



Article (refereed) - postprint

Boxall, A.B.A.; Keller, V.D.J.; Straub, J.O.; Monteiro, S.C.; Fussell, R.; Williams, R.J. 2014. Exploiting monitoring data in environmental exposure modelling and risk assessment of pharmaceuticals.

Copyright © 2014 Elsevier Ltd.

This version available http://nora.nerc.ac.uk/508154/

NERC has developed NORA to enable users to access research outputs wholly or partially funded by NERC. Copyright and other rights for material on this site are retained by the rights owners. Users should read the terms and conditions of use of this material at http://nora.nerc.ac.uk/policies.html#access

NOTICE: this is the author's version of a work that was accepted for publication in *Environment International*. Changes resulting from the publishing process, such as peer review, editing, corrections, structural formatting, and other quality control mechanisms may not be reflected in this document. Changes may have been made to this work since it was submitted for publication. A definitive version was subsequently published in *Environment International* (2014), 73. 176-185. 10.1016/j.envint.2014.07.018

www.elsevier.com/

Contact CEH NORA team at noraceh@ceh.ac.uk

The NERC and CEH trademarks and logos ('the Trademarks') are registered trademarks of NERC in the UK and other countries, and may not be used without the prior written consent of the Trademark owner.

Exploiting Monitoring Data in Environmental Exposure Modelling and Risk **Assessment of Pharmaceuticals** Boxall, A.B.A^{1*}, Keller, V.D.J.², Straub, J.O.³, Monteiro, S.C.⁴, Fussell, R.⁴, Williams, R.J.² ¹ – Environment Department, University of York, Heslington, York, UK, YO10 5DD ² – Centre for Ecology and Hydrology, Wallingford, UK, OX10 8BB ³ – F.Hoffmann-La Roche Ltd, CH–4070 Basle, Switzerland ⁴ – Food and Environment Research Agency, Sand Hutton, York, UK, YO41 1Z * - corresponding author: Telephone 01904 434791; email Alistair.boxall@york.ac.uk

Abstract

14

15

16

17

18

19

20

21

22

23

24

25

26

27

28

29

30

31

32

33

34

In order to establish the environmental impact of an active pharmaceutical ingredient (API), good information on the level of exposure in surface waters is needed. Exposure concentrations are typically estimated using information on the usage of an API as well as removal rates in the patient, the wastewater system and in surface waters. These input data are often highly variable and difficult to obtain, so model estimates often do not agree with measurements made in the field. In this paper we present an approach which uses inverse modelling to estimate overall removal rates of pharmaceuticals at the catchment scale using a hydrological model as well as prescription and monitoring data for a few representative sites for a country or region. These overall removal rates are then used to model exposure across the broader landscape. Evaluation of this approach for APIs in surface waters across England and Wales showed good agreement between modelled exposure distributions and available monitoring data. Use of the approach, alongside estimates of predicted no-effect concentrations for the 12 study compounds, to assess risk of the APIs across the UK landscape, indicated that, for most of the compounds, risks to aquatic life were low. However, ibuprofen was predicted to pose an unacceptable risk in 49.5% of the river reaches studied. For diclofenac, predicted exposure concentrations were also compared to the Environmental Quality Standard previously proposed by the European Commission and 4.5% of river reaches were predicted to exceed this concentration. While the current study focused on pharmaceuticals, the approach could also be valuable in assessing the risks of other 'down the drain' chemicals and could help inform our understanding of the important dissipation processes for pharmaceuticals in the pathway from the patient to ecological receptors.

36

37

38

35

Key words

Active pharmaceutical ingredient, inverse modelling, ibuprofen, diclofenac

Introduction

65

40 41 During the life cycle of a pharmaceutical product, Active Pharmaceutical Ingredients (APIs) 42 43 may be released to the natural environment (Daughton and Ternes, 1999; Boxall, 2004) and a 44 wide range of APIs have been detected in surface waters (Hirsch et al., 1999; Kolpin et al., 2002; Monteiro and Boxall, 2010). Even though the reported concentrations are generally low 45 46 (i.e. sub-µg/l), questions have been raised over the potential impacts of APIs in the 47 environment on flora and fauna and human health. Environmental risk assessments are also 48 now required in many regions as part of the marketing authorisation process of a new API 49 (Breton and Boxall, 2003). In order to establish the risks of APIs, it is essential to have a good understanding of the levels of exposure that occur in natural systems. 50 51 A range of exposure modelling approaches is currently being applied in the assessment of the 52 environmental risks of APIs. These include simple deterministic algorithms through to more complex models such as the GREAT-ER, PhATE and LF2000-WQX models (EMA, 2006; 53 Schowanek and Webb, 2002; Schwab et al., 2005; Williams et al., 2009) which use data on 54 55 flow in rivers to estimate how APIs will be distributed within river catchments. In order to accurately estimate concentrations in the environment, these models traditionally require 56 57 comprehensive information on the usage of an API within the system of interest, the extent of 58 metabolism of the API within treated humans and the degree of removal in wastewater 59 treatment processes and in receiving waters. 60 Many countries collate detailed information on the quantities of APIs used. For example, in 61 the UK, the National Health Service collect monthly information on the number of prescriptions made for different products in different regions. From this freely available 62 63 information, it is possible to determine the amounts of different APIs prescribed in an area over time. Similar systems are in place in Denmark, Germany and Australia. However, the 64

estimation of API usage, based on prescription volumes, may over-estimate what is actually

released to the environment. Over half of patients store unused medicines in their home as a consequence of dosage changes, discontinuation of the medication due to, for example, the occurrence of adverse side effects, or because the medications have reached their expiry date. It is estimated that anywhere between 3 and 65% of prescribed pharmaceuticals are not used and many of these will ultimately be returned to the pharmacist or disposed of to landfill (Seehusen and Edwards, 2006; Musson and Townsend, 2009). While numerous publications are available on the metabolism of APIs, the results of these studies can be highly variable. For example, for cyclophosphamide (one of the APIs investigated in the current study), amounts excreted are reported to range from 2 to 25% of the applied dose (Bagley et al., 1973). The observed differences are probably explained by genomically distinct metabolising capacities as well as differences in race, sex, age and health status of the studied subjects, all of which are known to affect the route and rate of metabolism (Dorne, 2010). The method of administration, previous exposure of a patient to the pharmaceutical and simultaneous exposure to other APIs and xenobiotics can also affect the degree of metabolism. For many APIs, no data exist on removal in wastewater treatment. In instances where data are available, variations can also be seen in the reported removal efficiencies (Sipma et al., 2010). These variations can be explained by differences in technologies used at different treatment works and differences in operating parameters. Some metabolites may also be reconverted back to the parent compound in wastewater treatment (Heberer et al., 2002). In large catchments it is likely that numerous treatment technologies will be in use and that these will vary in size and performance, so a variety of removal rates may need to be employed in the modelling. The fate of substances in the sewer system is also unknown. Finally, available data on dissipation of APIs in receiving waters is mostly generated under controlled laboratory conditions and dissipation in natural aquatic systems is often much

66

67

68

69

70

71

72

73

74

75

76

77

78

79

80

81

82

83

84

85

86

87

88

89

slower than in the laboratory (Fono et al., 2006). When all of these different factors are considered, it is perhaps not surprising that the selection of the input parameters for exposure modelling for APIs can be challenging and that, while some exposure modelling of this type has been successful for some contaminants (Ort et al., 2009), predictions do not always agree with observed measurements of APIs in the field (Metcalfe et al., 2008). One approach to overcome the problem of the parameter selection process is to use monitoring data alongside inverse modelling to derive model input parameters. In this approach, data on measured concentrations of APIs within a study system are used in the models to back calculate one or more model input parameter. The derived parameters can then be employed to model exposure in other scenarios. The advantage of this approach in API exposure modelling is that it accounts for variability in factors such as metabolism of APIs within the population in the catchment; dissipation in the sewer network; effects of different types of treatment technologies that are employed; and the different dissipation processes that occur in surface waters. Inverse modelling, based on data on environmental occurrence, has already successfully been used to estimate usage of illicit drugs for different regions around the world (Zuccato et al., 2011) and emissions and half-lives of selected APIs into/in European surface waters (Pistocchi et al., 2012). In this paper we present and evaluate a combined monitoring and modelling approach that uses prescription and monitoring data to estimate removal of pharmaceuticals between the point of use and emission into surface waters. We then show how the removal estimates can be used to estimate concentration distributions for API in water bodies at the landscape scale. We illustrate the utility of the approach by assessing the risks of 12 commonly used APIs across surface waters in England and Wales.

