lewis, E., Wallace, D.W.R., 2011. MATLAB program developed for CO<sub>2</sub> system calculations, ORNL/CDIAC-105b, Carbon Dioxide Int. Anal. Cent., Oak Ridge Natl. Lab., US DOE, Oak Ridge, Tenn.
- Venables, H.J., Meredith, M.P., Atkinson, A., Ward, P., 2012. Fronts and habitat zones in the Scotia Sea. Deep-Sea Res. II 59-60, 14-24. doi:10.1016/j.dsr2.2011.08.012.
- Watson, A.J., Orr, J.C., 2003. Carbon dioxide fluxes in the global ocean. in: Fasham, M.J.R. (Ed.), Ocean Biogeochemistry: The Role of the Ocean Carbon Cycle in Global Change. Springer-Verlag, Berlin.

- Weiss, R.F., Price, B.A., 1980. Nitrous-oxide solubility in water and seawater. Marine Chem. 8, 347-359. doi:10.1016/0304-4203(80)90024-9.
- Whitworth, T., Nowlin, W.D., Orsi, A.H., Locarnini, R.A., Smith, S.G., 1994. Weddell Sea Shelf Water in the Bransfield Strait and Weddell–Scotia Confluence. Deep-Sea Res. I 41, 629–641. doi:10.1016/0967-0637(94)90046-9.
- Whitehouse, M.J., Korb, R.E., Atkinson, A., Thorpe, S.E., Gordon, M., 2008. Formation, transport and decay of an intense phytoplankton bloom within the high-nutrient low-chlorophyll belt of the Southern Ocean. J. Mar. Sys. 70, 150-167. doi:10.1016/j.jmarsys.2007.05.003.
- Whitehouse, M.J., Atkinson, A., Korb, R.E., Venables, H.J., Pond, D.W., Gordon, M., 2012. Substantial primary production in the land-remote region of the central and northern Scotia Sea. Deep-Sea Res. 59-60, 47-56. doi:10.1016/j.dsr2.2011.05.010.
- Zhou, M., Zhu, Y., Dorland, R.D., Measures, C.I., 2010. Dynamics of the current system in the southern Drake Passage. Deep-Sea Res. I 57, 1039-1048. doi:10.1016/j.dsr.2010.05.012.

Figure 1. Map of the Scotia Sea showing the location of the seasonal meridional transect by large dashed line and summer 2008 extension as a dotted line. Important topographic features are: South Scotia Ridge (SSR), South Orkney Islands (SOI), South Georgia (SG), North Scotia Ridge (NSR), Georgia Basin (GB), Northeast Georgia Rise (NEGR) and Maurice Ewing Bank (MEB). The location of the Weddell-Scotia Confluence (WSC) is also marked. Schematic positions of the Southern Boundary (SB), Southern ACC Front (SACCF) and the Polar Front (PF) are shown, adapted from Meredith et al., (2003). Depth contours are at 1000 and 2000 m (GEBCO, 2001).

Figure 2. Seasonal satellite chlorophyll-a (chl-a, mg m<sup>-3</sup>) composite time series for the Scotia Sea region for each austral spring (September-November), summer (December-February) and autumn (March-May) during the 2006-2009 study period. Winter (June-August) images, mostly obscured by ice and/ or cloud cover and data limited by reduced light availability, are not shown. Data are from 2006 (SeaWiFS/MODIS merged product), 2007 (SeaWiFS), 2008 (MODIS) and 2009 (MODIS). Depth contours are at 1000 and 2000 m (GEBCO, 2001).

Figure 3. Areal extent (10<sup>3</sup> km<sup>2</sup>, bars) and duration (days, dashed lines) of the principal Scotia Sea (S Sea, dark blue) and Georgia Basin (GB, light blue) blooms for each austral spring (September-November), summer (December-February) and autumn (March-May) during the 2006-2009 study period (Table 1).

Figure 4. Monthly satellite derived sea ice cover (fraction) along the meridional transect for the Scotia Sea and Weddell-Scotia Confluence from September to August 2006/2007, 2007/2008 and 2008/2009.

