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# A foodweb model to explore uncertainties in the South Georgia shelf pelagic ecosystem.

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## **ABSTRACT**

Foodweb models provide a useful framework for compiling data on biomass, production,

consumption and feeding relationships. They are particularly useful for identifying gaps and

inconsistencies in the data, and for exploring plausible scenarios of change. We compiled

data on the pelagic foodweb of the South Georgia shelf, which is one of the most intensively

studied areas in the Southern Ocean. The data suggest that current average annual copepod

production is three times that of Antarctic krill and that flying seabirds and fish are

respectively responsible for 25% and 21% of local krill consumption. The most striking

inconsistency was that estimated consumption of fish was 5 times their estimated production. We developed a static mass balance model of the foodweb representing one of many possible solutions to the inconsistencies in the data. The model included sufficient fish biomass to balance the original consumption estimate, and consequently fish became the main krill consumers. Nonetheless, only 74% of local krill production was consumed by predators, suggesting that there are important mortality sources which we did not explicitly model. We developed further models to explore scenarios incorporating plausible climatedriven reductions in krill biomass. In scenarios with unchanged predator diets, an 80% reduction in krill biomass resulted in a 73% reduction in vertebrate biomass. However, when predators with diverse diets were able to switch to feeding on alternative zooplankton prey, total vertebrate biomass was maintained at current levels. Scenarios in which 80% of krill biomass was replaced with copepod biomass required 28% more primary production because the estimated consumption rate of copepods is higher than that of krill. The additional copepod biomass did not alter the consequences for vertebrates. These scenarios illustrate the wide range of potential consequences of a shift from a krill to a copepod dominated system in a warming climate. They suggest that both maintenance and dramatic reduction of vertebrate production are plausible outcomes, although the former requires major changes in predator diets.

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KEY WORDS: Foodweb model, climatic changes, competitors, primary production, ecosystem services.

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## 1. INTRODUCTION

Antarctic krill, *Euphausia superba*, plays a major role in the pelagic marine foodweb on the South Georgia shelf (Atkinson et al., 2001; Murphy et al., 2007a). It is an important prey item for many vertebrate predators including demersal and pelagic fish, mammals, and seabirds (Croxall et al., 1997; Main et al., 2009; Reid & Arnould, 1996; Shreeve et al. 2009). Krill are also one of the main metazoan grazers of phytoplankton and therefore a major regulator of production and nutrient flows (Atkinson and Whitehouse 2001; Schmidt et al., 2011a; Whitehouse et al., 2008, 2011a). In addition to these direct trophic interactions, krill might have indirect competitive interactions with other grazers. Observations at South Georgia suggest that high copepod abundance coincides with relatively low krill abundance (Atkinson et al., 1999). When krill are scarce, some ordinarily krill-feeding predators switch to carnivorous macroplankton which, in turn, feed mainly on copepods (Croxall et al., 1999).

The abundance of krill in the South Georgia shelf system is highly variable. This variability can include years of famine, such as the summer of 2008/09 when krill was virtually absent from the diets of many predators and there were no fishery catches (BAS unpublished data). Such events are almost certainly linked to climate variability. South Georgia is near the northern limit of krill's distribution, and the variability in its local and regional abundance is correlated with climatic indices (Murphy et al. 2007b; Whitehouse et al., 2008). These relationships, combined with decreases in krill recruitment and abundance within the Scotia Sea (Atkinson et al., 2004; Siegel and Loeb, 1995; Trivelpiece et al. 2011) have led to predictions that plausible climate change could remove most of the krill from the South Georgia shelf, causing a prolonged extension of the conditions observed in 2008/09 (Mackey et al., this issue; Murphy et al., 2007b).

Dramatic changes in the abundance of an important organism will inevitably affect the structure of the wider foodweb. Such changes could also affect critical aspects of foodweb operation including its resilience to further change, and therefore its ability to support ecosystem services including production of commercially harvested species, carbon cycling, and the biodiversity which underpins wildlife tourism. It is therefore important to understand the potential consequences of climate induced change for the structure and operation of the ecosystem.

The marine ecosystem around South Georgia is one of the most studied in the Southern Ocean. It was frequently surveyed during the *Discovery Expeditions* between 1928 and 1935 because of its importance to the whaling industry, and it is currently the focal area for many of the British Antarctic Survey's marine ecological studies. It would be valuable to bring the abundant available data together to produce a quantitative description of the ecosystem. A useful first step in this direction is to describe the foodweb. The widely used Ecopath foodweb modelling framework (Christensen and Pauly, 1992) provides a useful template for compiling relevant data on biomass, rate processes, and feeding relationships. This is particularly useful for identifying gaps and inconsistencies in the data. Ecopath can also be used to model the propagation of change (induced, for example, by harvesting and climate) through the foodweb to identify plausible consequences.

This study develops a quantitative description of the South Georgia shelf pelagic foodweb with the particular aim of identifying major inconsistencies in the data and evaluating the trophic roles of krill and copepods. It also uses foodweb models to explore how changes in krill abundance might affect both zooplankton and vertebrate predators, and how these impacts might be modulated by flexibility in predator diets.

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## 2. METHODS

#### 2.1 Database

We developed a foodweb model to investigate how changes to the zooplankton might impact the abundant vertebrate predators concentrated around South Georgia. These predators and their prey interact with other larger ecosystems (including the Scotia Sea to the South and the Antarctic Circumpolar Current system) at a variety of scales (Murphy et al., 2007a). The South Georgia shelf pelagic system is a pragmatic scale to develop a foodweb model to address these questions, partly because many of the available data are more applicable to this system than to the larger scale and partly because this system encloses both the breeding colonies of seabirds and seals on the South Georgia archipelago and the entire habitat of the shelf's demersal fish. The choice of scale and focus is reflected in the model structure including: the choice of functional groups, which are resolved to species level for many vertebrates but are more aggregated for invertebrates and basal groups; the approach to modelling interactions with neighbouring ecosystems, which is explained in detail below; and the inclusion of an aggregated general "benthos" which represents a boundary for the more explicitly modelled pelagic system.

We compiled available information on the pelagic foodweb of the South Georgia shelf to construct an Ecopath dataset. For the purposes of this study, the boundaries of this ecosystem are the shoreline of South Georgia and the 1,000m bathymetric contour, encompassing an area of 45,530 km² (Fig. 1). The dataset nominally represents the average state of the foodweb during the past decade.

Ecopath is a widely used framework for constructing internally consistent marine foodweb models (Christensen and Walters, 2004). It describes foodwebs in terms of the biomass, consumption, production rates and diets of species or life stages aggregated into functional groups on the basis of trophic similarity. These parameters are described in a consistent metric, which was wet mass km<sup>-2</sup> in this case. Ecopath is used to produce mass-balance models which obey the logical constraint that the consumption of any trophic group cannot exceed production by that group over some appropriate time period, which was one year in this case. Production can, however, exceed consumption and this difference is described in the "ecotrophic efficiency" (EE) parameter which we discuss later.

Following a review of the available information and consultation with the experts listed in the Supplementary Information (SI), we structured the model around 30 functional groups (Table 3). The vertebrates were grouped on the basis of taxonomy and similarity of adult diets, and invertebrates were aggregated on the combined basis of data availability and functional similarity. The names of these functional groups are given in italics throughout the text.

Compiling this information was a detailed process drawing on a range of sources (primary and grey literature, unpublished datasets, expert opinion, and proxies from other species and areas) and sometimes requiring subjective interpretation. Our summary of the available data is defensible but there is considerable uncertainty in this (and any pelagic foodweb dataset) which has not been possible to fully characterise. This could mean that there are alternative, equally defensible values for many of our input data. It is good practice to ensure that each value in the input data is traceable to its source and any manipulations are transparent and repeatable. This allows readers to assess the validity of

any value. Due to space constraints, this information is provided in the SI using a tabular format. A summary of this information for the zooplankton functional groups is given in Table 1.

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The detritus (DET) group in many Ecopath models represents all non-living organic material from dissolved organic matter to the carcasses of large animals. Any modelled production that is not assimilated by predators or otherwise explicitly accounted for (e.g. biomass accumulation, fishery removals, other exports) becomes detritus. The consumption or assimilation of non-living organic material by any functional group is modelled by including detritus in that group's diet (but see Pinkerton et al., 2008). This is largely true of the current study with two important caveats. Firstly, we used the model balancing process to estimate the primary production required in each model scenario (see below). Dead phytoplankton (phytodetritus) are important diet components for various organisms including krill (Schmidt et al., 2011a) and benthic deposit feeders. We used the approach of Pinkerton et al. (2008, 2010) and modelled this trophic interaction as direct feeding on primary producers (PHY). We also represented the diet of heterotrophic bacteria as 100% detritus which, while not technically accurate, facilitates the calculation of primary production required without impacting the other results and conclusions of the study. Secondly, when changes were made because the EE of a prey group was >1, the change made was to reduce its EE to approximately 1 (see section 2.2). Thus, almost all production of the prey group was consumed by predators in the resulting balanced model. This could potentially understate the contribution of these prey groups to the detritus pool and therefore to the diets of scavengers, deposit feeders etc.

The South Georgia shelf foodweb is part of an open ecosystem. Larval krill are uncommon, and local reproduction appears insufficient to maintain the krill stock (Tarling et al., 2007). The majority of the available krill are probably imported into the system on ocean currents (Murphy et al., 2004). Many of the air-breathing vertebrates that feed in this system spend much of their time on land, where they produce offspring, defecate and may die. Many of these animals are also highly migratory with ambits that may extend to thousands of kilometres, but which congregate at South Georgia to feed, breed or both. We scaled biomass by the fraction of the year that groups are resident, so that prey consumption within the system was also scaled appropriately. We distinguished between off-shelf krill (EIM1) and other off-shelf prey (EIM2) in predator diets. Off-shelf feeding results in an import of material into the modelled system. However, we defined EIM1 and EIM2 as explicit groups within the model, each of which fed entirely on "Import" (an Ecopath function to represent feeding outside the modelled system). We gave off-shelf krill the production to biomass ratio (P/B) and consumption to biomass ratio (Q/B) of krill, and other off-shelf prey the P/B and Q/B of pelagic fish, and we set their biomasses to satisfy predator demand for off-shelf prey in the base model. This structure allowed us to reduce the availability of off-shelf krill to explore scenarios incorporating a reduction in the krill biomass available to predators both on and off the South Georgia shelf. These import groups were excluded from the calculation of statistics (e.g. total production) for the modelled system. Baleen whales barely feed when they migrate out of the Southern Ocean to breed, but they can migrate extensively while foraging in the Southern Ocean. We represented baleen whales using mean abundances km<sup>-2</sup> and feeding rates for the Scotia

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Sea (Reilly et al. 2004) so there was no need to explicitly represent import consumption, or rescale biomass for this group.

