

1 **Phytoplankton dynamics from the Cambrian Explosion to the onset of the Great**
2 **Ordovician Biodiversification Event: a review of Cambrian acritarch diversity**

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16
17 **Abstract:** Most early Palaeozoic acritarchs are thought to represent a part of the marine
18 phytoplankton and so constituted a significant element at the base of the marine trophic chain
19 during the 'Cambrian Explosion' and the subsequent 'Great Ordovician Biodiversification Event.'
20 Cambrian acritarch occurrences have been recorded in a great number of studies. In this paper,
21 published data on Cambrian acritarchs are assembled in order to reconstruct taxonomic diversity
22 trends that can be compared with the biodiversity of marine invertebrates. We compile a database
23 and calculate various diversity indices at global and regional (i.e. Gondwana or Baltica) scales. The
24 stratigraphic bins applied are at the level of the ten Cambrian stages, or of fourteen commonly used
25 biozones in a somewhat higher resolved scheme. Our results show marked differences between
26 palaeogeographical regions. They also indicate limitations of the data and a potential sampling

27 bias, as the taxonomic diversity indices of species are significantly correlated with the number of
28 studies per stratigraphic bin. The total and normalized diversities of genera are not affected in the
29 same way. The normalized genus diversity curves show a slow but irregular rise over the course of
30 the Cambrian. These also are the least biased. A radiation of species and to a lesser extent of genera
31 in the ‘lower’ Cambrian Series 2 appears to mirror the ‘Cambrian Explosion’ of metazoans. This
32 radiation, not evident on Gondwana, is followed by a prominent low in species diversity in the
33 upper Series 3 and lower Furongian. Highest diversities are reached globally, and on both Baltica
34 and Gondwana, in the uppermost Cambrian Stage 10, more precisely in the *Peltura* trilobite Zone,
35 preceding a substantial phase of acritarch species extinction below and at the Cambrian/Ordovician
36 boundary. Nearly all the genera present in Stage 10 survived into the Ordovician. The forms that
37 emerged during the Cambrian therefore became the foundation for the more rapid radiation of
38 acritarchs during the ‘Great Ordovician Biodiversification Event’.

39
40 **Keywords:** phytoplankton; acritarchs; Cambrian; biodiversity; paleobiogeography

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43 **1. Introduction**

44 The Cambrian fossil record is marked by the well-known ‘Cambrian Explosion’ (or radiation),
45 which is characterized by the appearance of most metazoan phyla in a seemingly short interval (e.g.
46 Conway Morris, 2000). Although molecular clock estimates now indicate a Proterozoic origin for
47 many lineages (Erwin et al., 2011), the Cambrian Explosion is still considered to be one of the most
48 important periods in the history of life. It was followed in the Ordovician by the ‘Great Ordovician
49 Biodiversification Event’ (GOBE), an episode of rising taxonomic diversity in most marine
50 organisms and of increasingly complex ecosystems that were more diverse and differentiated than
51 those of the Cambrian, with a wider range of ecological niches (Webby et al., 2004; Bambach et al.,
52 2007). Whether these two episodes were in fact separate, or should be considered as one long

53 period of diversification, is currently debated (Alroy et al., 2008), as are their possible causes and
54 mechanisms.

55

56 Phytoplankton is an important constituent at the base of the foodweb today and in this capacity may
57 have played a key role in the early Palaeozoic diversification events (Debrenne and Zhuravlev,
58 1997; Butterfield, 1997). It has been argued that a higher concentration of phytoplankton in the
59 early Palaeozoic oceans triggered the major evolutionary events and had an important impact on
60 metazoan diversification. Butterfield (1997), for example, argued that the interaction between
61 phytoplankton and metazoans fuelled the Cambrian Explosion after the appearance of
62 mesozooplankton. Similarly, Servais et al. (2008, 2010) argued that the evolution of the
63 phytoplankton triggered a 'plankton revolution' through the Cambrian–Ordovician boundary
64 interval at the beginning of the GOBE. The expanding and increasingly diverse phytoplankton
65 could have served as food for the developing zooplankton, but also for various clades of suspension
66 feeders and detritus feeding organisms. Detailed knowledge on the development of biomass,
67 taxonomic and morphological diversity of phytoplankton would be useful to test this hypothesis.
68 Here we concentrate on taxonomic diversity.

69

70 The phytoplankton of early Palaeozoic oceans is generally considered to be present among the
71 acritarchs, which are defined as organic-walled microfossils of uncertain biological affinity (Evitt,
72 1963). Consequently, the acritarchs are a polyphyletic group and have a long stratigraphical range.
73 A number of organisms originally considered to be acritarchs now have established affinities with
74 other groups, including prasinophycean and zygnematacean green algae, cyanobacteria,
75 dinoflagellates, multicellular algae (Butterfield, 2004; Mendelson, 1987), fungi (Butterfield, 2005)
76 and even metazoans (e.g. schizomorphae: Van Waveren, 1992; Van Waveren and Marcus, 1993;
77 *Ceratophyton*: Kiryanov in Volkova et al. 1979; Fatka and Konzalová, 1995; large spinose
78 Ediacaran microfossils: Cohen et al., 2009). Although technically no longer acritarchs (Evitt,

79 1963), they are still occasionally referred to as such. Nevertheless, most of the remaining
80 Palaeozoic acritarchs are considered to be cysts of marine, generally planktonic unicellular algae,
81 which are often globally distributed. We here use the term ‘acritarchs’ to include all organic-walled
82 microfossils of possible phytoplankton origin, including acritarchs *s.s.* and organic microfossils that
83 are now attributed to the green algae and cyanobacteria. Where preserved, they are often recovered
84 in large numbers, even from small samples, resulting in an abundant record and frequent application
85 in biostratigraphy (e.g. Martin and Dean, 1981; 1988; Moczyłowska, 1991; 1998; Vanguetaine
86 and Van Looy, 1983).

