



## Article (refereed) - postprint

Adams, Jessica L.; Tipping, Edward; Thacker, Sarah A.; Quinton, John N. 2018. An investigation of the distribution of phosphorus between free and mineral associated soil organic matter, using density fractionation.

© Springer International Publishing AG 2017

This version available http://nora.nerc.ac.uk/518569/

NERC has developed NORA to enable users to access research outputs wholly or partially funded by NERC. Copyright and other rights for material on this site are retained by the rights owners. Users should read the terms and conditions of use of this material at http://nora.nerc.ac.uk/policies.html#access

This is a post-peer-review, pre-copyedit version of an article published in Plant and Soil, 427 (1-2). 139-148. The final authenticated version is available online at: https://doi.org/10.1007/s11104-017-3478-4.

There may be differences between this version and the publisher's version. You are advised to consult the publisher's version if you wish to cite from this article.

> Contact CEH NORA team at noraceh@ceh.ac.uk

The NERC and CEH trademarks and logos ('the Trademarks') are registered trademarks of NERC in the UK and other countries, and may not be used without the prior written consent of the Trademark owner.

1	Revision submitted to Plant and Soil August 2017			
2				
3	An investigation of the distribution of phosphorus between free and mineral associated soil organic			
4	matter, using density fractionation			
5				
6	Jessica L Adams · Edward Tipping · Sarah A Thacker · John N Quinton			
7				
8	Jessica L Adams · Edward Tipping · Sarah A Thacker			
9	Centre for Ecology and Hydrology, Lancaster Environment Centre, Lancaster, LA1 4AP, UK			
10	e-mail jesams@ceh.ac.uk			
11				
12				
13	John N Quinton			
14	Lancaster Environment Centre, Lancaster University, Lancaster LA1 4YQ, UK			
15				
16				
17	Acknowledgements			
18	This work received National Capability funding from the UK Natural Environment Research Council			
19	(CEH project number NEC04841). We are grateful to Alan Lawlor, Patrick Keenan, Manisha Patel and			
20	Binoti Tanna for help with analysis, and to Aidan Keith (CEH) and two anonymous referees for their			
21	constructive comments.			
22				
23				
24				
25				
26				
27				
28				
29				
30				
31				
32				
33				

- 35 Abstract
- 36
- 37 *Aims* We investigated whether density fractionation can be used to determine the distribution of 38 organic phosphorus (OP) between free and mineral-associated soil organic matter (SOM).
- 39 Methods We performed density fractionations using sodium polytungstate solution (specific gravity
- 40 1.6 g cm<sup>-3</sup>) on 20 soils from UK semi-natural and pasture ecosystems, to obtain a light fraction (LF) and
- 41 a heavy fraction (HF) for each soil. The fractions were quantified by weight, and analysed for organic
- 42 carbon (OC), total N (TN), total P (TP), inorganic P (IP), and OP (by difference).
- 43 Results Good recoveries of soil mass (96%), OC and TN (both ~ 90%) were obtained, but recovery of 44 OP only averaged 56%. The average P:C ratio of HF SOM exceeded that of LF SOM by a factor of six, 45 greater than the factor of two obtained for TN:OC. For the soils studied, the elements of SOM were
- 46 predominantly in the HF, with averages of 75% for C, 82% for N, and 90% for P.
- 47 *Conclusions* The incomplete recovery of OP demands further work. Nonetheless, the results show that
- 48 HF SOM is much richer in P than LF SOM.
- 49
- 50
- 51
- 52 Keywords
- 53 Carbon  $\cdot$  Density fractionation  $\cdot$  Nitrogen  $\cdot$  Phosphorus  $\cdot$  Soil organic matter  $\cdot$  Stoichiometry

- 54 Introduction
- 55

The N (nitrogen) and P (phosphorus) contents of bulk SOM (soil organic matter) vary appreciably; C:N 56 57 ratios can be as low as 8 and higher than 30, while C:P ratios range from c. 20 to 1000 (Tipping et al. 2016). Such variation arises from differences in the element stoichiometry of input litter, the 58 59 processing of elements during decomposition, stabilisation processes, and within-soil transport. Accounting for the variation is necessary in order to understand and quantify the interlinked 60 61 biogeochemical cycles of the elements. From analysis of data obtained with the Hedley fractionation 62 procedure (Hedley et al, 1982), Yang and Post (2011) found that C and N in SOM were closely linked, 63 but that P was correlated to neither C nor N, and they concluded that OP is decoupled from OC and 64 ON. However, from an analysis of data for c. 2000 soils, including topsoils and subsoils under both 65 natural and agricultural vegetation, with %OC ranging from 0.1 to 50%, Tipping et al. (2016) found a 66 strong positive relationship between the N:C and P:C ratios of SOM. This was attributed to the 67 preferential adsorption by mineral matter (i.e. accumulation at mineral surfaces owing to physical and chemical interactions) of N-rich and P-rich organic compounds. Therefore, further insight might be 68 69 gained by fractionating soils according to density, then analysing the fractions for organic phosphorus.