1. Methods

91

92

93

94

95

96

97

98

99

100

101

102

103

104

105

106

107

108

109

110

111

112

113

114

115

1.1.Monitoring data

The measured data on concentrations of APIs in surface waters was taken from a recent study into the occurrence of APIs in surface and drinking waters in England and Wales (Boxall et al., 2012a). The twelve study APIs (Table 1) covered a range of chemical classes and varied in terms of their physico-chemical properties. The study was carried out at four catchments, which varied in terms of the population served and in the type of wastewater treatment technologies employed (Table 2). Triplicate samples of surface water (2.5 L) were taken from a single point in each catchment every 4 weeks for a period of 12 months. Following collection, these were immediately transported back to the laboratory where they were extracted onto HLB solid phase extraction cartridges before being analysed by LC-MS/MS using a Waters Acquity Ultra-Performance Liquid Chromatography (UPLC) system (Waters, Milford MA, US) fitted with a UPLC HSS T3 C18 column. Gradient elution was used with mobile phases consisting of 5 mM ammonium acetate in water and methanol. Concentrations were determined by comparison of peak areas with those of known matrix-matched standards. For a number of analytes (atenolol, carbamazepine, fluoxetine and ibuprofen), internal standards, comprising the deuterated form of the compound, were used to correct for losses during the extraction process and/or suppression or enhancement of the MS signal. In the event that internal standards were not available, sample over-spiking at a range of concentrations was used to assess recovery of a compound. Seven of the 12 study compounds were detected in surface waters at sub-µg/l concentrations. Mean concentrations and concentration ranges are shown in Table 1.

136

137

138

139

140

135

116

117

118

119

120

121

122

123

124

125

126

127

128

129

130

131

132

133

134

1.2.Model Description

The modelling was carried out using the LF2000-WQX model (Williams et al., 2009), which is a spatially-based modelling framework that has been widely applied to a number of chemicals discharged down-the-drain (Williams *et al.*, 2009; Rowney *et al.*, 2009; Price *et al.*,

2010a and b; Janna et al., 2011) and so only the parts salient to this analysis will be described here. LF2000-WOX is the water quality extension model to the Low Flows 2000 (LF2000) software system (Young et al., 2003; Environment Agency, 2004), which is a geographical information system (GIS) based decision support tool designed to estimate river flows at ungauged sites. It combines hydrological models estimating the magnitude and variability of flows across a catchment with a water quality model. The water quality model is driven by discharges from sewage treatment plants (STPs), the locations of which are preset in the model along with data describing the population served, treatment type and dry weather flow of each works. The outputs of the model are mean and 90th and 95th percentile concentrations for each river reach within the catchment being modelled. Calculation of concentrations in river reaches is based on a simple mass balance mixing equation which is applied in an iterative Monte Carlo simulation using the method of combining distributions proposed by Warn and Brew (Warn and Brew, 1980). Point-source effluent emissions are combined with reach-specific flow statistics to calculate in-river concentrations after mixing at the point of discharge, allowing for upstream concentrations of the pharmaceutical. Flow in the river and flow volume from the sewage works are described as distributions. The other parameters are held constant. The river flow is characterised as log-normal and the sewage works flows as normal. Changes in concentration with 'flow time' due to dilution, from e.g. inputs from tributaries, and degradation also are calculated. The emissions of an API for a given STP are typically derived from prescription data and STP characteristics. The STP inflow concentration (C_i) is estimated from the projected/actual per capita mass of chemical used/excreted (M, µg/cap/day), and the STP dry weather flow (*DWF*, L/day), using Equation 1:

164

141

142

143

144

145

146

147

148

149

150

151

152

153

154

155

156

157

158

159

160

161

162

$$C_i = \frac{M \cdot P}{DWF}$$
 Equation 1

Where *P* is the population served by the works and was obtained from the water utilities operating each works for all works across England and Wales. A normal distribution is assumed for DWF. The other parameters are fixed for each sewage treatment works.

The model allows removal in treatment efficiency to be considered using a global removal rate (r) including sewer removal, primary treatment and secondary treatment. The value of r can be specific to each STP modelled and can be varied according to the type of plant and levels of treatment applied. The final concentration in the effluent (C_{eff}) , is thus

174
$$C_{eff} = C_i \cdot (1-r)$$
 Equation 2

Equations 1 and 2 describe the STP process. They are calculated in turn for each of the Monte

Carlo iterations, so that the point source emission is expressed as a distribution.

Within LF2000-WQX, the whole catchment is structured as a network of interconnected model reaches. Reaches are defined as river stretches between model features, which are usually defined by significant tributaries, confluence of model reaches and STPs. Within the river, the model can simulate either conservative (no in-stream removal) or degradable (instream removal) substances. Modelling a degradable determinand, the concentration downstream (C_{DS}), after in-stream removal is defined with a first order exponential decay:

$$C_{DS} = C_{EP} e^{-kt}$$
 Equation 3

Where C_{EP} is the concentration in the river at the point of entry of an STP discharge, k (day⁻¹) is the decay rate, and t (day) is the time of travel along a reach defined as the reach length divided by the velocity of the river.

1.3. Estimation of removal of APIs using inverse modelling and comparison with removal estimates using standard modelling approach

Inverse modelling was used to estimate the mean, maximum and minimum removal of the study APIs between the point of use and the point at which surface water was sampled for the four study sites. Estimates of use of APIs were based on UK usage in 2009 (IMS Health, 2012) and were expressed as a per capita consumption per day (derived using the estimated UK population of 61,126,832; Eurostat, 2012). The LF2000-WQX model was run for all of the monitoring study sites, using only the per capita usage data (Table 1). It was assumed that all the prescribed drugs were consumed and excreted and that there was no removal in the STPs. For each API, percentage effective removals were calculated by dividing the measured value for an API at each of the study sites by the predicted mean value for the specific site. To allow comparison of the inverse modelling removal estimates with removal estimates from the 'standard' forward approach to API exposure modelling, removal percentages were also calculated for each of the monitoring study sites based on published data on metabolism, removal in treatment and dissipation in surface waters (Table 3). Where a range of values were reported for these input parameters, lowest and highest values were used to produce 'worst' and 'best' case estimates of removal. For use in broader modelling, a correction was made, using dissipation data from Table 3, to the inverse modelled removal rates to account for the in stream-dissipation of a study compound between the points of emission to the catchments and the monitoring points.

211

212

213

214

215

191

192

193

194

195

196

197

198

199

200

201

202

203

204

205

206

207

208

209

210

1.4. Evaluation of modelling approach against monitoring data

To evaluate the performance of the approach, predictions of concentrations in river catchments in England and Wales were compared with measured environmental concentrations from a range monitoring studies that have been performed in the UK over the

past eleven years (Boxall *et al.*, 2012b; Hilton *et al.*, 2003; Ashton *et al.*, 2004; Thomas and Hilton, 2004; Bound and Voulvoulis, 2006; Roberts and Bersuder, 2006; Roberts and Thomas, 2005; Kasprzyk-Hordern *et al.*, 2007 and 2009; Kasprzyk-Hordern and Baker, 2012; Zhang and Zhou, 2007; Zhou *et al.*, 2009; Table 4). Mean concentrations were then obtained for each sampling point and these mean MECs were then collated into one single distribution using the approach described by Straub (2008) and Metcalfe *et al.* (2008). Median and upper and lower quartiles were derived for the concentration distributions. Concentrations of monitored APIs were then estimated for all river reaches in the monitored catchments using the mean, minimum and maximum removal rates that were derived from the inverse modelling and corrected for in stream dissipation. Concentration distributions and associated summary statistics from the monitoring data analyses and the modelling were then compared.

1.5. Assessment of pharmaceutical risks to aquatic systems in England and Wales

The average, maximum and minimum removal estimates and corrected for in-stream dissipation data were then used in the LF2000-WQX model to predict concentrations of the 12 study APIs in 3117 river reaches distributed across 22 large catchments in England and Wales serving a population of 21 million people. Annual mean predicted environmental concentrations (PECs) were obtained for each pharmaceutical for every reach in each catchment.

To assess the implications of the predicted exposure distributions in terms of ecological risks, data on the acute and chronic (growth and reproduction) toxicity of the study APIs to algae, invertebrates and fish were extracted from the literature (Table 5). With the exception of naproxen, these data were used to derive predicted no-effect concentrations (PNECs) for each study pharmaceutical using assessment factors recommended by the European Chemicals Agency (ECHA, 2010). Studies reporting non-regulatory endpoints (e.g. biomarker,

histological and behavioural responses) were not considered in the derivation of PNECs. For naproxen, the Environmental Reference Concentration (ERC) proposed by Murray-Smith *et al* (2012) was used. Risk characterisation ratios (RCRs) were then calculated for each river reach using equation 4.