87

88 Biomarker evidence suggests that Palaeozoic acritarchs may include forms that have a close
89 biological affinity with (or be evolutionary precursors of) dinoflagellates (Moldowan and Talyzina,
90 1998; Talyzina et al., 2000). However, the oldest unambiguous dinoflagellates in the fossil record
91 are from the Triassic (e.g. Fensome et al., 1993) and an alternative view is that the Palaeozoic
92 acritarchs represent a “green” phytoplankton lineage (Martin et al., 2008). Observations of recent
93 dinoflagellates have shown that only some species form organic-walled cysts with the potential to
94 fossilise (Dale, 1976), and may do so at more than one stage in their life cycle (Fensome et al.,
95 1996a). If, by analogy, Palaeozoic acritarchs are also cysts, perhaps of various algal groups, there is
96 a clear implication that they only represent a part of the phytoplankton.

97

98 A further difficulty is that acritarch taxa are morphospecies rather than true biological species. In
99 some instances, morphological gradations from one Palaeozoic acritarch taxon to another have been
100 demonstrated, suggesting ecophenotypic variation rather than speciation (Servais et al., 2004b). If
101 so, the number of taxa recorded would overestimate the number of biological species. Conversely,
102 for the rather featureless sphaeromorph acritarchs, often identified simply as ‘sphaeromorphs’ or
103 ‘*Leiosphaeridia* spp.’, the number of taxa recorded probably underestimates biological diversity.
104 Despite these limitations, acritarchs remain the main source of information for phytoplankton in the

105 Cambrian, long before the appearance of planktonic algae forming calcitic and siliceous skeletons
106 during the Mesozoic (Bown, 2005; Sims et al., 2006).

107

108 Various Palaeozoic acritarch diversity curves have been published previously. Detailed global
109 curves are available for the Carboniferous (Mullins and Servais, 2008), Lochkovian to Tournaisian
110 (Klug et al. 2010) and the Permian (Lei et al., 2013). Regional diversity curves have been
111 published for the Ordovician on the Yangtze Platform (South China) and North Africa (Algeria,
112 Tunisia and Libya; also including the latest Cambrian; Servais et al. 2004a), for ‘northern
113 Gondwana’ (including North Africa, Turkey, Saudi Arabia and several peri-Gondwanan
114 microcontinents; Vecoli and Le Hérisse, 2004) and for Baltica (Hints et al., 2010). Li et al. (2007)
115 published a diversity curve for genera in the Ordovician of South China, North China and Tarim.

116

117 Acritarch diversity curves that include the Cambrian have been published by Tappan and Loeblich
118 (1972; 1973; genera, Precambrian to present), Vidal and Knoll (1982; species, upper Riphean to
119 lower Cambrian), Knoll (1994; species, Proterozoic to lower Cambrian), Strother (1996;
120 Precambrian and Phanerozoic, using data of Downie, 1984, and Fensome et al., 1990), Vidal and
121 Moczyłowska (1997; species, Proterozoic to Cambrian), Moczyłowska (1998; species, Cambrian
122 in Upper Silesia; and 2011; species, Ediacaran to basal Ordovician, global and Baltica), Zhuravlev
123 (2001; genera, Cambrian to Tremadocian), Katz et al. (2004; species, Proterozoic to Neogene, and
124 genera, Cambrian to Neogene), Huntley et al. (2006a; 2006b; genera, Proterozoic to Cambrian),
125 Michaud in Strother (2008; genera, Phanerozoic), and Servais et al. (2008; species, middle
126 Cambrian to Llandovery). Katz et al. (2004) and Michaud in Strother (2008) used the Palynodata
127 database (see Fensome et al., 1996b) to create their diversity curves. Palynodata had been compiled
128 by a consortium of oil companies and scientific institutions over the course of three decades and
129 was discontinued in 2006. Mullins et al. (unpublished) compiled the PhytoPal database of acritarch
130 occurrences covering the interval from the Cambrian to the Triassic (mainly Ordovician to

131 Devonian), data that were partly used by Servais et al. (2008), Klug et al. (2010) and
132 Moczyłowska (2011).

133

134 All these publications present a (usually global) total diversity (see 2.3). Knoll (1994) and Knoll et
135 al. (2006) also discussed the diversity of species in single assemblages through time between the
136 Proterozoic and the early Cambrian. The resolution varies between a single value for the whole
137 Cambrian (Tappan and Loeblich, 1972; 1973; Strother, 1996) and more than 20 intervals
138 (Zhuravlev, 2001).

139

140 Acritarch diversity analyses that include the Proterozoic-Cambrian transition at sufficient resolution
141 show fairly high global total diversities in the late Neoproterozoic, followed by decreasing diversity
142 towards the end of the Ediacaran (Vidal and Knoll, 1983; Knoll, 1994; Vidal and Moczyłowska,
143 1997; Knoll et al., 2006). Several studies found a diversification during the early Cambrian to a
144 peak around the late early/early middle Cambrian (Strother, 1996; Vidal and Moczyłowska, 1997;
145 Moczyłowska, 1998; Zhuravlev, 2001). The Ordovician, when it is included, is marked by
146 generally higher diversity than the Cambrian.

147

148 Diversity studies on Cambrian acritarchs published so far have had limitations in the choice of
149 sources and in the analytical methods they applied, which may have allowed possible biases to go
150 undetected, making them vulnerable to a certain degree of criticism. The goal of this study is to
151 produce a robust database of acritarch occurrences in the Cambrian by compiling as much data as is
152 feasible, in order to approximate their standing diversity using various methods, to identify
153 evolutionary trends, and to test the results for biases. In particular, we address whether the
154 Cambrian Explosion is reflected in acritarch diversity and whether the GOBE was preceded by a
155 diversification of acritarchs, a temporal relation which would be a primary requirement in support
156 of a cause-and-effect hypothesis. If phytoplankton had an effect on or was affected by these
157 radiations, it might be expected that this would be reflected in acritarch diversity, and that, in turn,

158 acritarch diversity might indicate general trends in phytoplankton biodiversity, assuming that other
159 effects could be ruled out.

160

161

162 **2. Materials and Methods**

163 *2.1. Source data*

164 The literature on Cambrian acritarchs includes over 500 publications, but many studies do not
165 include descriptions or precise biostratigraphical data. Occurrence data for acritarchs in this study
166 are derived from 103 studies (see supporting material S1, S2). All available publications with
167 consistent information on the presence of acritarch taxa in the Cambrian were included in the
168 database, but only 72 of these studies reported assemblages with stratigraphic information that
169 could be correlated precisely to any of the stratigraphic units we use (see 2.2., Fig. 1). We only
170 included the latter in our subsequent diversity analysis.