70 The physicochemical fractionation of SOM involves the separation of SOM through flotation, 71 sedimentation and aggregate disturbance. During fractionation, the organic debris including plant and 72 animal material, referred to here as the light fraction (LF), but sometimes referred to as particulate 73 organic matter (Zimmermann et al. 2007), is separated from organic material bound to mineral 74 matter, referred to as the heavy fraction (HF), using a dense solution. A range of fractionation 75 methods are available (Sohi et al. 2001; Kirkby et al. 2011; Zimmermann et al. 2007), and there is no 76 standardised procedure, therefore research into methodological aspects continues (Cerli et al, 2012). 77 For well-drained soils, the HF is generally regarded as the stable SOM pool, on the basis of stable 78 isotope ( $\delta^{13}$ C) and radiocarbon ( $^{14}$ C) analyses (Trumbore 1993; Swanston et al. 2005; Tan et al. 2007; 79 Kögel-Knabner et al. 2008). Almost all studies report a low C:N ratio within the HF whereas the LF 80 mostly has a high C:N ratio. We conducted a thorough literature search for reports of measurements of the distribution of OP between the HF and LF, and found none, although several studies have 81 82 reported the OP content of light material (Rodkey et al. 1995; O'Hara et al. 2006; Wick & Tiessen 2008). 83

In this study, we investigated the feasibility of using a density fractionation method to determine the distribution of OP in soils, so as to separate the heavy mineral-rich material from the lighter free organic matter, and analysing the fractions for organic carbon (OC), total nitrogen (TN),

- 87 total phosphorus (TP) and inorganic phosphorus (IP), and obtaining organic phosphorus (OP) by
- 88 difference.
- 89 Abbreviations are listed in Table 1.

- 91 Methods
- 92

Soil samples had been collected in a survey of the catchments of the Rivers Avon (Hampshire),
Conwy (N Wales), Dee (NE Scotland), and Ribble (NW England) carried out between 2013 and 2015
(Toberman et al. 2016). Samples had been bulked from 6 or 10 separate cores, and included both
topsoils and subsoils. We chose 20 soils to provide a range of SOM contents, and with sufficient light
material to analyse. Most soils were under seminatural vegetation, three were from improved
grassland. Arable soils were not analysed owing to their low contents of light material.

99 We applied a physicochemical density fractionation method based on the procedure of 100 Schrumpf et al. (2013), which in turn was derived from those of Golchin et al. (1994) and Sohi et al. (2001). Fig 1. is a schematic of the fractionation method. We distinguished non-occluded and 101 102 occluded light fractions (NLF, OLF), which were combined to make the light fraction (LF), and the heavy 103 fraction (HF). One fractionation was performed for each soil. Twenty-five g subsamples of sieved soil 104 were placed in 400 mL centrifuge bottles, with 250 mL sodium polytungstate (NaPT; Sometu, Belgium) at a density of 1.6 g cm<sup>-3</sup> (Cerli et al. 2012). The bottles were gently shaken by hand, then centrifuged 105 106 at 5500 rpm for 30 minutes. If the quantity of floating material (NLF) was low, it was removed using a 107 wide-tipped pipette and placed into 60 µm nylon mesh bags. For samples of heathland and forest soil 108 with higher quantities of NLF, material was removed using a spatula and placed in 60 µm nylon mesh 109 bags. The remaining suspension was brought back to its initial volume with fresh NaPT (this required 110 c. 20 mL) re-centrifuged, and then residual light fraction was removed, this procedure being repeated (no more than twice) until all NLF was accounted for. The material in the mesh bags was rinsed with 111 112 deionised water, and the leachate repeatedly measured for conductivity using a Jenway 4510 probe; complete removal of excess NaPT was assumed when the conductivity fell below 50 µs cm<sup>-1</sup>, except 113 that for calcareous soils conductivities  $< 200 \ \mu s \ cm^{-1}$  were considered acceptable, because of 114 115 dissolution of carbonates (Schrumpf et al. 2013). The rinsed samples were weighed, oven dried at 40 116 °C, weighed again, and once completely dried they were stored in a desiccator until further analysis.

117 Extraction of OLF was carried out using sonication (Sonics Vibracell CV18 probe). To avoid 118 aggregate breakdown of the HF, a pilot test for each of the soil types, based on bulk soil texture 119 (Toberman et al. 2016; Table S1), was carried out to find the optimal sonication energy input, following the procedure of Schrumpf et al. (2013). For sandy and silty soils (there were no clay rich soils), target 120 energy inputs of 100 and 300 J mL<sup>-1</sup> respectively were used. The samples were periodically checked 121 for complete aggregate disruption using a 0.1 mL subsample observed under a microscope at 100x 122 123 magnification. Complete disruption was assumed when no further OLF material could be seen 124 attached to minerals under the microscope. During sonication, the bottle was submerged in an ice

bath and the temperature of the sample was measured and maintained at < 40 °C (Schrumpf et al. 2013). Once fully sonicated, samples were left to stand for 1 hour and then centrifuged again at 5500 rpm for 30 minutes and the OLF extracted by pipette; if necessary, further centrifugation was performed (once or twice) to maximise the capture of OLF material. The OLF was added to the NLF in the 60  $\mu$ m mesh bags, the resulting LF was rinsed again until conductivity was < 50 or < 200  $\mu$ s<sup>-1</sup>, dried at 40 °C, weighed and ground to a fine powder using a Retsch MM400 mixer mill.