Figure 5. The latitudinal distribution of sea surface (a)  $CO_2$  disequilibrium ( $\Delta f CO_2$ ,  $\mu atm$ ), (b) chlorophyll-a (chl-a, mg m<sup>-3</sup>), (c) salinity, (d) sea surface temperature (sst, °C), (e) mixed layer depth (mld, m) and (f) sea ice cover (fraction) during spring 2006, summer 2008, autumn 2009 and winter ( $\theta_{min}$ ). The vertical grey bars mark the location of the sea ice edge in spring ( $\sim$ 59°S), summer ( $\sim$ 62°S) and winter ( $\sim$ 57°S) as determined by the northern limit of 0.5 fractional sea ice cover in (f). The sea ice edge in autumn was located further south than the transect southern limit and is hence not shown. The latitudinal limits of the Weddell-Scotia Confluence (WSC), Southern Boundary (SB), Southern ACC Front (SACCF), high-nutrient low-chlorophyll (HNLC) waters, northern Scotia Sea (nSS), North Scotia Ridge (NSR), Georgia Basin (GB) and the Polar Front (PF) are indicated (black arrows).

Figure 6. Seasonal distribution of  $CO_2$  disequilibrium ( $\Delta fCO_2$ ,  $\mu atm$ ) along the meridional transect across the Weddell-Scotia Confluence, Scotia Sea and Georgia Basin during (a) spring 2006, (b) summer 2008 and (c) autumn 2009. Bathymetry contours as for Fig. 4.

Figure 7. Seasonal variability of sea surface  $fCO_2$  (µatm) as a function of (a) temperature (°C), (b) salinity, (c) chlorophyll-a (chl-a, mg m<sup>-3</sup>) and (d) mixed layer depth (mld, m) in austral spring 2006, summer 2008, autumn 2009 and winter ( $\theta_{min}$ ). Regression lines are shown for significant correlations. The average  $fCO_2$  in air for 2006-2009 is shown for reference (black dashed line) where data below the line represent oceanic  $CO_2$  undersaturation.

Figure 8. Seasonal biological and temperature  $CO_2$  signals, following Takahashi et al. (2002): (a) biological ( $\Delta f CO_2$  bio), temperature ( $\Delta f CO_2$  sst) and temperature–biological ( $\Delta f CO_2$  sst-bio)  $CO_2$  disequilibrium as a mean per sub-region where zero  $\Delta f CO_2$  (black dashed line) marks the division between net seasonal  $CO_2$  increase, above, and net seasonal  $CO_2$  decrease, below; (b)  $CO_2$  disequilibrium difference vs. ratio of temperature and biological signals as a mean per sub-region with the balance between both controls (zero  $\Delta f CO_2$  difference and  $\Delta f CO_2$  ratio of 1) marked by horizontal and vertical black dashed lines, respectively.

SSR SOI SB SACCF SG NSR GB NEGR MEB PF

**WSC** 

Scotia Sea

Table 1. Areal extent and duration of the principal Scotia Sea (S Sea) and Georgia Basin (GB) blooms for each austral spring (September-November), summer (December-February) and autumn (March-May) during the 2006-2009 study period. Areal extent (10<sup>3</sup> km<sup>2</sup>) and duration (days) for blooms (chl-a exceeds 0.75 mg m<sup>-3</sup>) are determined from SeaWiFS/MODIS composite 8-day images. Areal extent is defined as the seasonal maximum limit of chlorophyll values exceeding 0.75 mg m<sup>-3</sup> within the Scotia Sea (55-59.3°S, 25-55°W) and Georgia Basin (49.5-53.0°S, 25-45°W). Duration is defined as the length of time a bloom is present in the images, in factors of 8 days up to a maximum of 88 days (i.e., the whole season). No blooms were detected during winter (June, July, August) 2006-2009. Each bloom is ranked in terms of extent (1 = largest area) and duration (1 = longest duration). The relative 'strength' of each bloom is expressed as a product of area and duration ranking. The average relative strength of all blooms is 52. The four strongest blooms are highlighted in bold. Timing of the seasonal meridional transects is indicated (\*).