171

172 Of the 72 source publications, 39 describe data from Gondwana (including various microcontinents
173 on its periphery, most importantly 12 from Avalonia), 31 from Baltica, 11 from Laurentia, 11 from
174 South China, 5 from Bruno-Silesia (or the Brunovistulicum), 4 from the Holy Cross Mountains (or
175 the Małopolska 'Block'), 2 each from Tarim and Siberia, 1 from Kara and 1 from North China.

176 Figure 2 shows the palaeogeographical positions of study areas of publications included in our
177 analysis, using the base maps of Torsvik and Cocks (2013). The only distinct regions with
178 sufficiently extensive records to allow individual diversity analyses for most of the Cambrian are
179 Baltica and that part of (peri-)Gondwana that includes northwestern Africa, Avalonia, Iberia and
180 Sardinia and which was positioned near the South Pole during the Cambrian (Hartz and Torsvik,
181 2002).

182

183 Many published studies only list selected, biostratigraphically useful taxa or those typical of an

184 assemblage. As a consequence, it is possible that we have undersampled the full range of species
185 and, by the nature of the fossil record, their actual stratigraphical ranges in the Cambrian.

186 Nevertheless, we consider our database to be representative of the current state of research.

187

188 2.2. Database

189 Our database (supplementary material S1) contains exactly 6000 entries reporting the
190 presence/absence (not abundance) of acritarch taxa within the studied stratigraphic intervals. Each
191 entry contains information on the identification of the taxon in question, the entry's source study,
192 study area and stratigraphic position (with lower and upper limits). Taxonomic information
193 includes genus name; species epithet or placeholder; qualifiers such as '?', 'cf.' and 'aff.', where
194 applicable, for either the genus or species; authorship; and the original identification if it has been
195 revised. Generally, we relied on the identifications and age-assignment in the original studies or
196 published later revisions. Obvious and reported synonymies were taken into account (see
197 supplementary material S3). The applied names follow the latest usage, without any intention on
198 our part to validate or revise them. This approach purely serves to create a taxonomic framework
199 that reflects the current state of research with as much consistency and as little redundancy as
200 possible.

201

202 We use two schemes of stratigraphic subdivisions or bins in the database. One scheme corresponds
203 to the ten stages of the International Chronostratigraphic Chart (state 2014; Cohen et al., 2013,
204 updated), and the other to fourteen biozones (Fig. 1). The latter are mainly trilobite biozones. The
205 Terreneuvian ('pre-trilobite' Cambrian) stages are dated by trace fossils or small shelly fossils
206 (Peng et al., 2012). The first appearance of the trace fossil *Trichophycus* (or alternatively
207 *Treptichnus*, *Phycodes* or *Manykodes*) *pedum* marks the base of the Cambrian System, the
208 Terreneuvian Series and the Fortunian Stage (Brasier et al., 1994); the base of the upper stage in the
209 Terreneuvian Series (Cambrian Stage 2) has not yet been defined. The choice of stratigraphic bins

210 was a compromise guided by the resolution required to address our research question, the available
211 data, and the maximal precision with which data can be attributed to a given stratigraphic interval in
212 the Cambrian. Due to imperfect correlation, some data sets could be assigned to biozones but not
213 stages or *vice versa*.

214

215 In order to calculate origination and extinction (see section 2.3.) for the basal and top Cambrian
216 bins, we had to consider the presence of acritarch taxa during Proterozoic and post-Cambrian times.
217 Data were derived from some of the sources mentioned above (section 2.1., Figs. 1, 2), the acritarch
218 and prasinophyte index of Fensome et al. (1990), and, for the post-Cambrian, the unpublished
219 PhytoPal database by Mullins et al.

220

221 2.3. Methods

222 Diversity can be measured in different ways and for any taxonomic rank. In the case of acritarchs,
223 treated as *incertae sedis*, only species and genera have a formal rank. Both are included in our
224 analysis. Acritarch genera counted herein include reports of species assigned under open
225 nomenclature (e.g. as ‘sp.’) to a genus or with an uncertain specific identification, but exclude
226 species that are only tentatively assigned to a genus (see supplementary material S3). Species
227 counted exclude those with uncertain identification, but include records that are only tentatively
228 assigned to any particular genus (see supplementary material S6-S11). After this selection process
229 and under the stratigraphic constraints discussed above (see section 2.2.), our dataset records the
230 ranges of 173 genera and 404 species.

231

232 Taxa are counted as present in a stratigraphic interval in one of four ways (Fig. 3): as crossovers
233 ranging through the entire interval; ranging into and going extinct in that interval; originating within
234 the interval and ranging beyond its upper boundary; and as singletons, with a range confined to an
235 interval, meaning that they originate and go extinct within the interval (Foote, 2000).

237 The actual number of coexisting taxa is the standing diversity (Sepkoski, 1975), and can only be
238 estimated for palaeontological data. The sampled-in-bin diversity (Figs. 4, 5) gives the total count
239 of taxa reported from a given stratigraphic interval. This index often underestimates diversity due
240 to the incompleteness of the fossil record and sampling bias. Total diversity (Figs. 4, 5, 8, 9) is
241 calculated by interpolating discontinuous taxon ranges from established occurrences (Foote, 2000),
242 i.e. taxa not recorded from an interval are counted as being present if they occur in intervals below
243 and above. With complete data, total diversity is likely to exceed the standing diversity at any
244 specific point in time, because it does not account for extinctions within an interval (Cooper, 2004).
245 Normalized diversity (Figs. 4, 5, 9) is calculated as the number of species ranging through an
246 interval (crossovers), plus half the number of taxa that originate and/or become extinct in that stage
247 (Sepkoski, 1975; Cooper, 2004). According to Cooper (2004), normalized diversity is a good
248 approximation of the mean standing diversity in a stratigraphic bin.