The centrifuge bottles containing the remaining material (HF) were refilled with ultra-pure deionised water and centrifuged at 5500 rpm for 10 minutes. After each centrifugation, the supernatant was decanted into plastic beakers and measured for conductivity. This process was repeated until the waste water had a conductivity of < 50 or < 200  $\mu$ s<sup>-1</sup>. The samples were then transferred into aluminium trays, oven dried at 40 °C and weighed. The dried HFs were ground to a fine powder using a ceramic pestle and mortar.

We tested for displacement of P forms from soil by NaPT by suspending 25 g subsamples of four of the sieved soils in 250 mL NaPT at a density of 1.6 g cm<sup>-3</sup> in 400 mL centrifuge bottles, as in the density fractionations. After sonication and centrifugation, the clear supernatant solution beneath the suspended light fraction was removed with a pipette and filtered (Whatman GFF). The solution was analysed for soluble reactive P (SRP) and total dissolved P (TDP) as described below.

Soil OC and TN were determined by the procedures given by Emmett et al. (2008). Before analysis for C and N, any samples that might have contained inorganic carbonate (bulk soil pH > 5.5) were treated with 0.1 M HCl and observed under microscope until all CO<sub>2</sub> release had occurred. These samples were then re-dried at 40 °C. Single determinations of total organic carbon (TOC) and total nitrogen (TN) in milled subsamples were made with a Vario EL elemental analyser. Repeated determinations by this method on three representative UK soils over the period of this study gave relative standard deviations of between 2.1 and 3.6 % for TOC and between 1.7 and 3.1% for TN.

149 Total P (TP) was determined by the ignition-extraction method as described in Olsen and 150 Sommers (1982). First, 0.5 g subsamples were ignited in a Pyrotherm muffle furnace at 550 °C for 1-2 151 hours, placed in 50 mL centrifuge bottles with 25 mL 0.5 M sulphuric acid and shaken for 16 hours. 152 These were then centrifuged at 10000 rpm for 30 minutes, filtered using Whatman 1573 1/2 (12-25 153 μm) filter papers and refrigerated at 4 °C until further analysis. The extracts were analysed for soluble 154 reactive phosphorus using the molybdate method (Olsen and Sommers, 1982). Measurements on a 155 reference sample (ISE sample 921 from Wageningen University, Netherlands) gave an average TP 156 value that was 96.9% (sd 1.1%, n = 4) of the expected value. Inorganic P (IP) was determined by 157 extracting 0.5 g of soil with 25 mL of 0.5 M sulphuric acid, then analysing the extract with molybdate. 158 Organic P was obtained as the difference between TP and IP. These analyses were replicated fourfold. The molybdate method was used to measure SRP in the supernatants of soil/NaPT suspensions (see above), and concentrations of TDP were also determined with molybdate after digestion with acid persulphate (Rowland and Haygarth 1997). The supernatants were diluted 100 times with deionised water before making the measurements, and at the resulting concentrations of NaPT, no interference with the molybdate method was found for SRP. However, acid persulphate digestion of the dilute NaPT solutions reduced the sensitivity of the molybdate assay, and this was taken into account in estimating TDP concentrations.

Bulk analyses of the soils were reported by Toberman et al. (2016), using the same methods for C, N and IP, but with a different method for TP, involving treatment of the samples with aqua regia and microwave digestion. Resource limitations meant that we were unable to determine soil TP by the same method for both bulk and fractionated soils. However tests on six bulk soil samples showed that results from the two TP methods were in agreement; the ratio of TP values from the ignitionextraction method to those from the aqua regia-microwave method ranged from 0.94 to 1.18, with a mean of 1.02 (not significantly different from 1.00, p > 0.05).

173 Statistical analyses (t-tests and linear regressions) were performed with Microsoft Excel. 174 Before conducting linear regression analyses, data were tested for normality using quantile–quantile 175 plotting. For t-testing the D'Agostino-Pearson test was used to check for normality. Non-normal data 176 were transformed using log transformations where necessary.

- 177 Results and discussion
- 178

179 Performance of the fractionation method

180

Good recoveries of soil mass from the density fractionation procedure were achieved for all the 181 182 samples, with an average of 96% and a range over the 20 soils of 90 – 105% (Table S2). Regression analyses indicated that recovery depended upon neither the amount of material in the heavy fraction, 183 184 nor the carbon content of the bulk soil (data not shown). The average recovery fell between the averages of 100% obtained by Swanston et al. (2002) for 7 soils, and 83% obtained by Schrumpf et al. 185 186 (2013) for 48 samples; we used essentially the same method as these previous studies. The loss of 187 some material in these types of methods is probably from some soluble compounds dissolving into 188 the NaPT solution and some solid material was probably lost during rinsing and collection of the 189 separate fractions (Cerli et al. 2012). We found no measurable SRP in the supernatants of four soils 190 that had been suspended in NaPT and the suspensions sonicated, but small amounts of TDP were 191 detected, corresponding to between 3 and 8% (average 4.6%) of the soil TP. The fraction of soil mass 192 in HF ranged from 78.6 to 98.5 % (Table S2).

