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Contact UKCEH NORA team at
noraceh@ceh.ac.uk

Title: Mechanistic effect modeling of earthworms in the context of pesticide risk assessment: Synthesis of the FORESEE Workshop.

Authors: *Forbes Valery E^a, Agatz Annika^b, *Ashauer Roman^{c,d}, Butt Kevin R^e, *Capowiez Yvan^f, Duquesne Sabine^g, *Ernst Gregor^h, Focks Andreasⁱ, *Gergs Andre^h, Hodson Mark E^j, Holmstrup Martin^k, Johnston Alice SA^l, *Meli Mattia^m, Nickisch Dirkⁿ, Pieper Silvia^g, *Rakel Kim J^o, Reed Melissa^p, Roembke Joerg^q, *Schäfer Ralf B^r, *Thorbek Pernille^s, Spurgeon, David J^t, Van den Berg Erikⁱ, Van Gestel Cornelis AM^u, Zorn Mathilde I^v, *Roeben Vanessa^o

Affiliations:

^aDepartment of Ecology, Evolution and Behavior, University of Minnesota, St. Paul, MN, 55108, USA

^bibacon GmbH, Roßdorf, Germany

^cSyngenta Crop Protection AG, Rosentalstrasse 67, Basel, CH-4002, Switzerland

^dDepartment of Environment and Geography, University of York, Wentworth Way, Heslington, York, YO10 5NG, UK

^eSchool of Forensic and Applied Sciences, University of Central Lancashire, Preston, PR1 2HE, UK

^fINRAE, UMR 1114 EMMAH, INRA/université d'Avignon, Domaine Sait paul, Agroparc, 84914 Avignon, France

^gUBA Umweltbundesamt, FGIV- 1.3, Section "Plant Protection Products", Wörlitzer Platz 1, 06844 Dessau, Germany

^hBayer AG, CropScience Division, Alfred-Nobel-Straße 50, 40789 Monheim, German

ⁱWageningen Environmental Research, P.O. Box 47, 6700 AA Wageningen, The Netherlands

^jDepartment of Environment and Geography, University of York, Heslington, York, YO10 5NG, UK

^kDepartment of Bioscience, Aarhus University, Vejlsovej 25, DK-8600 Silkeborg, Denmark

^lSchool of Biological Sciences, University of Reading, Reading, RG6 6AH, UK

^mAdama Agricultural Solutions Ltd, 33 rue de Verdun, 92156 Suresnes Cedex, France

ⁿRIFCON GmbH, Goldbeckstraße 13, 69493 Hirschberg, Germany

^ogaiac, Kackertstr. 10, 52074 Aachen, Germany

^pHealth and Safety Executive, Mallard House, Kings Poll, York, UK

^qECT Oekotoxikologie GmbH, Boettgerstr. 2-14, D-65439 Floersheim, Germany

^rInstitute for Environmental Sciences, University of Koblenz and Landau, Landau, Germany

^sBASF SE, APD/EE, APD/EE, Speyerer Strasse 2, 67117 Limburgerhof, Germany

^tUK Centre for Ecology and Hydrology, MacLean Buidling, Wallingford, Oxon, OX10 8BB, UK

^uDepartment of Ecological Science, Faculty of Science, Vrije Universiteit Amsterdam, De Boelelaan 1085, 1081 HV Amsterdam, The Netherlands

^vCTGB, Bennekomseweg 41, 6717 LL Ede, The Netherlands

*Indicates member of the Organizing Committee.

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1 **Abstract:** Earthworms are important ecosystem engineers, and assessment of the risk of plant
2 protection products towards them is part of the European environmental risk assessment (ERA). In the
3 current ERA scheme, exposure and effects are represented simplistically and are not well integrated,
4 resulting in uncertainty when applying the results to ecosystems. Modeling offers a powerful tool to
5 integrate the effects observed in lower tier laboratory studies with the environmental conditions under
6 which exposure is expected in the field. This paper provides a summary of the FORESEE Workshop
7 ((In)Field Organism Risk modeling by coupling Soil Exposure and Effect) held January 28-30, 2020
8 in Düsseldorf, Germany. This workshop focussed on toxicokinetic-toxicodynamic (TKTD) and
9 population modeling of earthworms in the context of environmental risk assessment. The goal was to
10 bring together scientists from different stakeholder groups to discuss the current state of soil
11 invertebrate modeling, explore how earthworm modeling could be applied to risk assessments, and in
12 particular how the different model outputs can be used in the tiered ERA approach. In support of these
13 goals, the workshop aimed at addressing the requirements and concerns of the different stakeholder
14 groups to support further model development. The modeling approach included four submodules to
15 cover the most relevant processes for earthworm risk assessment: Environment, Behavior (feeding,
16 vertical movement), TKTD, and Population. Four workgroups examined different aspects of the model
17 with relevance for: Risk assessment, earthworm ecology, uptake routes, and cross-species
18 extrapolation and model testing. Here, we present the perspectives of each workgroup and highlight
19 how the collaborative effort of participants from multidisciplinary backgrounds helped to establish
20 common ground. In addition, we provide a list of recommendations for how earthworm TKTD
21 modeling could address some of the uncertainties in current risk assessments for plant protection
22 products.

23

24 **Key Words:** cross-species extrapolation; plant protection products; population modeling; soil
25 organisms; uptake routes

26

27 **Background**

28 Earthworms are important ecosystem engineers that increase soil fertility and provide a wide
29 range of ecosystem services (Blouin et al. 2013). They are included in the safety assessment of
30 pesticides in the European Union (EU), which is prescribed by European legislation (Regulation (EC)
31 No. 1107/2009 and guidance document SANCO/10329/2002). In the EU, pesticides can only be
32 authorized if no unacceptable effects on non-target organisms, biodiversity, or the ecosystem will
33 occur. In the risk assessment procedures, testing representative species of earthworms and assessing
34 the risks to this group is deemed to cover soil-inhabiting Oligochaeta, belonging to the families
35 Lumbricidae (earthworms) and Enchytraeidae (potworms). Species from both groups are used in
36 standard ecotoxicological tests, but only tests with lumbricid earthworms need to be submitted
37 according to EU data requirements for pesticides (283/2013 and 284/213). The Annex to the data
38 requirement mentions as relevant OECD guidelines for testing the genus *Eisenia* (e.g., *E. fetida*, *E.*
39 *andrei*).