249

250 Indices of origination and extinction represent, respectively, the number of species originating and
251 becoming extinct in a stratigraphic interval. Turnover in an interval is the sum of origination and
252 extinction events. These indices of taxonomic change can be calculated as including or excluding
253 singletons. In Figure 6, singletons are excluded.

254

255 The use of stages and trilobite zones as the stratigraphic divisions in our analysis means that bins in
256 both schemes are inevitably of different durations, and longer bins may have a higher diversity than
257 shorter bins simply because they are sampling longer intervals, while at the same time obscuring
258 short-term excursions. As the Cambrian stages vary in duration from about three million years (e.g.
259 Guzhangian, Paibian) to twelve million years (Fortunian), a considerable effect may be expected.
260 To correct for a possible bias resulting from these unequal interval lengths, rates of origination,
261 extinction and turnover have been computed. They are equal to the origination, extinction and

262 turnover indices divided by the stage duration in Myr (from Peng et al., 2012; Fig. 6 B, D).

263

264 Vidal and Moczyłowska (1997) first applied poly-cohort survivorship to Cambrian acritarchs.

265 Poly-cohort survivorship measures the percentage of species in a cohort, i.e. species present in a

266 given stratigraphic interval that are still present in later intervals (Fig. 7A). In contrast, poly-cohort

267 pre-nascence (also called backward survivorship) measures the proportion of a cohort present in

268 earlier intervals (Fig. 7B; e.g. Raup, 1978; Foote, 2001). Poly-cohorts of survivorship and

269 pre-nascence respectively are calculated for all stratigraphic bins considered herein and plotted on a

270 logarithmic scale. The slope of the cohort curves then represents the rate of extinction or

271 origination for each cohort, with a linear curve (on a logarithmic scale) implying constant rates,

272 while changes over time or between cohorts can reveal changes in rates or biases.

273

274 We evaluate the effect of sampling bias and test for evolutionary links between acritarchs and

275 marine invertebrates (diversity values for marine invertebrate taken from the supporting information

276 of Na and Kiessling, 2015) by calculating Spearman's ρ (*rho*) or r_s – a non-parametric rank order

277 correlation coefficient (e.g. Press et al., 1992) – for correlations between diversity indices and the

278 number of studies per interval (Fig. 9) and between the sampled-in-bin genus diversities of

279 acritarchs and animals (Fig. 11), respectively. This metric is commonly used to identify bias in the

280 fossil record (e.g. Dunhill et al., 2012; Na and Kiessling, 2015). The value of r_s varies between -1

281 and 1 for perfect negative and positive correlation, respectively, with 0 indicating absence of

282 correlation. The statistical significance is given by the probability value p , which is derived from

283 permutation tests. A correlation is accepted as strong if the coefficient is high (> 0.75) and

284 significant if the probability value is low (< 0.05). In this case, a bias is possible. The results are

285 here rounded to two and to three significant digits for r_s and p , respectively. We also investigate the

286 changing strength of correlation through the studied stratigraphic intervals by applying a moving-

287 window approach (Fig. 11B).

288

289 Subsampling methods are often used in biodiversity analyses in order to address sampling bias.
290 These methods standardize for number of occurrences, number of studies, taxon frequency, or rock
291 volume (Alroy, 2010; Hannisdal and Peters, 2011). The resulting trends are usually less biased than
292 diversity based on raw counts, but they can introduce new distortions, depending on the premise of
293 the respective approach (Alroy, 2010). The applicability of such methods on the Cambrian
294 acritarch record might be explored in the future. Here we test for the existence of biases with other
295 methods.

296

297 The analyses are performed with a customised program using the statistical environment R (version
298 3.0.2; R Core Team 2013).

299

300

301 **3. Results**

302 *3.1. Global*

303 *3.1.1. Diversity by stages*

304 Global diversity curves through the ten Cambrian stages are shown in Figures 4A and 4B, for
305 species and genera respectively. Relatively few species range from the Proterozoic into the
306 Cambrian and global total species diversity (Fig. 4A) is at its lowest in the Terreneuvian Series
307 (Fortunian Stage and Stage 2). It is high in Stages 3, 4 and 5 and in Stage 10. The upper Series 3
308 (Drumian and Guzhangian stages) and the lower Furongian Series (Paibian and Jiangshanian stages)
309 are marked by low global species diversity. Normalized species diversity peaks are present in
310 Stages 5 and 10. The number of crossover species rises unsteadily over the course of the Cambrian.
311 Singletons are mostly minor constituents, except in Stage 10 where they constitute more than a third
312 of the total diversity.

313

314 Sampled-in-bin genus diversity (Fig. 4B) follows trends similar to those of the sampled-in-bin
315 species diversity (Fig. 4A). The total genus diversity, incorporating a high number of range-
316 through extensions for genera that are not present in a stage, but which are present in underlying
317 and overlying stages, rises more or less steadily from Stage 2 onwards, with a minor drop between
318 Stage 5 and the Drumian, barely hinting at the main trends of the species curve. This effect is even
319 more conspicuous in the normalized diversity curve. The highest point of global genus diversity is
320 reached in Stage 10. The proportion of crossovers is high in relation to the total genus count and in
321 comparison to taxonomic changes. Singletons are rare or absent. Perhaps somewhat counter-
322 intuitively, total genus diversity is higher than total species diversity in the Terreneuvian stages and
323 the Guzhangian Stage (see Discussion). They are equal in the Jiangshanian Stage.

324

325 3.1.2. Diversity by biozones

326 The biozonal curves are similar to the stage curves. Using trilobite biozones as stratigraphic bins
327 above Stage 2 (Fig. 5), total and sampled-in-bin species diversity rises from the base of the
328 Cambrian to highs between the *Holmia* and *Paradoxides paradoxissimus* zones, followed by low
329 values between the *Paradoxides forchhammeri* and *Leptoplastus* zones, with a peak in the *Olenus*
330 Zone, and reaching another peak in the *Peltura* Zone. Diversity drops slightly from the *Holmia*
331 Zone to the *Protolenus* Zone, in the Jiangshanian between the *Parabolina spinulosa* and
332 *Leptoplastus* zones, and from the *Peltura* Zone to the *Acerocare* Zone in Stage 10. The first two
333 falls in diversity are also evident on the stage curves, although not so pronounced; the last, in Stage
334 10, is not. The high number of singletons in Stage 10 on the stage scale is reduced to moderate
335 numbers on the biozone scale.