We did not explicitly model advective import of krill into the South Georgia shelf ecosystem but we estimated krill production rates based on growth alone (i.e. without a contribution from recruitment). The assumption of zero net imports is pragmatic in the absence of information about the relative magnitude of imports and exports and is useful for assessing whether krill local production is sufficient to meet local demand (e.g. Gilpin et al. 2002; Trathan et al. 1995) but there are plausible alternative scenarios (see Discussion).

The South Georgia shelf ecosystem is characterised by high inter-annual variability and some particularly extreme events have occurred in the last decade. The mass balance constraint is unlikely to apply over any one year in a highly variable system. The balanced models do not therefore represent any specific year but the average state during a longer period over which the assumption of mass balance is likely to be valid. Ecosystems may also exhibit long term trends in addition to interannual fluctuations. Ecopath models can include a biomass accumulation term to account for such changes over time. However, the available data are not sufficient to describe the dynamics of the whole foodweb so we used the traditional modelling approach of a steady-state approximation for simplicity.

The South Georgia shelf is a fished ecosystem. Average catches in the wider South Georgia area (FAO statistical area 48.3) were 43,565 t.yr<sup>-1</sup> in the period 2001 to 2009 (CCAMLR 2011). These included 37,305 t.yr<sup>-1</sup> of krill and 1,941 t.yr<sup>-1</sup> of mackerel icefish, which were respectively caught mainly and entirely in the modelled area. We did not model these removals which are equivalent to 1% and 7% of the *krill* and *mackerel icefish* 

production in the input (base) data, and 1% of the *mackerel icefish* production in the balanced (base) model.

## 2.2 Balancing

The Ecopath approach (Christensen and Pauly 1992) is based on the following equations for each functional group, *i*:

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$$B_i.P_i/B_i.EE_i - \sum_{j=1}^n B_j.Q_j/B_j.DC_{ji} - Y_i - E_i - BA_i = 0$$

211 and  $Q_i = P_i + R_i + GS_i Q_i$ 

where  $B_i$  is biomass and  $P_i/B_i$  is the production to biomass ratio of group i. The sum term is the total predation on group i, where  $Q_j/B_j$  is the consumption to biomass ratio of predator j and  $DC_{ji}$  is the proportional contribution of prey i to the diet of predator j. Additional terms account for fishery catches  $(Y_i)$ , net import  $(E_i)$  and biomass change  $(BA_i)$ .  $Q_i$ ,  $P_i$  and  $R_i$  are, respectively, the consumption, production and respiration of group i over the model time step.  $GS_i$  is the fraction of consumption that is not assimilated by the predator because it is lost through messy eating, defectaion, urination etc  $(GS_i)$  was set to the Ecopath default value of 0.8 for each group in this study).  $EE_i$ , the ecotrophic efficiency, is the consumption of group i by predators divided by the production by group i  $(B_i,P_i/B_i)$ . Therefore a dataset that satisfies the constraint that consumption of any functional group over some time period cannot exceed production by that group over the same period will allow a solution to the system of linear equations, with  $EE_i$  in [0,1], for each functional group.

Ecopath input datasets rarely satisfy these conditions, so it is necessary to adjust some of the values in the input (base) dataset to produce a balanced (base) model. We used manual balancing, which is the most common approach. We adjusted input values one at a time without the aid of an automated procedure. There have been various attempts to make this an entirely objective process, sometimes with the uncertainty in a particular value governing the magnitude of permitted changes (Kavanagh et al., 2004; Pinkerton et al. 2008, 2010). However, all approaches carry the risk that any revised value will be a less accurate reflection of reality than the original input value. We made changes within a clearly defined set of rules and provide a record of each step in the balancing process to allow readers to assess the validity of these changes and their consequences (see SI). The key rules were:

- (1) The base model (representing the "current" foodweb) was balanced primarily by increasing the production of prey groups (normally by increasing biomass) to meet the estimated consumption requirements of predators.
- (2) Further models (scenarios 1 to 4) were balanced by reducing predator demand (either by reducing biomass or by switching prey types depending on the scenario and scope for switching) to match prey availability.
- (3) All changes were calculated to result in an EE of 1 for the affected prey group. The exceptions were *rays, toothed whales, leopard seals* and *predatory-scavenging birds* which are unlikely to have EE=1 because they are unlikely to be consumed by predators and *juvenile toothfish*, for which the shelf is a nursery area and which therefore export biomass from the shelf. Because we did not explicitly model this export, the models recycle *juvenile toothfish* production thereby overestimating flows to detritus by no more than 0.02% in the

base model. Subsequent alterations to predators of affected prey groups could result in EEs <1.

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We produced an initial balanced (base) model to represent the average state of the South Georgia shelf pelagic foodweb in the last decade. This became the starting point for exploring four scenarios which are summarised in Table 2 and which we modelled by adapting the initial balanced (base) model, rather than the input dataset. The base model and the four scenarios are "snapshots" of possible foodwebs which obey the mass balance constraint. All four scenarios include an 80% reduction in krill biomass, which is the difference between the average and minimum estimates of krill density near South Georgia from annual acoustic surveys conducted during the summer predator feeding season (Hill et al., 2005) and is a plausible long-term consequence of future climate change (Murphy et al., 2007b). As explained above, we modelled a large scale reduction in krill biomass so it affected the availability of krill both on and off-shelf. The scenarios also explore the possibility that copepod biomass could increase as krill biomass is reduced, which might be expected if krill is indeed competitively dominant to copepods (Atkinson et al. 1999). Also they explore the possibility that krill predators could switch to feeding on other zooplankton groups, which were the only alternative prey for which substantial unused production was available (indicated by EE<1) in our balanced (base) model. Together, the four scenarios consider all combinations of copepod expansion versus no expansion and predator switching versus no switching.

Diet switching was implemented using an iterative process to identify the appropriate proportions of the *krill* and *off-shelf krill* components of predator diets to switch to *copepods* or *carnivorous macroplankton*:

- If a predator's diet included *krill* and one of *copepods* or *carnivorous*macroplankton, the *krill* component of the diet was reduced to (1-X) and the *off-shelf krill*component was reduced to (1-X1) of its initial amount where X and X1 are in [0,1].
- If a predator's diet included *krill* and *copepods*, the *copepod* component of the diet was increased by X\*the *krill* component plus X1\*the *off-shelf krill* component of the diet.
  - If a predator's diet included *krill* and *carnivorous macroplankton* but not *copepods*, the *carnivorous macroplankton* component of the diet was increased by X\*the *krill* component plus X1\*the *off-shelf krill* component of the diet.
  - These steps were applied simultaneously across all predators to identify values of X and X1 resulting in EE for *krill* of 1. We used a similar procedure to determine the proportion of the *carnivorous macroplankton* component of predator diets to switch to *copepods*.

The SI gives full details of how we implemented each of these scenarios, including each step in the balancing process.

## 3. RESULTS

## 3.1 Base data and model balancing

Table 1 gives details of the derivation of the base dataset values for the zooplankton and krill groups while the SI provides details for all functional groups (summarised in Table 3). The SI also provides full details of the balancing process for each model, which is summarised below.

The base dataset implied a total biomass consumption of 5,634 t.km<sup>-2</sup>.yr<sup>-1</sup> in the modelled system, but only 3,058 t.km<sup>-2</sup>.yr<sup>-1</sup> of production of which about half was primary production (Table 5). The demersal fish groups in particular were out of balance.

Consumption of these groups was 508% of their estimated production. To balance the base model we increased demersal fish biomass to 697% of the value suggested by the base dataset and we reduced the importance of demersal fish in the diets of other demersal fish (Table 4). We increased the biomass of *pelagic fish* (to 289% of the base value), *heterotrophic bacteria* (428%) and *primary producers* (176%) to match the consumption estimates. We increased the P/B ratio of *cephalopods* (123%) and *heterotrophic bacteria* (333%) to satisfy demand. These were the only alterations we made to P/B or Q/B values. The former was mainly to break a cycle that existed because *cephalopds* and demersal fish (specifically *juvenile toothfish*) prey on each other, meaning that it is impossible to balance this part of the system by changing biomass alone. The latter is a substantial increase to the base value, highlighting uncertainty in bacterial production estimates.

Overall, the balanced (base) model had higher total biomass, and consequently total production and consumption, than the base dataset. To balance models S1 and S3 we reduced the biomass of some predator groups from the levels suggested by the input data (base model with relevant changes to *krill* and *copepods*). This affected predators of *krill*, *cephalopods*, *toothfish* and *diverse flying birds*. To balance models S2 and S4 we altered the diets of predators which consume *krill* and at least one of *copepods* or *carnivorous macroplankton* using the switching rule defined in the Methods. A switch of 81% of the *krill* fraction (and 79% of the *off-shelf krill* fraction) of the diet to alternative prey resulted in full utilisation of krill production (i.e. EE for *krill*=1). However, this increased EE for *carnivorous* 

macroplankton above 1, and a switch of 92% of the *carnivorous macroplankton* fraction of the resulting diets to *copepods* was necessary to restore EE to 1. We also increased the biomass of *heterotrophic microplankton*, *heterotrophic bacteria* and *primary producers* to balance models S3 and S4. An 80% reduction in *krill* biomass (compared to the base model) reduced the primary production required to support the system by only 4% (models S1 and S2) whereas a corresponding increase in *copepod* biomass resulted in a 23% increase in the primary production required (models S3 and S4), alongside increased requirements for microplankton and bacteria, which are also components of *copepod* diets. This increased demand resulted from the higher consumption rate of *copepods* compared to *krill*.

Models S1, S2 and S3 had slightly lower overall (biomass-weighted average) EE than the base model, whereas S4 had slightly higher EE (Table 5). The switching models had higher overall EEs than their non-switching counterparts (S2>S1 and S4>S3) and the increased *copepod* models (S3 and S4) had higher overall EEs than their no-increase counterparts (S3>S1 and S4>S2). These differences reflect the balancing process: We calculated the biomass reduction in the switching scenarios to result in full utilisation of *krill* production, while increased consumption by *copepods* increased the consumption of the substantial *heterotrophic microplankton* production.

# 3.2 The trophic role of krill

*Krill* consumed 9% of the primary production in the base dataset and 5% in the base model. *Krill* were also major consumers of *heterotrophic microplankton* (15% of ZHT production in the base dataset and base model). The 314 t.km<sup>-2</sup>.yr<sup>-1</sup> estimated consumption by krill translated into 79 t.km<sup>-2</sup>.yr<sup>-1</sup> of krill production. According to the base dataset the EE

of krill is 0.34. Increased *krill* predation in the base model, due mainly to higher fish biomass, increased EE for krill to 0.74 (Table 6). This value was maintained in the non-switching scenarios (S1 and S3) whereas in switching scenarios (S2 and S4) the balancing process resulted in an EE of 1 for *krill*.