40 In 2017, the European Food Safety Authority (EFSA) requested an opinion from the Panel on
41 Plant Protection Products and their Residues (PPR) on the science behind the risk assessment of plant
42 protection products for in-soil organisms in preparation for a guidance update (EFSA PPR 2017). In
43 this opinion, a review was presented on the current risk assessment scheme, and proposals were made
44 for further progress. The use of mechanistic effect models was suggested in this opinion, for example,
45 models clarifying the relationships between internal concentrations and toxicological effects over time
46 for endogeic earthworms. However, gaps in the currently available models were identified as well as a
47 need for research on their applicability domains in soil risk assessment. Therefore, a workshop was
48 organized to clarify these issues. This synthesis provides a summary of the workshop findings and
49 recommendations.

50 The current soil risk assessment follows a tiered approach starting with simple assumptions
51 with effects characterized in standardized laboratory studies. In addition, whereas the exposure
52 assessment can take into account spatiotemporal variability of pesticides in soil, abiotic parameters
53 (such as soil temperature and moisture), and soil composition, the effect assessment for earthworms is
54 based on the outcome of a reproduction test that does not take such factors into account. In the first

55 tier, a chronic earthworm study (GD 222, OECD 2017) is required, which has the aim of assessing the
56 intrinsic toxicity of the tested substance. Exposure is assumed to vary in time, since the pesticides
57 interact with the soil and degrade, and hence a constant exposure cannot necessarily be maintained. In
58 this type of study, adult *E. fetida* are exposed to a series of pesticide concentrations, and the relevant
59 endpoints are assessed only once during the study period, i.e., after 28 days for survival and growth,
60 and after 56 days for the number of juveniles. Therefore, only a limited mechanistic understanding of
61 the underlying effect is provided. The results of this test, expressed in terms of a No Observed Effect
62 Concentration (NOEC) for mortality, reproduction, and growth or a ten percent Effect Concentration
63 (LC₁₀/EC₁₀) for mortality and reproduction is then related to a worst-case Predicted Environmental
64 Concentration (PEC) for soil to obtain a Tier 1 estimate of risk. Should the ratio of toxicity to
65 exposure (TER) be below a defined trigger value (currently 5 for chronic risk assessment in the EU;
66 Regulation (EU) 546/2011, 2011), a risk is indicated, and a higher tier assessment (intended to include
67 more realism in exposure and/or effects; Solomon et al. 2008), such as a field study with earthworms
68 (ISO 11268-3) can be performed to refine the risk.

69 The limited conceptual integration of exposure and effect assessments in Tier 1 soil risk
70 assessments leads to uncertainty when extrapolating the results to different ecosystems. Furthermore,
71 there is a large gap between a simple Tier 1 laboratory study and a full field study, which suggests the
72 need for intermediate tiers between these options. Key uncertainties that could be reduced through the
73 use of intermediate tier assessments include addressing actual exposure profiles arising from
74 earthworm vertical movement and spatiotemporal variability in pesticide concentration, and
75 extrapolation of the results from field studies to other environmental and/or agricultural situations
76 beyond actual conditions in the field study. A critical source of uncertainty is the possible difference in
77 sensitivity between species tested in the laboratory and species found in the field. The low field
78 relevance of *Eisenia* species has been recognized, and suggestions to use *Aporrectodea caliginosa* as
79 an additional test species are underway (e.g., Bart et al. 2018). Decreasing the uncertainties in the risk
80 assessment requires a better understanding of the risks of pesticides to different species of earthworms,
81 including whether there is any relationship between quantifiable traits and toxicological sensitivity.

82 Mechanistic effect modeling offers a potentially powerful tool to integrate pesticide exposure
83 and effects and extrapolate results observed in lower-tier laboratory studies to exposure scenarios that
84 are expected in the field (EFSA PPR 2017). In particular, individual-based models (IBMs) are of
85 interest, as they allow for a high degree of realism, can help to quantify uncertainties, and can integrate
86 processes that occur across multiple scales (DeAngelis and Grimm, 2014). Populations are represented
87 as consisting of discrete individuals, and population-level behavior and effects emerge from
88 interactions of the individuals with each other and with their environment (DeAngelis and Grimm,
89 2014). Johnston et al. (2014a, 2014b, 2015, 2018) developed several earthworm IBMs that incorporate
90 realistic earthworm behavior, address spatiotemporal variability in pesticide exposure, and integrate
91 exposure and effects using an energy budget approach. Given the growing recognition of the power of
92 mechanistic effect models for use in environmental risk assessment (ERA) (Hommen et al., 2016) and
93 recent regulatory guidance on their development, testing, and documentation (EFSA PPR 2014, 2017),
94 there are clear opportunities to address specific issues identified in ERA approaches by developing
95 dedicated models aiding at appropriate tiers of the ERA process. With that said, it is clearly
96 impractical to develop and apply unique effect models and related behavior and exposure scenarios for
97 every single species. Instead, an approach in which selected earthworm species can represent broader
98 ecological groups of earthworms is needed. In addition, acceptance of models and exposure scenarios
99 for ERA will be facilitated through consistent and transparent procedures for the development and use
100 of effect models, species behavior, and exposure scenarios. The acceptance can be further promoted by
101 evaluation and documentation and through broad stakeholder buy-in (Forbes et al. 2019).

102

103 **Objective and Rationale**

104 The FORESEE ((In)Field Organism Risk modeling by coupling Soil Exposure and Effect)
105 Workshop was held January 28 – 30, 2020, in Düsseldorf, Germany. The overall focus of the
106 workshop was to bring together scientists from different stakeholder groups (i.e., regulatory
107 authorities, industry, and contract research organizations (CROs)) and academic scientists to discuss
108 the current state of earthworm modeling. The workshop aimed to identify research gaps and explore

109 how mechanistic effect modeling of earthworms could be applied to soil organism risk assessments. In
110 particular, we considered how the different model outputs could be used in the regulatory framework
111 and in the tiered approach prescribed by the recent EFSA Scientific Opinion addressing the state of the
112 science on risk assessment of plant protection products for in-soil organisms (EFSA PPR 2017). In
113 support of these goals, the workshop aimed to discuss a common modeling framework for earthworms
114 and address the requirements and concerns of the involved stakeholder groups at an early stage of
115 model development.