336

337 In six of fourteen biozones (corresponding to the Terreneuvian stages and parts of Series 3 and the
338 Furongian), total genus diversity is higher than total species diversity (Fig. 8). In two more (the
339 *Schmidtiellus mickwitzi* and *Olenus* zones), it is barely lower.

340

341 *3.1.3. Taxonomic changes*

342 Origination indices for species and genera reach peaks in Stages 3 and 10 (Figs. 6A, C). The rate of
343 origination per Myr is only moderate for Stage 3, and that for Stage 10 appears higher in
344 comparison (Figs. 6B, D). The number of originating genera is also high in the Fortunian, while the
345 rate of origination is relatively low. Otherwise, the rates of taxonomic changes per Myr are similar
346 to the raw indices. Species extinctions are by far highest in Stage 5, discounting the extraordinarily
347 high number of singletons in Stage 10 (Fig. 4A). Almost no extinctions of species or genera occur
348 in the Fortunian, Stage 3 and Guzhangian stages. Stages 5 and 10 contain the highest number of
349 species turnovers from the raw data (Fig. 6A). Corrected for stage duration, turnover is also high in
350 the Paibian Stage (Fig. 6B).

351

352 Poly-cohorts of survivorship and pre-nascence produce almost parallel and relatively straight curves
353 (Fig. 7), indicating that rates of origination and extinction are relatively constant throughout the
354 Cambrian. The poly-cohorts of survivorship for the Terreneuvian stages have a gentler slope than
355 younger ones, indicating a greater number of long-ranging species. Otherwise the poly-cohorts
356 show no drastic differences in diversification or extinction trends between cohorts or over time.

357

358 *3.2. Regional trends*

359 The regional diversity curves are incomplete due to the limited sources and difficulties in
360 correlations. The curves for genera are not figured or discussed separately, as they are very similar
361 to the species curves, albeit with lower values (supplementary material S12, S13).

362

363 *3.2.1. Baltica*

364 The diversity curve for Baltica (Figs. 4C, 5C) mostly mirrors the global trends. Notable differences
365 from the global curve are the overall lower diversity, its exceptionally low and unchanging values in

366 the Fortunian Stage and Stage 2, and a slight rise in the Jiangshanian Stage, preceding the ultimate
367 diversity maximum in Stage 10. No data could be assigned to the Drumian. In the biozonal curve,
368 the *Olenus* Zone marks a minor diversity peak and is separated by a gap in the record in the
369 *Parabolina spinulosa* Zone and a minor diversity low in the *Leptoplastus* Zone from the point of
370 maximum diversity in the *Peltura* Zone. This is followed by a lowered diversity in the latest
371 Cambrian *Acerocare* Zone.

372

373 3.2.2. Gondwana

374 There are no published data from the Terreneuvian Series of Gondwana, so acritarch diversity at
375 that level cannot be evaluated. Also, we could not assign any data with certainty to Stage 3, the
376 *Schmidtellus mickwitzii* Zone and the *Leptoplastus* Zone. Species diversity on the Gondwana
377 margin (Figs. 4D, 5D) is very low in Stages 4 and 5, in contrast to the high diversity of the Baltic
378 and global curves. Above Stage 5, Gondwanan diversity is similar to, or higher than that on
379 Baltica. Diversity peaks in the Drumian Stage (Fig. 4D) or in the *Paradoxides paradoxissimus*
380 Zone (Fig. 5D), but corresponds to declining diversity in the global and Baltica curves. In parallel
381 to the situation on Baltica, the Guzhangian and Paibian stages are marked by relatively low
382 diversities, followed by a slight rise in the Jiangshanian Stage and a strong rise to an overall
383 maximum in Stage 10 (Fig. 4D). The same trend is seen on the biozonal curve (Fig. 5D), except
384 that the late Cambrian peak in the *Peltura* Zone is followed by a fall in the *Acerocare* Zone.

385

386 3.3. Distribution of studies over time

387 The number of studies in each of the stratigraphic intervals considered is very irregular and
388 particularly low for Stage 2, the *Agnostus pisiformis* Zone, the *Leptoplastus* Zone and the
389 *Acerocare* Zone (Fig. 8). The distribution of studies over the Cambrian has a conspicuous
390 correlation with species diversity (Fig. 8), as shown by the strong and significant correlation for
391 both total ($r_s = 0.95$; $p < 0.001$) and normalized ($r_s = 0.93$; $p < 0.001$) species diversity based on

392 stages (Figs. 9A, B). The correlation is weaker, but significant for species on a biozonal level (total:
393 $r_s = 0.76$; $p = 0.002$; normalized: $r_s = 0.57$; $p = 0.032$, Fig. 9C) and for genus total diversity based
394 on stages ($r_s = 0.78$; $p = 0.008$). In contrast, the correlation is both weak and insignificant for
395 normalized diversity of genera by stage ($r_s = 0.52$; $p = 0.121$) and total diversity of genera by
396 biozone ($r_s = 0.35$; $p = 0.214$). The Gondwanan total species diversity based on biozones ($r_s = 0.55$;
397 $p = 0.098$) and normalized species diversity based on stages ($r_s = 0.58$; $p = 0.306$) also only show
398 weak and not significant correlations. Global normalized genus diversity by biozone shows no
399 correlation ($r_s = -0.11$; $p = 0.708$, Fig. 9D).

400

401

402 **4. Discussion**

403 *4.1. Global diversity*

404 The total species diversity curve presented here is strikingly similar to that published by Vidal and
405 Moczyłowska (1997), although the latter was based on fewer sources and diversity levels are
406 mostly higher in our curves (Fig. 8). The same basic trends – rising diversity in the ‘lower’
407 Cambrian followed by low diversity and subsequent rise at the end of the Cambrian – also appear in
408 Strother's (1996) and Zhuravlev's (2001) diversity curves. This does not exclude the possibility of
409 biases, but would at least imply consistent biases, such as the concentration of productive studies in
410 certain areas and stratigraphic intervals. In particular, the northwestern part of Baltica is
411 disproportionately well studied and studies on Gondwana are essentially limited to the western and
412 southern margins (Fig. 2). Also, studies on the pre-trilobite Cambrian are usually poorly dated. In
413 fact, the uneven availability of index fossils in general might present a bias, since an important
414 amount of data remains simply unusable due to a lack of stratigraphic constraints.

