

**A high resolution palynozonation for the Pennsylvanian to Lower Permian  
Al Khlata Formation, south Oman**

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**ABSTRACT**

Palynology is the main method of correlating the subsurface glaciogenic Al Khlata Formation  
of Oman, due to extreme lateral variability of facies, and poor seismic resolution. The chief

operating company in Oman, Petroleum Development Oman, has developed a robust in-house palynozonation through almost 40 years of exploration and production based on thousands of samples and hundreds of well sections. In this paper the formal definitions of the biozones are published, and the biozones are correlated in detail with faunally-calibrated palynological biozones in Western Australia, thereby allowing correlation with the standard Russian stages. Seven biozones are distinguished. The oldest, biozones 2159A and B, of probable late Pennsylvanian age, are characterised by low diversity assemblages of *Punctatisporites* and monosaccate pollen, with Biozone 2159A having a lower proportion of monosaccate pollen than Biozone 2159B. Biozone 2165A, of probable Asselian age, is characterised by common cingulicamerate spores and *Microbaculispora* Group; while Biozone 2165B contains in addition to the above, common *Horriditriletes* Group, and is likely to be Asselian-Sakmarian in age. Biozone 2141A is characterised by common taeniate and non-taeniate bisaccate pollen and *Cycadopites cymbatus*, particularly toward the top; and the succeeding 2141B by common *Microbaculispora* Group and *C. cymbatus*. Biozones 2141A and B are believed to be Sakmarian in age. The youngest, Biozone 2141C, is characterised by *Microbaculispora* and *Horriditriletes* Group with the former being more abundant. The seven-fold subdivision is a considerable improvement on previous palynozonations of the Al Khlata Formation, which only allowed three or four-fold subdivisions; however, there still remain considerable uncertainties in dating the lowest biozones, 2159A and B. Improvements in dating of Al Khlata Formation sediments are most likely to come from palynological correlation with radiometrically- or faunally dated sequences in South America, eastern Australia and southern Africa.

## INTRODUCTION

Pennsylvanian to Lower Permian rocks crop out over relatively small areas of central Saudi Arabia, the Wajid outcrop area in southwestern Saudi Arabia, and in the Haushi-Huqf outcrop area in Oman (Text-Figure 1), but are present widely in the subsurface, where they form important hydrocarbon reservoirs (McGillivray and Hussein, 1992; Hughes Clarke, 1988). The main formations of the region containing Pennsylvanian to Lower Permian sequences are the Al Khlata and Gharif formations in Oman, and the Unayzah Formation in Saudi Arabia.

The Al Khlata Formation consists of a complex package of clastic lithologies that range from conglomerates through diamictites, gravels, pebbly sandstones, siltstones to silty shales (Braakman et al., 1982; Hughes Clarke, 1988; Levell et al., 1988; Dubreuilh et al., 1992; Roger et al., 1992; Al-Belushi et al., 1996; Angiolini et al., 2003). The glaciogenic nature of the Al Khlata Formation was established from studies of the outcrop area on the western flank of the Haushi-Huqf Uplift where striated pavements of the Precambrian Khufai Formation have been found (Braakman et al., 1982; Al-Belushi et al., 1996). Hughes Clarke (1988) credited Levell et al. (1982) with the definition of the type section of the Al Khlata Formation, which is located at Wadi Al Khlata near the Haushi-Huqf Uplift (57°25'46"E, 19°46'43"N; Text-Figure 1) where it is about 100 m thick. The subsurface reference section of the Al Khlata Formation was defined in Rahab-2 well, south Oman (55°06'32"E, 18°01'09"N) where the formation is more than 246 m thick (Hughes Clarke, 1988; Mohammed et al., 1997). In the subsurface of south Oman, the thickness of the Al Khlata Formation varies from about 100 to 800 m (Levell et al., 1988; Love, 1994; Knight and Hartkamp-Bakker, 1998). The variation of thickness is largely the result of syn-depositional subsidence and erosional palaeorelief on the pre-Al Khlata unconformity (Levell et al., 1988; Knight and Hartkamp-Bakker, 1998). During its deposition, the infra-Cambrian Ara salt in the South Oman Salt Basin moved diapirically to form salt domes, and developed a series of NS-oriented salt pods.

As a result, thick Al Khlata sediments are found in synclines formed by salt withdrawal. On the Eastern Flank, thick Al Khlata Formation deposits are also found where salt was apparently syndepositionally dissolved by groundwater (Heward, 1990; Osterloff et al., 2004).

The Al Khlata Formation is present throughout the central interior of Oman south of the Oman Mountains, and west of the Huqf Axis high (Text-Figure 1). It rests with angular unconformity on Devonian and older rocks (Hughes Clark, 1988). The upper boundary of the subsurface Al Khlata Formation is described by Mohammed et al. (1997) as broadly conformable with the overlying Gharif Formation. This lithological boundary is generally picked at the base of the lowermost Gharif sandstone, overlying the upper Rahab Member of the Al Khlata Formation. On electric logs this boundary is marked by a downhole increase in gamma ray, sonic, and the separation of the density and neutron logs down-hole (Osterloff et al., 2004). In north-central Oman, the Rahab Member is not distinct and the boundary is mainly identified via palynostratigraphical proxy.

Petroleum Development Oman has used palynology extensively for correlating the subsurface Al Khlata Formation, such that at present it is the chief method used due to extreme lateral variability of facies (and therefore of the wireline logs), and poor seismic resolution.

Historically PDO have identified important palynomorphs by code numbers rather than Linnaean binomials, for example the colpate pollen taxon *Cycadopites cymbatus* is known by the code 2141. Where a taxon is taken as marker or index for a biozone, its code is used also to denote that biozone. Hence the original 2141 Biozone in its broadest terms was characterised by the common occurrence of *Cycadopites cymbatus*. The system in various forms has been in operation for nearly 40 years in PDO, and has proved to be extremely robust having been developed from a database of thousands of samples and hundreds of well sections. However, although its basic characteristics have been illustrated (Osterloff et al.

2004, fig. 7), formal definitions of the biozones have not been published. The biozones have also not been correlated in detail with other biozones of Gondwana. Thus the main purpose of this paper is to define and correlate the biozones. For continuity and simplicity, the nomenclature for biozones used internally in PDO is retained.

## PALYNOLOGICAL SUCCESSION

Generally the diversity and yield of palynomorphs increases upsection through the Al Khlata Formation, probably in response to climatic amelioration, related to global post glacial warming and the northward movement of the Arabian Plate between the Pennsylvanian and Early Permian (Stephenson and Filatoff 2000; Stephenson et al., 2005; 2007). The earlier assemblages are characterised by common monosaccate pollen and *Punctatisporites* Group, while later assemblages are of greater diversity including cingulicamerate and cheilocardioid spores, and colpate and bisaccate pollen. The biozones are of the acme type with bases defined by quantitative changes in a palynomorph group or taxon. In this paper we will list the constituents of the groups and then show how the patterns of occurrence of groups define the biozones. The composition of the groups and authors of taxa are given in Table 1. Along with groups, certain taxa are also important, for example *Anapiculatisporites concinnus*, *Cycadopites cymbatus*, and *Kingiacolpites subcircularis*. Key taxa and groups are illustrated in Plates 1 and 2.

## BIOZONATION

**Methodology.** Percentage proportions of taxon groups and species are central to the method of biozonation, and proportions quoted in this paper are averages gained from many

hundreds of well analyses involving thousands of samples (Table 2). PDO in-house procedures suggest that counts of approximately 150 to 200 specimens per slide are adequate to represent true proportions. Three reliability categories are applied: ‘poor’ (total count 50 - 74 specimens), ‘fair’ (75 – 99 specimens) and ‘good’ (over 100 specimens). Samples with counts of less than 50 are designated non-diagnostic and therefore are not used in biozonation, except in the case of 2159, where a tentative assignment is allowed with counts of between 25 and 50. Samples from cored well sections are the main basis of the biozonation, though sidewall core samples are used in important sections where core is not available.