193 Light fraction element concentrations of OC showed only modest variation (relative standard 194 deviation, RSD, 12%), with a range of 26.5 to 45.5% and a mean of 36.1% (Table S3). This indicates 195 that the LF was predominantly but not entirely SOM (%C ~ 55%), i.e. some mineral matter was present. 196 Crow et al. (2007) reported values of 27 and 29% OC in two soils, and Swanston et al. (2002) obtained 197 a mean of 25% OC from 7 soils. Cerli et al. (2012) observed decreasing OC content in the light fraction 198 with increasing sonication time and intensity, suggesting a higher content of mineral matter through 199 aggregate breakdown. Thus the fractionation procedure certainly concentrates SOM in the LF, but 200 some mineral matter is retained.

201

202 Concentrations of P forms in the LF and HF

203

Concentration data for TP, IP and OP (by difference) are presented in Fig. 2 and Table S4. Based on
relative standard errors, the average reproducibility was ± 14% for the LF forms of P, and ± 6% for the
HF forms, which can be considered satisfactory, bearing in mind the several steps that are involved in
the analytical procedure.

For the majority of HF samples, most of the P is organic (range 50 to 97%, average 79%), whereas in the LF OP and IP are similar (the OP range is 24 to 77%, average 50%). The IP content of the LF is surprising, given that this material is thought to consist mainly of plant residues (Six et al.

2002). One possible explanation is that the strong acid reagent used to extract IP caused hydrolysis 211 212 of some of the LF SOM, releasing IP; however, Turner et al. (2005) considered this to apply to only a 213 small fraction of OP. To explore this further, we compared the results for the LF with data for "natural 214 LF", i.e. the organic horizons of Swiss forest soils (Walthert et al. 2004; Blaser et al. 2005, Zimmermann 215 et al. 2006) for which IP was analysed by the same method that we used here. We took data for 16 F 216 (Oe) and 16 H (Oa) horizons, each dominated by SOM. For the F horizons the mean IP was 8% of the total (range 0 - 30%), for the H horizons it was 16% (range 0 - 53%). Therefore the LF material isolated 217 218 by density fractionation in the present study appears to possess a higher fraction of its P in the 219 inorganic form than high-SOM bulk soils. One possible explanation is that IP owes its presence in the 220 LF to the coordination of inorganic phosphate with Al and Fe complexed by the SOM, which occurs to 221 different extents in the Swiss forest soils and the soils studied here. Another possibility is that IP is 222 associated with mineral matter, present at a higher concentration in the LF compared to the F and H 223 horizon soil samples.

224

## 225 Element recoveries in the fractionation process

226

227 Recoveries were calculated by combining the mass data with measured element concentrations in 228 bulk soil and in the two density-separated fractions. The results are summarized in Table 2 and 229 detailed in Tables S5 and S6. Average recoveries of OC and TN were each 91% (Table 2). The results 230 for OC fall within the range of published values, 72-101%, which come from data reported for two soils by Crow et al. (2007), one soil by Cerli et al. (2012), and 48 soils by Schrumpf et al. (2013), all 231 232 fractionated by a similar method to that used here; the overall average recovery for all 51 soils was 233 94%. Our average recovery of 91% for TN exceeds those of 85% reported for one soil by Cerli et al. 234 (2012), and 74% reported for two soils by Crow et al. (2007). Therefore our processing of the soils 235 with respect to mass, OC and TN achieved similar levels of recovery to those of previous studies.

Average recoveries of TP, IP and OP for individual soils were 62%, 117% and 56% respectively 236 237 (Table 2). In each case the variability in the recoveries is appreciably greater than for OC and TN 238 (Tables S5 and S6). However, regressions of the sums of the recovered forms of P in LF and HF against 239 the starting (bulk) values (Fig. S1) gave highly significant slopes, suggesting some consistency in the 240 behaviours of the P forms during the fractionation and analytical procedure. It appears that on average not much IP was lost, whereas definite losses of TP and OP occurred. The absolute losses of 241 TP and OP were similar (OP loss was equal to 90% of TP loss on average), and highly correlated ( $r^2$  = 242 0.90, p <0.001), indicating that most of the loss of TP was due to loss of OP; this follows because OP 243

was obtained as the difference between TP and IP, because IP was a minor part of TP in HF (see above),and because overall IP losses were minor.

- 246
- 247 Assessment of the methodology for phosphorus
- 248

249 Two aspects of the results obtained give cause for concern about the methodology, the loss of appreciable amounts of OP from some soils in the fractionation procedure (Table S6), and the high 250 251 variability in percentage recoveries (Tables 2 and S6, Fig. S1). Since these problems were not found 252 for OC and TN, they are specific to phosphorus forms, and are presumably due either to the behaviour 253 of OP during the density fractionation procedure or to errors in the analysis of TP and/or IP (OP is derived by difference). Experimental tests for the solubilisation, and therefore loss, of P forms during 254 255 extraction revealed only small losses of TP (see above), not at all sufficient to explain the low 256 recoveries of TP and OP. This rules out any major displacement of sorbed IP or OP by NaPT, which 257 seemed plausible in view of the fact that monotungstate can displace inorganic phosphate from 258 ferrihydrite (Gustafsson 2003). Therefore it seems unlikely that the fractionation procedure is at fault.