246
$$RCR = \frac{PEC}{PNEC \text{ or } ERC}$$
 Equation 4

Estimated RCRs for all the reaches in all the 22 catchments were then combined in order to develop risk distributions for each pharmaceutical. An RCR \geq 1 was considered as indicative of an unacceptable risk posed by an API to the aquatic population in a reach. In the past, diclofenac has been identified as a potential priority substance under the European Water Framework Directive and an environmental quality standard (EQS) of 0.1 μ g/l was proposed for this API. Therefore, in addition to deriving RCR distributions for diclofenac, we also compared exposure predictions to the proposed EQS value to see what the implication of the EQS would have been had it been introduced.

2. Results

2.1. Comparison of removal using forward with removal based on monitoring data at the study sites

Mean inverse modelling-based estimates of removal between the point of prescription/sale and the point of monitoring into the surface waters for the monitoring study sites ranged from 90.63 (carbmazepine) to 99.86% (ibuprofen) (Table 6). Concentrations of cyclophosamide, fluoxetine, ketoprofen, orlistat and simvastatin were below detection limits in the monitoring study, so it was only possible to estimate a minimum removal rate for these substances — these were all greater than 95.5% (Table 6). In comparison, estimates of effective removal,

based on forward modelling using data on usage, metabolism and dissipation in wastewater treatment and surface waters resulted in 'worst' case estimates of between 4 (atenolol) and 97.1% (naproxen) and 'best' case estimates of between 70.2 (trimethoprim) and 99.8% (ibuprofen) removal between use by the patient and the sampling points for the four study sites (Table 6). Mean percentage removal values for carbamazepine, diclofenac, fluoxetine, furosemide and trimethoprim, obtained from usage, metabolism and wastewater and surface water dissipation data were lower than removal values obtained using inverse modelling of the monitoring data and for selected compounds (e.g. trimethoprim), there was a large difference between the two approaches. Due to a lack of data, it was not possible to estimate removal percentages for cyclophosphamide, ketroprofen, orlistat and simvastatin using the forward modelling approach. Correction of the inverse modelling data for in-stream dissipation of the study compounds indicated that on average between 90.01 (atenolol) and 99.84% (ibuprofen) is removed between the point of prescription/use and the points of emission from treatment plants within the monitoring study catchments (Table 6). With the exception of atenolol where on average 3% of the compound was estimated to have dissipated in the rivers within the catchment, instream dissipation was found to actually play a negligible role in the overall dissipation of the study compounds and the monitoring sites. Never-the-less, the corrected values were employed in the subsequent landscape scale exposure modelling.

285

286

287

288

289

290

266

267

268

269

270

271

272

273

274

275

276

277

278

279

280

281

282

283

284

2.2. Comparison of exposure predictions against monitoring data

It was possible to obtain good datasets on mean concentrations of atenolol, carbamazepine, diclofenac, ibuprofen and trimethoprim in surface waters at different points within 10 catchments allowing concentration distributions to be derived (Figure 1). Mean, minimum and maximum estimated removal rates between the point of use and emission to surface

waters were used in the LF2000-WQX model to estimate mean concentrations of the study compounds for every river reach in the 10 monitored catchments in England and Wales. Summary statistics for the distributions of mean predicted concentrations for all river reaches in the monitored catchments, obtained from the modelling, are shown in Table 7. With the exception of ibuprofen, there was good agreement between the monitored and modelled distributions (Table 7; Figure 1). For ibuprofen, the modelled median concentrations and upper and lower quartiles for the distributions were substantially smaller than the summary statistic values obtained from monitoring studies.

2.3. Assessment of risks of APIs to surface waters in England and Wales

To assess the implications of the distributions of concentrations of the APIs, concentrations were used alongside ecotoxicity data to characterise the level of risk posed by each API in each of the river reaches modelled. Risk characterisation ratios for the study APIs for the 22 catchments (Figure 2) show that, for trimethoprim, furosemide, diclofenac and atenolol, RCRs were 0.008 or lower, indicating that these substances pose a very low risk to aquatic systems in England and Wales (Figure 2A). While the maximum RCRs of greater than one were obtained for carbamazepine, fluoxetine and simvastatin in one of the 3312 river reaches and for orlistat in 12 river reaches, simvastatin exceeded one, in the vast majority of reaches a RCRs were lower than one indicating that these substances generally pose an acceptable risk to the aquatic environment. For orlistat, simvastatin and fluoxetine, exposure estimates are based on limits of detection so in reality RCRs will be lower still. However, the maximum RCR for ibuprofen was 174 and, for this compound, 49.5% of river reaches across the 22 catchments were predicted to be at risk. When the catchments were considered individually (Figure 2B), nine of the catchments were found to have median concentrations for ibuprofen greater than the PNEC. The proposed EQS for diclofenac is 320-times lower than the

calculated PNEC. Comparison of this value with the exposure data suggested that 4.5% of river reaches in the 22 catchments would have concentrations higher than the proposed standard (Figure 2A), while none would have a concentration higher than the PNEC.

319

320

321

322

323

324

325

326

327

328

329

330

331

332

333

334

335

336

337

338

339

340

316

317

318

3. Discussion

A number of studies over the past few years have applied modelling approaches to predict the occurrence and risks of APIs in surface waters in different regions of the world (Williams et al., 2009; Hannah et al., 2009; Letzel et al., 2009). Typically, the modelling uses information on the usage of an API in an area, metabolism, fate in wastewater treatment systems and fate in receiving waters to estimate surface water concentrations. Comparison of estimates of removal, obtained using these information, with estimates of total removal, obtained using inverse modelling based on monitoring data for the four study sites, indicates that the standard modelling approach can either over- or under-estimate removal of pharmaceuticals in the real environment. This is probably one reason why previous studies have often shown little correlation between measured and modelled data (e.g. Metcalfe et al. 2008). The mismatch between the inverse modelled removal rates and rates obtained from usage, metabolism and dissipation data are likely explained by a number of factors, including: not all of a prescribed API is released to the wastewater system; metabolism in the actual population is greater or lower than indicated by literature studies on a few individuals; and variability in the types and performance of wastewater treatment works. When ranges of values are available for a particular model input parameter, there were also large differences between the maximum and minimum removal percentages for some compounds. This was particularly true for atenolol and furosemide, where removal was estimated to range from 4.0 to 97.9% and from 10 to 77.5%, respectively, highlighting the difficulty in selecting model input data. Smaller variation was seen for the inverse-modelling derived total removal rates

across the four study sites. Differences in demographic characteristics and the treatment technologies across the sites may contribute to the differences between the forward and inverse modelling derived values across the sites. For six of the seven study compounds, where extensive datasets were available on concentrations in rivers in England and Wales, there was close agreement between the results of the exposure modelling and the monitoring data. The disagreement between modelled and measured distribution statistics for ibuprofen may be partly explained by the fact that the ibuprofen monitoring dataset was dominated by measurements made in 2003 during periods of low precipitation when dilution of effluent through the wastewater treatment plants and dilution by receiving waters would be small. In addition, IMS Health data indicate that usage of ibuprofen in 2003 was higher than in 2009 when our study was performed (i.e. 293,802 kg in 2003 compared with 277,465 kg in 2009). There have also been significant advances in analytical methodologies since 2003, which have reduced the occurrence of analytical artefacts such as matrix interferences. In a recent large-scale monitoring study involving analysis of around 8,000 samples of undiluted wastewater effluents at over 160 treatment works, median concentrations for ibuprofen have been reported to be 330 ng/l (95 percentile concentration = $2.48 \text{ ug } \text{ }^{-1}$) suggesting the monitoring data from the earlier study are likely not typical of concentrations across the broader landscape (UKWIR, 2012). The inverse model results better reflect the much smaller values measured for ibuprofen in the most recent monitoring studies performed in 2006, 2007 and 2009. PNECs were derived from available acute and chronic ecotoxicity studies using standard endpoints such as mortality, reproduction and growth and assessment factors recommended by ECHA. Numerous studies have also explored effects of APIs on non-standard endpoints such as behaviour, histology and biochemical effects, sometimes at concentrations much lower than the standard endpoints (Hoeger et al., 2005; Ankley et al., 2007; Stanley et al.,