It should be noted that biostratigraphy of this type, based mainly on quantitative trends is not an exact science, and judgement is sometimes required in the positioning of biozonal boundaries. This is also true in the case of reworking of palynomorphs, which is common in glaciogenic sediments. The Al Khlata Formation contains palynomorphs derived by glacial erosion of consolidated hinterland rocks (‘long-cycle’ reworked palynomorphs, for example from the Devonian) as well those derived from younger unconsolidated glacial sediments (‘short-cycle’ reworked palynomorphs; Stephenson, 2008). The latter, which are difficult to distinguish from *in situ* palynomorphs, make reliable palynozonation difficult without reference to sedimentary facies and appropriate experience concerning which data to use or reject.

**2159A Biozone.** The lowest biozone of the Al Khlata Formation has a cored reference section in Marmul-6H1 well between 920.39 to 902.77m (Text-Figure 2). The primary diagnostic criterion is the abundance of the *Punctatisporites* Group which constitutes up to 100% of assemblages. Taxa of the Monosaccate Group constitute less than 5% of assemblages. Below the base of the biozone, samples contain non-diagnostic assemblages or

are barren with sparse reworking of Devonian to Lower Palaeozoic spores, cryptospores or acritarchs. Rarely *Anapiculatisporites concinnus*, *Aratrisporites saharaensis*, *Brevitriletes* spp., and members of the Cingulicamerate Group such as *Vallatisporites arcuatus* occur as do algal palynomorphs such as *Botryococcus*, *Tetraporina* spp. and *Tasmanites*. Apart from the reference section, this biozone is well represented in the cored interval from Qata-9 (Text-Figure 2). However, in uncored sections it is usually identified from spot sidewall samples such as exemplified in Rima-65 (see also Text-Figure 2).

There is a transition between 2159A and the succeeding Biozone 2159B, with the result that the boundary between the zones is difficult to position precisely. In the transition section, the percentage of monosaccate pollen fluctuate on average between 5% and 10% with the balance of the assemblage consisting mainly of the *Punctatisporites* Group (Qata-9, Text-Figure 2).

**2159B Biozone.** The reference section is the cored Marmul-6H1 interval from 901.8 to 870.5m (Text-Figure 2), and the main feature that distinguishes 2159B from 2159A is the increase in the proportion of the Monosaccate Group to greater than 10% of assemblages with a proportional decrease in *Punctatisporites* Group. 2159B Biozone is also more diverse than 2159A in containing *Anapiculatisporites concinnus* (usually less than 5% of the assemblage), *Aratrisporites saharaensis*, *Apiculiretusispora* spp., *Brevitriletes* spp., *Cyclogranisporites* spp., *Densosporites* spp. (including *D. rotundidentatus*), *Dibolisporites*, *Wilsonites australiensis*, *Spelaeotriletes* spp. (including *S. triangulus*), *Vallatisporites arcuatus* and *Verrucosisporites andersonii*. Overall there is an increase in the Monosaccate Group upsection through the biozone and this group may be locally very abundant (up to 70% of

assemblages) at the top of the biozone (Rima-65, Text-Figure 2). Apart from the reference section, this biozone is well represented in cored intervals within Al Burj-36 and Rima-65.

**2165A Biozone.** The reference section is the Wadi Al Khlata Borehole-5 between 27.0 and 17.75m. The primary diagnostic criteria are (1) the Cingulicamerate Group constitutes up to approximately 30% of the assemblages (mainly *Vallatisporites arcuatus*, *Lundbladispora braziliensis* and *Cristatisporites* spp.); and (2) the *Microbaculispora* Group constitutes approximately 5% of assemblages, although it can be rare or absent locally (Text-Figure 3). The *Punctatisporites* and Monosaccate groups may also be abundant. *Anapiculatisporites concinnus* is present, usually comprising less than 5% of the assemblage, and the *Horriditriletes* and Bisaccate groups are usually rare or absent. Diversity is relatively high in comparison with 2159A, with *Ahrensia cristatus* 1979, *A. saharaensis*, *Apiculiretusispora* spp., *Cyclogranisporites* spp., *Densosporites* spp., *D. disfacies*, *W. australiensis* and *V. andersonii* present (Text-Figure 3). Apart from the cored reference section, this biozone is well represented in sidewall sampled intervals within Dimeet-1 and Al Burj-23 (Text-Figure 3).

**2165B Biozone** The reference section is Wadi Al Khlata Borehole-5 from 16.73 to 14.45m. The primary diagnostic criteria are (1) *Horriditriletes* Group constitutes between 5 and 10%, or up to 20% of assemblages; and (2) *Microbaculispora* Group constitutes usually around 3 to 5% of assemblages but can be occasionally absent. The Cingulicamerate Group may constitute 20 to 30% or more of assemblages. (mainly *Lundbladispora* spp., *Cristatisporites* spp. and *Vallatisporites arcuatus*). The *Punctatisporites* Group is usually less frequent than in the underlying biozones. *Cycadopites cymbatus*, *Granulatisporites*



*confluens*, *Converrucosisporites grandegranulatus*, *Marsupipollenites* spp. and the *Vittatina* and Taeniate Bisaccate groups appear towards the top of this biozone (Text-Figure 3, 5).

Apart from the reference section, this biozone is well represented in the cored interval within Amal-9 and sidewall sampled sections from Dimeet-1 and Al Burj-23 (see also Text-Figure 3).

**2141A Biozone.** The reference section is Nimr-46 cored interval between 934.59 to 897.1m. The primary diagnostic criteria are: (1) non-taeniate and taeniate bisaccate groups together represent more than 10% of the assemblages; and (2) *Cycadopites cymbatus* is rare towards the base but increases to 5 to 10% of assemblages towards the top. The proportions of the Cingulicamerate Group vary considerably but are usually lower than those of 2165A or B biozones. The proportion of *Punctatisporites* Group continues to decrease so that percentages are lower than in 2165B. *Horriditriletes* Group usually represents 10% but can be up to 20% of assemblages, and the *Microbaculispora* Group is approximately 3 to 5% of assemblages but can be occasionally absent. Taxa or groups that occur rarely include *Kingiacolpites subcircularis* and the *Vittatina* Group, *Brevitriletes cornutus*, *Dibolisporites disfacies*, *Densosporites* spp. (including *Densosporites rotundidentatus*), *Verrucosisporites* spp., *Botryococcus*, *Deusilites tentus*, *Tetraporina* spp. and *Tasmanites* (Text-Figure 4). Apart from the reference section, this biozone is well represented in the cored intervals of Rahab-2 and Wadi Al Khlata Borehole-2 (Text-Figure 4).

**2141B Biozone.** The reference section is Rahab-2 between 1008.32 and 992.14 m. The primary diagnostic criteria are (1) The *Microbaculispora* Group represents greater than 10% and can constitute up to 60% of assemblages, in addition the *Microbaculispora* Group is

more abundant than the *Horriditriletes* Group; (2) *Cycadopites cymbatus* makes up 5 to 10% of assemblages; and (3) the Cingulicamerate Group are extremely rare or absent. The Non-Taeniate Bisaccate Group exceeds the Taeniate Bisaccate Group. Taxa that occur rarely include *Kingiacolpites subcircularis*, *Deusilites tentus*, *Tetraporina* spp. and *Tasmanites*. Apart from the reference section, this biozone is also well represented in the cored interval of Rahab-28 (Text-Figure 4).

**2141C Biozone.** The reference section is Rahab-2 between 982.38 and 964.7m. The primary diagnostic criterion is that the *Microbaculispora* Group represents more than 10% of assemblages, but the *Horriditriletes* Group exceeds it in proportion, reaching 30% or more. *Cycadopites cymbatus* and the Non-Taeniate Bisaccate Group are variable but usually common to abundant, reaching abundances of 60% or more of assemblages. The Taeniate Bisaccate Group represents 5 to 20% of assemblages but is always less abundant than the non-taeniate taxa. The accessory taxa are broadly the same as those in 2141B. Working downhole, however, the first appearances of *Vittatina* cf. *scutata* and *Converrucosisporites grandegrnulatus* are characteristic of this biozone. Apart from the reference section, this biozone is also well represented in the cored interval of Rahab-28 (Text-Figure 4).

## **CORRELATION WITH ARABIA AND WESTERN AUSTRALIA**

Besems and Schuurman (1987) and Love (1994) produced palynostratigraphical schemes for Oman (Text-Figure 6). The former authors, working on outcrops of the Al Khlata Formation in east-Central Oman, recognised two distinct palynological assemblages. Assemblage Group A is dominated by zonate, trilete spores and Assemblage Group B is characterised by taeniate

and non-taeniate bisaccate pollen. The quantitative data of Besems and Schuurman (1987; text-figures 3-7), which comes from a very small number of sample horizons suggests that Assemblage Group A correlates with 2165A and B, and Assemblage Group B with 2141A. The boundary between Assemblage Groups A and B coincides with that between 2165B and 2141A, because both are defined on the increase of bisaccate pollen to approximately 10%.