116 Johnston et al. (2014a, 2014b, 2015, 2018) developed and validated a suite of earthworm
117 models that integrate exposure, effects, energy budgets, behavior (movement), and life cycles. Based
118 on these models, Roeben et al. (2020) initiated the development of a modular framework for
119 earthworm modeling (FORESEE) that aims to cover most of the relevant earthworm ecological
120 categories (i.e., ecotypes). FORESEE is mechanistic and aims to provide spatiotemporal realism in
121 earthworm behavior, as well as exposure and effects of pesticides. The workshop was based on the
122 FORESEE modeling approach containing four submodules to cover relevant aspects of earthworm
123 modeling: Environment, Behavior (Feeding and Movement), Toxicokinetics/Toxicodynamics
124 (TKTD), and Population Dynamics. In practical terms, the Environment module is linked to an IBM
125 containing movement, TKTD, and population submodels from which earthworm population dynamics
126 emerge. The Environment module utilizes outputs from pesticide exposure models (e.g., PEARL,
127 PELMO, HYDRUS), providing spatially and temporally explicit information on soil moisture,
128 temperature, organic matter content, bulk density, and total and porewater pesticide concentrations.
129 The behavior module simulates the feeding and vertical movement of different species representing
130 four major ecological categories of earthworms using a trait-based approach. The TKTD module
131 covers the toxicity of pesticides to earthworms using the General Unified Threshold model of Survival
132 (GUTS, Jager et al. 2011; Jager and Ashauer 2018) for lethal effects and Dynamic Energy Budget
133 ((DEB)-TKTD, Jager et al. 2006; Jager 2019) models for sublethal effects. The population module
134 incorporates existing population models of different species (e.g., Johnston et al. 2014a, 2014b, 2015,
135 2018). Before use in regulatory risk assessments, all modules should be evaluated independently and
136 be designed to allow for updating when additional knowledge becomes available (EFSA 2014).

137 Preferably, the evaluation, release, and version control of the effect model versions could take place
138 within the already existing framework for the version control of pesticide fate models, i.e., the EFSA
139 Chaired FOCUS Version Control Group.

140 During the workshop, participants from academia, regulatory authorities, CROs, and industry
141 were divided into four workgroups. These workgroups examined different parts of FORESEE and
142 addressed various questions relevant to earthworm mechanistic effect modeling: risk assessment,
143 earthworm ecology, uptake routes, extrapolation and testing against experimental datasets, and
144 ecotoxicological study needs and data gaps.

145 Workgroup 1 focused on how model outputs could fit into future risk assessment procedures
146 for earthworms. Participants discussed how the ecotoxicity assessment and fate inputs fit into the
147 modeling approach. Furthermore, they explored how FORESEE outputs could be used to refine the
148 risk assessment of earthworms in different ways and how the modeling approach fits into the tiered
149 ERA employed under EU regulatory requirements.

150 Workgroup 2 focused on earthworm ecology. The group discussed the main factors governing
151 the behavior of important ecological groups of earthworms in arable soils and whether their movement
152 could be described by a set of behavioral traits. Furthermore, the workgroup looked at other traits,
153 such as reproduction, vertical distribution, and feeding type, and how such traits likely influence the
154 movement of earthworms. The modelers confronted model assumptions with knowledge on
155 earthworm ecology provided by the rest of the group, and in this way, tested whether the model is
156 sufficiently realistic while being generally applicable to different earthworm species.

157 Workgroup 3 focused on exposure of earthworms, uptake routes, and TKTD modeling.
158 Participants discussed relevant pesticide exposure routes (dermal vs. oral) and concentrations
159 (porewater vs. total soil or litter concentrations) of pesticides for earthworms. Their discussions
160 included the influence on exposure of different soil properties (e.g., organic matter), model calibration
161 with laboratory toxicity tests, and multiple pesticide applications.

162 Workgroup 4 focused on species extrapolation and testing to increase the validation status of
163 the model. The species typically used in laboratory experiments to evaluate pesticide risks often differ
164 from those characteristic of relevant field habitats. Therefore, species extrapolation and model testing
165 to increase the validation status were considered within the same workgroup. The workgroup
166 discussed models to extrapolate ecotoxicological sensitivity across species and how to address data
167 gaps. In addition, issues related to data availability and requirements for model evaluation were
168 discussed.

169 In this workshop synthesis, we present the perspectives of each workgroup and highlight how
170 the collaborative effort involving multiple stakeholders and representing a diversity of scientific
171 expertise was able to reach consensus on a suite of recommendations and priorities for future work to
172 develop FORESEE into an implementable tool for pesticide risk assessment in the EU.

173

174 **Key Findings**

175 *Risk Assessment (WG 1)*

176 The current risk assessment scheme (SANCO/10329/2002) has a gap between the risk
177 assessment tiers. There are currently only a few intermediate refinements of risk (e.g., laboratory tests
178 using natural soils or additional test species) between the Tier 1 risk assessment using the chronic
179 laboratory study and the higher tier assessment based on a field study. We identified several levels of
180 the risk assessment in which modeling tools can be used. In the lower tier risk assessment, a model
181 could be used to understand the impact of soil properties and bioavailability on the toxicity to soil
182 organisms. Likewise, a model combining the realistic movement of earthworms (e.g., in relation to
183 soil moisture or food availability) with a spatiotemporal exposure profile could help to generate
184 refined exposure endpoints. Those endpoints could be used to calculate a refined TER, based on the
185 simulated movement and resulting exposure. At the next tier, a potential advance would be to combine
186 this spatiotemporal exposure pattern with TKTD modeling following the principles outlined in
187 EFSA's scientific Opinion on TKTD modeling (EFSA 2018) to predict risk at the level of individuals.