341

342

343

344

345

346

347

348

349

350

351

352

353

354

355

356

357

358

359

360

361

362

363

364

2007; Boxall et al., 2012b). However, in this study we did not use these data to inform the PNEC derivation. This approach is consistent with the recent Technical Guidance Document for deriving Environmental Quality Standard (EQSs) in the scope of the European Water Framework Directive (EQS-TGD, 2011). With the exception of diclofenac, these nonstandard effects are seen at concentrations higher than the estimated PNECs, providing reassurance that the PNEC values for most of the study substances are protective against more subtle effects. For diclofenac, the EQS value is slightly higher than concentrations where histological and biochemical effects have been reported. A recent study (Memmert et al., 2103) has questioned the reliability of the conclusions on the histopathology studies on which the EQS is based and it is possible that the EQS is overly conservative. With the exception of ibuprofen, where a risk was identified for 49.5% of river reaches, RCRs for the other study compounds in river reaches in England and Wales were generally lower than one (for carbamazepine, orlisat, fluoxetine and simvastatin a risk was identified in one, one and 12 of the 3312 river reaches respectively), indicating that the other compounds pose an acceptable risk to the UK environment. The findings for ibuprofen agree with conclusions from other studies into the risks of APIs in aquatic systems, where ibuprofen has been highlighted as a drug of potential concern in river systems (Christensen et al., 2009; Lienert et al., 2007). We would therefore advocate that further work is carried out to explore the wider occurrence of ibuprofen in surface waters in England and Wales and to explore whether effects are occurring in the catchments where a significant proportion of river reaches are predicted to be at risk; the PNEC may also need to be re-assessed as well. Comparison of exposure predictions for diclofenac with the previously proposed EQS for diclofenac, indicated that 4.5% of river reaches would have exceeded the EQS had it been adopted. This percentage is in agreement with the value of 3-5% previously predicted for

366

367

368

369

370

371

372

373

374

375

376

377

378

379

380

381

382

383

384

385

386

387

388

391 2013). 392 Overall, this study demonstrates the potential of using inverse modelling alongside 393 monitoring data to generate model input data in exposure and risk assessment. As a total 394 removal rate is estimated for a broad scale, the approach offers a number of advantages, i.e. it 395 takes into account factors such as the non-use of prescribed drugs by patients; it addresses 396 differences in metabolism across the population; it accounts for dissipation processes in the 397 local sewerage network and it accounts for differences in effectiveness of different 398 wastewater treatment technologies in a catchment. The four study sites used in this study 399 were based in four different counties located in the South East and Midland regions of 400 England so the approach appears to be effective at estimating exposure for different regions 401 in a country the size of England. This is backed up by the comparisons of exposure 402 predictions for the catchments that have been monitored in England and Wales with the 403 experimentally-derived data. 404 The approach is, however, reliant on the availability of good quality monitoring data and 405 cannot be applied to compounds that are not yet in use. Cultural and demographic differences 406 might mean that the total removal predictions from this study cannot be applied to other 407 countries. However, there is no reason why a similar monitoring and modelling strategy to 408 that employed in the current study could not be applied elsewhere in order to generate state-409 or country-specific removal rates and hence assess the broad scale exposure risks of APIs in 410 other regions of the world. The concept could also be applied at different stages in the 411 pathway of a pharmaceutical from the patient to environmental receptors to better understand 412 key dissipation processes for pharmaceuticals to inform future modelling initiatives.

EQS exceedences in rivers in England using the forward modelling approach (Johnson et al.,

413

414

390

Acknowledgements

We are grateful to the Drinking Water Inspectorate for funding the monitoring component of this work. Many thanks to Mathieu Guillaume (F. Hoffmann-La Roche, Basle) for help with the IMS Health data. IMS Health is acknowledged for the use of API sales data; however, analysis of IMS Health data was arrived at independently by the authors of the present paper on the basis of the data and other information and IMS Health is not responsible for any reliance by recipients of the data or any analysis thereof. We are grateful to an anonymous reviewer who provided constructive comments on an earlier version of this manuscript.

422

423

References

- 424 Anderson PD, D'Aco VJ, Shanahan P, Chapra SC, Buzby ME, Cunningham VL, et al.
- 425 Screening analysis of human pharmaceutical compounds in US surface waters.
- 426 Environmental Science and Technology 2004; 38: 838–849.
- 427 Ando T, Nagase H, Eguchi K, Hirooka T, Nakamura T, Miyamoto K, et al. A novel method
- 428 using cyanobacteria for ecotoxicity test of veterinary antimicrobial agents. Environ Toxicol
- 429 *Chem* 2007; 26(4): 601–606.
- 430 Andreozzi R, Raffaele M, Paxeus N. Pharmaceuticals in STP effluents and their solar
- photodegradation in aquatic environment. *Chemosphere* 2003; 50: 3019-3030.
- 432 Ankley GT; Brooks BW; Huggett DB; Sumpter JP. Repeating history: Pharmaceuticals in
- 433 the environment. *Environ. Sci. Technol.* 2007; 41: 8211-8217.
- 434 Araujo L, Villa N, Camargo N, Bustos M, Garcia T, de Jesus Prieto A. Persistence of
- 435 gemfibrozil, naproxen and mefenamic acid in natural waters. Environ. Chem. Lett. 2011;
- 436 9:13-18.
- 437 Ashton D, Hilton M, Thomas KV. Investigating the environmental transport of human
- pharmaceuticals to streams in the United Kingdom. Sci. Tot. Environ. 2004; 333, 167–184.

- Bagley CM, Bostick FW, DeVita VT. Clinical pharmacology of cyclophosphamide. Cancer
- 440 Res. 1973; 33, 226-233.
- Bound JP, Voulvoulis N. Predicted and measured concentrations for selected pharmaceuticals
- in UK rivers: Implications for risk assessment. Water Res. 2006; 40, 2885–2892.
- Boxall ABA. The environmental side effects of medication. EMBO Reports. 2004; 5(12),
- 444 1110-1116.
- Boxall ABA, Fogg LA, Kay P, Blackwell PA, Pemberton EJ, Croxford A. Veterinary
- 446 medicines in the environment. Reviews in Environmental Contamination and Toxicology.
- 447 2004; 180: 1-91.
- Boxall ABA, Rudd MA, Brooks BW, Caldwell DJ, Choi K, Hickmann S. Pharmaceuticals
- and personal care products in the environment: What are the key questions. *Environ. Health*
- 450 *Persp.* 2012a; 120(9):1221-1229.
- Boxall ABA, Monteiro SC, Fussell R, Williams RJ, Bruemer J, Greenwood R, et al. *Targeted*
- 452 Monitoring for Human Pharmaceuticals in Vulnerable Source and Final Waters. Department
- 453 for Environment, Food and Rural Affairs, London, England 2012b.
- Breton R, Boxall ABA. Pharmaceuticals and personal care products in the environment:
- regulatory drivers and research needs. *QSAR Comb. Sci.* 2003; 22, 399-409.
- 456 Brooks BW, Richards SM, Weston J, Turner PK, Stanley JK, La Point TW, et al. Aquatic
- ecotoxicology of fluoxetine: a review of recent research. In: Dietrich D., Webb S., Petry T.,
- Eds. Hot spot pollutants: pharmaceuticals in the environment. Academic/Elsevier, New York,
- 459 USA, 2005.
- 460 Brun GL, Bernier M, Losier R, Doe K, Jackman P, Lee H-B. Pharmaceutically active
- 461 compounds in Atlantic Canadian sewage treatment plant effluents and receiving waters, and
- 462 potential for environmental effects as measured by acute and chronic toxicity. Environ.
- 463 *Toxicol. Chem.* 2006; 25(8): 2163-2176.