Love (1994) described four palynological assemblages from the entire subsurface Haushi Group. The *Potonieisporites* Assemblage is reportedly low in diversity containing simple trilete spores and monosaccate pollen such as *Potonieisporites*. His succeeding *Microbaculispora* Assemblage was correlated to Besems and Schuurman's Assemblage Group A (Text-Figure 6) and his third assemblage, the *Cycadopites cymbatus* Assemblage to Besems and Schuurman's Assemblage Group B. Based on the data of Besems and Schuurman (1987) which show percentages of approximately 10% Cingulicamerate Group in Assemblage Group B, we correlate that biozone with the 2141A Biozone proposed here. Thus only the lower part of the *Cycadopites cymbatus* Assemblage of Love (1994) correlates with Assemblage Group B. As a whole the *Cycadopites cymbatus* Assemblage correlates with 2141A to C because the eponymous taxon and the Bisaccate Group are common throughout those biozones. Love's *Kingiacolpites subcircularis* Assemblage, which is not dealt with here, is equivalent to zones immediately overlying 2141C.

The field scale biozonation of Stephenson and Osterloff (2002) was primarily aimed at high resolution correlation of the Lower Gharif Member and Rahab Shale. The lowest *Microbaculispora tentula* Biozone correlates with 2141B because it is primarily characterised by high numbers of that taxon (>10% of assemblages). The succeeding *Converrucosisporites* sp. A - *Microbaculispora grandegrnulata* Biozone correlates to the 2141C Biozone because

it is based on the presence of common coarsely ornamented cheilocardioid spores reaching approximately 5% (Text-Figure 6).

OSPZ1 the lowest of the Arabian regional biozones (Stephenson et al. 2003) is characterised by assemblages dominated by *Punctatisporites*, *Retusotriletes* and bilaterally and radially symmetrical monosaccate pollen, which also contain *Anapiculatisporites concinnus*, thus it correlates with 2159A and B biozones. The base of OSPZ2 is defined on the first occurrence of *Microbaculispora tentula* and *Horriditriletes* spp. thus this horizon correlates approximately with the base of 2165A. The upper parts of OSPZ2 are characterised by rising numbers of *C. cymbatus*, *M. tentula* and coarsely ornamented cheilocardioid spores and thus correlate with the interval 2165B to 2141C.

In this paper we correlate with Western Australia primarily because these sequences have been documented in detail including quantitative data, allowing correlation with the present biozones which are defined with quantitative criteria. Also Western Australia has the most comprehensive marine faunal record which has allowed the palynological succession to be correlated with the standard Russian sequence (Archbold 1999; Stephenson 2008).

The palynostratigraphy of Australasian pre-glacial and periglacial sediments is known in less detail than that of later Australasian sequences (Kemp et al. 1977). Amongst published studies, Kemp et al. (1977) and Powis (1984) give generalised details on Australian Stage 1 and 2 biozones (Text-Figure 6). Data from single sequences or small groups of sequences within this interval were supplied by Truswell (1978), Backhouse (1991, 1993) and Jones and Truswell (1992). The unpublished doctoral thesis of Powis (1979) concerning the pre-glacial and periglacial sediments of the Canning Basin, Western Australia provides perhaps the most comprehensive survey of the palynostratigraphy of such sediments. Powis (1984) defined the

base of Stage 2 as the level of the collective first appearance of *Microbaculispora tentula*, *Horriditriletes ramosus* and *Horriditriletes tereteangulatus*, and Jones and Truswell (1992) correlated their *Microbaculispora tentula* Biozone with Stage 2 of Powis (1984). Thus, the base of 2165A correlates with the bases of Stage 2 of Powis (1984) and the *Microbaculispora tentula* Biozone of Jones and Truswell (1992).

Backhouse (1991, 1993) reported that *C. confluens* occurs first toward the top of eastern Australian Stage 2, and this order of first occurrence is also evident in Oman, where *C. confluens* occurs stratigraphically well above the first occurrences of *M. tentula* and *H. ramosus*. Hence, 2165A and B correlate in part with 'Stage 2' sensu Backhouse.

*Converrucosisporites confluens* occurs first between 2165B and 2141B thus the *C. confluens* Biozone of Backhouse (1991) correlates with those south Oman biozones. The presence of large, heavily-ornamented cheilocardioid spores such as *Converrucosisporites* sp. A and *Verrucosisporites* cf. *naumovae* in 2141C may suggest that this biozone correlates with the *Pseudoreticulatispora pseudoreticulata* Biozone of Backhouse (1991).

An advantage of correlation between south Oman biozones and Western Australia is that the calibration by marine fauna available in the latter area can be applied in south Oman.

Backhouse (1991) considered his Stage 2 to be Asselian in age, while the *Pseudoreticulatispora pseudoreticulata* biozone was considered Sakmarian. The *Converrucosisporites confluens* Biozone appears to straddle the boundary between Asselian and Sakmarian (see Backhouse, 1991, fig. 10). By correlation therefore, 2165A and B are likely to be Asselian in age, while 2141A to C are likely Sakmarian. A single faunal calibration is available in Oman, since the Haushi limestone (of the Lower Gharif Member) which overlies the Al Khlata formation, contains definite Sakmarian fusulinids (Angiolini et al. 2006; Text-Figure 6). Age assessments for biozones 2159A and B are less certain since

Western Australian faunal calibration does not reach below Stage 2 of Powis (1984) or Backhouse (1991). However, Jones and Truswell (1992) considered their *Asperispora reticulatispinosus* Biozone to be uppermost Westphalian D (Late Moscovian based upon the assessment of Davydov et al., 2004) to Autunian or Early Asselian in age, and a similar age is therefore adopted for 2159A and B for the present.

## CONCLUSIONS

Palynology is the main method of correlating the subsurface glaciogenic Al Khlata Formation of Oman, due to extreme lateral variability of facies, and poor seismic resolution. Over many years a robust seven-fold in-house PDO palynozonation has been developed and in this paper the formal definitions of the biozones are published, and the biozones are correlated. Ages applied to the upper five biozones are the most precise that can presently be offered, through correlation with recently faunally-dated palynological biozones in Western Australia. The dates thereby gained are consistent with the single faunal tie-point given by fusulinids in the Haushi limestone of the Lower Gharif Member, which shows that the entire Al Khlata Formation is Sakmarian or older. Age assessments for biozones 2159A and B are less certain since Western Australian faunal calibration does not reach below Stage 2 of Western Australia. Further improvements in dating of Al Khlata Formation sediments are most likely to come from palynological correlation with radiometrically- or faunally dated sequences in South America, eastern Australia and southern Africa (see Stephenson, 2008).

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### FIGURE CAPTIONS

Fig. 1. South Oman study area indicating locations of 2159, 2165 and 2141 palynozonal correlation lines.

Fig. 2. South Oman 2159 palynozonal correlation example; (left to right) Marmul-6 well reference section and Qata-9, Al Burj-36 and Rima-65 well sections.

Fig. 3. South Oman 2165 palynozonal correlation example; (left to right) Dimeet-1, Al Burj-23, Amal-9 well sections and Wadi Al Khlata (short) Borehole-5 reference section.

Fig. 4. South Oman 2141 palynozonal correlation example; (left to right) Rahab-28 well section, Rahab-2 reference section, Nimr-46 and Wadi Al Khlata Borehole-2 well sections.

Fig. 5. Haushi Palynozonation scheme displaying palynomorphs groups, key and main accessory taxa.

Fig. 6. Correlation chart with Arabia and Australia.

Table 1. Composition of taxon groups used in palynozonation.

Table 2. Summary of main biozone characteristics.

Plate 1. Representative palynomorphs from the Cingulicamerate, *Horriditriletes*, *Microbaculispora*, *Punctatisporites* groups and other 2159 biozonal taxa from the Al Khlata Formation, South Oman. Magnification X600. All slides are held in the collection of Petroleum Development Oman, Muscat.