188 Another use of the models could be to compare the Tier 1 assessment with the field study to check on
189 the degree of conservatism of the Tier 1 assessment. However, the current standard chronic earthworm
190 laboratory study (OECD 2017) is mainly suitable for setting NOECs and/or EC₁₀s but is not adequate
191 for parameterizing effect models. The standard study does not provide information on the time course
192 of effects and cannot differentiate between reproductive effects and mortality of newly hatched
193 juveniles. An option could be a modified test to allow counting of cocoons and assessment of hatching
194 rate, in addition to the direct measurement of juvenile production. Moreover, the results of the Tier 1
195 laboratory chronic test are based on nominal pesticide concentrations, as there is no requirement to
196 measure soil or tissue concentrations. So for modeling to be used to refine risk assessments, new study
197 designs that increase the number of recorded parameters are needed.

198 Furthermore, environmental conditions and farming practices vary across regions and crops.
199 Modeling could facilitate the extrapolation of the findings from the conditions under which the field
200 studies were conducted to other conditions. As for Tier 1 data, measuring the exposure profile in the
201 field is necessary. Following successful validation, the model could then be used to extrapolate to the
202 relevant untested conditions such as other regions, crops, good agricultural practices (GAPs), or across
203 multiple years. Modeling could also be used to inform the revision of the risk assessment scheme. For
204 instance, it could be used in conjunction with field studies to calibrate the lower tiers of the risk
205 assessment or assess the relevant soil depth at which to apply the PEC to be used in standard risk
206 assessments. Finally, modeling could be used for interpretation of field study results and exploration
207 of mitigation and compensation options.

208 If exposure models are used to provide input for the modeling, the resolution of data has to be
209 considered. Some current EU regulatory pesticide fate models include sufficient temporal and spatial
210 resolution (e.g., FOCUS, PEARL) but are only suitable for simulating uniform application scenarios,
211 such as spray applications to the crop or soil surface, or applications by injection and incorporation
212 into the soil (Van den Berg et al. 2016). If the application is not homogenous (e.g., drip application,
213 tree row application, or precision farming), the fate models will need a higher spatial resolution to
214 produce outputs useful for higher-tier ERAs. Although high-resolution two-dimensional fate models

215 exist, such as HYDRUS (Šimůnek et al. 2012) or 2DROPS (Agatz and Brown 2017), they are not yet
216 open access.

217 As is the case for all models used for ERA, it is critical that FORESEE is evaluated and
218 documented thoroughly following the principles of Good Modeling Practice as recommended by
219 EFSA (2014). Evaluation options include testing against additional laboratory and field studies,
220 sensitivity and robustness analysis (i.e., pushing the limits of the model and testing its domain of
221 applicability), evaluation of submodels in fit-for-purpose studies, and using results of control and toxic
222 standard treatments from field studies. In addition, an uncertainty analysis of assessments based on the
223 model, model assumptions, and parameterizations would need to be included – also in comparison to
224 standard assessment procedures (EFSA 2018; 2019).

225 Scenarios need to be clearly defined to represent relevant environmental conditions, and the
226 fate models must provide the necessary inputs for temperature and soil moisture. However, scenarios
227 that have been chosen to be worst-case from a pesticide fate perspective may not be worst-case from
228 an ecological perspective (e.g., if dry conditions during the exposure window keep the earthworms in
229 deeper soil layers). Thus, the fate scenarios need to be evaluated to ascertain whether they are
230 sufficiently worst-case from an ecological perspective to determine whether new scenarios are needed.
231 Models are acknowledged as a useful tool for understanding processes or simulating effects that
232 cannot be tested in the laboratory, such as effects of repeated exposure over multiple years or
233 extrapolation to other GAPS. Furthermore, they can help to calibrate the risk assessment, e.g., from
234 Tier 1 to reference tier (field), as well as for refining the risk estimates and addressing uncertainties
235 associated with realistic conditions when Tier 1 ERA identifies a non-acceptable risk.

236 *Earthworm Ecology and Behavior (WG 2)*

237 Earthworm vertical movement plays an important role in population-level exposure to
238 pesticides in the field. Understanding how different earthworm species move in response to
239 environmental changes is crucial for effective risk assessments of pesticides in soils. In general,
240 earthworm movement is determined by the ecological category to which they belong and various
241 abiotic and biotic factors (Roeben et al. 2020).

242 Earthworms are often categorized into three ecotypes: epigeic, endogeic, and anecic (Bouché
243 1977; Bottinelli et al. 2020). Epigeic (surface-living) and anecic (vertical burrowing) earthworms rely
244 on leaf litter at the soil surface for habitat (epigeic only) and food, whereas geophagous endogeic
245 earthworms live in temporary horizontal burrows in the mineral soil (Jégou et al. 1998; Capowiez et
246 al. 2014). Distinct patterns of movement and surface activity across earthworm ecological groups,
247 together with the environmental fate of different pesticide applications, can strongly influence
248 pesticide exposure through the soil profile. Accurate assessments of pesticide effects on earthworm
249 populations necessitate the consideration of each ecological group (Tomlin 1992). Ecotypes might not
250 always explain the behavior observed in the field, but are currently the most accepted concept and
251 therefore chosen as model categories. Differences in reaction to changes in environmental parameters
252 might also be observed not only between species – but also between juvenile and adult worms of one
253 species, leading to different movement ranges and distribution patterns over time.

254 *Eisenia fetida*, *Aporrectodea caliginosa*, and *Lumbricus terrestris* are often mentioned as
255 representative species of epigeics, endogeics, and anecics, respectively (Lee 1985). However, the
256 position of *L. terrestris* within these ecological categories, as defined by Bouché (1977), has been
257 questioned. Many authors refer to *L. terrestris* as epi-anecic rather than anecic (e.g., Hoeffner et al.
258 2019) due to differences in diet and behavior. Epi-anecic species first build a burrow or shelter and
259 subsequently use it to forage at the soil surface, whereas anecic species (also referred to as “true
260 anecic”) burrow more continuously in the soil and thus ingest more soil (Ferrière 1980; Bastardie et al.
261 2005). To fully represent earthworm ecotypes (and thus communities) currently found in agricultural
262 lands, the workgroup decided that for a model to be applied in soil risk assessment, four main ecotypes
263 are needed: epigeic, endogeic, epi-anecic, and anecic. Different species can be used to represent these
264 ecotypes, and good examples are: *Lumbricus castaneus* for epigeics, *A. caliginosa* for endogeics, *L.*
265 *terrestris* for epi-anecics, and *Aporrectodea nocturna* or *Aporrectodea longa* for anecics. *E. fetida* is
266 also used to represent epigeics, primarily because so much data are available for this species.