259 The analysis procedures for soil P differ from those for OC and TN in two respects. Firstly, the 260 concentrations of the P forms are relatively low, being about an order of magnitude less than that of 261 TN, and two to three orders less than that of OC. Secondly, the combined analytical method for OC 262 and TN is simple and reproducible, comprising full combustion and gas analysis ( $CO_2$  and  $NO_2$  in the method that we used here). In contrast, our analytical method for TP involved combustion in a muffle 263 furnace, extraction of inorganic P into H<sub>2</sub>SO<sub>4</sub>, and then determination of the resulting SRP, while 264 265 determination of IP omits the combustion step. Therefore there is more scope for errors to arise. As 266 shown by the results in Fig. 2 we obtained quite good reproducibility in the P measurements on 267 individual soils, which may suggest that the problems arise from variability in the extraction steps, i.e. 268 the combustion of OM and conversion of OP to IP, or the extraction of the so-formed organic P into H<sub>2</sub>SO<sub>4</sub>. However, to account for the low recoveries and variabilities, difficulties with the combustion 269 270 and extraction would have to apply only to fractionated soil, since full recovery of TP in bulk soils was 271 achieved (see Methods) and we cannot see an obvious reason why that should occur.

Further work is clearly needed to improve the yields from the density fractionation procedure.
The incomplete recoveries must be borne in mind when interpreting our results, in particular the
consequences of different relative losses of OP from HF and LF.

275

276 Element relationships in LF and HF

To explore element relationships in SOM, we assume TN (Table S3) is all organic. According to Stevenson (1986), inorganic N comprises 10% of TN on average, while Schulten and Schnitzer (1998) estimated only 5%. The inorganic contribution is highest in deeper soils and soils poor in SOM, opposite circumstances to our relatively SOM-rich topsoils. In this section, for simplicity and clarity we use N:C, P:C, C:N and C:P to refer to ratios of organic forms of the elements when discussing SOM compositions.

284 Table 3 shows averaged OC concentrations and element ratios (g g<sup>-1</sup>) for LF and HF. The 285 average N:C ratio of the HF is significantly (p < 0.001) higher than that of the LF, the HF N:C ratio 286 exceeding the LF ratio for 19 of the 20 soils. Such a difference also applies for P:C (p < 0.001), again 287 with 19 of the 20 soils fitting the pattern. However, the difference is considerably greater for P:C, since there is nearly six times as much P per unit C in the HF compared to the LF, whereas the factor 288 289 for N:C is only 1.9. Higher N:C ratios (lower C:N ratios) in the HF have been reported before, by Sollins 290 et al. (2006), Crow et al. (2007), Cerli et al. (2012) and Schrumpf et al. (2013), but we could not find 291 published information for P:C ratios in density-fractionated soil. Therefore this appears to be the first 292 time that the difference between HF and LF has been demonstrated for P:C.

293 A comparable study is that of Kirkby et al. (2011), who used a dry sieving and winnowing 294 method to separate light fractions from six Austrailian soils, two natural or semi-natural and four 295 agricultural. They obtained an average OC content of 15.1% and average TN:OC and TP:OC ratios of 296 0.057 and 0.0039 respectively. The higher element ratios, compared to the values for LF in Table 3, 297 can be explained in terms of the lower OC concentration, as discussed below. The remaining soils had 298 an average OC content of about 3%, and TN:OC and OP:OC ratios of 0.085 and 0.0053 respectively, 299 similar to the values in Table 3. Another relevant study is by O'Hara et al. (2006), who used 300 fractionation with water to obtain LFs from several native eucalypt forest soils in Australia, and determined their OC and total P concentrations. The average OC concentration was 45.5% (SD 0.9%), 301 302 and the average TP:OC ratio 0.0011 (SD 0.0003) g g<sup>-1</sup>. Although the TP:OC ratio must be considered a 303 maximum estimate of the SOM P:C (since some of the TP could have been IP), the key point is that the 304 P:C ratio of SOM in these LFs was low, similar to the values in Table 3. Also relevant are data for the 305 "natural LF" of Swiss forest soils, considered above in relation to IP contents. Data for the Oe and Oa 306 horizons gave an average OC concentration of 36.4 (SD 6.3) % and an average TN:OC ratio of 0.045 307 (SD 0.007), both very similar to our values for LF shown in Table 3. For OP:OC, the Swiss forest soils average was 0.0019 (SD 0.0009), which is about double our LF value (Table 3), but still substantially 308 309 lower than the average of 0.0049 that we find for the HF. The similarities between these literature 310 data and our values for density-fractionated LF strongly suggest that although losses of OP from LF 311 material may have contributed to the overall losses of OP in the fractionations, these losses were not

disproportionate, i.e. did not bias the estimated distributions of OP between LF and HF. In view of the
 relatively low levels of OP in the LF, this suggests that most of the OP losses were from the HF.