1 - 4. *Microbaculispora* Group. 1,2 *Microbaculispora tentula* Habhab-2 1200.41m core, M39/4 and X31/3 ; 3 *Converrucosisporites grandegrnulatus* Habhab-2 1205.42m core, P28/4; 4 *Converrucosisporites confluens* Habhab-2 1205.42m core, C24/3.

5, 8 and 9 Other 2159 Zonal Taxa. 5 *Anapiculatisporites concinnus* Habhab-2 1207.50m core, K21/4; 8 *Wilsonites australiensis*; 1202.70m core, N39/4; 9 *Dibolisporites* sp. 1202.70m core, H25.

6, 7 *Horriditriletes* Group; 6 Habhab-2 1197.60m core, P32 and 7 Nimr-38H1 914.48m core, N30.

10 – 15 Cingulicamerate Group. 10 *Lundbladispota* sp. Wadi Al Khlata Borehole #5 17.70m; 11, 13 and 15 *Cristatisporites* spp. Habhab-2 1207.50m core, B37/4 and 1202.70m core, O17/3, Nimr-38 914.48m core, D29/3 ; 12 *Lundbladispota* sp. Habhab-2 1200.41m core, W34/1 ; 14 *Vallatisporites arcuatus* Nimr-38 914.48m core, P32.

16 – 18        *Punctatisporites* Group. 16, 17 *Calamospora* spp. Habhab-2 1213.52m core, T26/0; 1197.60m core, C44/1. 18 *Punctatisporites gretensis* Habhab-2 1216.50m core, K31/1.

Plate 2. Representative palynomorphs from the Non-Taeniate Bisaccate, Taeniate Bisaccate and Monosaccate Groups, colpate pollen and algal taxa. Magnification X600,

Slide repository as for plate 1.

1 – 6    Colpate pollen taxa: 1 and 2 *Cycadopites cymbatus* Habhab-2 1200.41m, R35/4 and 1200.41m, K46/0; 3 *Striasulcites* sp.; Habhab-2, 4 and 5 *Marsupipollenites* spp. Habhab-2H1 1213.52m , Q26 and Saih Rawl-8 2961.38m, V44/4; 6 *Kingiacolpites subcircularis* Amal South-5 1450.05m, C25/3.

8, 9    Algae: ?*Tetraporina* spp., 8 Habhab-2 1207.50m, E21/3; 9 Wadi Al Khlata Borehole #2, 14.40m M39/4.

7, 10    Non-Taeniate Bisaccate Group: 7 *Alisporites* sp. Habhab-2 1200.41m, X38; 10 *Alisporites indarraensis* Amal-9 1298.45m, K39/1.

11 – 14        Taeniate Bisaccate Group: 11 *Striatoabieites multistratiatus* Rima-69 975m, O38/4; 12 *Vittatina saccata* Habhab-2 1207.50m, Q35/2; 13 *Protohaploxypinus amplus* Habhab-2 1205.42m, B23/3; 14 *Strotersporites* sp. cf. *indicus* Habhab-2 1200.41m, F29/4.

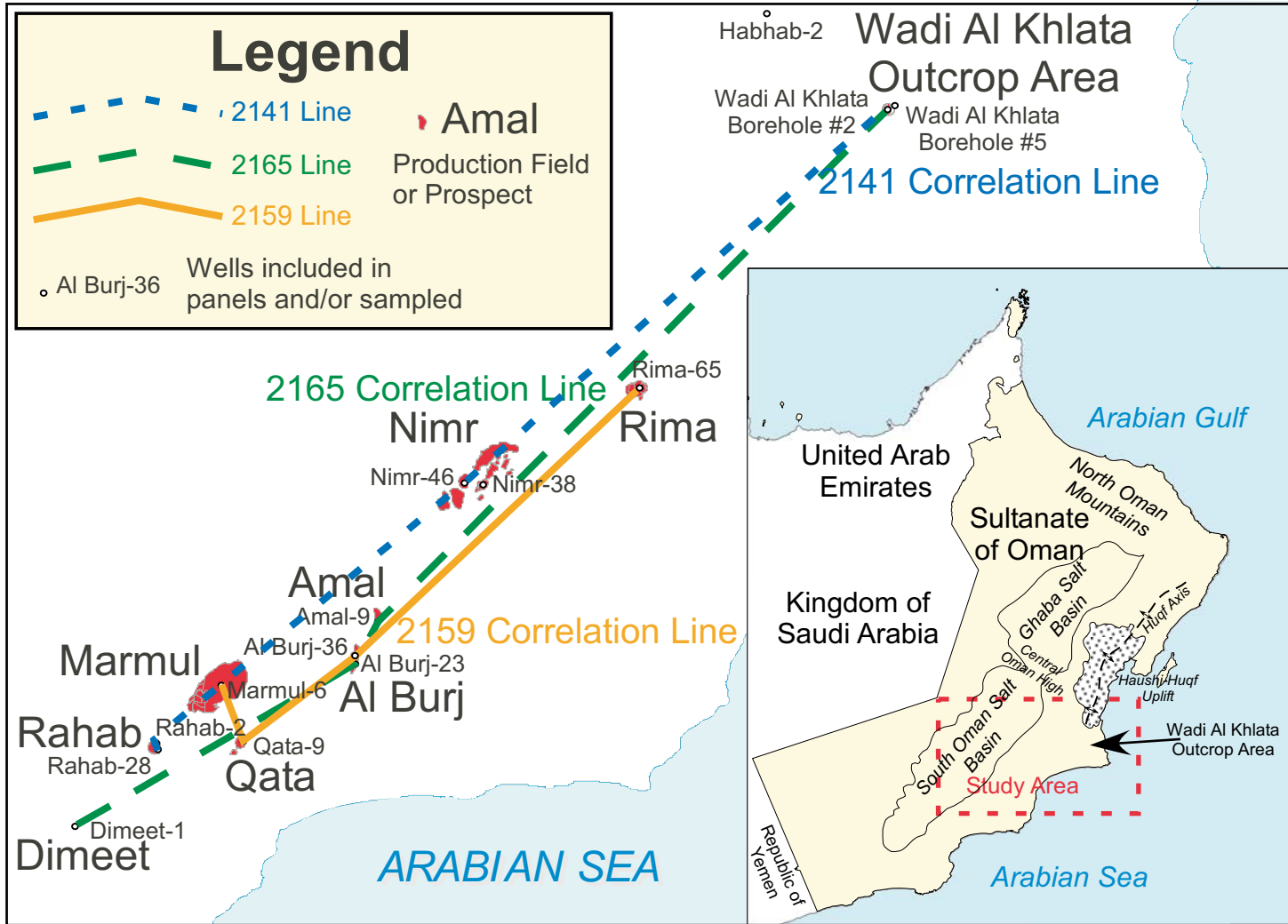
15, 16    Monosaccate Group: 15 *Caheniasaccites* cf. *ovatus* Nimr-38 901.03m, S36/4; 16 *Potonieisporites* sp. Nimr-38 901.03m, D28.

17, 20, 21        Taeniate Bisaccate Group: 17 *Strotersporites indicus* Nimr-39, 914.48m, M24; 20 *Protohaploxypinus* cf. *goraiensis* Nimr-38, 913.68m, O34/2; 21 *Complexisporites polymorphus* Habhab-2 1205.42m, M22/3.

18, 19, 22, 23 Monosaccate Group: 18 ?*Divarisaccus* sp. Habhab-2 1216.50m, C19/2; 19  
*Florinites flaccidus* Habhab-2 1205.42m, R24/2; 22 *Plicatipollenites malabarensis* Habhab-2  
1197.60m, D39/4; 23 *Barakarites* cf. *rotatus* Habhab-2 1218.82, W26/3.