According to the base dataset, the *diverse flying birds* were the most important krill consumers, accounting for 25% of total *krill* consumption (Fig 2). As a result of biomass increases in the balancing process, demersal fish became the main krill consumers in the base model. *Mackerel icefish* and *minor shelf fish* together accounted for 47% of krill consumption, the remaining demersal fish groups accounted for 6% and pelagic fish a further 10%, making fish more important krill consumers than air breathing vertebrates.

Baleen whales included both krill and copepods in their diets; gentoo penguins, macaroni penguins, mackerel icefish and other icefish included both krill and carnivorous macroplankton, and diverse flying birds, small rock cod, other shelf fish, pelagic fish and cephalopods included all three. These predators therefore switched diets in scenarios S2 and S4.

Demersal fish were the main *krill* consumers in the non-switching scenarios (S1 and S3) and the second most important in switching scenarios (S2 and S4), accounting for 68% and 36% of krill consumption respectively. This reduction in switching scenarios was because many fish switched diets whereas *fur seals* and most birds did not. *Pelagic fish* were also important in non-switching scenarios (13%) whereas *Antarctic fur seals* became important (37%) in switching scenarios.

## 3.3 Trophic role of copepods

Copepods were about three times more important consumers of primary production than krill, removing 25% in the base dataset and 14% in the base model. They also removed 50% of *heterotrophic microplankton* production in both the base data and base model. The 950 t.km<sup>-2</sup>.yr<sup>-1</sup> estimated consumption by *copepods* translated into 237 t.km<sup>-2</sup>.yr<sup>-1</sup> of *copepod* production. According to the base dataset the EE of *copepods* is 42%. This rose slightly to 50% in the base model and was in the range 23% to 77% in the scenario models.

Carnivorous macroplankton were the most important copepod consumers in the base dataset (89%: Fig 3) and all balanced models (87% in the base model, 57% in switching scenarios and 97% in no-switching scenarios). Biomass increases during the balancing process increased the importance of pelagic fish as copepod consumers in the base model (4%) compared to the base data (2%). In the switching scenarios, vertebrates which normally feed mainly on krill became important copepod consumers, with flying birds accounting for 19% of copepod consumption, and pelagic (19%) and demersal (2%) fish increasing consumption compared to the base model

## 3.4 Comparing scenarios

A reduction in *krill* biomass without predator switching reduced the biomass and production of the many krill-eating groups. Compared to the base model, S1 and S3 had 82% less mammal biomass, 50% less penguin biomass, 94% less flying bird biomass, a 73% reduction in both pelagic and demersal fish and an 83% less *cephalopods*. The high *copepod* and *carnivorous macroplankton* production suggested by the base dataset was sufficient to make up the shortfall of krill in predator diets in each of the switching scenarios.

Consequently, introducing more copepods into the system (S3 and S4) had no effect on higher trophic levels (compared to the base model) due to the balancing approach (i.e. higher trophic level biomass was not expanded to take advantage of increased prey availability).

Production by all zooplankton groups including *krill* (but excluding *heterotrophic microplankton*) was reduced by 15% as a result of an 80% reduction in *krill* biomass, but the net increase was 38% when the missing *krill* biomass was replaced with *copepods* (scenarios S3 and S4), because the P/B for *copepods* was 358% of that for *krill*.

In the base data, production by *pelagic fish* was slightly higher than that by demersal fish but the adjustments made in balancing increased demersal fish production above that of *pelagic fish* (Fig 4). This ranking was preserved in all scenarios. In the switching scenarios (S2 & S4), the production of all vertebrates was maintained at the levels of the base model while in the non-switching scenarios (S1 and S3), production by flying birds, mammals, cephalopods and demersal and pelagic fish was reduced by 73 to 95% compared to the base model. Penguin production was reduced by 42%.

Table 6 gives the trophic levels of each functional group in the various models. The initial balancing had affected the trophic levels of several demersal fish groups, particularly that of *other icefish*, which was reduced by 0.17 compared to the base data. The switching scenarios resulted in increased trophic levels for most krill-eating groups whereas the trophic level of some groups, notably *pelagic fish* and *king penguins* fell, due to the switch from *carnivorous macroplankton* to *copepods*.

3.5 The role of imports

The air breathing predators represented in the models obtain a considerable part of their diet from outside the modelled system (Table 4). This accounted for 1.2% of the total consumption in the base dataset (83 t.km<sup>-2</sup>.yr<sup>-1</sup>; 70% of all consumption by birds and mammals). The relative importance of import consumption was reduced to 1.1% of total consumption in the base model. Import consumption fell to 6 t.km<sup>-2</sup>.yr<sup>-1</sup> in models S1 and S3 but accounted for only slightly less (69%) of the total consumption by the reduced populations of birds and mammals in these models. In the switching scenarios (S2 and S4), predators could replace *off-shelf krill* in their diets with on-shelf *carnivorous macroplankton* and *copepods*. This reduced import consumption to 55 t.km<sup>-2</sup>.yr<sup>-1</sup> (47% of consumption by birds and mammals). Nonetheless, the unused copepod production, (1-EE)\*P, exceeded import consumption in all models.

# 3.6 Summary

These results provide a quantitative description of trophic relationships on the South Georgia shelf and highlight the uncertainties associated with the current dataset, which required ten-fold increases in the biomass estimates for some groups to balance the model. The base model is one of many possible models of the state of the foodweb over the past decade. The scenario models suggest that unchanged copepod production could support the currently observed levels of upper trophic level biomass if predators can switch most of their krill consumption to copepods and carnivorous macroplankton.

## 4. DISCUSSION

We have compiled a detailed dataset characterising the foodweb of one of the most important and intensively studied pelagic ecosystems in the Southern Ocean. This data compilation and the balanced foodweb model quantify the relative importance of a range of feeding relationships and highlight some of the key uncertainties. The whole foodweb context provides a broader perspective than many previous studies (e.g. Croxall et al., 1984; Hill et al., 2007a). We begin the Discussion by exploring the caveats and uncertainties of the approach before considering what it reveals about the structure and operation of the foodweb. We conclude by discussing what the scenario models suggest about responses to change.

#### 4.1 Caveats and uncertainties

The South Georgia pelagic ecosystem is often described as a krill-based system, characterised by iconic krill predators such as fur seals and penguins (e.g. Croxall et al., 1985). The structure of our models and the precision in our estimates reflects the way this view has focused studies of the system. There is a mismatch between the distribution of research effort amongst trophic groups and their importance in total energy flow. Information is particularly scarce for basal groups including *primary producers* and *heterotrophic bacteria*. In our models, these uncertainties have minimal effect on the upward propagation of changes to krill and zooplankton. Nonetheless it is appropriate to acknowledge that their potential magnitude might outweigh many of the other uncertainties that we discuss.

Variance does not fully describe the uncertainty in a dataset because variance does not indicate bias. Bias can arise from methods that produce over- or underestimates, do not

encompass the full range of spatial and temporal variability, or rely on assumed parameter values or conversion factors. Other issues arise from model structures. Particular problems with this type of foodweb model are due to simplification. Dynamic processes operate on many different time scales (Murphy et al., 1988) and converting the data to a common time scale (in this case one year) often requires some extrapolation. This is an important issue for the South Georgia ecosystem, which is highly variable both within and between seasons, whereas most of the available data were collected over relatively short periods, usually during the summer when measured biomass, production and consumption might not be representative of annual averages. Furthermore, aggregation into functional groups causes problems with averaging, especially when data availability varies between the members of a group. It can also lead to a high degree of apparent cannibalism which can make it difficult to establish mass balance. These are just some of the potential sources of uncertainty in this dataset and the thousands of foodweb models for other regions that have already been published.

For the reasons above it is not possible to fully quantify the uncertainty in the base dataset, but some of the major uncertainties became apparent because of the differences between the base dataset and the balanced (base) model. Furthermore, the scenario exploration, which considers a range of responses, is a form of uncertainty analysis (Hill et al., 2007b), which suggests the range of the potential consequences of krill reduction. A systematic exploration of additional scenarios would be necessary to establish bounds on these potential consequences with confidence.

Even after increasing fish biomass fivefold to balance the model and assuming conservatively that there is no net krill input onto the shelf, 26% of krill production was still

unaccounted for. This result merits some discussion of the high P/B value of 2.4 that we used for krill. Krill growth rates are sensitive to temperature, food availability and krill size (Atkinson et al., 2006; 2009) and there is substantial variability in observed krill production rates (Siegel and Nicol, 2000) and considerable uncertainty in annualised extrapolations of these. Our value is based on measurements of daily growth rate at South Georgia (Atkinson et al. 2006) extrapolated over a 4 month growth period. For comparison, a recent circumpolar-scale study derived conservative krill production estimates of 342-536 million tonnes per year, based on a biomass of 379 million tonnes (Atkinson et al., 2009). Previous P/B estimates for krill (Ross and Quetin, 1986; Siegel, 2000) are also lower than our estimate. We consider that an appropriate value for South Georgia is likely to be higher than these wider scale estimates firstly because the South Georgia bloom is the largest in the Antarctic Circumpolar Current and one of the longest lasting, extending for 4-5 months of the year (Atkinson et al., 2001; Korb et al. this issue; Murphy et al., 2007a). Secondly Krill continue to feed throughout the year at South Georgia (Schmidt et al., this issue) with a growth season starting as early as October and probably lasting 6 months (Reid, 2000).

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The role of advective krill imports is a related issue. The South Georgia shelf is widely considered to be a "semi-open system" where the krill population is not self-replacing but is maintained by a net influx (Atkinson et al. 2001; Gilpin et al. 2002; Shreeve et al. 2005; Tarling et al., 2007; Trathan et al., 1995). Previous studies have compared estimates of krill consumption at South Georgia with estimates of local krill production and either reached the conclusion that consumption is greater than production (Shreeve et al., 2002; Trathan et al., 1995) or standing stock (Boyd and Croxall, 1996; Boyd, 2002; Croxall et al 1984; Croxall and Prince 1987) or conversely that local production is sufficient to support consumption

(Atkinson et al 2001; Gilpin et al 2002). These studies differed both in the methods they used to extrapolate estimates of production and consumption to greater temporal and spatial scales and the organisms that were included in the suite of consumers. Our study suggests that local krill production is more than adequate to account for known krill consumption. Indeed a much lower P/B value (0.82) would have been sufficient to support the consumption estimate from the base dataset. However, it is possible that there are krill consumers that we have not accounted for (see section 4.2). The relative importance of import versus local production merits further investigation with specific exploration of the uncertainties affecting production, consumption and import estimates. Scenario analysis using a foodweb models, as in this study, is a valuable method for this type of investigation.