- 464 Bürge IJ, Poiger T, Müller MD, Buser H-R. Caffeine, an anthropogenic marker for
- 465 wastewater contamination of surface waters. Environmental Science and Technology 2003;
- 466 37: 691–700.
- Buser H-R, Poiger T, Müller MD. Occurrence and environmental behaviour of the chiral
- 468 pharmaceutical drug ibuprofen in surface waters and in wastewater. Environmental Science
- 469 and Technology 1999; 33: 2529–2535.
- 470 Christensen AM, Markussen B, Baun A. Halling-Sorensen, B. Probabilistic environmental
- 471 risk characterization of pharmaceuticals in sewage treatment plant discharges. *Chemosphere*
- 472 2009; 77(3): 351-358.
- 473 Cleuvers M. Mixture toxicity of the anti-inflammatory drugs diclofenac, ibuprofen, naproxen,
- and acetylsalicylic acid. *Ecotox. Environ. Safe.* 2004; 59: 309–315.
- Daughton C, Ternes T. Pharmaceuticals and personal care products in the environment:
- agents of subtle change? *Environ. Health Persp.* 1999; 107: 907–937.
- 477 De Lorenzo ME, Fleming J. Individual and mixture effects of selected pharmaceuticals and
- 478 personal care products on the marine phytoplankton species Dunaliella tertiolecta. Arch.
- 479 Environ. Contam. Toxicol. 2008; 54:203–210
- Dorne JLCM. Metabolism, variability and risk assessment. Toxicology 2010; 268, 156-164.
- 481 ECHA. Guidance on information requirements and chemical safety assessment. Chapter
- 482 R.10: Characterisation of dose [concentration]-response for environment. ECHA, Helsinki,
- 483 Finland, 2010.
- 484 EMA. Guideline on the environmental risk assessment of medicinal products for human use.
- European Medicines Agency, London, U.K., 2006.
- 486 Environment Agency. Development of the integrated water resources and water quality
- 487 *modelling system*. Environment Agency, Bristol, England, 2004.

- 488 EQS-TGD, Common Implementation Strategy for the Water Framework Directive
- 489 (2000/60/EC). Technical guidance for deriving environmental quality standards: Guidance
- 490 *Document No.* 27. European Commission, Brussels, 2011.
- 491 Eurostat. *Population in Europe*. European Union Statistics Office, Brussels, 2012.
- 492 Ferrari B, Mons R, Vollat B, Fraysse B, Paxeus N, Lo Giudice R. et al. Environmental risk
- 493 assessment of six human pharmaceuticals: are the current environmental risk assessment
- 494 procedures sufficient for the protection of the aquatic environment? *Environ. Toxicol. Chem.*
- 495 2009; 23(5): 1344-1354.
- 496 Ferrari B, Paxeeus N, Lo Giudice R, Pollio A, Garric J. Ecotoxicological impact of
- 497 pharmaceuticals found in treated wastewaters: study of carbamazepine, clofibric acid, and
- 498 diclofenac. Ecotox. Environ. Safety 2003; 55: 359-370.
- 499 Fono LJ, Kolodziej EP, Sedlak DL. Attenuation of wastewater-derived contaminants in an
- effluent-dominated river. Environ. Sci. Technol. 2006; 40, 7257-7262.
- Garcia-Ac A, Segura PA, Gagnon C, Sauvé S. Determination of bezafibrate, methotrexate,
- 502 cyclophosphamide, orlistat and enalapril in waste and surface wtaers using on-line solid-
- 503 phase extraction liquid chromatography coupled to polarity-switching electrospray tandem
- mass spectrometry. *Journal of Environmental Monitoring* 2009; 11: 830–838.
- 505 Gros M, Petrovič M, Ginebreda A, Barceló D. Removal pf pharmaceuticals during
- 506 wastewater treatment and environmental risk assessment using hazard indexes. Environ Int
- 507 2010; 36: 15–26.
- 508 GSK. Safety datasheet: Alli. GSK, Brentford, Middlesex, UK, 2008.
- Han S, Choi K, Kim J, Ji K, Kim S, Ahn B, et al. Endocrine disruption and consequences of
- 510 chronic exposure to ibuprofen in Japanese medaka (Oryzias latipes) and freshwater
- 511 cladocerans *Daphnia magna* and *Moina macrocopa*. *Aquat. Toxicol*. 2010; 98: 256-264.

- Hannah R, D'Aco VJ, Anderson PD, Buzby ME, Caldwell DJ, Cunningham VL. Exposure
- assessment of 17α -ethinylestradiol in surface waters of the United States and Europe.
- 514 Environ. Toxicol. Chem. 2009; 28(12), 2725-2732.
- 515 Heberer, T. Occurrence, fate, and removal of pharmaceutical residues in the aquatic
- environment: a review of recent research data. *Toxicology Letters* 2002; 131: 5–17.
- Heberer, T.; Reddersen, K.; Mechlinski, A. From municipal sewage to drinking water: fate
- and removal of pharmaceutical residues in the aquatic environment in urban areas. Wat. Sci.
- 519 *Technol.* 2002; 46(3), 81–88.
- 520 Hilton, M.J.; Thomas, K.V.; Ashton, D. Targeted Monitoring Programme for
- 521 Pharmaceuticals in the Aquatic Environment. Environment Agency, Bristol, England, 2003.
- 522 Hirsch, R.; Ternes, T.; Heberer, K.; Kratz, K. Occurrence of antibiotics in the aquatic
- 523 environment. Sci. Total Environ. 1999; 225, 109-118.
- Hoeger, B.; Kollner, B.; Dietrich, D.R.; Hitzfeld, B. Water-borne diclofenac affects kidney
- and gill integrity and selected immune parameters in brown trout (Salmo trutta f. fario).
- 526 Aquat. Toxicol. 2005; 75, 53-64.
- 527 IMS Health. Global Product Strategy; IMS MIDAS in PADDS. Subscription database. IMS
- Health Ltd, Danbury, USA, 2012.
- 529 Isidori M, Lavorgna M, Nardelli A, Parrella A, Previtera L, Rubino M. Ecotoxicity of
- naproxen and its phototransformation products. Sci. Tot. Environ. 2005; 348: 93-101.
- Janna H, Scrimshaw MD, Williams RJ, Churchley J, Sumpter JP. From dishwasher to tap?
- 532 Xenobiotic substances benzotriazole and tolyltriazole in the environment. Environ. Sci.
- 533 Technol. 2011; 45(9): 3858-3864.
- Johnson AC, Dumont E, Williams RJ, Oldenkamp R, Cisowska I, Sumpter JP. Do
- 535 Concentrations of Ethinylestradiol, Estradiol, and Diclofenac in European Rivers Exceed

- Proposed EU Environmental Quality Standards? Environ. Sci. Technol. 2013; 47(21): 12297-
- 537 304.
- 538 Kasprzyk-Hordern B, Baker DR. Enantiomeric profiling of chiral drugs in wastewater and
- 539 receiving waters. *Environ. Sci. Technol.* 2012, 46; 1681–1691.
- Kasprzyk-Hordern B, Dinsdale RM, Guwy AJ. Multi-residue method for the determination of
- basic/neutral pharmaceuticals and illicit drugs in surface water by solid-phase extraction and
- 542 ultra performance liquid chromatography-positive electrospray ionisation tandem mass
- 543 spectrometry. J. Chromat. A. 2007; 1161, 132–145.
- Kasprzyk-Hordern B, Dinsdale RM, Guwy AJ. The removal of pharmaceuticals, personal
- 545 care products, endocrine disruptors and illicit drugs during wastewater treatment and its
- impact on the quality of receiving waters. *Water Res.* 2009; 43, 363–380.
- 547 Key PB, Hoguet J, Chung KW, Venturell JJ, Pennington PL, Fulton MH. Lethal and
- sublethal effects of simvastatin, irgarol, and PBDE-47 on the estuarine fish, Fundulus
- 549 heteroclitus. J. Environ. Sci. Hlth. B. 2009; 44(4): 379-382.
- 550 Kim Y, Choi K, Jung J, Park S, Kim P-G, Park J. Aquatic toxicity of acetaminophen,
- 551 carbamazepine, cimetidine, diltiazem and six major sulfonamides, and their potential
- ecological risks in Korea. *Environment International* 2007; 33: 370–375
- Kolpin D, Furlong E, Meyer M, Zaugg S, Barber L, Buxton H. Pharmaceuticals, hormones,
- and other organic wastewater contaminants in US streams, 1999 2000: A national
- 555 reconnaissance. *Environ. Sci. Technol.* 2002; 32, 1202 1211.
- Küster A, Alder AC, Escher BI, Duis K, Fenner K, Garric J. et al. Environmental Risk
- 557 Assessment of Human Pharmaceuticals in the European Union: A Case Study with the β-
- Blocker Atenolol. *Integrated Environmental Assessment and Management* 2010; 6(suppl 1):
- 559 514–523