- Fig. 3 shows how the organic forms of the elements are partitioned into the HF, in relation to the partitioning of soil mass. In 18 of the 20 soils C, N and P are predominantly in the heavy fraction, and the HF percentages are in the order P>N>C. The preferential occurrences of N and P in the HF result from both the HF:LF partitioning of organic matter per se, and also the enrichments of the two elements in HF SOM (see above). This leads to the especially strong partitioning of OP into the HF.
- 319 Stoichiometric relationships among the elements can also be seen in log-log plots of N:C and 320 P:C against %C (Fig. 4), following the approach of Tipping et al. (2016) in their analysis of bulk C-N-P-S 321 data for c. 2000 soils. Tipping et al. (2016) formulated a model of SOM stoichiometry in which the 322 SOM of a soil is considered to be a mixture of two end-members, nutrient-poor SOM (NPSOM) which 323 has low N:C and P:C ratios (0.039 and 0.0011 g g<sup>-1</sup> respectively), and nutrient-rich SOM (NRSOM) which has high ratios (0.12, 0.016 g g<sup>-1</sup>). All NRSOM is considered to be adsorbed to mineral matter, while 324 325 NPSOM may or may not be adsorbed. All unadsorbed SOM is NPSOM. Because mineral matter is the 326 obverse of the measured quantity OC%, the fraction of NPSOM increases linearly with log<sub>10</sub> %C, 327 between limits of 0.1% C and 50% C, and as a result the log<sub>10</sub> N:C and log<sub>10</sub> P:C values are predicted to 328 fall with %C as shown by the lines in Fig. 4. If it is assumed that the adsorption processes responsible 329 for SOM accumulation on mineral matter are unaffected by the physical fractionation of the soil then 330 the model should also hold for the results reported here, and the HF and LF N:C and P:C ratios should 331 follow the predicted relationships, but be separated according to the OC concentrations. As shown 332 by the plots in Fig. 4, the expected trends are indeed approximately followed. Apart from two outliers 333 (heathland soils), the N:C values fall close to the model line, and this is also true of the P:C values for 334 HF, while for LF the ratios are somewhat lower than expected. The heathland outlier HF results may 335 reflect the sandy nature of the soils, which may limit adsorption. Overall, we can conclude that the 336 present results support the Tipping et al. (2016) model.

337 The key result of this work is that the N:C ratios, and especially the P:C ratios, of the HF are 338 significantly and substantially higher than those of the LF. This is consistent with the preferential 339 adsorption by mineral matter of N- and P-rich compounds, proposed by Tipping et al. (2016) as a 340 principal mechanism by which NRSOM is formed. The C-N-P stoichiometry of NRSOM does not reveal 341 much about its molecular constituents, which could include recognisable biochemicals from plants and microbes, and their breakdown products. The material may also comprise larger molecules 342 produced by humification, perhaps by reactions occurring at the mineral surface (Collins et al. 1995; 343 Johnson et al. 2016). Interestingly, McLaren et al. (2015a) presented evidence that about two-thirds 344 345 of the OP in five differing topsoils occurred in high molecular weight material. At least some of the

346 mineral-associated SOM has accumulated over hundreds to thousands of years, and therefore must347 reflect the long-term supply of competing adsorbates, as well as post-adsorption modifications.

348 Our findings demonstrate that density fractionation is a promising approach to investigate the 349 interactions governing soil OP and its relationships to OC and ON. However, as already discussed, in 350 view of the incomplete recoveries that we obtained, there is a need to improve the basic 351 methodology. This might extend to the use of different analytical techniques for the determination 352 of different chemical forms, in view of recent evidence that the ashing-extraction technique used here 353 may underestimate TP (McLaren et al. 2015b). It is also important to recognise the different ways that 354 organic P might be measured (Olsen and Summers 1982; Turner et al 2005). Interesting possibilities 355 to obtain additional information are the coupling of density fractionation with subsequent chemical 356 fractionation, e.g. by the Hedley scheme (Hedley et al. 1982), and/or size fraction (Makarov et al. 357 2004).

- The density fractionation method yielded good recoveries of soil mass, OC and TN for 20 semi natural and pasture soils with OC concentrations ranging from 4.5 to 18%.
- 363 2. Average recoveries of TP and OP were relatively low, 62% and 56% respectively (c. 50%), and
  364 further work is need to improve them.
- 365 3. Organic matter of the heavy fraction was richer in N and P than that of the light fraction, on
  366 average by a factor of two in N, and by a factor of six in P.
- The elements of organic matter were predominantly in the heavy fractions of the soils, with
   averages of 75% for C, 82% for N and 90% for P.
- 369 5. The variations with soil %C of stoichiometric ratios (P:C, N:C) in HF and LF agree approximately

with the predictions of the two end-member mixing model of SOM advanced by Tipping et al.

- 371 (2016), in which organic molecules rich in P and N preferentially accumulate on mineral matter
- 372 surfaces through strong adsorption.

## 374 References

Blaser P, Zimmermann S, Luster J, Walthert L, Lüscher P (2005) Waldböden der Schweiz Vol 2, Hep
Verlag, Bern, 920 pp

- Cerli C, Celi L, Kalbitz K, Guggenberger G, Kaiser K (2012) Separation of light and heavy organic matter
   fractions in soil Testing for proper density cut-off and dispersion level. Geoderma 170: 403–
   416
- Collins MJ, Bishop AN, Farrimond P (1995) Sorption by mineral surfaces rebirth of the classical
   condensation pathway for kerogen formation. Geochim Cosmochim Acta 59: 2387–2391
- Crow SE, Swanston CW, Lajtha K, Brooks JR, Keirstead H (2007) Stabilization and destabilization of soil
   organic matter: a new focus. Biogeochem 85: 69-90
- Emmett BA, Frogbrook ZL, Chamberlain PM, Griffiths R, Pickup R, Poskitt J, Reynolds B, Rowe E,
   Rowland P, Spurgeon D, Wilson J. & Wood CM (2008) Countryside Survey Soils Manual.
   Countryside Survey Technical Report No. 04/07, Centre for Ecology & Hydrology, Wallingford.
- Golchin A, Oades JM, Skjemstad JO, Clarke P (1994) Study of free and occluded particulate organic
   matter in soils by solid state <sup>13</sup>C CP/MAS NMR spectroscopy and scanning electron microscopy.