There are assumptions associated with our scenario exploration. Our switching scenarios imply a competitive hierarchy. Krill predators which are known to include copepods or *carnivorous macroplankton* in their diets (e.g. *diverse flying seabirds*) were forced to switch while those which are not (e.g. *Antarctic fur seals*) were able to consume the krill that switching released. Also, those that fed on both copepods and *carnivorous macroplankton* (*diverse flying seabirds, rock cods, pelagic fish, cephalopods* and *benthos*) took *copepods* in preference to *carnivorous macroplankton*. In this case, predators which fed on *carnivorous macroplankton* but not *copepods* were able to consume the *carnivorous macroplankton* that switching released. This was a parsimonious way to implement switching scenarios but there is little information on real competitive hierarchies amongst these organisms. Consequently, the models should not be regarded as reliable predictions of the relative abundance of the different predator groups under change scenarios even

though the general conclusion that zooplankton production exceeds predator demand in all scenarios remains valid.

The top-down model structure (Steele, 2009) means that it was not necessary to account for the fate of all biomass production to balance the model (Pinkerton et al., 2008). Any production that is not assimilated by predators or explicitly removed as fishery catch or exports is recycled via detritus. Consequently the model does not require expansion of predator biomass in response to the increased zooplankton production in S3 and S4, although this outcome is plausible.

The food value of a biomass unit varies between prey types. Some modellers attempt to overcome this issue by formulating models in terms of a more consistently conserved currency such as organic carbon or energy rather than biomass *per se* (e.g. Pinkerton et al., 2010). This introduces additional uncertainties when carbon is estimated from biomass, and it does not resolve differences due to the costs of acquiring or digesting prey. Macaroni penguins are able to switch to amphipods when krill are scarce but they ingest less prey wet mass per unit foraging time, and raise smaller offspring (Croxall et al., 1999). This is evidence of a greater cost in foraging time per unit biomass acquired, and might also indicate a lower food value per unit biomass. Our modelling does not distinguish between functional groups on the basis of food value and acquisition costs.

# 4.2 Structure and operation of the South Georgia pelagic foodweb

Notwithstanding the above caveats, Ecopath provides a rational quantitative framework for synthesising voluminous, disparate foodweb data. This provides a powerful tool for assessing and exploring uncertainty. When we assembled the data, it became clear

that estimates of production by fish are much lower than estimates of consumption of fish. This corresponds with a previously observed discrepancy between mackerel icefish biomass estimates and potential consumption by fur seals (Reid et al., 2005). The inconsistency could arise because of inaccuracies in any (probably all) of the relevant parameter estimates. Hill et al. (2005) noted that the bottom trawl surveys used in stock assessment are likely to underestimate the abundance of these benthopelagic organisms, which provides support for our approach of adjusting fish biomass to balance the model. Nonetheless this is compelling evidence that the role of fish in the foodweb is poorly understood and merits further investigation.

The EE for *primary producers* in the base dataset was 1.76, indicating that our estimate of production was only sufficient to meet 57% of the estimated demand. The shortfall was even more severe (meeting only 46% of demand) in our increased *copepod* scenarios. These differences might be greater still if assimilation rates of consumed phytoplankton are less than the assumed 80%. Feeding on primary production accounts for a substantial fraction of the energy transfer in foodwebs (40% of all consumption in our balanced base model). Uncertainty in primary production estimates has important consequences for understanding foodweb properties such as their ability to resist and recover from perturbation and there is a clear need to investigate and reduce this uncertainty.

The relatively low EE values for many organisms indicate uncertainty about the fate of production. In the base dataset, 60% of meso and macroplankton production was not explicitly accounted for in the modelled feeding relationships. This fell to 40% in the base model. In our models this "missing production" enters a general detritus pool and the model

does not differentiate between the types of non-living organic material consumed by organisms as diverse as bacteria and vertebrate scavengers. Pinkerton et al. (2008) suggest an alternative approach in which it is possible to constrain EE to 1 by explicitly modelling feeding on non-living organic material originating from each group of larger organisms:

Scavengers are modelled as predators of the groups which are the source of this material.

At present we do not have sufficient information to apply this approach and the low ecotrophic efficiencies highlight the underlying uncertainties and the need for further studies to understand the fate of this "missing production".

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The fate of the "missing krill production" is the opposite of the problem suggested by Boyd and Croxall (1996) who calculated that complete replacement of the resident krill stock each month is necessary to satisfy the great demands of the penguins and fur seals. In contrast, our estimate of krill production exceeds our estimate of consumption of krill, suggesting that there may be other sources of mortality in addition to those that we have explicitly modelled. One candidate is high consumption by benthic organisms over the shelf. These organisms might consume either dead and dying krill, injured by surface-layer predators, or krill actively migrating to the seabed to feed. Main et al. (2009) found that krill comprised a large fraction of skate diets, and Schmidt et al. (2011a) found that krill visits to the seafloor were frequent, widespread and an integral part of their biology. There is uncertainty about the extent and importance of benthic predation on krill. The assumption of negligible predation in this study is one of the bounds on this uncertainty whereas a fuller exploration will also need to consider models with high predation. Given the potential importance of bentho-pelagic coupling in ecosystem resilience, this issue warrants further study.

Many existing studies of the predation pressure on krill concentrate on a limited range of predators. Croxall et al. (1984) considered birds that nest at South Georgia, and krill consumption across their entire foraging range during chick rearing. They identified macaroni penguins as the main krill consumers nesting at South Georgia with the members of our *diverse flying seabirds* second and *gentoo penguins* third. Hill et al. (2007a) included fish in their data compilation for a selection of important krill consumers at the Scotia Sea and Antarctic Peninsula scale. This compilation excluded flying seabirds, which were the main krill consumers on the South Georgia shelf in the base dataset. Hill et al. (2007a) suggested that, at the regional scale, fish consume more krill than baleen whales, penguins, and fur seals combined but that around South Georgia (within 126km of the coast) penguins were the main krill consumers, followed, in descending order of importance, by fur seals, demersal fish, pelagic fish, and whales.

The discrepancies between these studies and our own arise largely from the different scales considered, and the omission of significant consumers in previous studies. This highlights the current lack of comprehensive assessments of foodweb structure at the various scales that ecosystem models need to consider (Hill et al., 2006, 2007b). Although diverse flying birds apparently obtain 81% of their diet from off-shelf areas compared to 25% for macaroni penguins, the greater biomass of the former means that they are more important on-shelf krill consumers. Antarctic fur seals were also more important than macaroni penguins due to both a higher overall consumption and more on-shelf feeding. The balanced model presents a very different view of krill consumption due to the revised fish biomass needed to match estimates of fish consumption. This raises the possibility that

fish might, in fact, be more important krill consumers than the air breathing vertebrates that have dominated previous studies.

The data compilation suggests that copepods are potentially more significant consumers of primary production and producers of biomass than krill. This was also the conclusion of Shreeve et al. (2005), whose estimates of copepod production we included in the base data. Many of South Georgia's vertebrates feed mainly on krill and are affected by fluctuations in its availability (Croxall et al., 1997; Everson et al., 1997; Main et al., 2009; Reid & Arnould, 1996; Reid & Forcada, 2005; Shreeve et al., 2009). Production estimates suggest that there is abundant alternative biomass in copepods and their consumers. However, observations suggest that, in the short term at least, predators are not able to efficiently exploit this alternative biomass. It is currently unknown whether predator behaviour could adapt over the longer term to exploit alternative prey types.

The scenario exploration suggests that copepod biomass production exceeds the feeding requirements of upper trophic level predators in this ecosystem. Our switching scenarios required a substantial replacement of *krill* in predator diets with much smaller *copepods*. It was not possible to maintain predator biomass using larger alternative prey (fish and macroplankton) alone, as these food sources were fully utilised (*EE*=1). Predators could therefore be severely impacted by a plausible decline in krill biomass if they are not able to efficiently exploit copepods.

The scenario exploration suggests that complete replacement of krill biomass by copepod biomass is only possible if primary production is not limiting. Nonetheless this replacement is not necessary to maintain zooplankton production equivalent to

consumption by predators. Indeed this production, even with an 80% reduction in krill biomass, exceeds all modelled secondary consumption including off-shelf feeding.

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The integration of many previous studies gives an indication of the role of each predator group in the foodweb as a whole, which is summarised in the group's trophic level. Stowasser et al. (this issue) used stable isotope analysis to estimate the trophic levels of a range of organisms in the wider Scotia Sea ecosystem and comparison of the two studies provides a level of cross-validation. Trophic levels for comparable taxa were (simple average of Stowasser et al. this issue estimates, followed by our base data estimate): fur seals 3.9, 3.9; gentoo penguins 3.9, 4.2; macaroni penguins 4.0, 4.3; chinstrap penguins 3.5, 4.0; blackbrowed albatross 4.8, 4.2; grey-headed albatross 4.8, 3.9; predatory seabirds 5.2, 3.8; diverse flying birds 3.8, 3.5; pelagic fish 4.1, 4.2; cephalopods 3.7, 4.1; krill 2.5, 2.7; carnivorous macroplankton 3.2, 3.4; copepods 2.7, 2.8; salps 2.0, 2.8. There was reasonable agreement between the two studies, although there were significant discrepancies for most bird groups, especially predatory seabirds. The Stowasser et al. (this issue) estimate for predatory seabirds is consistent with the other apex predators in our study, suggesting that our estimate is not representative of this taxon. 80% of the diet of predatory seabirds was imports. This highlights a weakness in Ecopath based estimates of trophic levels for groups, including most of our bird groups, foraging outside the model arena. The stable isotope study also provides some guidance for revision of our functional groups. For example, we included Themisto gaudichaudii in carnivorous macroplankton whereas the trophic level estimated by Stowasser et al. (this issue) suggests that this species is functionally more similar to *herbivorous zooplankton*.

The scenarios and their associated caveats are an expression of uncertainties in the current knowledge that warrant further exploration. There is a need to better understand zooplankton dynamics at interannual timescales, as the response of the zooplankton foodweb to change will determine effects on the higher trophic levels. As discussed above, the plausibility of the scenarios depends to some extent on the ability of predators to switch between zooplankton prey and maintain previous levels of production per unit consumption. It also depends on whether an increase in copepod production would be limited by available primary production. More information on diet flexibility and its consequences would be useful for predicting future dynamics. Finally, the prevalence of offshelf feeding highlights the strong levels of ecological connectivity between areas and the somewhat arbitrary nature of a geographical definition of the foodweb. Future model development should consider the consequences of this connectivity.