- 560 Kwon J-W, Armbrust K. Laboratory persistence and fate of fluoxetine in aquatic
- 561 environments. Environ. Toxicol. Chem. 2006; 25(10): 2561-2568.
- Lam MW, Young CJ, Brain RA, Johnson DJ, Hanson MA, Wilson CJ. Aquatic persistence of
- eight pharmaceuticals in a microcosm study. Environ. Toxicol. Chem.2004; 23(6): 1431-
- 564 1440.
- Leclercq M, Mathieu O, Gomez E, Casellas C, Fenet H, Hillaire-Buys D. Presence and fate
- of carbamazepine, oxcarbazepine, and seven of their Metabolites at wastewater treatment
- plants. Archives of Environmental Contamination and Toxicology 2009; 56: 408-415.
- Lee H-B, Peart TE, Svoboda ML, Backus S. Occurrence and fate of rosuvastatin, rosuvastatin
- lactone, and atorvastatin in Canadian sewage and surface water samples. *Chemosphere* 2009;
- 570 77: 1285-1291.
- 571 Letzel M, Metzner G, Letzel T. Exposure assessment of the pharmaceutical diclofenac based
- on long-term measurements of the aquatic input. *Environ. Internat.* 2009; 35, 363-368.
- Lienert J, Güdel K, Escher BI. Screening Method for Ecotoxicological hazard assessment of
- 574 42 pharmaceuticals considering human metabolism and excretory Routes. Environ. Sci.
- 575 Technol. 2007; 41, 4471–4478.
- Lister A, Regan C, Van Zwol J, Van Der Kraak GJ. Inhibition of egg production in zebrafish
- 577 by fluoxetine and municipal effluents: a mechanistic perspective. Aquat Toxicol 2009; 95:
- 578 320–329.
- Memmert U, Peither A, Burri R, Weber K, Schmidt T, Sumpter JP, Hartmann A. Diclofenac:
- new data on chronic toxicity and bioconcentration in fish. *Environ. Toxicol. Chem.* 2013; 32:
- 581 442-452.
- Metcalfe CD. et al. Exposure assessment methods for veterinary and human-use medicines
- 583 in the environment: PEC vs MEC comparisons. In Pharmaceuticals in the Environment, 2nd
- 584 Edition; Kümmerer K Ed.; Springer, Berlin Heidelberg, Germany, 2008.

- 585 Monteiro SC, Boxall ABA. Occurrence and fate of human pharmaceuticals in the
- 586 environment. Rev. Environ. Contam. T. 2010; 202, 53-154.
- Murray-Smith RJ, Coombe VT, Grönlund MH, Waern F, Baird JA. Managing emissions of
- 588 active pharmaceutical ingredients from manufacturing facilities: An environmental quality
- standard approach. Integr. Environ. Assess. Man. 2012; 8(2), 320-330
- 590 Musson SE, Townsend TG. Pharmaceutical compound content of municipal solid waste
- 591 pharmaceutical compound content of municipal solid waste. J. Haz. Mater. 2009; 162, 730–
- 592 735
- Nalecz-Jawecki G.: Evaluation of the in vitro biotransformation of fluoxetine with HPLC,
- mass spectrometry and ecotoxicological tests. *Chemosphere* 2007; 70: 29–35.
- Oakes KD, Coors A, Escher BI, Fenner K, Garric J, Gust M. Environmental risk assessment
- 596 for the serotonin re-uptake inhibitor fluoxetine: Case study using the European risk
- 597 assessment framework. *IEAM* 2010; 6(1): 524-539.
- 598 Ort C, Hollender J, Schaerer M, Model-Based Evaluation of Reduction Strategies for
- 599 Micropollutants from Wastewater Treatment Plants in Complex River Networks,
- 600 Environmental Science Technology 2009; 43(9), 3214-3220.
- Paffoni C, Welte B, Gousailles M, Montiel A. Nouvellem molécules mises en cause par les
- directives européennes: de la station d'épuration à l'usine de traitement d'eau potable.
- 603 European Journal of Water Quality 2006; 37(1): 21–38.
- Park S, Choi K. Hazard assessment of commonly used agricultural antibiotics on aquatic
- 605 ecosystems. *Ecotoxicology* 2008; 17: 526–538.
- 606 Paxéus N. Removal of selected non-steroidal anti-inflammatory drugs (NSAIDs),
- 607 gemfibrozil, carbamazepine, β-blockers, trimethoprim and triclosan in conventional
- 608 wastewater treatment plants in five EU countries and their discharge to the aquatic
- environment. Water Science and Technology 2004; 50(5): 253-260.

- 610 Piecha M, Sarakha M, Trebse P, Kocar D. Stability studies of cholesterol lowering statin
- drugs in aqueous samples using HPLC and LC–MS. Environ. Chem. Lett. 2010; 8:185-191.
- 612 Pistocchi A, Marinov D, Pontes S, Gawlik BM. Continental scale inverse modeling of
- 613 common organic water contaminants in European Rivers. *Environ. Poll.* 2012; 162, 159-167.
- Price O.R, Williams RJ, van Egmond R, Wilkinson MJ, Whelan MJ. Predicting accurate and
- ecologically relevant regional scale concentrations of triclosan in rivers for use in higher-tier
- aquatic risk assessments. *Environ. Internat.* 2010; 36, 521-526.
- 617 Price OR, Williams RJ, Zhang Z, van Egmond R. Modelling concentrations of
- decamethylcyclopentasiloxane in two uk rivers using 1f2000-wqx. Environ. Pollut. 2010;
- 619 158(2), 356-360.
- 620 Roberts PH, Bersuder P. Analysis of OSPAR priority pharmaceuticals using high-
- performance liquid chromatography-electrospray ionisation tandem mass spectrometry. J.
- 622 Chromat. A. 2006; 1143, 143–150.
- Roberts PH, Thomas KV. The occurrence of selected pharmaceuticals in wastewater effluent
- and surface waters of the lower Tyne catchment. Sci. Tot. Environ. 2005; 356, 143–153.
- Rowney NC, Johnson AC, Williams RJ. Cytotoxic drugs in drinking water: A prediction and
- 626 risk assessment exercise for the thames catchment in the united kingdom. *Environ. Toxicol.*
- 627 *Chem.* 2009; 28(12), 2733-2743.
- 628 Runkel R, Chaplin M, Boost G, Segre E, Forchielli E. Absorption, distribution, metabolism
- and excretion of naproxen in variouslaboratory animals and human subjects. J. Pharm. Sci.,
- 630 1972; 61, 703-708.
- 631 SCHER. Opinion on 'Chemicals and the water framework directive: draft environmental
- quality standards' Diclofenac. SCHER, Brussels, Belgium, 2011.
- 633 Schowanek D, Webb S. Exposure simulation for pharmaceuticals in European surface waters
- 634 with GREAT-ER. *Toxicol. Lett.* 2002; 131, 39–50

- 635 Schwab BW, Hayes EP, Fioric JM, Mastrocco FJ, Roden NM, Cragin D. Human
- pharmaceuticals in US surface waters: A human health risk assessment. Regulat. Toxicol.
- 637 *Pharmacol.* 2005; 42(3), 296-312.
- 638 Seehusen DA, Edwards J. Patient practices and beliefs concerning disposal of medications. J.
- 639 Am. Board. Fam. Med. 2006; 19, 542–7.
- 640 SFT. Initial assessment of eleven pharmaceuticals using the EMEA guideline in Norway.
- Statens forurensningstilsyn (SFT), Strømsveien, Norway, 2006.
- 642 Sipma J, Osuna B, Collado N, Monclús H, Ferrero G, Comas J. Comparison of removal of
- pharmaceuticals in MBR and activated sludge systems. *Desalination* 2010; 250, 653–659.
- 644 Stanley JK, Ramirez AJ, Chambliss CK, Brooks BW. Enantiospecific sublethal effects of the
- antidepressant fluoxetine to a model aquatic vertebrate and invertebrate. Chemosphere 2007;
- 646 69(1), 9-16.
- 647 Straub JO, Stewart KM. Deterministic and probabilistic acute-based environmental risk
- assessment for naproxen for western Europe. *Environ. Toxicol. Chem.* 2007; 26(4):795-806.
- 649 Straub JO. Deterministic and probabilistic environmental risk assessment for diazepam. In
- 650 Pharmaceuticals in the Environment; Sources, Fate, Effects and Risks; Kümmerer K, ed.
- 651 Springer, Heidelberg, Germany, 2008.
- 652 Ternes T, Bonerz M, Schmidt T. Determination of neutral pharmaceuticals in wastewaters
- and rivers by LC-ES-tandem MS. *Journal of Chromatography A* 2001; 938: 175–185.
- Thomas KV, Hilton MJ. The occurrence of selected human pharmaceutical compounds in
- 655 UK estuaries. *Mar. Pollut. Bull.* 2004; 49, 436–444.
- 656 Upton RA, Buskin JN, Williams RL, Holford NH, Riegelman S. Negligible excretion of
- unchanged ketoprofen, naproxen, and probenecid in urine. J. Pharm. Sci. 1980; 69(11):1254-
- 658 7.
- 659 Warn AE, Brew JS. Mass balance. Wat. Res. 1980; 14(10), 1427-1434.