267 The reliability of population models fundamentally depends on the availability of data, and the
268 suitability of different earthworm species for population models depends on the ecotypes most at risk.

269 Individual-based models, for instance, require detailed information on the biology and behavior of the
270 modeled species at the individual- and population levels, both for model development and model
271 validation. Across earthworm ecotypes, abiotic factors play an important role in driving movement
272 behavior, and thus also the possible exposure to a pesticide. Four environmental variables have been
273 identified as critical and feasible to be used for simulating the behavior and vertical movement of
274 earthworms: soil water potential (Gerard 1967; Holmstrup 2001), soil organic matter content (Le
275 Couteulx et al. 2015; Frazao et al. 2019), temperature (Eriksen-Hamel and Whalen 2006), and bulk
276 density of the soil (Kretzschmar 1991). For anecic and epi-anecic species, light is an additional factor
277 to be considered (Nuutinen et al. 2014). Roeben et al. (2020) provide a detailed review of the effects of
278 biotic and abiotic factors on the vertical movement of earthworms.

279 IBMs have the advantage of allowing interaction between an individual and its virtual
280 environment. The identified environmental variables that influence the movement of earthworms are
281 stored in the modeling environment, which is represented through patches in a spatially-explicit
282 setting. One approach to incorporate the simultaneous influence of these four environmental variables
283 on earthworm movement in an IBM is a patch quality index. This index is also part of the modeling
284 environment and determines the movement decisions of individuals in the movement module. The
285 index scales the attractiveness of each soil patch according to the four variables from 1 (attractive) to 0
286 (not attractive). In this way, the quality index can account for the combined effects of temperature, soil
287 water potential, organic matter content, and bulk density on earthworm movement. To be able to
288 represent the different ecotypes and their preferences realistically, the workgroup suggested a dynamic
289 trade-off between the following factors:

- 290 • If temperature and water potential are within a defined performance range, organic matter
291 content should be most important.
- 292 • If temperature and water potential are outside this range, organic matter content has no
293 importance.
- 294 • The last factor is the bulk density, which inhibits the movement of earthworms with increasing
295 density.

296 The performance ranges, slopes, and threshold values depend on the ecotype/species modeled and
297 should be fitted to laboratory and field data individually. Thereby, the patch quality index can cover
298 the different importance of factors for different ecotypes.

299 Besides the abiotic factors listed above, other factors, including exposure to pesticides, food
300 availability, avoidance behavior, inter-species interactions, spatial competition, and intra-species
301 interactions, can also affect the movement of earthworms and, therefore the risk of exposure (Uvarov
302 2017; Capowiez and Belzunces 2001). The workgroup discussed the possible extent of the different
303 influences. Most of the workshop participants concluded that these features do not necessarily need to
304 be included in the model, depending on the level of realism required to address the specific question at
305 hand and considering trade-offs between generality and realism. However, some participants
306 recommended that the relevance of these features ought to be analyzed in a sensitivity analysis prior to
307 considering trade-offs between simplification and realism. If identified as important, they should be
308 considered for inclusion in the model to increase reliability of model outputs.

309 In some cases, there may be a lack of available data, which has to be acknowledged when
310 choosing a modeling approach. For the epigeic and endogeic ecotypes, it is assumed that mating takes
311 place when earthworms meet another individual randomly within the soil. For epi-anecic species,
312 foraging is the primary driver for movement on the soil surface, and if two adult individuals meet, the
313 earthworms may mate and reproduce, depending on the season.

314 The workgroup concluded that the development of a trait-based approach for the movement of
315 earthworms is possible but data-intensive. A list of necessary data and existing knowledge gaps for
316 representative earthworms from the given ecological categories can be found in Table 1. Ideally, an
317 energy-budget model is available for the representative species of each ecotype. Furthermore, data on
318 mortality and longevity of the species are needed and how these traits are influenced by abundance.
319 Moreover, information about preferences towards the four environmental factors determining
320 movement is essential. Finally, information on behavioral aspects is necessary, such as the percentage
321 of time spent on different activities. This includes the time spent foraging at the surface, burrowing,
322 moving in existing burrows, and being inactive. It is crucial to be aware that these traits and

323 preferences can change with developmental stage and exposure, and juveniles will likely have
324 different traits than adults. For the four ecotypes, knowledge gaps that have to be filled for a trait-
325 based movement model to be implemented have been identified. For some categories, the data gaps
326 are greater than others (Table 1), but population models are available for three of the four groups,
327 whereas a model for “true” anecics still needs to be developed (Johnston et al. 2014a, 2014b, 2015,
328 2018).

329 *Uptake Routes (WG 3)*

330 Extrapolation of effects from standardized laboratory toxicity tests to effects in the
331 environment is challenging because it requires several extrapolation steps. Using a TKTD framework,
332 in combination with soil fate modeling, allows the use of mechanistic modeling to facilitate the
333 required extrapolations. At first, the exposure to a substance with specific physicochemical
334 characteristics in the artificial soil used in the laboratory toxicity tests has to be translated to different
335 real soil types in the environment. In comparison to current methods, this translation can be made
336 more accurate by explicitly modeling the fraction of active ingredient in porewater and sorbed to
337 particles in both systems (i.e., in the laboratory toxicity test and in the environment) (Li et al. 2020).

338 Relevant chemical properties include the partitioning coefficient K_D (or the organic-carbon
339 normalized variant K_{OC}), which describes the partitioning of a chemical between the water and soil
340 phase and, therefore its availability for transport, uptake, and subsequent effects. This partitioning is
341 influenced by soil composition, for example, the amount of organic carbon, but also by the actual soil
342 water content. For ionizable chemicals, also the pH and speciation information, such as pKa values,
343 are informative. Ultimately, biodegradation rate constants need to be considered as they capture the
344 decline of chemicals.