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TABLE 1: Base data values for krill and zooplankton groups for the South Georgia shelf system. Abbreviations introduced in this table: DM = dry mass, WM = wet mass, C = carbon, SG = South Georgia, GGE = gross growth efficiency, defined as growth divided by ingestion, chl = chlorophyll. We consistently converted dry mass to wet mass using a conversion factor of 4. We also consistently used a GGE estimate of 25% (Straile 1997). The exact meaning of a measured growth rate can vary depending on the methodology used, which is why we used either simple or compound scaling depending on the details. In either case the resulting production value was compared with a value derived from consumption and GGE to ensure consistency. Scaling of daily values by growing season was achieved by multiplying the daily value by 365 \* (growing season in months)/12.

| Group                      | Biomass (B: g wet mass m <sup>-2</sup> )   | Annual production/mean biomass (P/B)   | Annual consumption/ mean biomass (Q/B)  | Diet % composition  |
|----------------------------|--|--|---|---|
| ZKR (Euphausia<br>superba) | Based on a synoptic krill survey in 2000 (Hewitt et al. 2004) which produced coastal and oceanic estimates of krill biomass density at South Georgia using the Stochastic distorted wave Born approximation methodology (BAS unpublished data). This is the average of those two values weighted by the proportion of the modelled area that fell into the two survey strata | 2.4 Compound mean daily mass growth rate(1.024% per day) at low and high summer chl a sites at South Georgia (Table 7 of Atkinson et al. 2006), over a 4 month growing The assumption of a 4 month growing season is conservative, based on unpublished seasonal fisheries-derived data by Atkinson (growth), and published data in Reid (2000) (growth) and Schmidt et al. (this issue) (feeding at South Georgia occurs year-round). | 9.6 P/B / GGE of 25% (Straile 1997). This value fits well with a daily C ration of 5% measured by Pakhomov et al. (1997a) at South Georgia in summer, if this rate is sustained for half of the year. | ZCA 0.5, ZHE 0.5 ZCO 2.0, ZHT 45, PHY 42, DET 10  Based on volumetric gut content analysis from 16 krill on SG shelf in summer bloom (Schmidt et al. 2006), and finding of important role of benthic feeding on detritus (Schmidt et al. 2011a) Note that seabedderived phytodetritus component is included in PHY group here.  Metazoan contribution is based mainly on volumetric analysis of krill stomachs from Scotia Sea in spring (Schmidt et al. this issue).  Importance of non-phytoplankton food at South Georgia supported by Atkinson and Snyder (1997) and Pakhomov et al. (1997a). |
| ZCA (Carnivorous           | 8.4  | 4.87   | 19.5  | ZCO 54, ZHE 15, ZSA 1, ZHT 10, PHY  |
| macroplankton:             | Mean DM of 2.1 g m <sup>-2</sup> based   | Based on growth rates measured on  | Based on the production values  | 10, DET 10  |
| amphipods,                 | on 3 mainly summer studies   | dominant amphipod <i>Themisto</i>  | and a GGE of 25% (Straile 1997).  | Based on SG diet data for <i>Themisto</i>   |
| chaetogantahs,             | (Ward 1990, Atkinson et al.  | gaudichaudii on inner SG shelf by  | Alternatively based on a daily  | gaudichaudii and carnivorous  |

| small euphausiids,<br>fish larvae, mysids,<br>large predatory<br>copepods such as<br>Euchaeta spp.)   | 1996, Pakhomov et al.<br>1997b). Biomass assumed<br>essentially constant<br>throughout season based on<br>winter survey data (Atkinson<br>et al. 1988, Atkinson and<br>Peck 1990)  | Watts and Tarling (in press, this volume) which gives daily mean P/B of 0.0133 multiplied by 365 to represent a 12 month growing sea. The P/B for mysids also exceeds the value for krill at South Georgia. (Ward 1985).              | carbon ration of 7.1% (for Themisto gaudichaudii in summer at SG; Pakhomov and Perssinotto, 1996) the value would be 25.9, assuming a year-round growing season. This broadly supports the estimate given.          | copepods and fish larvae (Pakhomov and Perissinotto 1996, Øresland and Ward 1993, North and Ward 1989, 1990), plus diet and trophic level data for Antarctic chaetognaths, small euphausiids in Scotia sector (Øresland 1990, Hopkins and Torres 1988, Hopkins et al. 1993a,b, Stowasser et al. in review, this volume) |
|---|--|---|---|---|
| ZHE (Mainly herbivorous non-copepod mesozooplankton: pteropods, ostracods, appendicularians, and meroplankton of benthic organisms)           | Mean total mesozooplankton biomass is 8.63 g DM m <sup>-2</sup> , based on 7 studies (Ward et al. , 1995, 2006, Ward and Shreeve, 1999, Atkinson et al. 1996, Atkinson & Whitehouse 2001, Pakhomov et al. 1997b). ZHE comprises 20% of this, based on Atkinson et al. (1996), Ward & Shreeve (1999), remainder being copepods. Winter biomass is similar to that in summer (Atkinson and Peck 1990). | 8.6 In absence of other data we used the ZCO P/B as below.  | In the absence of SG grazing data across the non-copepod herbivores their Q/B is derived from the P/B and a GGE of 0.25 (Straile 1997)  | PHY 40, ZHT 50, BACT 8, DET 2 In the absence of SG grazing data across the non-copepod herbivores their diets are assumed the same as that of the ZCO (see below)   |
| ZCO (Copepods that<br>are mainly small<br>particle feeders) for<br>example<br>Rhincalanus gigas,<br>Calanoides acutus,<br>Calanus propinquus, | 27.6<br>See row above for source of<br>this value  | 8.6 This is based on a P/B for stages CIV and CV of <i>Calanoides acutus</i> , a biomass-dominant copepod measured during 2001/2002 summer by Shreeve et al. (2005). This daily P/B estimate (in C) of 0.0566 was scaled to an annual | Q/B is derived from the P/B and a GGE of 0.25 (Straile 1997). This corresponds to a daily C ration of 22% sustained over a period of 5 months where chl a levels at SG are > 1 mg m <sup>-3</sup> (see P/B column). | PHY 40, ZHT 50, BACT 8, DET 2 Based on study at SG and Bellingshausen on major species of both large and small copepods (Atkinson 1994, 1995, Atkinson and Shreeve 1995). Diatoms comprised 39 and 49% of identified food C,  |

| C. simillimus,     |  | value assuming a growing season of 5             | This ration is within the range    | remainder being motile, mainly           |
|--------------------|--|--|------------------------------------|--|
| Metridia spp.      |  | months, based on the period when                 | measured for a range of copepod    | members of ZHT group. Importance         |
| Ctenocalnus spp.   |  | mean chl a concentration is at bloom             | sizes at SG (Atkinson et al. 1992, | of motile taxa to diets of many          |
| Microcalanus spp., |  | levels of >1 mg m <sup>-3</sup> at South Georgia | Atkinson 1994)                     | species supported by SG study of         |
| Oithona spp.,      |  | (Whitehouse et al. in review, this               |                                    | Atkinson et al. (1996). We ascribed a    |
| Oncaea spp.        |  | volume). This is also the period in              |                                    | nominal 10% to detritus colonised by     |
|                    |  | which the biomass-dominants are in               |                                    | bacteria, a known food for <i>Oncaea</i> |
|                    |  | the upper water layers (Atkinson 1998)           |                                    | sppand <i>Oithona</i> spp                |
| ZSA (salps)        | 1                                      | 7.6  | 30.4                               | PHY 45, ZHT45, BAC 6, ZHE 1, ZCO 2       |
|                    | A nominal value that reflects          | In the absence of direct data from               | Based on a daily C ration of 25%   | Based on a roughly 50:50 ratio of        |
|                    | the low abundance of this              | South Georgia we used the Q/B value              | (Pakhomov et al. 2006) and a 4     | diatoms versus protozoans plus           |
|                    | taxon in the SG area (Ward             | multiplied by a GGE of 25% (Straile              | month feeding season (von          | small metazoans (Hopkins and             |
|                    | et al. 1995, Atkinson et al.           | 1997)  | Harbou et al. in press)            | Torres 1988). A nominal remaining        |
|                    | 1996, Pakhomov et al.                  |  |                                    | 10% is divided into bacteria             |
|                    | 1997b) particularly over its           |  |                                    | (associated with detrital aggregates     |
|                    | shelf                                  |  |                                    | and incidental capture of larger         |
|                    |  |  |                                    | metazoans                                |
| ZHT (Heterotrophic | 20.3                                   | 47.2   | 163                                | PHY 60, PBA 40                           |
| microplankton <    | Biomass of protists is 1.138           | This is derived from a daily P/B ratio           | Based on the P/B ratio divided by  | This is simply a nominal value. Like     |
| 200 microns:       | gC m <sup>-2</sup> based on integrated | for SG system from Spring, summer                | a GGE of 0.29, a value             | the other values for heterotrophic       |
| crustacean larvae, | water column value from 2              | and autumn cruises (Korb et al. this             | representative of protozoans       | microplankton it is very poorly          |
| ciliates,          | stations at NW shelf and               | volume). Chl a was thus converted to             | (Straile 1997). This equates to a  | constrained). These values are           |
| dinoflagellates,   | shelfbreak at SG (Priddle et           | carbon using SG-derived C:chl ratio of           | daily C ration of 67% per day,     | poorly constrained because this ZHT      |
| microflagellates)  | al. 1995).                             | 75 (Priddle et al. 1995). This mean              | sustained over a 8 month season    | box contains up to 3 trophic levels      |
|                    | The assumed C to WM                    | daily P/B ratio for autotrophs of 0.194          | of elevated chl a (see P/B column) | (including, heterotrophic                |
|                    | conversion factor was 10.              | was assumed as similar to that of                |                                    | nanoflagellates, large dinoflagellates,  |
|                    | The above value for protists           | autotrophs which are of similar size in          |                                    | small crustaceans)                       |
|                    | is added to a micrometazoan            | SG system and unlike atotrophs can               |                                    |  |
|                    | value of 2.23gDM m <sup>-2</sup> based | grow round the clock. Production is              |                                    |  |
|                    | on the difference between              | based on 8 months of year where chl              |                                    |  |
|                    | 53 μm and 200 μm catches               | levels are elevated (Whitehouse et al.           |                                    |  |
|                    | in northern Scotia Sea (Ward           | in review, this volume)                          |                                    |  |
|                    | et al. in review this volume).         |  |                                    |  |

TABLE 2 Summary of the scenarios explored.