- 660 Williams RJ, Keller VDJ, Johnson AC, Young AR, Holmes MGR, Wells C. A national risk
- assessment for intersex in fish arising from steroid estrogens. Environ. Toxicol. Chem. 2009;
- 662 28(1), 220-230.
- Winter MJ, Lillicrap AD, Caunter JE, Schaffner C, Alder AC, Ramil M. Defining the chronic
- impacts of atenolol on embryolarval development and reproduction in the fathead minnow
- 665 (Pimephales *promelas*). *Aquatic Toxicology* 2008; 86: 361–369.
- Wu Y, Chen D-H, Kookana R. Aqueous photodegradation of selected antibiotics under
- different conditions. *Energy Procedia* 2011; 11: 2098–2103.
- Yamamoto H, Nakamura Y, Moriguchi S, Nakamura Y, Honda Y, Tamura I. Persistence and
- 669 partitioning of eight selected pharmaceuticals in the aquatic environment: Laboratory
- photolysis, biodegradation, and sorption experiments. Water Research 2009; 43: 351-362.
- Yang L-H, Ying G-G, Su H-C, Strauber JL, Adams MS, Binet MT. Growth-inhibiting effects
- of 12 antibacterial agents and their mixtures on the freshwater microalga Pseudokirchneriella
- 673 subcapitata. *Environ Toxicol Chem* 2008; 27(5): 1201–1208.
- Young AR, Grew R, Holmes MGR. Low flows 2000: A national water resources assessment
- and decision support tool. *Wat. Sci. Technol.* 2003; 48(10), 119-126.
- 276 Zhang ZL, Zhou JL. Simultaneous determination of various pharmaceutical compounds in
- water by solid-phase extraction-liquid chromatography-tandem mass spectrometry. J.
- 678 *Chromat. A.* 2007; 1154, 205–213.
- 679 Zhi J, Melia AT, Eggers H, Joly R, Patel IH. Review of limited systemic absorption of
- orlistat, a lipase inhibitor, in healthy human volunteers. J. Clinical Pharmacology 1995;
- 681 35:1103-1108.
- Zhou JL, Zhang ZL, Banks E, Grover D, Jiang JQ. Pharmaceutical residues in wastewater
- treatment works effluents and their impact on receiving river water. J. Haz. Mater. 2009;
- 684 166, 655–661.

685	Zounkova R, Odradka P, Dolezalova L, Hilscherova K, Marsalek B, Blaha L. Ecotoxicity and
686	genotoxicity assessment of cytistatic pharmaceuticals. Environ. Toxicol. Chem. 2007; 26(10):
687	2208-2214.
688	Zuccato E et al. Environmental loads and detection of pharmaceuticals in Italy. In Kümmerer
689	K, (ed) Pharmaceuticals in the Environment. Springer, Berlin, 2001.
690	Zuccato E, Castiglioni S, Tettamanti M, Olandese R, Bagnati R, Melis M. et al. Changes
691	inillicit drug consumption patterns in 2009 detected by wastewater analysis. Drug Alcohol
692	Dependence 2011; 118(2-3), 464-469.
693	
694	

Table Legends

Table 1. Usage and mean concentrations (expressed in ng l⁻¹) measured at the four study sites. Values in parentheses for the study sites indicate the measured concentration range over the 12 month study period (Measured data taken from Boxall *et al*, 2012a).

Table 2. Characteristics of study catchments in terms of population served, types of sewage treatment plants present and average residence time between discharge points and the monitoring points.

Table 3. Information on percentage of API excreted, percentage API removed in wastewater treatment removal rates, and half lives for in stream dissipation used in the modelling of the study APIs.

Table 4. Summary of the monitoring studies used in the evaluation of the modelling approach.

Table 5. Ecotoxicological data used alongside the model predictions to establish the level of risk of the study compounds across the 18 study catchments in England and Wales.

Table 6. Summary of removal percentages for the 12 study APIs at the monitoring sites, obtained using the inverse modelling approach and the traditional forward modelling approach.

Table 7. Comparison of summary statistics for modelled and measured distributions of mean concentrations of APIs for river reaches in catchments that have been monitored for APIs in England and Wales.

Figure legends

Figure 1. Cumulative distributions of mean concentrations of pharmaceuticals in river reaches in catchments in England and Wales derived from the LF2000-WQX model or from

monitoring data. Modelled distributions are developed for the catchments where a pharmaceutical has been monitored and are based on mean, maximum and minimum inverse-modelled rates of removal. Monitoring data were taken from: Hilton *et al.*, 2003; Ashton *et al.*, 2004; Thomas and Hilton, 2004; Bound and Voulvoulis, 2006; Roberts and Bersuder, 2006; Roberts and Thomas, 2006; Kasprzyk-Hordern *et al.*, 2007, 2009, 2012; Zhang and Zhou, 2007; Zhou *et al.*, 2009; Boxall *et al.*, 2012. In instances where a number of samples were taken from sites in a catchment, mean concentrations were estimated for each site. If a measured concentration was reported as < LOD then half of the LOD was used in the calculation of means.

Figure 2. Box and whisker plots (indicating median, upper and lower quartile and maximum and minimum values) of PEC/PNEC ratios for A) a range of APIs, obtained from LF-2000-WQX exposure predictions for 2950 river reaches across 22 large river catchments in England and Wales and PNEC values derived from published literature on the effects of the study compounds on aquatic organisms. Removal rates for the grey bars were based on non-detect data so are a 'worst' case indication of risks; and B) for ibuprofen in the individual 18 large river catchments modelled in England and Wales.

Table 1.

Compound	Class	Use in UK in 2009	Site 1	Site 2	Site 3	Site 4
		(Kg/yr)				
Atenolol	β-blocker	32944	43.8 (18.1 – 66.3)	54.2 (31.2 – 91.2)	26.7 (8.2 – 67.6)	41.1 (19.6 – 114)
Carbamazepine	anti-epileptic	49781	103.6 (49.4 – 199)	138.8(45.0 - 277)	272.3 (34.3 – 555)	182.6 (16.4 – 480)
Cyclophosphamide	chemotherapy agent	281	<1	<1	<1	<1
Diclofenac	non steroidal anti-inflammatory	34720	10.3 (<10 – 24.3)	18.1 (<10 – 39.0)	20.5 (<10 – 76.3)	15.9 (<10 – 47.1)
Fluoxetine	antidepressant	6377	<5	<5	<5	<5
Furosemide	diuretic	20872	10.2 (<5 - 28.9)	19.8 (6.59 – 43.1)	9.1 (<5 - 36.0)	17.4 (5.34 – 63.5)
Ibuprofen	non steroidal anti-inflammatory	277466	17.6 (6.33 – 30.8)	11.8 (<2 - 38.4)	8.4 (< 2 - 21.5)	18.2 (< 2 - 38.2)
Ketoprofen	non steroidal anti-inflammatory	1878	<1	<1	<1	<1
Naproxen	non steroidal anti-inflammatory	67672	18.2 (10.2 - 26.4)	18.1 (6.93 – 42.2)	13.6 (4.85 - 28.9)	23.1 (11.1 – 44.4)
Orlistat	anti-obesity	16669	<10	<10	<10	<10
Simvastatin	hypolipidemic	50070	< 50	< 50	< 50	< 50
Trimethoprim	antibiotic	13094	10.0 (<5 - 13.8)	9.5 (<5 – 13.8)	4.1 (<5 – 8.27)	8.9 (<5 - 26.4)

Table 2.

Site	Location	Population	Number of treatment plants upstream of sampling point	Types of STPs [†]	Average water residence between discharge and sampling (d)
1	Midlands	402227	17	SAS 1	1.27
				SB 5	
				TA 3	
				TB 8	
2	Southern England	2071445	81	SAS 12	2.33
				SB 22	
				TA 19	
				TB 28	
3	Southern England	395581	27	SAS 2	2.42
				SB 11	
				TA 8	
				TB 6	
4	South East	177801	14	SAS 2	1.46
				SB 3	
	England			TA 4	
				TB 5	

†General classification of sewage treatment plants: SAS - Secondary Activated Sludge; SB - secondary biological filter; TA - Activated Sludge with tertiary treatment; TB - Biological filter with tertiary treatment.

Table 3.