345 The second extrapolation requires accounting for different uptake routes (e.g., via skin, via
346 gut), which are of different relative importance for different earthworm species (e.g., different
347 movement patterns, different food sources including litter). Currently, this is not explicitly accounted
348 for in the risk assessment, though some would argue that differentiating between uptake routes in the
349 standard ERA may not be needed if it is sufficiently conservative. This limitation could be overcome

350 by using the internal pesticide concentrations in the earthworm. The extrapolation can be made more
351 accurate by a two-step TKTD approach (Ashauer and Escher 2010). In the first step, the different
352 uptake routes are simulated to calculate the time-variable internal exposure (approximated as whole
353 body residues). This is proposed to be done when analyzing the laboratory toxicity study, and when
354 simulating effects in the environment. In a second step, the effects (toxicodynamics) are simulated
355 using the internal pesticide concentration as the forcing variable. This can be done when analyzing the
356 laboratory toxicity study to calibrate the TKTD model and when predicting effects in the environment.

357 For the endpoint survival, this approach is termed full-GUTS (Jager et al. 2011; Ashauer et al.
358 2016), and the same principle can be applied to DEB-TKTD to account for sublethal effects. GUTS is
359 considered ready to be used in risk assessment in the EFSA scientific opinion on TKTD for aquatic
360 organisms, and although the DEB-TKTD modeling approach is currently limited to research
361 applications, its potential for future use in ERA for pesticides is recognized (EFSA 2018). Key aspects
362 of TKTD modeling can be transferred from the aquatic to the terrestrial risk assessment, in particular
363 the calculation of exposure multiplication factors (Ashauer et al. 2013, EFSA 2018) as well as many
364 recommendations for model calibration. This same document (EFSA 2018) recommends strict
365 requirements for the validation of models.

366 For uptake of pesticides into soil organisms, it is essential to consider bioavailability as there
367 are multiple compartments of the soil in which the pesticide can be present (in porewater and sorbed to
368 soil organic matter and soil mineral particles) and bioavailable to different extents. Pesticide
369 properties, such as partitioning coefficients or biodegradation rate constants, and soil properties,
370 including water content, pH values, and organic carbon content, can influence partitioning of the
371 pesticide in soil. These properties can result in different concentrations in porewater, sorbed to soil
372 organic and soil mineral particles, and in the soil pore airspace and, therefore, in differences in
373 bioavailability. Thus, pesticide exposure depends on local conditions, pesticide properties, and
374 earthworm ecology (e.g., movement, food sources). The pesticide distribution in the soil can be
375 modeled using fate models, but these need to be extended to include the additional effects of soil
376 properties that influence bioavailability if internal concentrations are to be predicted. Modeling uptake

377 from porewater and soil particles via skin and uptake from particles via the gut accounts for
378 bioavailability (relative contributions of the different compartments) in pesticide uptake and effects.

379 The model by Jager et al. (2003) is a good starting point for accounting for general uptake via
380 dermal exposure versus feeding. We are not aware of any alternative, however additional experimental
381 work will be required to underpin the tentative relationship between $\log K_{ow}$ and uptake rate constants
382 established by Jager et al. (2003) with a larger database and to account for confounding factors related
383 to bioavailability. This is important because the rate constants acquired from experiments as described
384 by Jager et al. (2003) may depend on environmental variables, e.g., soil properties, soil water content
385 or temperature. Thus, new experimental and data analysis protocols are needed to disentangle the
386 influence of environmental variables and substance properties on rate constants. The approach of
387 acquiring uptake and elimination rate constants for both exposure routes (dermal and oral) via the
388 partition coefficient, $\log K_{ow}$, is based on only three example compounds with a rather high $\log K_{ow}$
389 and is subject to the limitations described above (dependency on experimental variables). Until this
390 relationship is made more robust with more data covering a wider range of $\log K_{ow}$ values, and
391 disentangled from experimental variables, it is better to measure the actual uptake rates via gut and
392 skin for each compound under investigation and in each soil type of interest. The limitations described
393 here can be overcome by modeling the fate and distribution of test substances in the soil of the
394 laboratory experiment in combination with TKTD modeling. Such data analysis may be able to
395 disentangle the influence of environmental variables, and bioavailability, on TK rate constants from
396 their relationship with substance properties.

397 Specifically, there is a need for toxicity tests for more and specifically low sorbing compounds
398 to evaluate the usefulness of the whole approach, i.e., the combination of soil fate modeling with two-
399 step TKTD modeling. Validation experiments can include laboratory toxicity tests with different soils
400 and compounds (to evaluate if bioavailability is properly accounted for) as well as experiments with
401 different exposure patterns and field studies (see also next section).

402 Understanding the bioavailability and actual exposure in the toxicity test used for model
403 calibration is essential because it enables better extrapolation to different soils in the environment.

404 Including measurements and/or model simulations of pesticide fate in the chronic earthworm study
405 (OECD 2017) would be a step towards providing a more relevant exposure estimate within current
406 testing schemes.

407 *Cross-Species Extrapolation and Model Testing (WG 4)*

408 For earthworms, as for most taxa, a major issue hampering between-species extrapolation is
409 that the relevant field species are not tested in the laboratory on a routine basis, and species may differ
410 in their sensitivity and traits. As a consequence, the evaluation of TKTD models and population
411 models based on laboratory data, by comparing them with field studies, is associated with additional
412 challenges. The suitability of models developed for a laboratory test species, such as *E. fetida*, for field
413 species remains uncertain and may be inaccurate if species vary in inherent sensitivity and traits.

414 From previous studies, it has been shown that earthworms can have different inherent
415 sensitivities to chemicals, including pesticides (Ma and Bodt 1993; de Lima e Silva et al. 2017;
416 Römbke et al. 2017). In a meta-analysis of species sensitivity, Pelosi et al. (2014) found that reported
417 LC₅₀ values for more widespread and ecologically relevant earthworm taxa were, on average,
418 significantly lower than for *E. fetida*. This finding is indicative of a systematic lower sensitivity of this
419 widely tested species that needs to be considered in any modeling framework. Whereas this difference
420 in sensitivity has been observed for lethal effects, little is known regarding which traits explain the
421 differences in inherent sensitivity, and sublethal endpoints for non-standard test species are difficult to
422 determine.