| Scenario  | Mid-trophic levels  | Predator response  |
|-----------|---|--|
| S1        | On-shelf <i>krill</i> biomass reduced by 80% (26 t.km <sup>-2</sup> ). Availability of <i>off-shelf krill</i> also reduced by 80%.  | Diets unchanged.   |
| S2        | On-shelf <i>krill</i> biomass reduced by 80% (26 t.km <sup>-2</sup> ). Availability of <i>off-shelf krill</i> also reduced by 80%.  | Diet switching from <i>krill</i> and <i>off-shelf krill</i> to on-shelf <i>carnivorous macroplankton</i> and <i>copepods</i> . |
| <b>S3</b> | On-shelf <i>krill</i> biomass reduced by 80% (26 t.km <sup>-2</sup> ). Availability of <i>off-shelf krill</i> also reduced by 80%. <i>Copepod</i> biomass increased by 26.17 t km <sup>-2</sup> | Diets unchanged.   |
| \$4       | On-shelf <i>krill</i> biomass reduced by 80% (26 t.km <sup>-2</sup> ). Availability of <i>off-shelf krill</i> also reduced by 80%. <i>Copepod</i> biomass increased by 26.17 t km <sup>-2</sup> | Diet switching from krill and off-shelf krill to on-shelf carnivorous macroplankton and copepods.                              |

TABLE 3: Input (base data) and balanced (model) values of non-diet input variables for all functional groups represented in the models.

|     |                                     |              |               | E       | 3       |         |         | P,           | Q/B           |                                 |
|-----|-------------------------------------|--------------|---------------|---------|---------|---------|---------|--------------|---------------|---------------------------------|
|     | Group                               | base<br>data | base<br>model | S1      | S2      | S3      | S4      | base<br>data | all<br>models | base<br>data &<br>all<br>models |
| MTW | Toothed Whales                      | 1.3E-02      | 1.6E-02       | 1.8E-03 | 1.4E-02 | 1.8E-03 | 1.4E-02 | 4.0E-02      | 4.0E-02       | 7.2E+00                         |
| MBW | Baleen whales                       | 3.0E-01      | 3.0E-01       | 8.1E-02 | 3.0E-01 | 8.1E-02 | 3.0E-01 | 2.0E-02      | 2.0E-02       | 3.4E+00                         |
| MFS | Antarctic fur seals                 | 2.0E-01      | 2.0E-01       | 1.0E-02 | 2.0E-01 | 1.0E-02 | 2.0E-01 | 2.7E-01      | 2.7E-01       | 7.7E+01                         |
| MES | Southern elephant seals             | 4.1E-01      | 4.1E-01       | 7.4E-02 | 4.1E-01 | 7.4E-02 | 4.1E-01 | 2.2E-01      | 2.2E-01       | 3.4E+01                         |
| MLS | Leopard seals                       | 1.8E-04      | 1.8E-04       | 2.7E-05 | 1.8E-04 | 2.7E-05 | 1.8E-04 | 2.0E-01      | 2.0E-01       | 3.4E+01                         |
| ВКР | King penguins                       | 1.5E-01      | 1.5E-01       | 1.5E-01 | 1.5E-01 | 1.5E-01 | 1.5E-01 | 2.1E-01      | 2.1E-01       | 6.9E+00                         |
| BGP | Gentoo peguins                      | 4.5E-02      | 4.5E-02       | 1.2E-02 | 4.5E-02 | 1.2E-02 | 4.5E-02 | 2.6E-01      | 2.6E-01       | 7.6E+01                         |
| ВМР | Macacroni penguins                  | 1.5E-01      | 1.5E-01       | 7.3E-03 | 1.5E-01 | 7.3E-03 | 1.5E-01 | 1.3E-01      | 1.3E-01       | 8.2E+01                         |
| ВСР | Chinstrap penguins                  | 4.7E-04      | 4.7E-04       | 2.3E-05 | 4.7E-04 | 2.3E-05 | 4.7E-04 | 4.6E-01      | 4.6E-01       | 8.2E+01                         |
| BBA | Black-browed albatross              | 1.7E-02      | 1.7E-02       | 4.5E-03 | 1.7E-02 | 4.5E-03 | 1.7E-02 | 6.8E-02      | 6.8E-02       | 4.8E+01                         |
| BGA | Grey-headed albatross               | 1.8E-02      | 1.8E-02       | 4.8E-03 | 1.8E-02 | 4.8E-03 | 1.8E-02 | 9.8E-02      | 9.8E-02       | 9.5E+01                         |
| ВРВ | Predatory-scavenging birds          | 2.5E-03      | 2.5E-03       | 1.6E-04 | 2.5E-03 | 1.6E-04 | 2.5E-03 | 1.8E-01      | 1.8E-01       | 7.5E+01                         |
| BDF | Diverse flying seabrids             | 2.5E-01      | 2.5E-01       | 7.6E-03 | 2.5E-01 | 7.6E-03 | 2.5E-01 | 1.3E-01      | 1.3E-01       | 2.8E+02                         |
| FAM | Mackerel icefish                    | 1.6E+00      | 1.1E+01       | 2.9E+00 | 1.1E+01 | 2.9E+00 | 1.1E+01 | 4.1E-01      | 4.1E-01       | 1.9E+00                         |
| FOI | Other icefish                       | 2.1E-01      | 1.8E+00       | 4.9E-01 | 1.8E+00 | 4.9E-01 | 1.8E+00 | 4.1E-01      | 4.1E-01       | 1.7E+00                         |
| FJT | Juvenile Patagonian toothfish       | 2.6E-01      | 2.6E-01       | 2.0E-02 | 2.6E-01 | 2.0E-02 | 2.6E-01 | 5.6E-01      | 5.6E-01       | 2.0E+00                         |
| FRO | Small rock cod                      | 2.3E-01      | 2.2E+00       | 6.0E-01 | 2.2E+00 | 6.0E-01 | 2.2E+00 | 2.4E-01      | 2.4E-01       | 2.0E+00                         |
| FOS | Other shelf fish                    | 1.5E+00      | 1.2E+01       | 3.2E+00 | 1.2E+01 | 3.2E+00 | 1.2E+01 | 2.4E-01      | 2.4E-01       | 2.5E+00                         |
| FRA | Rays                                | 3.6E-02      | 3.6E-02       | 1.8E-03 | 3.6E-02 | 1.8E-03 | 3.6E-02 | 1.6E-01      | 1.6E-01       | 1.0E+00                         |
| FPE | Pelagic fish                        | 3.2E+00      | 9.2E+00       | 2.5E+00 | 9.2E+00 | 2.5E+00 | 9.2E+00 | 4.4E-01      | 4.4E-01       | 4.3E+00                         |
| ICE | Squid                               | 5.8E-02      | 5.8E-02       | 9.9E-03 | 5.8E-02 | 9.9E-03 | 5.8E-02 | 6.7E+00      | 8.2E+00       | 2.2E+01                         |
| IBE | Benthos                             | 7.2E+01      | 7.2E+01       | 7.2E+01 | 7.2E+01 | 7.2E+01 | 7.2E+01 | 5.4E-01      | 5.4E-01       | 2.2E+00                         |
| ZKR | Euphausia superba                   | 3.3E+01      | 3.3E+01       | 6.5E+00 | 6.5E+00 | 6.5E+00 | 6.5E+00 | 2.4E+00      | 2.4E+00       | 9.6E+00                         |
| ZCA | Carnivorous zooplankton             | 8.4E+00      | 1.0E+01       | 1.0E+01 | 1.0E+01 | 1.0E+01 | 1.0E+01 | 4.9E+00      | 4.9E+00       | 1.9E+01                         |
| ZHE | Herbivorous zooplankton             | 6.9E+00      | 6.9E+00       | 6.9E+00 | 6.9E+00 | 6.9E+00 | 6.9E+00 | 8.6E+00      | 8.6E+00       | 3.4E+01                         |
| ZCO | Hebivorous & detritivorous copepods | 2.8E+01      | 2.8E+01       | 2.8E+01 | 2.8E+01 | 5.4E+01 | 5.4E+01 | 8.6E+00      | 8.6E+00       | 3.4E+01                         |
| ZSA | Salps                               | 1.0E+00      | 1.0E+00       | 1.0E+00 | 1.0E+00 | 1.0E+00 | 1.0E+00 | 7.6E+00      | 7.6E+00       | 3.0E+01                         |
| ZHT | Heterotrophic<br>microplankton      | 2.0E+01      | 2.0E+01       | 2.0E+01 | 2.0E+01 | 2.4E+01 | 2.4E+01 | 4.7E+01      | 4.7E+01       | 1.6E+02                         |
| PBA | Heterotrophic Bacteria              | 5.1E+00      | 2.2E+01       | 2.2E+01 | 2.2E+01 | 2.6E+01 | 2.6E+01 | 2.0E+01      | 2.0E+01       | 6.7E+01                         |
| PHY | Primary producers                   | 2.6E+01      | 4.5E+01       | 4.3E+01 | 4.3E+01 | 5.5E+01 | 5.5E+01 | 6.0E+01      | 6.0E+01       | 0.0E+00                         |

TABLE 4a: Diet matrix for the base model (vertebrate predators). The table demonstrates the method used to distinguish off-shelf feeding on krill (EIM1) from off-shelf feeding on other prey (EIM2): The off-shelf prey were modelled as discrete functional groups, each feeding each feeding entirely on "Import".

