

## Effects of silver nanoparticles on freshwater microbial communities



**METHODS** 

SWIFT periphyton assay according to

Porsbring et al., 2007

Colonisation of glass discs by natural

days in a stream.

temperature).

part of the community:

physiological processes.

reference to AgNO<sub>3</sub>

Tedpella 20 nm, citrate coated,

communities of algae and bacteria for 7-10

☐ Exposure to the toxicant in the lab for 96

(light:dark cycle 16:8 hours & ambient

to metabolise different carbon sources.

biomass indicator & changes in pigment

patterns indicating changes in species

hours under controlled semi-static conditions

☐ Structural analysis of the algal & the bacterial

Catabolic profiling & physiological activity of

HPLC pigment profiling: Chl a content as a

composition and interferences with specific

Analytical verification of the exposures with

& NTA (agglomeration, NP behaviour, size).

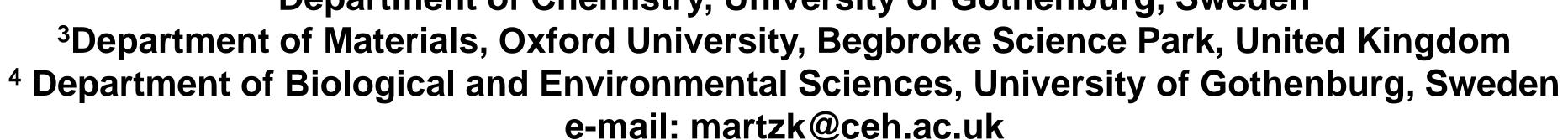
ICP-MS/ultrafiltration (total and dissolved silver)

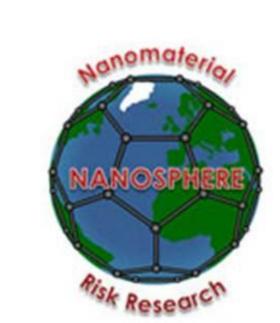
Testing of two different spherical AgNPs in

the bacteria using the Ecolog approach = ability



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### INTRODUCTION

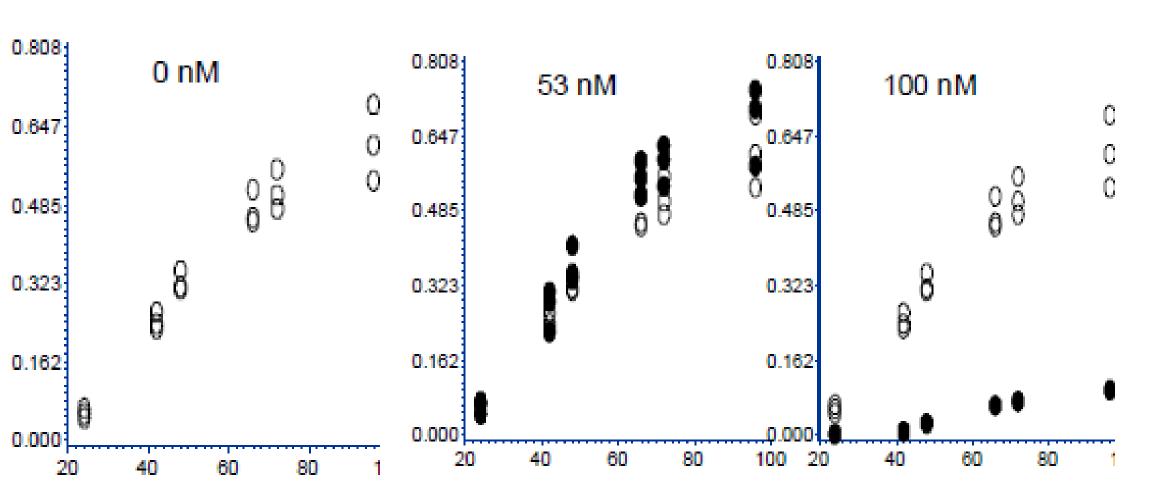
- ☐ Silver nanoparticles (AgNPs) are one of the particles types with the highest production volume.
- ☐ Widespread use of AgNPs will lead to environmental release and their beneficial/wanted microbiocidal effect might become problematic in the natural aquatic environment
- ☐ Especially algae & bacteria might be put at risk.
- ☐ Predicted concentrations of AgNPs in surface water range from 0.1 to 1 nmol/L (Gottschalk et al. 2009, 2010, Mueller & Nowack 2008).

## **AIM** - to answer the following questions...

- > Effects on microbial freshwater communities at environmentally relevant concentrations?
- > Are there any nano-specific effects?
- > Are their differences in the sensitivity of biofilm algae and bacteria?
- > Toxicant induced changes in the community structure?