415 Considering the low number of studies that deal specifically with Stage 2 and the *Agnostus*
416 *pisiformis*, *Leptoplastus* and *Acerocare* zones, all of which coincide with decreased diversity,
417 results from these units should be regarded with some caution. The conspicuous correlation

418 between total species diversity and the number of studies per stratigraphic bin (Figs. 8, 9) suggests
419 the influence of an important sampling and/or preservational bias.

420

421 The same would be true for the sampled-in-bin genus diversity, but in this case the interpolation of
422 ranges in the total diversity curve changes the picture profoundly. A high number of genera that are
423 missing from the low diversity intervals of Stage 2 and from the Drumian to Jiangshanian stages
424 (and the corresponding biozonal units) are present in the preceding and succeeding higher diversity
425 intervals. These genera are inferred to range through the low diversity intervals and so are also
426 inferred to contribute to total generic diversity in those intervals. The result is a decoupling of the
427 total and sampled-in-bin diversity curves (Figs. 4B, 5B), indicating a significant incompleteness of
428 the record. This would certainly also affect species diversity and perhaps even more severely.

429

430 As a consequence of the obvious incompleteness of the record, it is conceivable that the total genus
431 diversity in the affected intervals is still underestimated and that the diversification of genera
432 actually continued throughout the whole Cambrian at a more or less constant rate (Figs. 4B, 5B). A
433 further consequence of the interpolation of generic occurrences to construct the total global genus
434 diversity curves is that the total genus diversity actually exceeds total species diversity in the less
435 extensively studied intervals of the Terreneuvian Series and parts of Series 3 and the Furongian
436 Series (Fig. 8). This is an artefact of the data, as well as highlighting some of the limitations, and is
437 explained by the generally longer ranges of genera being more likely to span gaps in the record. It
438 might be argued that if a genus is inferred to be present in a low diversity interval, based on its
439 interpolated range, then at least one species of that genus should also be present. The anomaly of
440 genus diversity exceeding species diversity would be removed by inferring the presence of at least
441 one species per genus, but the true number of missing species would be unknown.

442

443 The normalized genus diversity curve also indicates a more or less steady, if slow, diversification

444 (Fig. 4B). Regression analysis indicates that normalized genus diversity is the least biased (Fig.
445 9D). Genera are typically easier to identify than species since they are defined by broader
446 characteristics, and can often be determined when the species is unknown to the author or the
447 preservation is too poor to identify it. Taxonomic inflation may also be less of a problem at the
448 generic level for the same reasons. This means that the record of genera is potentially more
449 complete and the ranges more accurate than those of species. Generic diversity can therefore be
450 considered more robust than species diversity. However, it cannot show the full range of diversity
451 and may be a poor proxy for species diversity, because the number of species in a genus varies
452 greatly and is not constant over time. Furthermore, the possibility of polyphyly needs to be
453 considered for genera even more so than for species. Even so, based on current knowledge, the
454 generic record hints at an increasing diversification trend through the Cambrian. Prenascence
455 analysis of species also suggests an overall stable origination pattern (Fig. 7 B).

456

457 Global sea level is generally considered to have risen more or less constantly throughout the
458 Cambrian (Miller et al., 2005; Haq and Schutter, 2008), a plausible cause being the formation of
459 mid-ocean ridges following the breakup of Pannotia in the late Neoproterozoic (Miller et al., 2005;
460 Scotese, 2009). A correlation between long-term sea-level changes on Palaeozoic and Mesozoic
461 phytoplankton diversity has previously been reported, based on diversity curves for various groups,
462 including acritarchs (Katz et al., 2004). During transgressions, flooded continental areas, which are
463 the most favorable habitats for phytoplankton, increase and so provide more niche space and
464 opportunities for diversification, following a species-area relationship (e.g. Sepkoski, 1976; Katz et
465 al., 2004). The Cambrian acritarch record would be consistent with that idea. Flooding of
466 continental shelves also controls the accumulation of sedimentary rocks and provides a general
467 explanation for the correlation between biodiversity in the fossil record and available rock volume
468 (Hannisdal and Peters, 2011).

469

470 Bambach et al. (2004) noted that proportional extinction among metazoan genera during most of the
471 Cambrian and Early Ordovician was unusually high compared to the rest of the Phanerozoic. They
472 identified two phases of declining diversity due to low origination; the late Botomian and the early
473 late 'Middle' Cambrian. Na and Kiessling (2015) found that per-capita rates of extinction exceeded
474 origination in Stage 3, the Drumian, and Stage 10. Acritarch genera show such a signal only in the
475 Guzhangian and Drumian. The sampled-in-bin diversities of Cambrian animal (based on Na and
476 Kiessling, 2015) and acritarch genera are neither strongly nor significantly correlated ($r_s = 0.55$; $p =$
477 0.098 ; Fig. 11A), which means there currently is no evidence for a continuous, strong evolutionary
478 link between the two groups.

479

480 4.2. Regional trends

481 As shown in this paper and previously by others (Yin and Xue, 2002; Molyneux et al., 2013) from
482 more limited sources, regional differences do exist in the currently available data. Even for
483 acritarchs in the Cambrian, the global diversity does not necessarily reflect regional trends (or *vice*
484 *versa*), as for example on Gondwana in stages 4 and 5, where diversity is particularly low, in
485 contrast to the high contemporary diversity on Baltica (Figs. 4, 5, supplementary material S15).
486 Such differences are not evident above Stage 5. In a similar fashion, Moczydłowska's (1998) curve
487 for Upper Silesia shows diversification in the lower part of the Cambrian up to the *A. oelandicus*
488 biochron, with no earlier peak (see supplementary material S15). As pointed out by Molyneux et al.
489 (2013), important taxa were present on both Baltica and Avalonia. The primary difference between
490 Gondwana and Baltica, at least for the Cambrian, therefore lies not in the differentiation of
491 microfloras. The differences in diversity are quite probably at least enhanced by poor sampling of
492 the lower Cambrian of Gondwana. They may also indicate differences in the local
493 palaeoenvironment, rather than discrete provinces (see e.g. Lei et al., 2012, and references therein).