 389
 Aust J Soil Res 32: 285-309

- Gustafsson JP (2003) Modelling molybdate and tungstate adsorption to ferrihydrite. Chem Geol 200:
   105-115
- Hedley MJ, Stewart JWB, Chauhan BS (1982) Changes in the inorganic and organic soil phosphorus
   fractions induced by cultivation practices and by laboratory incubations. Soil Sci Soc Am J 46:
   970–976
- Johnson K, Purvis G, Lopez-Capel E, Peacock C, Gray N, Wagner T, März C, Bowen L, Ojeda J, Finlay N,
   Robertson S, Worrall F, Greenwell C (2016) Towards a mechanistic understanding of carbon
   stabilization in manganese oxides. Nature Comm 6:7628, DOI: 10.1038/ncomms8628
- Kirkby CA, Kirkegaard JA, Richardson AE, Wade LJ, Blanchard C, Batten G (2011) Stable soil organic
   matter: A comparison of C:N:P:S ratios in Australian and other world soils. Geoderma 163:
   197–208
- Kögel-Knabner I, Guggenberger G, Kleber M, Leinweber P (2008) Organo-mineral associations in
   temperate soils: Integrating biology, mineralogy, and organic matter chemistry. J Plant Nutr
   Soil Sci 171: 61–82
- 404Makarov MI, Haumaier L, Zech W, Malysheva TI (2004)Organic phosphorus compounds in particle-405size fractions of mountain soils in the northwestern Caucasus. Geoderma 118: 101–114

- McLaren TI, Smernik RJ, McLaughlin MJ, McBeath TM, Kirby JK, Simpson RJ, Guppy CN, Doolette AL,
   Richardson AE (2015a) Complex forms of soil organic phosphorus–a major component of soil
   phosphorus. Environ Sci Technol 49: 13238–13245
- McLaren TI, Simpson RJ, McLaughlin MJ, Smernik RJ, McBeath TM, Guppy CN, Richardson AE (2015b)
   An assessment of various measures of soil phosphorus and the net accumulation of
   phosphorus in fertilized soils under pasture. J Plant Nutr Soil Sci 178: 543–554
- 412 O'Hara CP, Bauhus J, Smethurst PJ (2006) Role of light fraction soil organic matter in the phosphorus
  413 nutrition of *Eucalyptus globulus* seedlings. Plant Soil 280: 127–134
- Olsen SR, Sommers LE (1982) Phosphorus. In: Page AL, Miller RH, Keeney DR (eds) Methods of soil
  analysis: Part 2. Chemical and microbiological properties. Agron. Mongr. 9. 2nd ed. ASA and
  SSSA, Madison, WI, pp 403-430
- Rodkey KS, Kaczmarek DJ, Pope PE (1995) The distribution of nitrogen and phosphorus in forest floor
  layers of oak-hickory forests of varying productivity. Proc 10th Central Hardwood Forest
  Conference, Gen Tech: 94–108
- Rowland AP, Haygarth PM (1997) Determination of total dissolved phosphorus in soil solutions. J
   Environ Qual 26: 410–415
- Schrumpf M, Kaiser K, Guggenberger G, Persson T, Kögel-Knabner I, Schulze E-D (2013) Storage and
  stability of organic carbon in soils as related to depth, occlusion within aggregates, and
  attachment to minerals. Biogeosci 10: 1675–1691
- Schulten H-R, Schnitzer M (1998) The chemistry of soil organic nitrogen: a review. Biol Fertil Soil 26:1–
  15
- 427 Six J, Conant RT, Paul EA, Paustian K (2002) Stabilization mechanisms of soil organic matter:
   428 Implications for C-saturation of soils. Plant Soil 241: 155–176
- Sohi SP, Mahieu N, Arah JRM, Powlson DS, Madari B, Gaunt JL (2001) A Procedure for isolating soil
  organic matter fractions suitable for modeling. Soil Sci Soc Am J 65: 1121-1128
- Sollins P, Swanston C, Kleber M, Filley T, Kramer M, Crow S, Caldwell BA, Lajtha K, Bowden R (2006)
   Organic C and N stabilization in a forest soil: evidence from sequential density fractionation.
   Soil Biol Biochem 38: 3313–3324
- 434 Stevenson FJ (1986) Cycles of soil: carbon, nitrogen, phosphorus, sulfur, micronutrients. Wiley, New
  435 York
- 436 Swanston CW, Caldwell BA, Homann PS, Ganio L, Sollins P (2002) Carbon dynamics during long-term
  437 incubation of separate and recombined density fractions from seven forest soils. Soil Biol
  438 Biochem 34: 1121–1130