## RESULTS

- Bacteria reacted more sensitive than algae.
- > Bacterial part (effects based on average well colour development (AWC), Fig. 1): **AgNP NM-300K** (20 nm) > 0.5 nM, **AgNP TP** (20 nm) > 22 nM, **AgNO**<sub>3</sub> between 53 nM - 100 nM.
- > Algal part (effects based on Chl a content, Fig. 2): **AgNO**<sub>3</sub> effects > 150 nM, **AgNP TP** (20 nm) > 300 nM and **AgNP NM-300K**: > 1300 nM.
- $\triangleright$  Comparing AgNO<sub>3</sub> with AgNPs: AgNO<sub>3</sub> more toxic for the algae, AgNPs more toxic for the bacteria.
- > Analytics suggest differences between nominal and real concentrations in terms of total and dissovled silver.

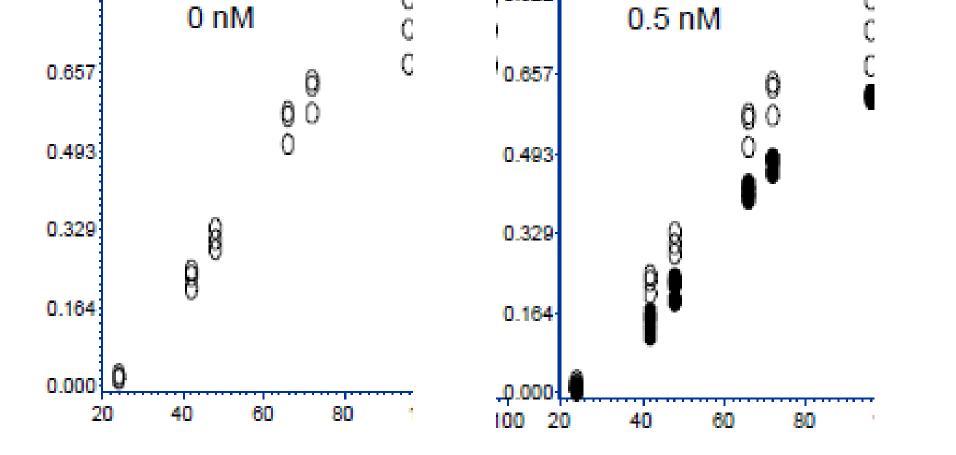


a)  $AgNO_3$ 

22 nM 0 nM0.661 0.331 0.331

# aqueous suspension (AgNP TP)

### (AgNP NM-300K) b) AgNP Tedpella (20 nm, citrate coated) **CONCLUSIONS & OUTLOOK** ✓ Bacteria more sensitive than algae with AgNP NM-300K AgNP Tedpella AgNO<sub>3</sub> effects for the NM-300K in the range of (20 nm)(20 nm)



## c) AgNP NM-300K (20 nm)

Fig 1: Average well colour (AWC) development in the Ecologolates over time for selected concentrations of a) AgNO3 b) AgNPs TP c) AgNPs NM-300K, x-axis gives time in hours, y-axis OD of the colour development at 595 nm