season	year/s	location	extent		duration		rank product
			$10^3 \text{ km}^2$	rank	days	rank	
spring*	2006	S Sea	256	5	48	4	20
		GB	8	18	32	6	108
summer	2006/2007	S Sea	463	1	88	1	1
		GB	279	4	88	1	4
autumn	2007	S Sea	46	13	8	9	117
		GB	30	15	8	9	135
spring	2007	S Sea	50	12	32	6	72
		GB	165	9	64	3	27
summer*	2007/2008	S Sea	41	14	32	6	84
		GB	325	2	88	1	2
autumn	2008	S Sea	26	17	16	8	136
		GB	83	11	72	2	22
spring	2008	S Sea	210	6	72	2	12
		GB	182	8	72	2	16
summer	2008/2009	S Sea	196	7	72	2	14
		GB	298	3	88	1	3
autumn*	2009	S Sea	29	16	24	7	112
		GB	102	10	40	5	50

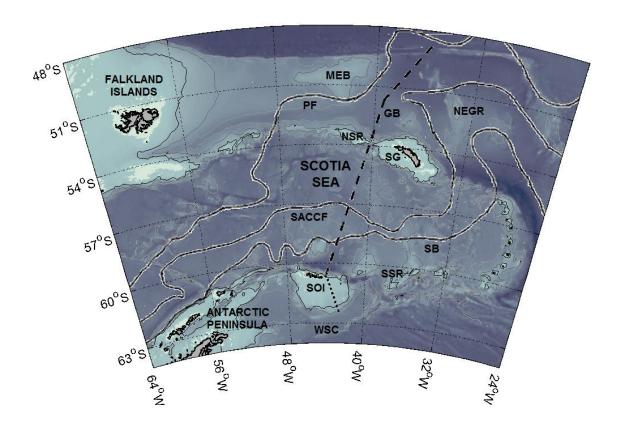


Figure 1

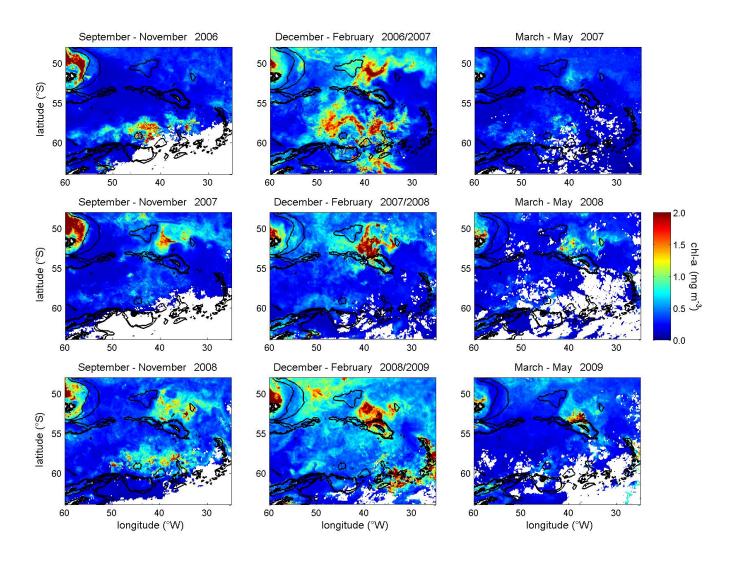


Figure 2

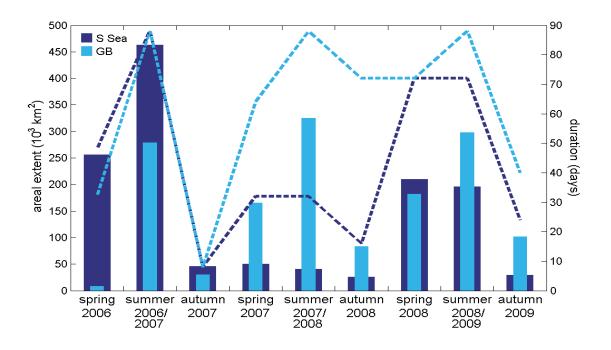


Figure 3