494

495 4.3. Cambrian Explosion

496 The apparent rapid diversification of acritarch species in Stage 3 parallels the radiation of
497 metazoans (Sepkoski, 1997, Na and Kiessling, 2015; Fig. 10). Such a correlation could point to a
498 link between phytoplankton and metazoans. For the Cambrian in total, the correlation between
499 metazoans and acritarchs is moderate ($r_s = 0.55$) and not significant ($p = 0.098$) (Fig. 11A). In
500 contrast, applying a moving window to each set of eight successive stages results in a good and
501 significant correlation ($r_s = 0.88$, $p = 0.004$) for the interval from Stage 2 to the Jiangshanian Stage
502 (9), inclusive (Fig. 11B). This correlation thus supports the presence of a more or less partial, but
503 important link between acritarch and metazoan diversities during the time of the Cambrian
504 Explosion. This result may be more or less modulated by a possible sampling bias, because most
505 acritarch diversity indices are strongly and significantly correlated with the distribution of studies
506 over time (Fig. 9). The dramatic increase in diversity in Stage 3 follows the particularly poorly
507 covered Stage 2, one of the stages where total genus diversity exceeds species diversity (Fig. 8).
508 The latter proves that this unit is critically undersampled and the following increase in diversity at
509 least exaggerated. The normalized genus curve, which we find to be the least biased, indicates that
510 acritarchs diversified without a notable peak. These trends, supported by the prenascence cohorts
511 (Fig. 7B), suggest that there is no evidence for a distinct or sudden origination phase in the first half
512 of the Cambrian. This does not negate the possibility of important interactions between certain
513 groups of acritarch-producing organisms and metazoans, but the relation between the Cambrian
514 Explosion and the diversity of the acritarchs as a whole remains unclear.

515

516 4.4. Furongian diversification and the GOBE

517 Servais et al. (2008) perceived a rise in phytoplankton diversity and disparity following the SPICE
518 $\delta^{13}\text{C}_{\text{carb}}$ event, which broadly coincides with the Paibian Stage and the *Olenus* trilobite Zone
519 (Saltzman et al., 2011). In our results, the Jiangshanian Stage, which follows the SPICE event, is
520 indeed marked by a slight rise in diversity in the regional species curves for both Baltica and
521 Gondwana (Fig. 4 C, D). This rise is not reflected on the global curves (Fig. 4A), which show a
522 decline in diversity from the Paibian Stage to the Jiangshanian Stage, but it is in the origination

523 index, which exceeds the extinction even at a global scale (Fig. 6A). It does not show up in the
524 total or sampled-in-bin diversity curves on a global scale because more species went extinct within
525 or at the end of the Paibian but are still counted in that stage, and because these are minor
526 fluctuations (Figs. 4A, 5A). There is no evidence that the Jiangshanian was a time of great
527 innovation, the amount of originating genera being very low. Many new genera with innovative
528 morphologies that reach into the Ordovician do appear later in Stage 10, several million years after
529 the SPICE event and without any obvious connection. The biozonal curve even shows a time of
530 exceptionally low origination (and extinction) in the upper Jiangshanian *Leptoplastus* Zone,
531 separating the minor lower Jiangshanian diversification from the massive origination phase in Stage
532 10, although this is certainly related to the very low number of studies treating the *Leptoplastus*
533 Zone as such (Fig. 8). By contrast, the diversity maximum in Stage 10 appears globally and on both
534 Baltica and Gondwana (Figs. 4C, D), which makes this short-lived phenomenon credible. It is
535 tempting to see the sudden rise in diversity as a precursor of the GOBE, but at the finer scale of
536 trilobite biozones, the diversity is already seen to fall from the *Peltura* Zone to the *Acerocare* Zone
537 (Fig. 5B, C). Only about half of the species recorded from Stage 10 range into the Tremadocian.
538 Thus the Cambrian seems to end with a phase of species extinction. However, few genera go
539 extinct at the Cambrian/Ordovician boundary. Animal genera do show a comparatively high per-
540 capita extinction rate in Stage 10 (Na and Kiessling, 2015). Metazoan and acritarch genus
541 extinctions seem to be generally unrelated in the Cambrian (see 4.1.). Among the acritarch genera
542 surviving the transition, several bore morphological traits that emerged during the late Cambrian
543 (Servais et al., 2008) and would be important contributors to later assemblages (such as the
544 ‘galeate’ acritarchs: Servais and Eiserhardt, 1995; Servais et al., 2004). The latest Cambrian can be
545 seen as a time of phytoplankton turnover, preceding but distinct from the radiation of acritarchs
546 during the Ordovician.

547

548

549 **5. Conclusions**

550 Our new, more complete dataset on Cambrian acritarchs provides new insights into the global and
551 regional diversification of phytoplankton in the Cambrian oceans. Global acritarch species
552 diversity starts on a comparatively low level at the base of the Cambrian, rises in parallel to the
553 radiation of metazoans to a high in stages 3 to 5 (or the *Holmia* to *Paradoxides paradoxissimus*
554 zones), then drops considerably to an extended low until it peaks sharply in Stage 10 (more
555 precisely in the *Peltura* Zone). Genera show an irregular increase in total and normalized diversity
556 throughout the Cambrian, indicating that the low species diversity in Stage 2 and the upper Series 3
557 to lower Furongian interval is at least in part due to an incomplete record. Correlation analysis
558 shows normalized genus diversity to be the least biased. The regional curves are markedly different
559 in Series 2 and Stage 5; while diversity is high on Baltica, it is low on Gondwana. This difference
560 may be attributed to the limited record on Gondwana and perhaps palaeoenvironmental differences
561 rather than distinct provinces. After Stage 5, the regional diversities are similar. A diversity
562 maximum in the *Peltura* Zone and subsequent fall in the *Acerocare* Zone is mirrored on both
563 Baltica and Gondwana. The end of the Cambrian is marked by an extinction of species, while
564 nearly all genera present in Stage 10 survived into the Ordovician.