423 Explicitly addressing differences in species sensitivity in mechanistic effect models ideally
424 involves the identification of potential underlying causes for cross-species differences. Several
425 characteristics of a species related to a) phylogeny, b) physiology, morphology, ecology, and c) gene
426 and protein expression, are likely to provide mechanistic explanations for sensitivity differences
427 among species. Species sensitivity can be represented by summary statistics, like LC₅₀ or NOEC, but
428 approaches for predicting TKTD model parameters are more likely to succeed as the model parameters
429 are biologically meaningful (Ashauer and Jager 2018; Gergs et al. 2019). Van den Berg et al. (2020)
430 hypothesize that models related to physiology, morphology, and ecology exhibit the highest prediction

431 power for TK parameters, whereas gene and protein expression models may exhibit the highest
432 prediction power for TD parameters.

433 As for TK parameters, a wide range of physical and ecological traits can potentially affect
434 exposure and uptake. These include skin and gut wall morphology and structure; the role of the gut
435 microbiome, which varies between species; body size in relation to passive diffusion (also applicable
436 for life stage sensitivity); gut residence times; lipid content and metabolic capacity (Phase I, II, and III
437 enzyme activities) of species. Some of these trait data are simple to measure and can be collected
438 fairly easily for widespread earthworm species, whereas others will be difficult to fully characterize. In
439 the latter cases, it may be more efficient to categorize traits relating to TK by assessment of rates based
440 on screening metabolism of different model compounds, rather than through detailed mechanistic
441 prediction that attempts to cover all substances.

442 TD traits that determine sensitivity include the presence, structure, and functional motif of
443 potential molecular targets for the chemical, the extent of damage resulting from a given level of
444 exposure, as well as repair mechanisms. Gene and protein expression-based approaches are available
445 for the assessment of these characteristics and can identify the presence of putative target orthologues,
446 such as with the ECOdrug tool (Verbruggen et al. 2018). For a more detailed target-specific sequence
447 analysis, the SEQUAPASS tool supports orthologue identification, as well as motif and specific
448 residue level analyses (LaLone et al. 2016). The underlying assumption inherent in these tools is that
449 the presence of an orthologue in a species is likely to be associated with higher sensitivity. In addition,
450 species that possess orthologues containing conserved ligand binding motifs and key residues
451 associated with strong ligand interactions will be more sensitive than species that lack strong ligand
452 binding domains or residues. These assumptions have been tested in a number of selected case studies
453 (Gunnarsson et al. 2008; LaLone et al. 2017). However, the complexities of genome evolution,
454 including gene family expansion and reduction as well as gene and even whole genome duplications,
455 mean that these tools are still far from being at a stage in which they fully capture all TD processes
456 that may influence sensitivity.

457 If sufficiently reliable models for the extrapolation of species sensitivity towards different
458 chemicals are available, or in cases for which laboratory toxicity data are available for the
459 parameterization of effect models for relevant field species, confidence in population models can be
460 increased based on field toxicity trials. An example of population-level testing is reported in Johnston
461 et al. (2018). The use of effect data for testing TKTD models derived from field studies is limited, but
462 field data could be used to partially (e.g., initial decline in abundance) validate predictions from TKTD
463 models such as GUTS. It is recognized that both the range and ranking of species sensitivities may
464 vary considerably among compounds, and pragmatic approaches for dealing with this are needed.

465 Any move to apply mechanistic models for modeling pesticide impacts in earthworms will
466 require a change in current testing procedures. The current chronic earthworm test involves the
467 assessment of survival on day 28 only and measurement of reproduction at test termination (day 56).
468 Development of process-based approaches such as TKTD models, however, requires data at a higher
469 temporal resolution. Designs that include measurement of survival at regular times for
470 parameterization of TKTD models, such as GUTS, are potentially easy to conduct by extending
471 exposure time and increasing observations of mortality to at least four time points. A challenge for
472 such studies with earthworms is simply that soil, unlike water, is not transparent. Consequently, each
473 measurement involves disturbing the test system (e.g., by hand sorting), raising the issue of stress and
474 potential mechanical damage. Alternatively, a destructive sampling design could be used, though this
475 would require additional replicates. For DEB model application, the slow rate of earthworm
476 development and the extended timescale of reproduction mean that life-cycle tests measuring juvenile
477 and adult traits over time are unlikely to be feasible, and if conducted, would need to take compound
478 fate into consideration. It should also be noted that for many field-relevant species, following
479 individuals from birth through adulthood and reproduction is not practical. Approaches that separately
480 measure juvenile growth and adult reproduction have been proposed and could form the basis of a
481 suitable method for time series data collection (Van Gestel et al. 1991; Spurgeon et al. 2003). Given
482 the intricacies of TKTD model development and parameterization for a long-lived soil-dwelling
483 species, a further challenge is how to validate model predictions for those species. One approach is to
484 use mechanistic measurements, such as internal concentrations or measurements of tissue “damage”

485 (although difficult to define) for the testing of model components. However, the targeted nature of
486 such measurements means that they may only validate one parameter, rather than the output of the
487 model as a whole. Therefore, validation of TKTD model predictions on the level of survival or
488 reproduction based on laboratory validation tests is recommended.

489 Ideally, the conditions for experiments for validation purposes should be different from those
490 in the calibration experiment and reflect the (regulatory) question to be addressed by the model.
491 Examples include variation of exposure duration, spatial variation of exposure, variation in time scale
492 (temporal extrapolation), or variation in environmental conditions such as soil properties affecting
493 chemical fate and exposure, and earthworm movement and population structure.

494

495 **Priorities for Future Work**

496 We conclude that a mechanistic modeling approach, linking appropriate environmental
497 variables, reflecting defined scenarios, TKTD processes, and movement behavior, can provide realistic
498 individual- and population-level predictions. This approach offers promise for improving scientific
499 understanding and informing pesticide risk assessment for earthworms in the EU regulatory
500 framework. With that said, we have identified several areas in which more work is needed to allow
501 FORESEE to reach its full potential. Moreover, we provide several recommendations for moving this
502 initiative forward. Filling the remaining data gaps identified by workshop participants would enable
503 FORESEE to achieve its full potential as a tool for refining risk and to address uncertainties in the
504 present risk assessment for earthworms exposed to pesticides. These are shown in Table 2.