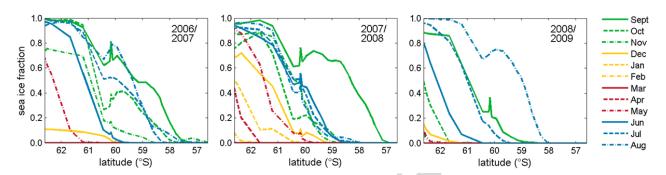


Figure 4

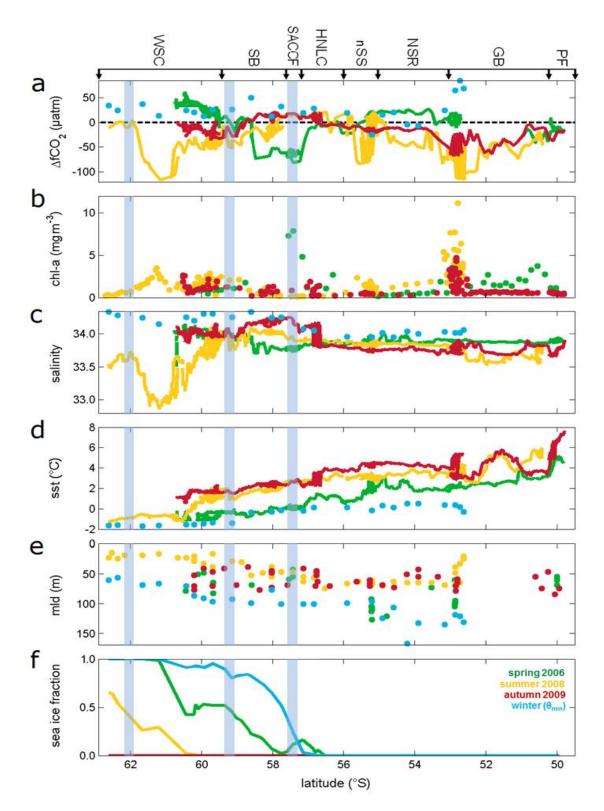


Figure 5

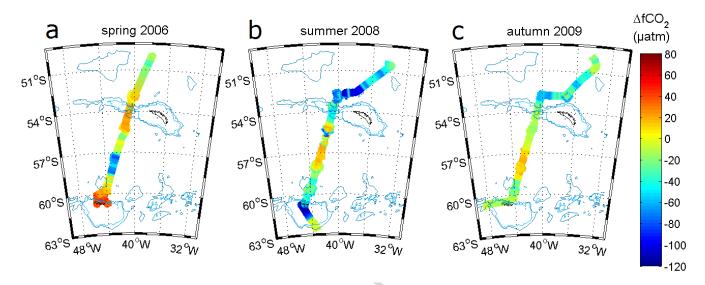


Figure 6

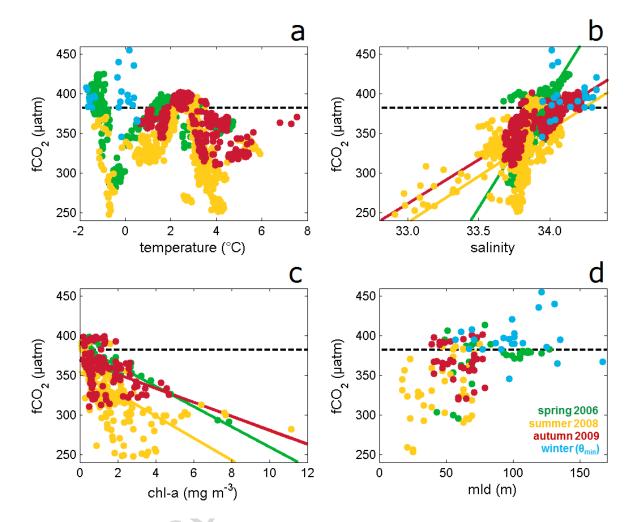


Figure 7

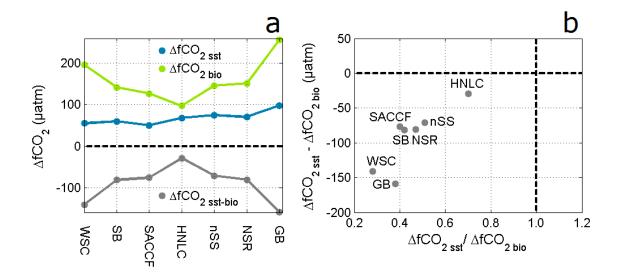


Figure 8



### Research highlights

A first seasonally-resolved oceanic CO<sub>2</sub> time series in the Scotia Sea is presented. Substantial biological CO<sub>2</sub> uptake occurs in the iron-fertilised South Georgia bloom. Regions of seasonally strong CO<sub>2</sub> uptake are formed in blooms following sea ice melt. High-nutrient low-chlorophyll waters had the weakest biological control. Biological processes dominate seasonal controls on oceanic CO<sub>2</sub> in the Scotia Sea.