| Prey \ | MTW     | MBW | MFS     | MES     | MLS     | ВКР     | BGP     | ВМР     | ВСР     | BBA     | BGA     | ВРВ     | BDF     | FAM     | FOI     | FJT     | FRO     | FOS     | FRA     | FPE |
|--------|---------|-----|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|-----|
| MTW    |         |     |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |     |
| ИBW    |         |     |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |     |
| ИFS    | 5.9E-03 |     |         |         | 3.3E-01 |         |         |         |         |         |         | 3.9E-03 |         |         |         |         |         |         |         |     |
| ИES    |         |     |         |         | 3.0E-03 |         |         |         |         |         |         | 1.3E-03 |         |         |         |         |         |         |         |     |
| MLS    | 2.0E-05 |     |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |     |
| ВКР    | 2.0E-03 |     |         |         |         |         |         |         |         |         |         | 5.4E-02 |         |         |         |         |         |         |         |     |
| BGP    |         |     |         |         | 2.1E-01 |         |         |         |         |         |         | 2.6E-02 |         |         |         |         |         |         |         |     |
| ВМР    |         |     |         |         | 7.0E-03 |         |         |         |         |         |         | 5.1E-02 |         |         |         |         |         |         |         |     |
| ВСР    |         |     |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |     |
| BBA    |         |     |         |         |         |         |         |         |         |         |         | 2.6E-03 |         |         |         |         |         |         |         |     |
| BGA    |         |     |         |         |         |         |         |         |         |         |         | 1.3E-03 |         |         |         |         |         |         |         |     |
| ВРВ    |         |     |         |         |         |         |         |         |         |         |         | 1.0E-03 |         |         |         |         |         |         |         |     |
| BDF    |         |     |         |         |         |         |         |         |         |         |         | 8.4E-02 |         |         |         |         |         |         |         |     |
| FAM    | 4.1E-02 |     | 1.3E-02 | 5.0E-03 |         | 1.0E-02 | 2.0E-01 | 1.2E-01 | 1.1E-02 | 2.7E-02 | 8.0E-03 |         | 1.3E-04 |         | 1.1E-01 | 3.8E-02 |         | 5.2E-02 |         |     |
| =OI    | 5.5E-03 |     | 2.7E-03 | 3.5E-03 |         |         |         |         |         |         |         |         | 2.4E-05 | 2.0E-02 | 5.0E-03 | 2.9E-02 |         | 7.5E-03 | 3.9E-02 |     |
| FJT    | 6.6E-03 |     |         |         |         |         |         |         |         |         |         | 6.6E-03 | 4.8E-05 |         |         |         |         |         |         |     |
| RO     | 5.9E-03 |     | 1.4E-03 |         |         | 1.0E-02 | 4.5E-03 |         | 1.1E-02 |         |         |         | 1.1E-04 |         | 3.9E-02 | 3.1E-01 | 2.7E-02 |         | 1.4E-01 |     |
| FOS    | 4.0E-02 |     | 1.3E-02 | 2.0E-02 |         | 5.0E-03 | 1.6E-03 | 9.0E-02 | 1.7E-02 | 2.1E-02 | 6.1E-03 |         | 1.8E-04 | 3.0E-02 | 1.5E-01 | 1.3E-01 |         |         | 2.2E-01 |     |
| ₹RA    | 9.4E-04 |     |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         | 3.7E-05 |     |
| PE     | 7.3E-02 |     | 3.7E-02 | 3.0E-02 |         | 7.5E-02 | 5.0E-04 | 6.1E-02 | 3.9E-02 |         |         |         | 6.2E-03 |         | 1.0E-02 | 6.8E-02 |         | 5.6E-02 |         |     |
| CE     | 2.0E-02 |     | 7.7E-04 | 3.2E-02 |         |         |         | 1.3E-03 | 8.1E-04 |         |         |         | 1.3E-04 |         |         | 1.5E-02 |         |         | 8.3E-02 |     |
| BE     |         |     |         |         |         |         |         |         |         |         |         |         |         |         | 2.4E-01 | 7.6E-02 | 4.5E-02 | 2.0E-02 | 4.1E-01 |     |

| ZKR      | 8.0E-01 | 3.7E-01 | 4.5E-01 | 7.9E-01 | 4.2E-01 | 3.2E-01 | 8.2E-02 | 7.6E-02 | 7.9E-02 | 9.8E-02 | 7.7E-01 | 4.0E-01 | 3.0E-01 | 5.4E-01 | 3.8E-01 | 1.1E-01 | 1.4E-01 |
|----------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|
| ZCA      |         |         |         | 6.0E-04 | 5.9E-02 |         |         |         |         | 7.3E-03 | 1.8E-01 | 3.8E-02 | 4.0E-02 | 3.1E-01 | 3.7E-01 |         | 7.0E-01 |
| ZHE      |         |         |         |         |         |         |         |         |         |         |         |         |         | 0.0E+00 | 0.0E+00 |         | 1.5E-03 |
| zco      | 2.0E-01 |         |         |         |         |         |         |         |         | 4.9E-02 |         |         |         | 8.3E-02 | 0.0E+00 |         | 1.4E-01 |
| ZSA      |         |         |         |         |         |         |         |         |         |         |         |         |         | 6.0E-04 | 1.1E-01 |         | 1.7E-02 |
| ZHT      |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |
| PBA      |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |
| PHY      |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |
| EIM1     |         | 4.7E-01 |         |         | 1.4E-01 | 4.8E-01 | 5.5E-01 | 7.7E-01 | 1.3E-01 | 4.9E-01 |         |         |         |         |         |         |         |
| EIM2     | 8.0E-01 | 8.6E-02 | 9.1E-01 | 9.0E-01 | 1.1E-01 | 1.2E-01 | 3.2E-01 | 1.4E-01 | 3.9E-01 | 3.2E-01 |         |         |         |         |         |         |         |
| Detritus |         |         |         |         |         |         |         |         | 1.7E-01 | 3.1E-02 |         |         |         | 7.0E-04 |         |         |         |
| Import   |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |         |

TABLE 4b: Diet matrix for the base model (invertebrate predators and off-shelf goups).

| Prey\    | ICE     | IBE     | ZKR     | ZCA     | ZHE     | ZCO     | ZSA     | ZHT     | PBA     | EIM1    | EIM2    |
|----------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|
| predator |         |         |         |         |         |         |         |         |         |         |         |
| FJT      | 5.0E-02 |         |         |         |         |         |         |         |         |         |         |
| FRO      | 5.0E-02 |         |         |         |         |         |         |         |         |         |         |
| FOS      | 5.0E-02 |         |         |         |         |         |         |         |         |         |         |
| FRA      |         |         |         |         |         |         |         |         |         |         |         |
| FPE      | 5.0E-02 |         |         |         |         |         |         |         |         |         |         |
| ICE      |         |         |         |         |         |         |         |         |         |         |         |
| IBE      | 1.5E-01 | 8.9E-02 |         |         |         |         |         |         |         |         |         |
| ZKR      | 2.5E-01 |         |         |         |         |         |         |         |         |         |         |
| ZCA      | 1.0E-01 | 4.7E-04 | 1.0E-02 |         |         |         |         |         |         |         |         |
| ZHE      |         | 4.7E-04 | 5.0E-03 | 1.5E-01 |         |         | 2.0E-02 |         |         |         |         |
| ZCO      | 3.0E-01 | 2.4E-04 | 1.5E-02 | 5.4E-01 |         |         | 2.0E-02 |         |         |         |         |
| ZSA      |         |         |         | 1.0E-02 |         |         |         |         |         |         |         |
| ZHT      |         | 1.9E-01 | 4.5E-01 | 1.0E-01 | 5.0E-01 | 5.0E-01 | 4.5E-01 |         |         |         |         |
| PBA      |         | 2.0E-01 |         |         | 8.0E-02 | 8.0E-02 | 6.0E-02 | 4.0E-01 |         |         |         |
| PHY      |         | 5.1E-01 | 4.2E-01 | 1.0E-01 | 4.0E-01 | 4.0E-01 | 4.5E-01 | 6.0E-01 |         |         |         |
| EIM1     |         |         |         |         |         |         |         |         |         |         |         |
| EIM2     |         |         |         |         |         |         |         |         |         |         |         |
| Detritus |         | 1.5E-02 | 1.0E-01 | 1.0E-01 | 2.0E-02 | 2.0E-02 |         |         | 1.0E+00 |         |         |
| Import   |         |         |         |         |         |         |         |         |         | 1.0E+00 | 1.0E+00 |

TABLE 5: Summary statistics for the base dataset and balanced models. All estimates exclude *detritus* unless otherwise indicated. EE (overall) and mean trophic level are biomass weighted averages.

| Statistic          | base data | base model | S1    | S2    | S3    | S4    | units                                |
|--------------------|-----------|------------|-------|-------|-------|-------|--------------------------------------|
| B (total)          | 208       | 274        | 219   | 246   | 266   | 293   | t.km <sup>-2</sup>                   |
| B (UTL)            | 9         | 38         | 10    | 38    | 10    | 38    | t.km <sup>-2</sup>                   |
| B (ZOO)            | 77        | 78         | 52    | 52    | 78    | 78    | t.km <sup>-2</sup>                   |
| P (total)          | 3,058     | 4,578      | 4,400 | 4,409 | 5,624 | 5,635 | t.km <sup>-2</sup> .yr <sup>-1</sup> |
| P (UTL)            | 3         | 13         | 4     | 13    | 4     | 13    | t.km <sup>-2</sup> .yr <sup>-1</sup> |
| P (ZOO)            | 424       | 432        | 369   | 369   | 594   | 594   | t.km <sup>-2</sup> .yr <sup>-1</sup> |
| PP                 | 1,534     | 2,702      | 2,597 | 2,597 | 3,324 | 3,324 | t.km <sup>-2</sup> .yr <sup>-1</sup> |
| Q (total)          | 5,634     | 6,851      | 6,419 | 6,600 | 8,248 | 8,429 | t.km <sup>-2</sup> .yr <sup>-1</sup> |
| Q (of detritus)    | 414       | 1528       | 1501  | 1503  | 1835  | 1838  | t.km <sup>-2</sup> .yr <sup>-1</sup> |
| Q (of imports)     | 83        | 83         | 6     | 55    | 6     | 55    | t.km <sup>-2</sup> .yr <sup>-1</sup> |
| Q (of ZKR)         | 27        | 58         | 12    | 16    | 12    | 16    |                                      |
| Q (of ZCA)         | 16        | 49         | 13    | 49    | 13    | 49    | t.km <sup>-2</sup> .yr <sup>-1</sup> |
| Q (of ZCO)         | 100       | 120        | 108   | 184   | 108   | 184   | t.km <sup>-2</sup> .yr <sup>-1</sup> |
| EE (overall)       | 1.68      | 0.92       | 0.89  | 0.91  | 0.92  | 0.94  |                                      |
| Mean trophic level | 2.51      | 2.54       | 2.34  | 2.55  | 2.32  | 2.50  |                                      |

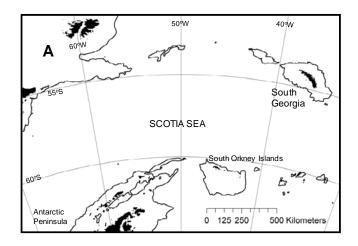
UTL= upper trophic levels (mammals, birds, fish and cephalopods); ZOO=zooplankton (excluding ZHT); B=biomass; P=production; PP=primary production; Q=consumption.