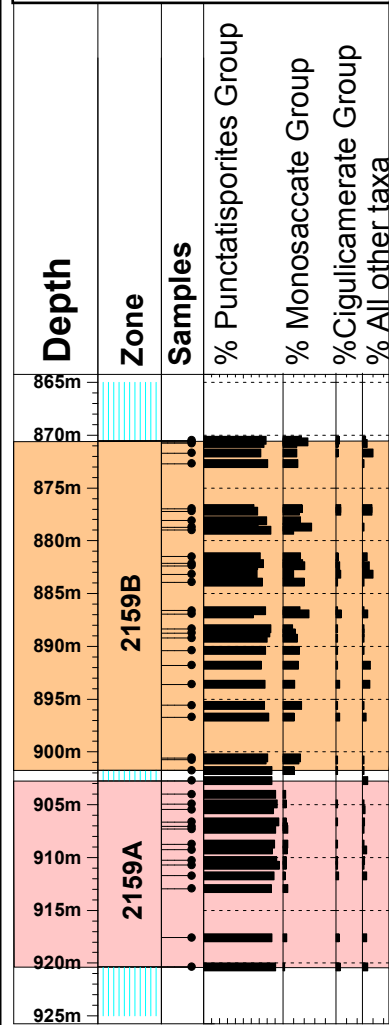
### Legend

- - - 2141 Line
- - - 2165 Line
- - - 2159 Line
- Amal
- Production Field or Prospect
- Wells included in panels and/or sampled