Compound	Proportion of administered compound excreted by patient (%)	Proportion of compound removed in wastewater treatment plants	Dissipation half-life in receiving water (d)
Atenolol	69 – 96	-93 - 97	3 - 30
Carbamazepine	26 - 31	-122 - 58	82 - 100
Cyclophosphamide	2.5 - 20	0	43
Diclofenac	6 - 26	-143 - 80	5
Fluoxetine	20 - 26	33	112 - 113
Furosemide	90	-119 – 75	-
Ibuprofen	11 - 47	52 – 99.7	~20
Ketoprofen	-	40–100	-
Naproxen	0.6 - 5.6	48 - 93	10.2 - 14.6
Orlistat	83.1	~90	-
Simvastatin	-	-17 – 91	7.8
Trimethoprim	50 - 70	-40 - 40.4	5.7 - 100

Data collated from: Anderson et al., 2004; Andreozzi et al., 2003; Araujo et al., 2011; Bagley et al., 1973; Boxall et al., 2002; Bürge et al., 2003; Buser et al., 1999; Garcia-Ac et al., 2009; Gros et al., 2010; Heberer et al., 2002; Kasprzyk Hordern, 2012; Kovalova et al., 2011; Kwon; & Armbrust, 2006; Küster et al., 2010; Lam et al., 2004, LeClerq et al., 2009; Lee et al., 2009; Lienert et al., 2007; Paffoni et al., 2006, Paxeus, 2004; Piecha et al., 2010; Roche, unpublished; Runkel et al., 1972; Sipma et al., 2010; Ternes et al., 2001; Upton et al., 1980; Wu et al., 2011; Yamamoto, 2009; Zhi et al., 1995; Zuccato et al., 2001.

Table 4.

Compound	River catchments	References
Atenolol	Blackwater, Derwent, Ely, Great Ouse,	1,2
	Taff, Thames	
Carbamazepine	Blackwater, Derwent, Ely, Great Ouse,	1-3
	Taff, Thames	
Diclofenac	Blackwater, Derwent, Ely, Great Ouse,	1-4
	Lea, Nene, Sussex Ouse, Taff, Thames,	
	Welland	
Furosemide	Blackwater, Derwent, Ely, Great Ouse,	1,2
	Taff, Thames	
Ibuprofen	Blackwater, Derwent, Ely, Great Ouse,	1-6
	Lea, Nene, Sussex Ouse, Taff, Thames,	
	Tyne, Welland	
Naproxen	Blackwater, Derwent, Ely, Great Ouse,	1,2
	Taff, Thames	
Trimethoprim	Blackwater, Derwent, Ely, Great Ouse,	1-3,6
	Lea, Nene, Taff, Thames, Tyne, Welland	

¹⁻Kasprzyk Hordern et al., (2008); 2- Boxall et al., (2012); Zhang and Zhou (2007); 4- Environment Agency (2003); 5 – Bound and Volvoulis (2006); 6 – Roberts and Thomas (2006)

Table 5.

	Test	EC50 (mg/l)	NOEC (mg/l)	AF	PNEC (µg/l)	Source
				(REACH)		
Atenolol	Pseudokirchneriella subcapitata 72 h growth		128.8			Küster et al., 2010
	Daphnia magna 21 d reproduction		8.872			Küster et al., 2010
	Daphnia magna 21 d second generation reproduction test started with 1 st gen. brood		1.48	10	148	Küster et al., 2010
	Pimephales promelas 32 d hatching, survival, growth		3.2			Winter et al., 2008
Carbamazepine	Daphnia magna 48 h	13.8				Ferrari et al., 2009
•	Pseudokirchneriella subcapitata 96 h growth		100			Ferrari et al., 2003
	Ceriodaphnia dubia 7 d reproduction		0.025	10	2.5	
	Danio rerio 10 d ELS		25			
Cyclophosphamide	Pseudokirchneriella subcapitata 96 h growth	930		100	560	Zounkova et al., 2007; SFT, 2006
	Daphnia magna 48 h immobilisation	>1000				
	Daphnia magna reproduction		56			
Diclofenac	Proposed EQS under the Water Framework				0.1	SCHER, 2011a
	Directive					
	Rainbow trout reproduction		0.32	10	32	Novartis (personal comm)
	Zebra fish reproduction		0.32	10	32	
Fluoxetine	Daphnia magna chronic		0.089			Brooks et al., 2005; Lister et al., 2009;
	Danio rerio reproduction		0.0032			Oaks <i>et al.</i> , 2010
	Desmodesmus subspicatus growth		0.0006	10	0.06	
	Thamnocephalus platyurus 24 h		0.76			Nalecz-Jawecki, 2007
Furosemide	Pseudokirchneriella subcapitata 96 h growth		70	1000	70	Isidori et al., 2006
	Daphnia magna 48 h immobilisation	60.62				
Ibuprofen	Scenedesmus subspicatus 72 h growth	342		10	0.01	Han et al., 2010; Cleuvers, 2004
	Daphnia magna 21 d reproduction		<1.23			
	Oryzias latipes 120 d (post hatch) survival		0.0001			
Naproxen	Daphnia magna 48 h	37				Straub and Stewart, 2007
	Lepomis macrochirus 96h	560				
	Pseudokirchneriella subcapitata 96 h growth		0.032			Brun et al., 2006
	Ceriodaphnia dubia 7 d reproduction		0.032	100		
	ERC value developed from chronic studies				4.2	Murray-Smith et al, 2012

	Test	EC50 (mg/l)	NOEC (mg/l)	AF (REACH)	PNEC (µg/l)	Source
Orlistat	Selenastrum capricornutum 10 d		1.92			GSK 2008
	Daphnia magna 21 d NOEC		0.0016	50	0.032	
	Oncorhyncus mykiss 96 h	>18.5				
Simvastatin	Dunaliella tertiolecta 96 h EC50	22.8				Delorenzo and Fleming, 2008
	Fundulus heteroclitus 96 h EC50	2.68				Key et al., 2009
	Palaemonetes pugio	1.18		1000	1.20	·
Trimethoprim	Pseudokirchneriella subcapitata 72 h		16			Yang et al., 2008
•	8 species of cyanobacteria 144 h		3.1-200	50	62	Ando et al., 2007
	Daphnia magna 21 d reproduction		6			Park & Choi, 2008
	Oryzias latipes 96 h	>100	20			Kim et al., 2007

Table 6.

Compound	Removal between point of use and	'Best' and 'Worst' case removal	Inverse modelled removal rate
	point of monitoring using inverse modelling (%)	between point of use and monitoring points based on published metabolism,	corrected for in-stream dissipation (%)
	moderning (70)	treatment and dissipation data (%)	
Atenolol	93.92 (86.94-97.27)	4.0 – 97.9	90.92 (81.87-95.28)
Carbamazepine	90.63 (85.20-96.07)	69.0 – 89.1	90.01 (84.89-93.95)
Cyclophosphamide	95.36 (93.75-96.73)	_	94.74 (93.75-95.38)
Diclofenac	98.24 (97.20-99.42)	74.0 - 95.2	97.64 (96.51-99.00)
Fluoxetine	98.97 (98.63-99.26)	82.6 - 86.6	98.87 (98.66-99.03)
Furosemide	98.18 (96.67-99.10)	10 - 77.5	97.67 (95.81-98.82)
Ibuprofen	99.86 (99.81-99.92)	77.4 – 99.97	99.84 (99.80-99.91)
Ketoprofen	99.31 (99.07-99.50)	_	99.25 (99.11-99.37)
Naproxen	99.18 (98.74-99.48)	97.1 - 99.6	99.01 (98.59-99.31)
Orlistat	98.11 (97.45-98.63)	_	97.94 (97.52-98.30)
Simvastatin	98.42 (97.86-99.02)	_	98.01 (97.52-98.38)
Trimethoprim	97.85 (96.39-99.07)	30-70.2	97.14 (95.54-98.73)

Table 7.

	Median concentration (ng/l)		Lower	_l uartile	Upper quartile concentration (ng/l)	
			concentrat	ion (ng/l)		
	Monitoring Modelled		Monitoring	Monitoring Modelled		Modelled
	data	data	data	data data		data
Atenolol	140	92	46	43	435	172
Carbamazepine	61	193	14.5	89	265	321
Diclofenac	20	24	6.4	11	61	46
Furosemide	10.4	19	3.1	8.9	34	32
Ibuprofen	125	14	25	6.4	475	26
Naproxen	13	24	4.2	11	42	46
Trimethoprim	6.7	11	2.1	4.9	21	21