TABLE 6: Trophic level and Ecotrophic efficiency (EE) for all trophic groups in the models. Trophic level is calculated as 1 + the mean trohic level of a predator's prey groups, weighted by their contribution to the predator's diet, where detritus and primary producers have a trophic level of 1. Bold values indicate "unbalanced" ecotrophic efficiencies (>1) in the base dataset.

|       |           | Troph      | ic level  |      |            |           | EE        |            |      |      |            |           |  |
|-------|-----------|------------|-----------|------|------------|-----------|-----------|------------|------|------|------------|-----------|--|
| Group | base data | base model | <b>S1</b> | S2   | <b>S</b> 3 | <b>S4</b> | base data | base model | S1   | S2   | <b>S</b> 3 | <b>S4</b> |  |
| MTW   | 5.20      | 5.10       | 5.10      | 5.10 | 5.10       | 5.10      | 0.00      | 0.00       | 0.00 | 0.00 | 0.00       | 0.00      |  |
| MBW   | 3.71      | 3.71       | 3.71      | 3.77 | 3.71       | 3.77      | 0.00      | 0.00       | 0.00 | 0.00 | 0.00       | 0.00      |  |
| MFS   | 3.92      | 3.91       | 3.91      | 3.89 | 3.91       | 3.89      | 0.06      | 0.06       | 0.16 | 0.06 | 0.16       | 0.06      |  |
| MES   | 5.18      | 5.11       | 5.11      | 5.06 | 5.11       | 5.06      | 0.00      | 0.00       | 0.00 | 0.00 | 0.00       | 0.00      |  |
| MLS   | 4.43      | 4.37       | 4.37      | 4.48 | 4.37       | 4.48      | 5.53      | 0.06       | 0.05 | 0.06 | 0.05       | 0.06      |  |
| BKP   | 5.16      | 5.13       | 5.13      | 4.89 | 5.13       | 4.89      | 0.32      | 0.32       | 0.02 | 0.32 | 0.02       | 0.32      |  |
| BGP   | 4.19      | 3.94       | 3.93      | 4.47 | 3.94       | 4.47      | 0.54      | 0.54       | 0.16 | 0.54 | 0.16       | 0.54      |  |
| BMP   | 4.26      | 4.23       | 4.22      | 4.58 | 4.23       | 4.58      | 0.51      | 0.51       | 0.65 | 0.51 | 0.66       | 0.51      |  |
| BCP   | 3.98      | 3.97       | 3.97      | 3.95 | 3.97       | 3.95      | 0.00      | 0.00       | 0.00 | 0.00 | 0.00       | 0.00      |  |
| BBA   | 4.21      | 4.17       | 4.17      | 4.29 | 4.17       | 4.29      | 0.45      | 0.45       | 0.10 | 0.45 | 0.10       | 0.45      |  |
| BGA   | 3.91      | 3.89       | 3.89      | 3.94 | 3.89       | 3.94      | 0.15      | 0.15       | 0.03 | 0.15 | 0.03       | 0.15      |  |
| BPB   | 3.84      | 3.80       | 3.80      | 3.87 | 3.80       | 3.87      | 3.85      | 0.42       | 0.42 | 0.42 | 0.42       | 0.42      |  |
| BDF   | 3.52      | 3.52       | 3.52      | 3.70 | 3.52       | 3.70      | 0.49      | 0.49       | 0.99 | 0.49 | 1.00       | 0.49      |  |
| FAM   | 4.03      | 3.88       | 3.88      | 4.33 | 3.88       | 4.33      | 4.45      | 1.00       | 0.69 | 1.00 | 0.69       | 1.00      |  |
| FOI   | 4.98      | 4.11       | 4.11      | 4.42 | 4.11       | 4.42      | 2.48      | 1.00       | 0.91 | 1.00 | 0.91       | 1.00      |  |
| FJT   | 5.03      | 4.49       | 4.49      | 4.64 | 4.49       | 4.64      | 0.49      | 0.49       | 0.99 | 0.49 | 0.99       | 0.49      |  |
| FRO   | 4.03      | 3.94       | 3.94      | 3.80 | 3.94       | 3.80      | 10.10     | 1.00       | 0.74 | 1.00 | 0.74       | 1.00      |  |
| FOS   | 4.18      | 4.11       | 4.11      | 4.33 | 4.11       | 4.33      | 7.92      | 1.00       | 0.57 | 1.00 | 0.57       | 1.00      |  |
| FRA   | 4.38      | 4.31       | 4.31      | 4.35 | 4.31       | 4.35      | 0.02      | 0.02       | 0.04 | 0.02 | 0.04       | 0.02      |  |
| FPE   | 4.19      | 4.19       | 4.19      | 3.81 | 4.19       | 3.81      | 1.82      | 1.00       | 0.65 | 1.00 | 0.65       | 1.00      |  |
| ICE   | 4.11      | 4.07       | 4.07      | 4.03 | 4.07       | 4.03      | 1.21      | 1.00       | 1.00 | 1.00 | 1.00       | 1.00      |  |
| IBE   | 2.60      | 2.60       | 2.60      | 2.60 | 2.60       | 2.60      | 0.36      | 0.40       | 0.37 | 0.40 | 0.37       | 0.40      |  |
| ZKR   | 2.69      | 2.69       | 2.69      | 2.68 | 2.69       | 2.68      | 0.34      | 0.74       | 0.74 | 1.00 | 0.74       | 1.00      |  |
| ZCA   | 3.39      | 3.39       | 3.39      | 3.39 | 3.39       | 3.39      | 0.40      | 1.00       | 0.26 | 1.00 | 0.26       | 1.00      |  |
| ZHE   | 2.78      | 2.78       | 2.78      | 2.78 | 2.78       | 2.78      | 0.45      | 0.53       | 0.51 | 0.51 | 0.51       | 0.51      |  |

| ZCO | 2.78 | 2.78 | 2.78 | 2.78 | 2.78 | 2.78 | 0.42  | 0.50 | 0.46 | 0.77 | 0.23 | 0.40 |
|-----|------|------|------|------|------|------|-------|------|------|------|------|------|
| ZSA | 2.76 | 2.76 | 2.76 | 2.76 | 2.76 | 2.76 | 0.30  | 0.78 | 0.40 | 0.78 | 0.40 | 0.78 |
| ZHT | 2.40 | 2.40 | 2.40 | 2.40 | 2.40 | 2.40 | 0.83  | 0.83 | 0.71 | 0.71 | 1.00 | 1.00 |
| PBA | 2.00 | 2.00 | 2.00 | 2.00 | 2.00 | 2.00 | 14.28 | 1.00 | 1.00 | 1.00 | 1.00 | 1.00 |
| PHY | 1.00 | 1.00 | 1.00 | 1.00 | 1.00 | 1.00 | 1.76  | 1.00 | 1.00 | 1.00 | 1.00 | 1.00 |
| DET | 1.00 | 1.00 | 1.00 | 1.00 | 1.00 | 1.00 | 0.00  | 0.80 | 0.78 | 0.84 | 0.82 | 0.87 |

Fig 1: The modelled area (South Georgia shelf to the 1000m isobath) in the regional (A) and (B) local context.



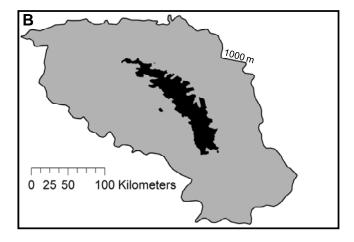


Fig 2: The trophic role of Antarctic krill showing flow rates from its prey and to its main predators (in the base dataset and, in parentheses, the base model).

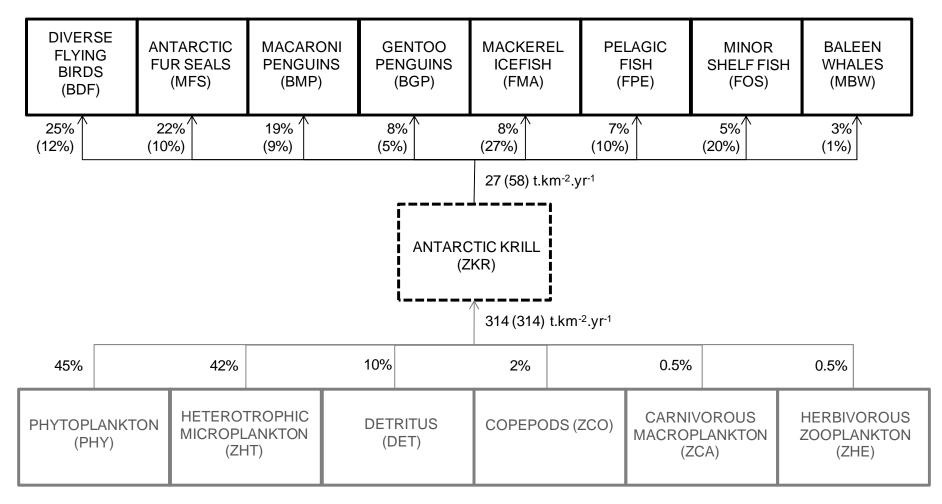


Fig 3: The trophic role of herbivorous and detritivorous copepods showing flow rates from their prey (in the base dataset) and to their main predators (in the base dataset and, in parentheses, the base model).

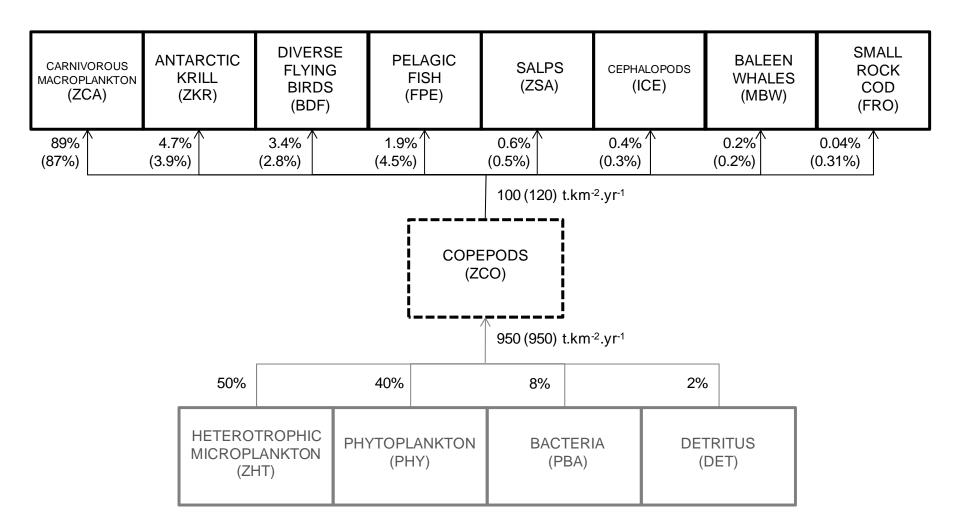


Figure 4: Production by higher trophic level taxa (M=mammals, A(P)=penguins, A(F)=flying seabirds, F(D)=demersal fish, F(P)=pelagic fish, C=cephalopods) in the base dataset, the base model, and switching (S2 & S4) and non switching (S1 & S3) scenario models.