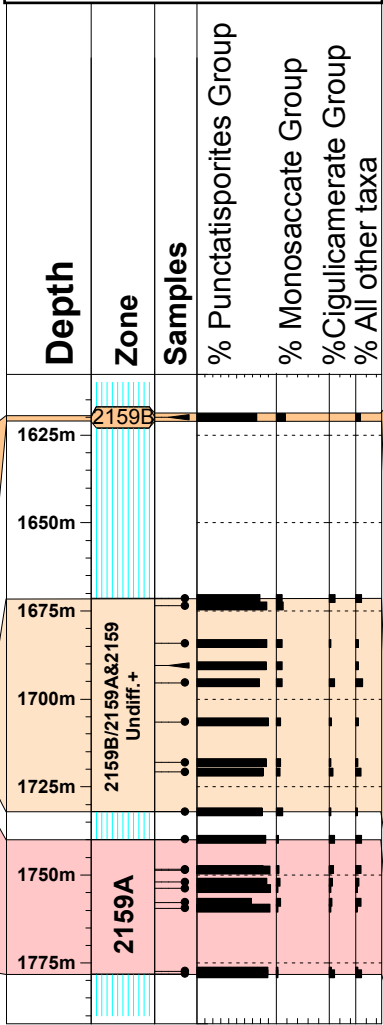




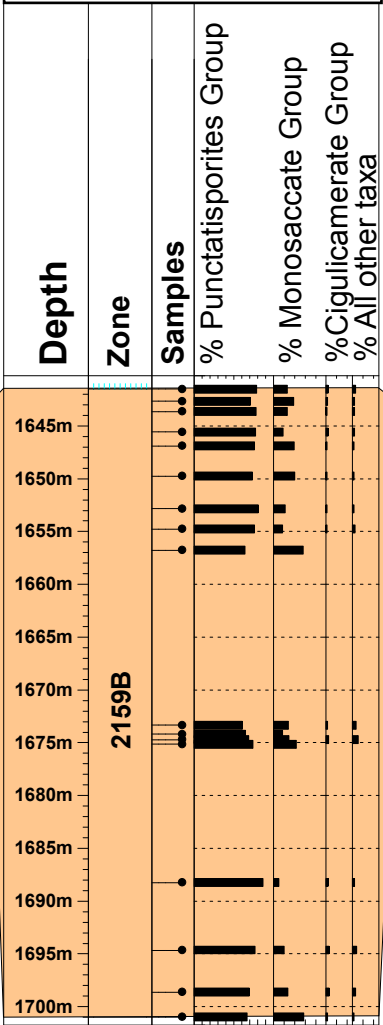
Marmul-6



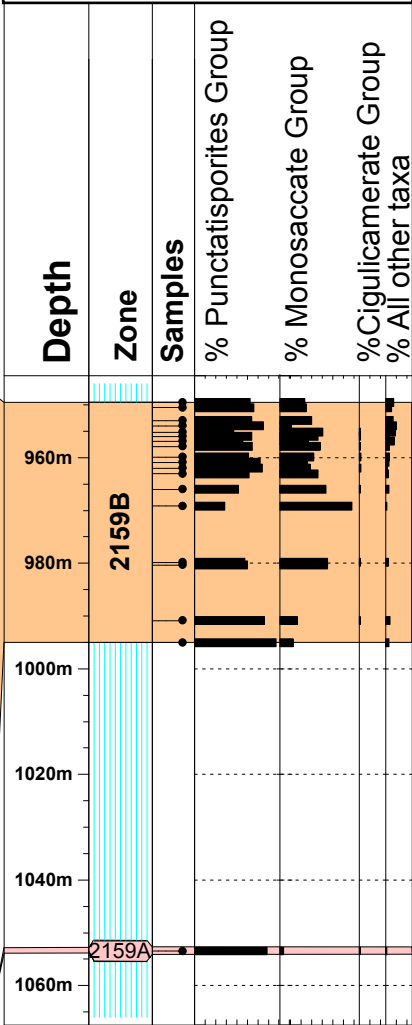
Qata-9



Al Burj-36



Rima-65



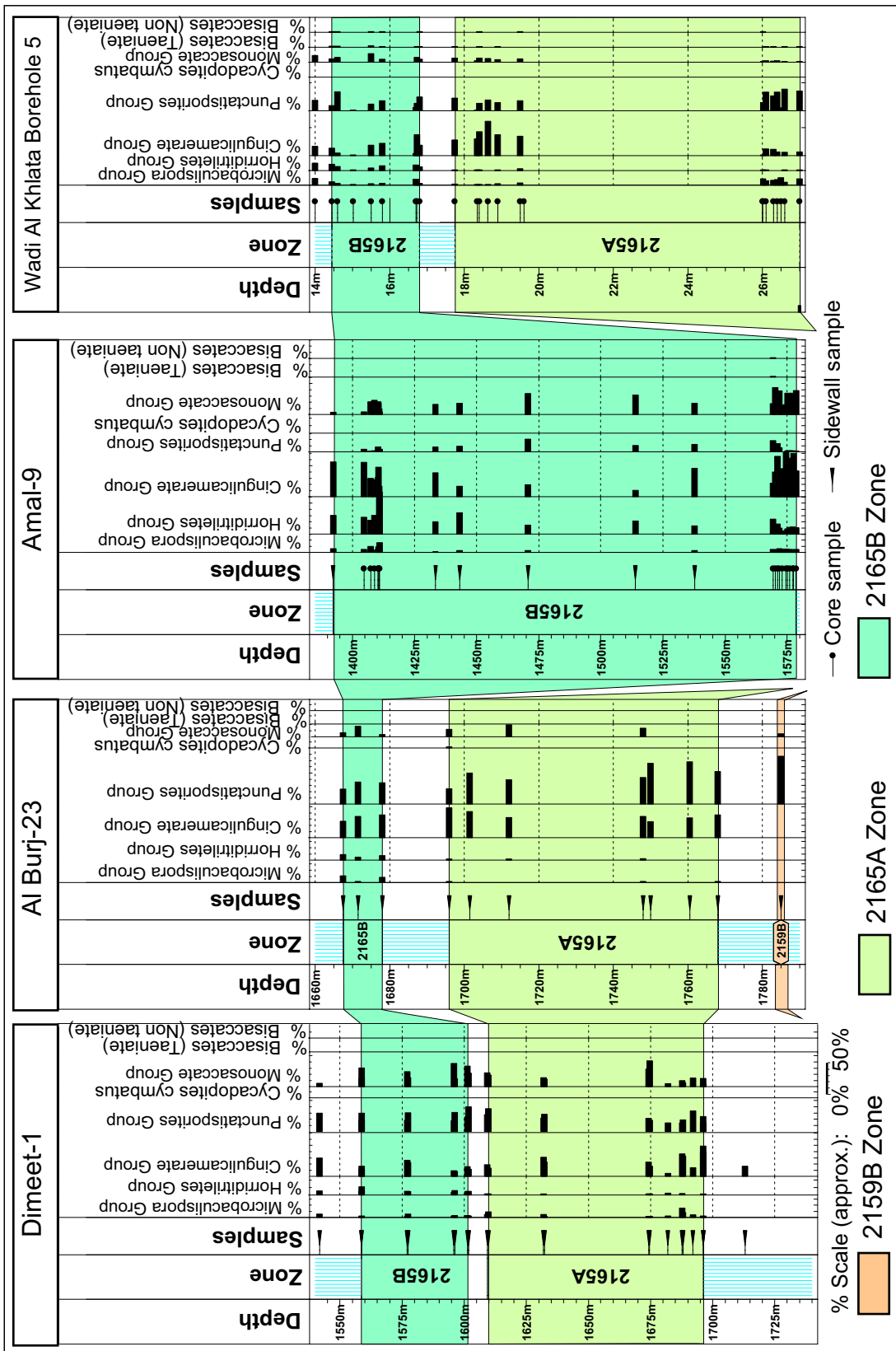
% Scale (approx.): 0% 50%

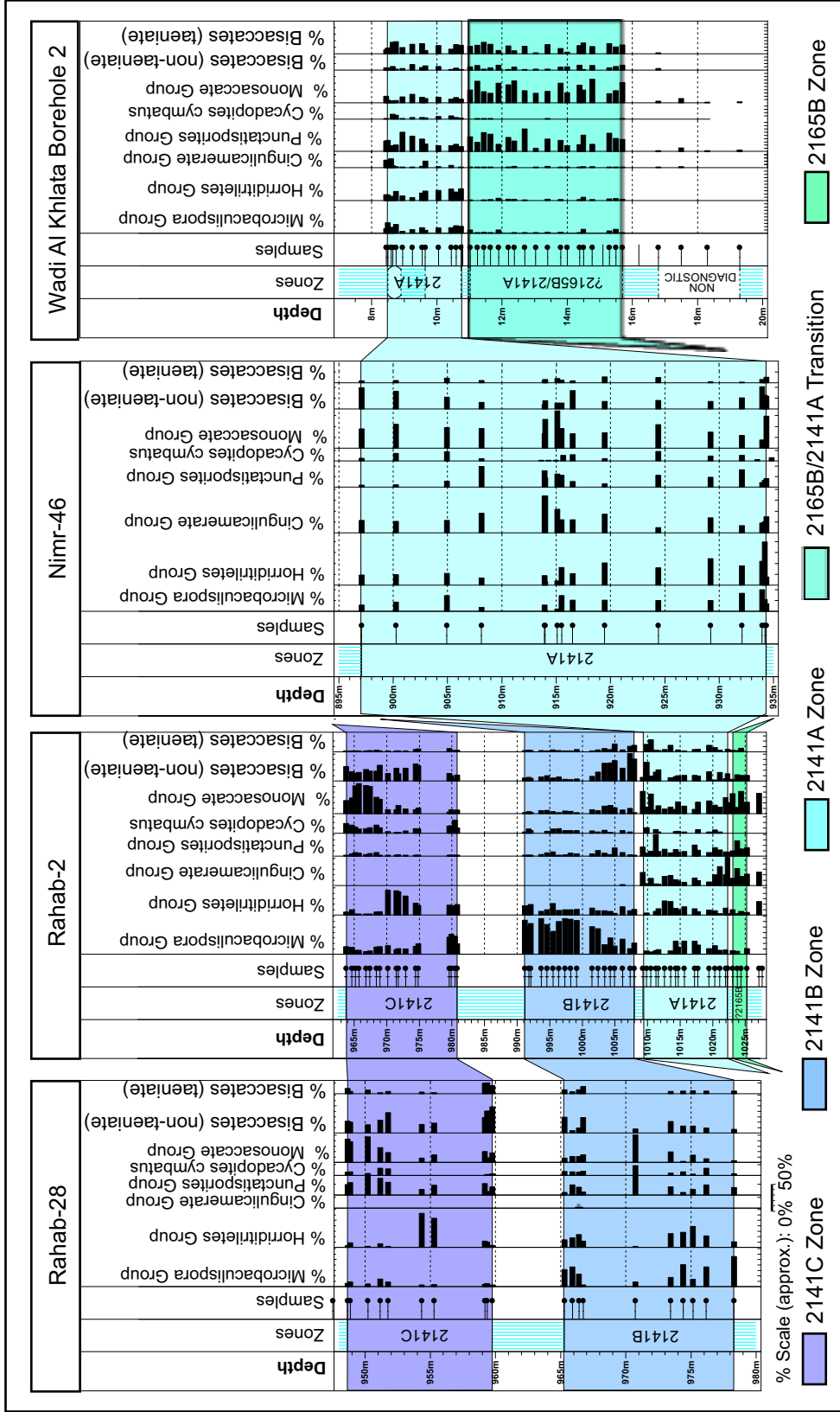
• Core sample — Sidewall sample

2159B Zone

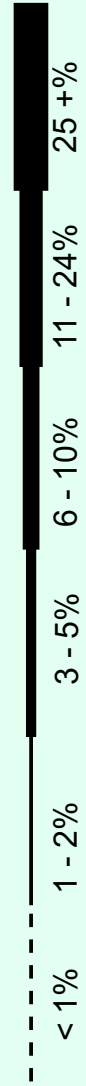
2159A/B Transitional Zone

2159A Zone





# Key



Non-Diagnostic	Palynomorph Zone (Spore/Pollen)						Palynomorph Groups & Key Taxa	Main Accessory Taxa
	2159		2165		2141			
	A	B	A	B	A	B	C	
~	~	~	~	~	~	~	~	<i>Punctatisporites</i> Group
~	~	~	~	~	~	~	~	Monosaccate Group
~	~	~	~	~	~	~	~	Cingulicamerate Group
~	~	~	~	~	~	~	~	<i>Anapiculatisporites concinnus</i>
~	~	~	~	~	~	~	~	<i>Microbaculispora</i> Group
~	~	~	~	~	~	~	~	Bisaccates (non-taeniatae)
~	~	~	~	~	~	~	~	Bisaccates (taeniatae)
~	~	~	~	~	~	~	~	<i>Horriditriletes</i> Group
~	~	~	~	~	~	~	~	<i>Cycadopites cymbatus</i>
~	~	~	~	~	~	~	~	<i>Vittatina</i> Group
~	~	~	~	~	~	~	~	<i>Wilsonites australiensis</i>
~	~	~	~	~	~	~	~	<i>Apiculiretusispora</i> spp.
~	~	~	~	~	~	~	~	<i>Ahrensisporites cristatus</i>
~	~	~	~	~	~	~	~	<i>Aratrisporites saharaensis</i>
~	~	~	~	~	~	~	~	<i>Cyclogranisporites</i> spp.
~	~	~	~	~	~	~	~	<i>Verrucosisporites andersonii</i>
~	~	~	~	~	~	~	~	<i>Brevitriletes</i> spp.
~	~	~	~	~	~	~	~	<i>Densosporites</i> spp.
~	~	~	~	~	~	~	~	<i>Dibolisporites disfacies</i>
~	~	~	~	~	~	~	~	<i>Spelaeotriletes triangularis</i>
~	~	~	~	~	~	~	~	<i>Marsupipollenites</i> spp.
~	~	~	~	~	~	~	~	<i>Convruccosisporites grandegranulatus</i>
~	~	~	~	~	~	~	~	<i>Kingiacolpites subcircularis</i>
~	~	~	~	~	~	~	~	<i>Convruccosisporites confluens</i>
~	~	~	~	~	~	~	~	<i>Deusilites tentus</i>
~	~	~	~	~	~	~	~	<i>Vittatina</i> cf. <i>scutata</i>

Pennsylvanian		Permian		System/stage
?Late Moscovian - Gzhelian	Asselian	Sakmarian		
Stratigraphic range of <i>P. ex gr. karapetovi</i> in Haushi limestone				Arabian faunal content
OSPZ1	Al Khlata Fm.	L. Gharif Mb.	OSPZ2	Stephenson et al., 2003, Arabian Peninsula
			a	
			b	
			c	
			<i>C. pox</i>	<i>K. subcircularis</i> Assemblage
			<i>U. oman,</i>	
			<i>A. indarra.</i>	
			<i>Conv. sp. A</i>	<i>C. cymbatus</i> Assemblage
			<i>M. grande</i>	
			<i>M. tentula</i> ?	<i>Microbaculispora</i> Assemblage
				Ass. Group B
				Assemblage Group A
				2159B
				2159A
				2141C
				2141B
				2141A
				2165B
				2165A
				<i>Microbaculispora tentula</i> Biozone = eastern Australian Stage 2
				<i>V. pseudo.</i>
				Stage 2
				Stage 1
				Stage 2
				Stage 2 <i>sensu</i> Backhouse (1991)
				<i>S. fusus</i>
				<i>P. pseudo.</i>
				<i>C. confluens</i>
				Stage 2 <i>sensu</i> Backhouse (1991)
				Western Australia zones, Backhouse, 1991
				Powis, 1984
				Eastern Australian zones, Jones and Truswell, 1992
				This study
				Oman, Besems and Schuurman, 1987
				Oman, Love, 1994
				Stephenson and Osterloff, 2002, Oman
				Arabian Peninsula
				Arabian faunal content
				System/stage