- 439 Swanston CW, Torn MS, Hanson PJ, Southon JR, Garten CT, Hanlon EM, Ganio L (2005) Initial
   440 characterization of processes of soil carbon stabilization using forest stand-level radiocarbon
   441 enrichment. Geoderma 128: 52–62
- Tan Z, Lal R, Owens L, Izaurralde RC (2007) Distribution of light and heavy fractions of soil organic
  carbon as related to land use and tillage practice. Soil Tillage Res 92: 53–59
- 444 Tipping E, Somerville CJ, Luster J (2016) The C:N:P:S stoichiometry of soil organic matter. Biogeochem
  445 130: 117-131
- Toberman H, Adams J, Tipping E, Schillereff D, Somerville C, Coull M, Helliwell R, Carter H, Guyatt H,
  Keenan P, Lawlor A, Pereira MG, Patel M, Tanna B, Thacker S, Thomson N, Owens J, Gibbs S,
  Smith D, Bryant C, Elliot F, Gulliver P (2016). Soil survey in England, Scotland and Wales carried
  out during 2013 and 2014 [LTLS]. NERC Environmental Information Data Centre.
  https://doi.org/10.5285/17bebd7e-d342-49fd-b631-841ff148ecb0
- Trumbore SE (1993) Comparison of carbon dynamics in tropical and temperate soils using radiocarbon
   measurements. Global Biogeochem Cycles 7: 275–290
- 453 Turner BL, Cade-Menun BJ, Condron LM, Newman S (2005) Extraction of soil organic phosphorus.
  454 Talanta 66: 294–306
- Walthert L, Zimmermann S, Blaser P, Luster J, Lüscher P (2004) Waldböden der Schweiz Vol 1, Hep
  Verlag, Bern, 768 pp
- Wick B, Tiessen H (2008) Organic matter turnover in light fraction and whole soil under silvopastoral
  land use in semiarid northeast Brazil. Rangeland Ecol Management 61: 275–283
- Yang X, Post WM (2011) Phosphorus transformations as a function of pedogenesis: A synthesis of soil
   phosphorus data using Hedley fractionation method. Biogeosci 8: 2907–2916
- Zimmermann S, Luster J, Blaser P, Walthert L, Lüscher P (2006) Waldböden der Schweiz Vol 3, Hep
  Verlag, Bern, 848 pp
- Zimmermann, M. Leifeld J, Schmidt MWI, Fuhrer J (2007) Measured soil organic matter fractions can
  be related to pools in the RothC model. Eur J Soil Sci 58: 658–667
- 465

- 466 Tables

## 468 Table 1. Abbreviations

Abbreviation	Full title
С	Carbon
HF	heavy fraction
IP	inorganic phosphorus
LF	light fraction
Ν	Nitrogen
NaPT	sodium polytungstate
NLF	non-occluded light fraction
NPSOM	nutrient-poor soil organic matter
NRSOM	nutrient-rich soil organic matter
OC	organic carbon
OLF	occluded light fraction
ON	organic nitrogen
OP	organic phosphorus
Р	Phosphorus
RSD	relative standard deviation
SD	standard deviation
SOM	soil organic matter
SRP	soluble reactive phosphorus
TDP	total dissolved phosphorus
TN	total nitrogen
тос	total organic carbon
TP	total phosphorus

474 Table 2. Summary of element recoveries.

Element	Range %	Mean %	SD %	Range %
IP	28-486	117	105	28-486
ТР	19-121	62	30	19-121
OP	17-124	56	29	17-124
OC	61-123	91	18	61-123
TN	70-110	91	12	70-110

Table 3. Concentrations of OC and ratios of N and P to C (g g<sup>-1</sup>) in SOM of the LF and HF, for the 20 soil
samples. The LF and HF ratios are significantly different in all cases (t-test; p < 0.001). C:N and C:P</li>
ratios are also shown.

	LF			HF		
	range	mean	SD	range	mean	SD
%OC	26.5-45.5	36.1	4.4	3.2-15.7	8.1	3.7
N:C	0.027-0.054	0.040	0.009	0.039-0.106	0.074	0.017
P:C	0.00029-0.0019	0.00084	0.00042	0.00080-0.0139	0.0049	0.0033
C:N	18.4-37.0	26.1	6.1	9.4-25.4	14.7	4.2
C:P	1260-3430	1490	730	72-1260	337	284

485	Figure captions	
-----	-----------------	--

Fig. 1 Schematic of the fractionation procedure. Key: NLF non-occluded light fraction, OLF occludedlight fraction, LF light fraction, HF heavy fraction.

489

Fig. 2 Concentrations of total, inorganic and organic phosphorus (TP, IP, OP) in the light and heavy
fractions (LF and HF) of the 20 soil samples. The error bars indicate standard errors. Key: B broadleaf
woodland, C conifer plantation, H heathland, IG improved grassland, R rough grassland; d subsoil, s
topsoil.

494

Fig. 3 Percentage of OC, TN and OP in the heavy fraction (HF) vs percentage of soil mass in the HF.
Data for two "outliers" (H1s, H4d) that do not fit the general pattern are indicated by dashed outlines.

Fig. 4 Variations of N:C and P:C in SOM with %C for the light fraction (LF, open circles) and the heavy fraction (HF, filled circles); ON is assumed equal to TN. The lines are predictions from the two endmember mixing model of Tipping et al. (2016). The full range of %C is plotted to show end-member ratios at  $\leq 0.1$  %C and  $\geq 50\%$  C. Data for two "outliers" (H1s, H4d) that do not fit the general pattern for HF are indicated by dashed outlines.



516 Fig. 1



518 Fig. 2





527 Fig. 4