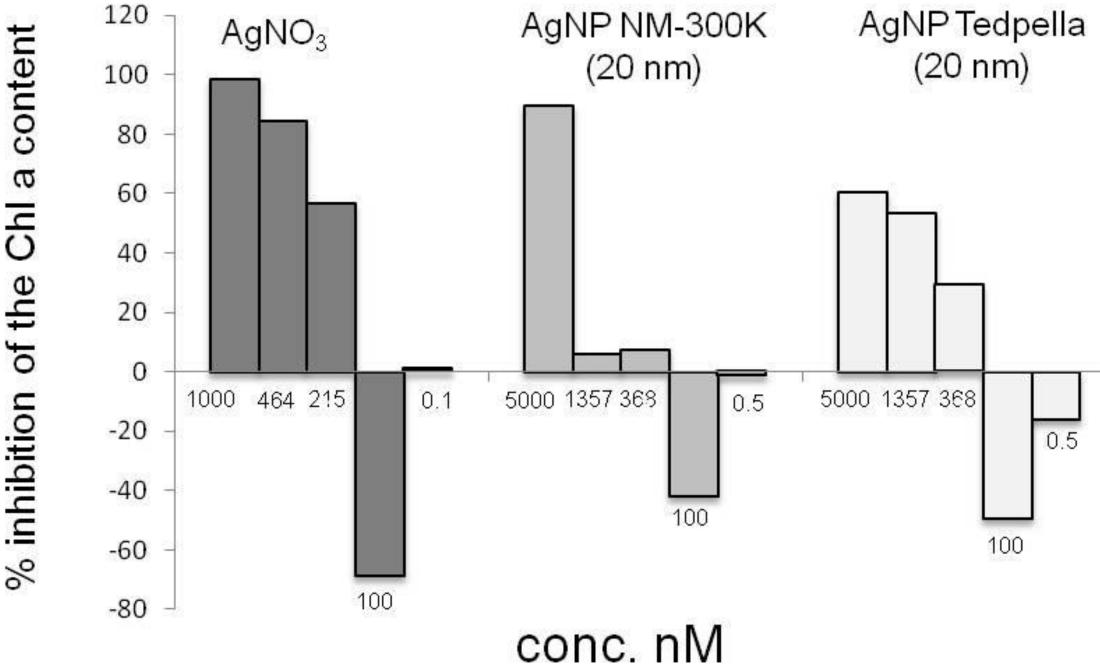
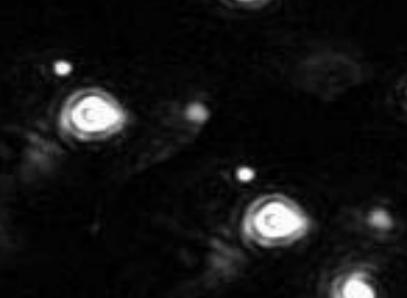
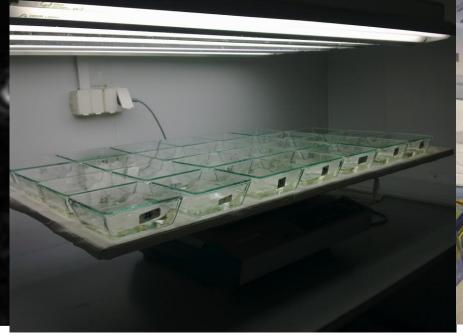


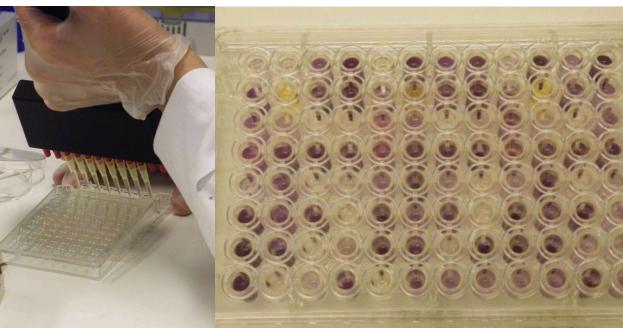
Fig. 2: % Inhibition of the Chlorophyll a (Chl a) content for 5 different concentrations of AgNO<sub>3</sub> and AgNPs

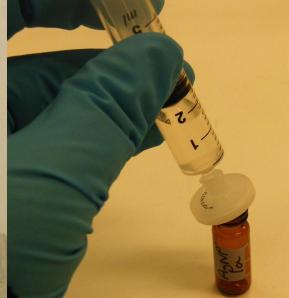
- environmental realistic concentrations.
- ✓ Analytics crucial for a correct interpretation of the ecotoxicological results— otherwise underestimation of theAgNPs effects and hazards.
- ✓ Indications for nanospecific effects: higher toxicity of the AgNPs than the AgNO<sub>3</sub> for the bacteria.
- Link of the ecotoxicological and analytical data (ICP-MS, NTA and MinteQ modelling).

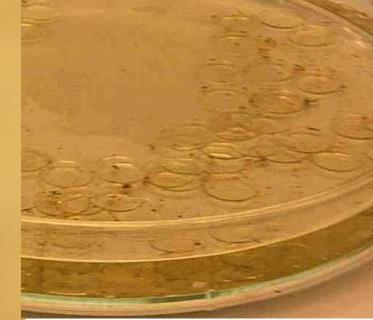












OECD particles NM-300K,

20 nm, aqueous suspension

with stabilizers (Tween 20)

Sampling of natural biofilm

0.822

Field site

NanoSight

Climate room for exposure

Ecolog plates (bacterial part of the community)

Filtering pigment samples for HPLC

Periphyton on glass discs

References

Gottschalk, F. et al., 2009. Modeled environmental concentrations of engineered nanomaterials (TiO(2), ZnO, Ag, CNT, Fullerenes) for different regions. Environmental science & technology, 43(24) Gottschalk, F. et al., 2010. Possibilities and limitations of modeling environmental exposure to engineered nanomaterials by probabilistic material flow analysis. Environmental toxicology and chemistry, 29(5) Mueller, N.C. & Nowack, B., 2008. Exposure modeling of engineered nanoparticles in the environment. Environmental science & technology, 42(12), pp.4447-53.