<b>Group</b>	<b>Main genera</b>	<b>Main species</b>
'Punctatisporites' Group	<i>Punctatisporites</i> , <i>Calamospora</i> , <i>Cyclogranisporites</i> , <i>Retusotriletes</i>	<i>Punctatisporites gretensis</i> , <i>Punctatisporites gretensis</i> forma <i>minor</i> , <i>Punctatisporites lucidulus</i>
'Cingulicamerate' Group	<i>Vallatisporites</i> , <i>Densoisporites</i> , <i>Lundbladispora</i> , <i>Cristatisporites</i> , <i>Indotriletes</i>	<i>Vallatisporites arcuatus</i> , <i>Vallatisporites</i> cf. <i>vallatus</i> , <i>Lundbladispora braziliensis</i> , <i>Densoisporites solidus</i> , <i>Cristatisporites crassilabratus</i> .
'Microbaculispora' Group	<i>Microbaculispora</i> , <i>Converrucosisporites</i> <i>Granulatisporites</i>	<i>Microbaculispora tentula</i> , <i>Converrucosisporites</i> <i>grandegrnulata</i> , <i>Converrucosisporites confluens</i> .
'Horriditriletes' Group	<i>Horriditriletes</i> , <i>Lophotriletes</i>	<i>Horriditriletes ramosus</i> , <i>Horriditriletes tereteangulatus</i> , <i>Horriditriletes uruguayensis</i> , <i>Lophotriletes sparsus</i>
'Monosaccate' Group	<i>Barakarites</i> , <i>Cannanoropollis</i> , <i>Plicatipollenites</i> , <i>Caheniasaccites</i> , <i>Potonieisporites</i>	<i>Barakarites rotatus</i> , <i>Caheniasaccites ovatus</i> , <i>Plicatipollenites malabarensis</i> , <i>Potonieisporites</i> <i>braziliensis</i> , <i>Potonieisporites novicus</i>
Bisaccate (Non taeniate) Group	<i>Alisporites</i> , <i>Limitisporites</i> , <i>Platysaccus</i> , <i>Sulcatisporites</i> , <i>Sahnites</i>	<i>Alisporites indarraensis</i> , <i>Sahnites gondwanensis</i>
Bisaccates (Taeniate) Group	<i>Complexisporites</i> , <i>Protohaploxypinus</i> , <i>Striatoabieites</i> , <i>Striatopodocarpites</i> , <i>Strotersporites</i>	<i>Complexisporites polymorphus</i> , <i>Protohaploxypinus</i> <i>limpidus</i> , <i>Protohaploxypinus amplius</i> , <i>Striatoabieites</i> <i>multistriatus</i> , <i>Striatopodocarpites cancellatus</i> , <i>Strotersporites</i> cf. <i>indicus</i> .
'Vittatina' Group	<i>Circumstriatites</i> , <i>Marsupipollenites</i> , <i>Pakhapites</i> , <i>Vittatina</i> , <i>Striasulcites</i>	<i>Vittatina subsaccata</i> , <i>Vittatina</i> cf. <i>scutata</i> , <i>Circumstriatites talchirensis</i> , <i>Marsupipollenites</i> <i>striatus</i> , <i>Pakhapites fusus</i> , <i>Striasulcites tectus</i>

Biozone	Main characteristics
2159A	Abundance of <i>Punctatisporites</i> Group which constitutes up to 100% of assemblages. Taxa of the Monosaccate Group constitute less than 5% of assemblages.
2159B	Increase in the proportion of the Monosaccate Group to greater 10% of assemblages such that <i>Punctatisporites</i> spp. does not exceed 90% of assemblages.
2165A	(1) the Cingulicamerate Group constitutes up to approximately 30% of the assemblages; and (2) the <i>Microbaculispora</i> Group constitutes approximately 5% of assemblages.
2165B	(1) <i>Horriditriletes</i> Group constitutes between 5 and 10%, or up to 20% of assemblages; and (2) <i>Microbaculispora</i> Group constitutes usually around 3 to 5% of assemblages.
2141A	(1) non-taeniate and taeniate bisaccate groups together represent more than 10% of the assemblages; and (2) <i>Cycadopytes cymbatus</i> is rare towards the base but increases to 5 to 10% of assemblages towards the top of the biozone.
2141B	(1) The <i>Microbaculispora</i> Group represents greater than 10% and can constitute up to 60% of assemblages, in addition the <i>Microbaculispora</i> Group is more abundant than the <i>Horriditriletes</i> Group; (2) <i>Cycadopytes cymbatus</i> makes up 5 to 10% of assemblages; and (3) the Cingulicamerate Group are extremely rare or absent.
2141C	The <i>Microbaculispora</i> Group represents more than 10% of assemblages, but the <i>Horriditriletes</i> Group exceeds it in numbers, reaching 30% or more.

## APPENDIX

The following is a complete listing of all taxa mentioned in the text of this paper including their full author citations.

*Ahrensia* *crustatus* Playford & Powis 1979  
*Alisporites indarraensis* Segroves 1970  
*Anapiculatisporites concinnus* Playford 1962  
*Apiculiretusispora* spp.  
*Aratrisporites saharaensis* Loboziak et al., 1986  
*Barakarites rotatus* (Balme & Hennelly) Bharadwaj & Tiwari 1964  
*Brevitriletes cornutus* (Balme & Hennelly) Backhouse 1991  
*Caheniasaccites ovatus* Bose & Kar 1966  
*Circumstriatites talchirensis* Lele & Makada 1972  
*Converrucosisporites confluens* (Archangelsky & Gamero) Playford & Dino 2002  
*Converrucosisporites grandegrulatus* (Anderson) Lindström 1995  
*Cristatisporites crassilabrus* Archangelsky & Gamero 1979.  
*Cycadopites cymbatus* (Balme & Hennelly) Segroves 1970  
*Densoisporites rotundidentatus* Segroves 1970  
*Densoisporites solidus* Segroves 1970  
*Densoisporites* sp. Jones & Truswell 1992  
*Deusilites tentus* Hemer & Nygreen 1967  
*Dibolisporites disfacies* Jones & Truswell 1992  
*Florinites flaccidus* Menéndez & Azcuy 1973  
*Horriditriletes ramosus* (Balme & Hennelly) Bharadwaj & Salujah 1964  
*Horriditriletes tereteangulatus* (Balme & Hennelly) Backhouse 1991  
*Horriditriletes uruguaiensis* (Marques-Toigo) Archangelsky & Gamero 1979  
*Kingiacolpites subcircularis* Tiwari & Moiz 1971  
*Lophotriletes sparsus* Singh 1964  
*Lundbladispora* spp.  
*Lundbladispora braziliensis* (Pant & Srivastava) Marques Toigo & Pons 1976  
*Marsupipollenites striatus* (Balme & Hennelly) Foster 1975  
*Microbaculispora tentula* Tiwari 1965  
*Pakhapites fusus* (Bose & Kar) Menéndez 1971  
*Plicatipollenites malabarensis* (Potonié & Sah) Foster 1975  
*Potonieisporites brasiliensis* (Nahuys et al.) Archangelsky & Gamero 1979  
*Potonisporites novicus* Bharadwaj 1954.  
*Protohaploxypinus amplus* (Balme & Hennelly) Hart 1964  
*Protohaploxypinus* cf. *goraiensis* (Potonié & Lele) Hart 1964  
*Punctatisporites gretensis* Balme & Hennelly 1956  
*Punctatisporites gretensis* forma *minor*, (Balme & Hennelly) Hart 1965  
*Punctatisporites lucidulus* Playford & Helby 1966  
*Sahnites gondwanensis* (Mehta) Pant 1955  
*Spelaotriletes triangulus* Neves & Owens 1966  
*Striatoabieites multistriatus* (Balme & Hennelly) Hart 1964  
*Striasulcites tectus* Venkatachala & Kar 1968  
*Strotersporites* cf. *indicus* Tiwari 1965  
*Vallatisporites arcuatus* (Marques-Toigo) Archangelsky & Gamero 1979  
*Vallatisporites* cf. *vallatus* Hacquebard 1957  
*Verrucosisporites andersonii* Backhouse 1988