505

506 **Recommendations**

507 We recommend:

- 508 • Further developing FORESEE as a mechanistic effect model that could be applied for
509 pesticide risk assessment and parameterized for relevant earthworm ecotypes represented in
510 European agricultural systems.
- 511 • Additional assessment of the differences in species sensitivity between standard test species
512 and more ecologically relevant earthworm species for different compounds, as species
513 sensitivity can vary between chemicals.
- 514 • Further investigation of the relevance of impacts of abiotic and biotic factors on the movement
515 of earthworms.
- 516 • Employing a data-informed, trait-based approach to simulate a set of representative earthworm
517 species using a framework considering four ecotypes, which we believe to be sufficient for
518 capturing earthworm behavioral traits regarding movement patterns in the soil. Traits to
519 include describe moving and burrowing behavior and niche characteristics (i.e., tolerance to
520 drought, temperature, soil bulk density, and food conditions).
- 521 • Modeling and/or measuring internal concentrations (body residues) as a step to account for
522 different routes of uptake (e.g., dermal, gut) as a refined option in the tiered risk assessment
523 scheme.
- 524 • Measurements of organism size, mortality, and reproductive output at intermediate time points
525 in laboratory toxicity studies to facilitate parameterization of TKTD modeling (GUTS and
526 DEB-TKTD). This will require a reassessment of the standardized approach currently used for
527 earthworm toxicity testing, especially for the measurement of reproduction for intermediate
528 time points. In addition, substantial additional effort will be involved as either soil will need to
529 be changed at each sampling, or additional replicates will be needed to allow destructive
530 sampling.
- 531 • Further developments of FORESEE for possible use in EU pesticide risk assessment following
532 EFSA's guidance for good modeling practice, including detailed and transparent model
533 documentation. This includes the consideration of model uncertainty.

- 534 • Version control of effect models in order for them to be used in the EU registration procedure.
535 Version control can be done within the existing EFSA-Chaired Version Control Workgroup
536 for pesticide fate models.
- 537 • Organization of a follow-up working group or targeted workshop to establish detailed
538 experimental designs for robust model calibration and evaluation.
- 539 • Broad stakeholder engagement to achieve agreement on the data sets that FORESEE should be
540 tested against, validation of study designs, and other criteria for model evaluation to increase
541 the validation status of the effect models.
- 542 • Broad scientific discussion to gain consensus on appropriate ecological scenarios in which to
543 assess risk using FORESEE given that scenarios used to derive worst-case pesticide fate
544 estimates may not be appropriate for modeling earthworm risk.
- 545 • EFSA to critically consider the key findings and recommendations from this workshop
546 together with other relevant reports or published scientific information during the revision of
547 their guidance for risk assessment of soil organisms to improve the linkage of exposure and
548 effects and address other knowledge gaps in current ERA practice.
- 549 • Establishing a formal and transparent mechanism to ensure that models for pesticide risk
550 assessment in the EU can be effectively and efficiently evaluated.

551 We acknowledge that current approaches to pesticide risk assessment include uncertainties
552 with regard to spatiotemporal variation in pedological, climatic, and biological conditions,
553 agronomical practices, and complexities occurring at the landscape scale (Topping et al. 2020).
554 However, we conclude that mechanistic effect modeling of the kind described here can help to
555 quantify and reduce uncertainties in ERA by providing improved integration of exposure and effects
556 and by incorporating different pesticide application scenarios and greater ecological realism.

557

558

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Table 1. Data availability, identified by work group 2, for the development of realistic population modules for different earthworm species representative of four earthworm ecotypes. Ticks represent available data, (lit.) indicates that the data are available from the scientific literature, - indicates not available, and question marks require a literature review to identify whether the data are available.

	<i>Lumbricus terrestris</i>	<i>Aporrectodea longa</i>	<i>Aporrectodea caliginosa</i>	<i>Eisenia fetida / Lumbricus castaneus</i>	
	epi-aneic	aneics	endogeics	epigeics	
Energy budget	√	√	√	√	-
Mortality rate	?	?	?	?	?
Temperature preference	√ (lit.)	?	√ (lit.)	√ (lit.)	?
Soil water potential preference	√ (lit.)	√ (lit.)	√ (lit.)	√ (lit.)	?
Soil organic matter preference	-	?	√	-	-
Bulk density preference	√	?	√	-	-
Mating as surface	√	-	-	√	√
% time at surface	√	?	√	√	√
% time burrowing	√	? (lit.)	√	√	√
% time displacing	√	? (lit.)	√	√	√
% time inactive	√	? (lit.)	? (lit.)	?	?

Table 2. Main data gaps for earthworms in the context of this workshop on soil organism pesticide risk assessments and how filling them would improve ERA.

Data Gap	Needed for
Definition of realistic worst-case environmental scenarios for modeling (spatial and temporal scales, number of spatial dimensions, soil and climate variables) and establishing link to existing exposure models	Relevant data for FORESEE's environment module
Intermediate measurements of survival, growth, and reproduction in chronic earthworm study	Time course data to parameterize GUTS or DEB-TKTD
Toxicity test results for different soils and chemicals with a range of Log K_{ow} values	Proof of concept with a short term benefit to the existing risk assessment as it could be used to replace the arbitrary correction factor of 2 when $\log K_{ow} > 2$
Measured dermal and oral uptake rate constants for a wide range of Log K_{ow} values disentangled from experimental variables (e.g. soil type, water content)	Establishing the relationship between uptake rate constants and substance properties (e.g. $\log K_{ow}$) whilst accounting for bioavailability
A few comprehensive studies with measurements of several state variables (e.g., concentrations in bulk soil, porewater and earthworms & toxicity, over time)	Better system understanding and evaluation if model complexity is appropriate
Ecological studies	Data on movement differences among earthworm ecological categories
Tests of inherent toxicity in multiple worm species	Data needed for cross-species extrapolation and to distinguish sensitivity differences from exposure differences