*Verrucosiporites* cf. *naumovae* Hart 1963  
*Vittatina* cf. *scutata* (Balme & Hennelly) Bharadwaj 1962  
*Vittatina subsaccata* Samoilovich 1953  
*Wilsonites australiensis* Playford & Helby 1968

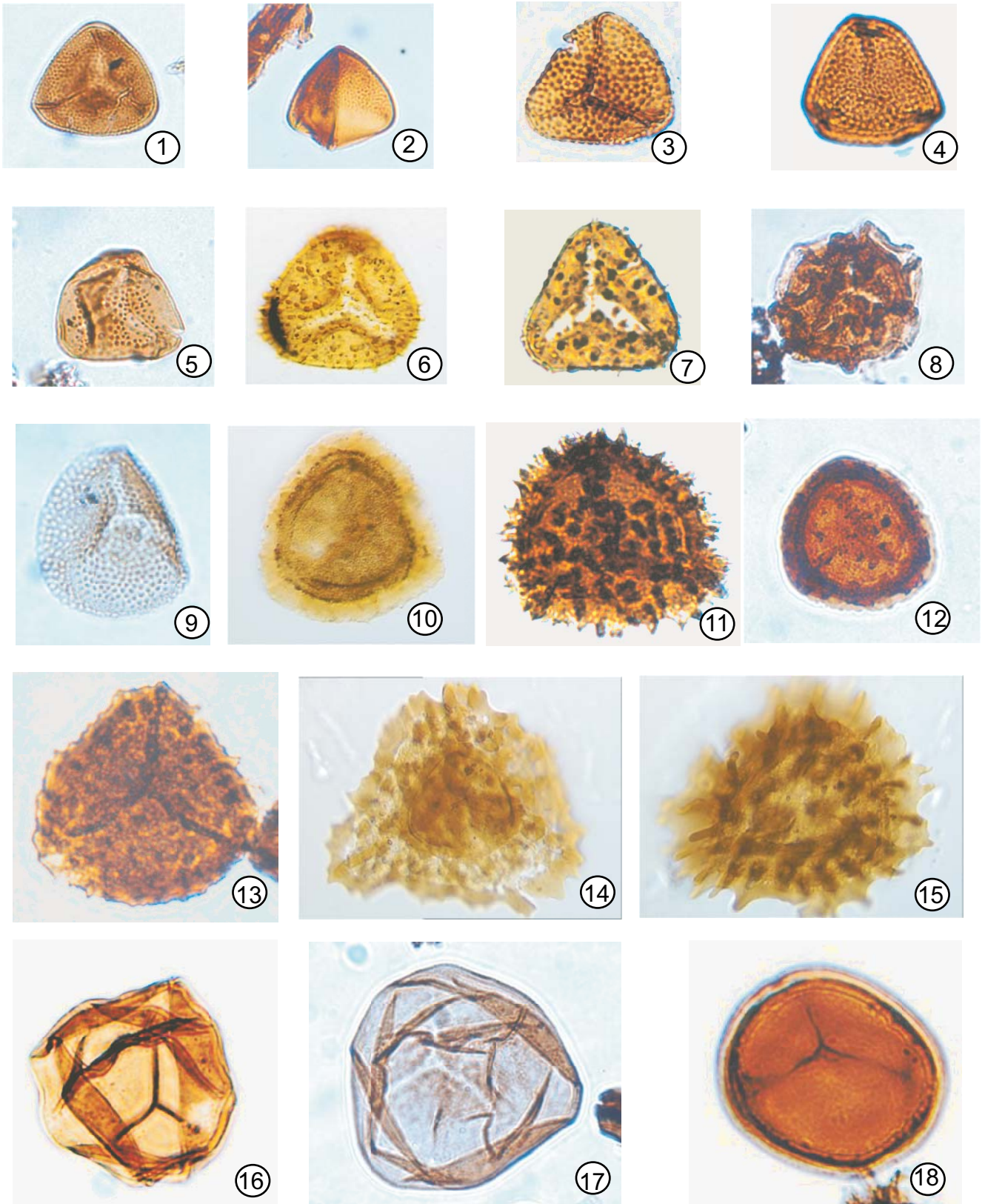


PLATE 1; Penney, Stephenson and Al Barram

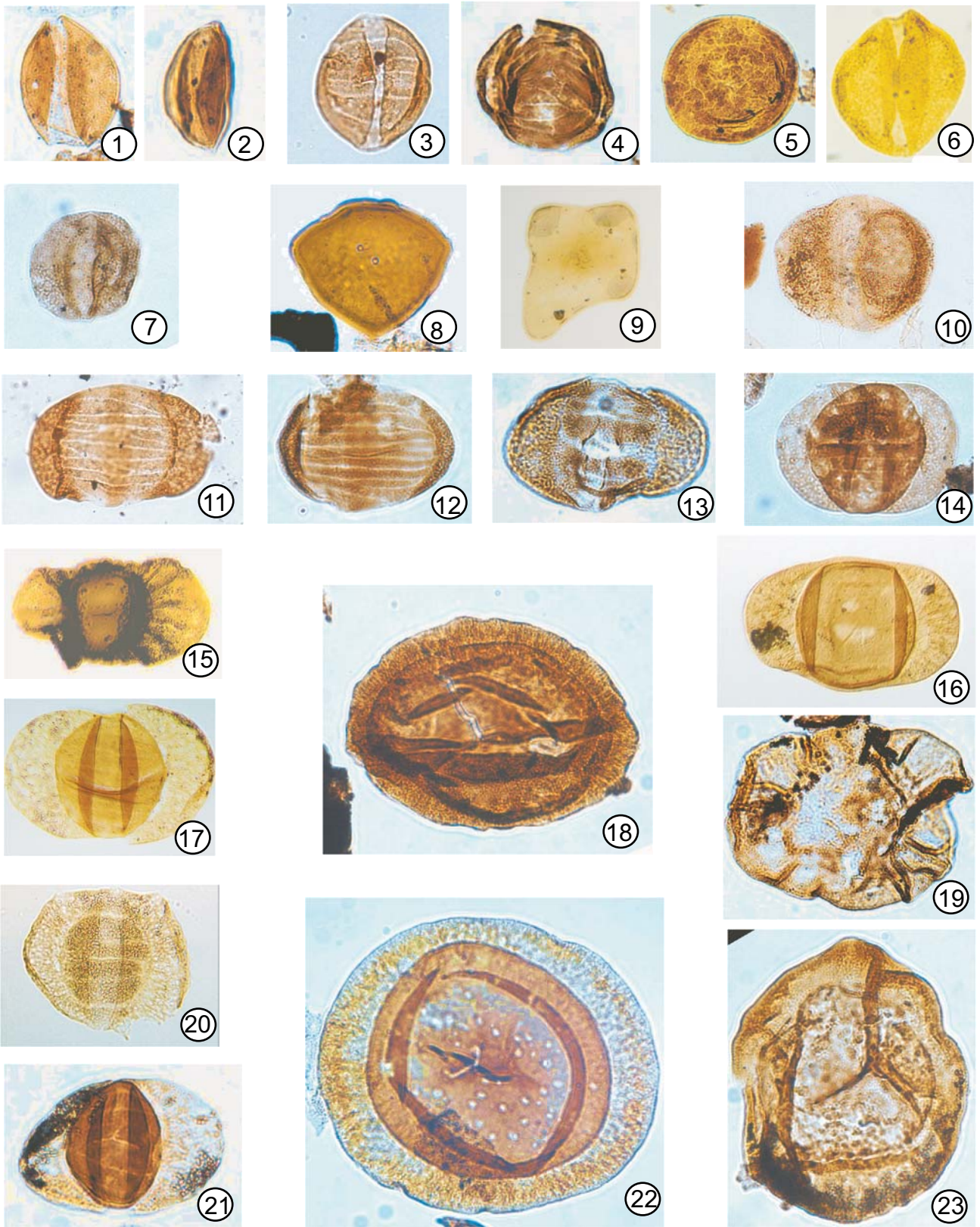


PLATE 2; Penney, Stephenson and Al Barram