565

566 Our acritarch diversity curves mirror the diversity curves of marine invertebrates to some extent.
567 The correlation between both diversities has statistical significance only when the first and last
568 stage of the Cambrian are excluded. Taken at face value, rising diversity in Stage 3 mirrors that of
569 the metazoans (Cambrian Explosion), but the acritarch diversity trends can mostly be attributed to
570 sampling bias. The relationship between the two groups remains uncertain. Similarly, although the
571 end of the Cambrian coincides with an extinction-led turnover of acritarch species, morphological
572 innovations from the Cambrian, particularly among genera, might well have paved the way for the
573 rapid diversification of acritarchs during the Ordovician that can be viewed as a part of the Great
574 Ordovician Biodiversification Event.

575

576

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585

586

587 **Appendix A. Supplementary data**

588

589 Supplementary material S1.

590 Database of Cambrian acritarch occurrences. (CVS)

591

592 Supplementary material S2.

593 List of source references, including bibliography, countries and palaeogeographical units

594 corresponding to study areas. (DOC)

595

596 Supplementary material S3.

597 List of acritarch genera and species reported from the Cambrian System, with marked known

598 synonymies and homonymies, including references. (DOC)

599

600 Supplementary material S4.

601 Global species range charts based on stages, blue = presence reported; green = presence inferred;
602 white = absence. (EPS)
603
604 Supplementary material S5.
605 Global species range charts based on biozones, blue = presence reported; green = presence inferred;
606 white = absence. (EPS)
607
608 Supplementary material S6.
609 Baltica species range charts based on stages, blue = presence reported; green = presence inferred;
610 white = absence. (EPS)
611
612 Supplementary material S7.
613 Baltica species range charts based on biozones, blue = presence reported; green = presence inferred;
614 white = absence. (EPS)
615
616 Supplementary material S8.
617 Gondwana species range charts based on stages, blue = presence reported; green = presence
618 inferred; white = absence. (EPS)
619
620 Supplementary material S9.
621 Gondwana species range charts based on biozones, blue = presence reported; green = presence
622 inferred; white = absence. (EPS)
623
624 Supplementary material S10.
625 Global genus range chart based on stages; blue = presence reported; green = presence inferred;
626 white = absence. (EPS)

627

628 Supplementary material S11.

629 Global genus range chart based on biozones; blue = presence reported; green = presence inferred;
630 white = absence. (EPS)

631

632 Supplementary material S12.

633 Values of diversity indices based on stages. (CVS)

634

635 Supplementary material S13.

636 Values of diversity indices based on biozones. (CVS)

637

638 Supplementary material S14.

639 Spearman's r_s and probability value p for correlations between acritarch diversity indices and the
640 number of studies per stage, and between global sampled-in-bin genus diversities of acritarchs and
641 marine invertebrates (values for marine invertebrates were taken from the supporting information of
642 Na and Kiessling, 2015). (CVS)

643

644 Supplementary material S15.

645 Comparison of regional species diversities based on biozones; Baltica and Gondwana: this study,
646 sampled-in-bin diversity; Upper Silesia: Moczydłowska (1998), total? diversity. (EPS)

647

648

649

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- 1028

1029 **Fig. 1. Cambrian stratigraphic scheme and stratigraphic ranges of source studies used for**
1030 **diversity analyses.**

1031 Chronostratigraphy according to Peng et al. (2012). *Ac.* = *Acadoparadoxides*. *Ag.* = *Agnostus*. *Pb.* =
1032 *Parabolina*. For complete references of source studies see supplementary information S2.

1033

1034 **Fig. 2. Palaeogeographical reconstruction for the mid Cambrian Period (510 Ma) with study**
1035 **areas marked.**

1036 Base map produced with BugPlates (Torsvik, 2009). 1 Nova Scotia, Canada (Palacios et al., 2009);
1037 2 Eastern Newfoundland, Canada (Martin and Dean, 1981; Martin and Dean, 1983; Martin and
1038 Dean, 1984; Martin and Dean, 1988; Parsons and Anderson, 2000); 3 County Wexford, Ireland
1039 (Brück and Vanguetstaine, 2004); 4 Wales (Young et al., 1994); 5 Belgium (Vanguetstaine, 1986); 6
1040 Sardinia, Italy (Ribecai et al., 2005); 7 Sierra Morena, Spain (Mette, 1989); 8 Cantabrian
1041 Mountains, Spain (Palacios and Vidal, 1992; Albani et al., 2006; Palacios, 2010); 9 N Algeria
1042 (Vecoli, 1996; Vecoli and Playford, 1997; Vecoli et al., 1999); 10 Ghadames Basin, Libya/Tunisia
1043 (Albani et al., 1991); 11 Barrandian area, Czech Republic (Fatka et al., 2004); 12 Upper Silesia
1044 (Bruno-Silesia), Poland (Jachowicz, 1994; Buła and Jachowicz, 1996; Moczydłowska, 1998;
1045 Jachowicz-Zdanowska, 2013); 13 Holy Cross Mountains (Małopolska High), Poland (Jagielska,
1046 1965; Lenzion et al., 1982; Kowalski, 1983; Żylińska and Szczepanik, 2009); 14 Central Alborz
1047 Mountain Range, Iran (Ghavidel-syooki, 2006); 15 Zagros Basin, Iran (Ghavidel-syooki and
1048 Vecoli, 2008); 16 South Australia (Jago et al., 2006); 17 Baltic Poland (Moczydłowska, 1981, 1988,
1049 1991; Moczydłowska and Vidal, 1986; Jankauskas and Lenzion, 1992; Szczepanik, 2000); 18
1050 Bornholm, Denmark and Skåne, Sweden (Vidal, 1981; Moczydłowska and Vidal, 1992); 19
1051 Lithuania (Jankauskas, 1972); 20 Östergötland and Öland, SE Sweden (Vidal, 1981; Di Milia et al.,
1052 1989; Eklund, 1990); 21 SW Sweden and SE Norway (Vidal, 1981; Vidal and Moczydłowska,
1053 1996); 22 Belarus (Jankauskas and Lenzion, 1992); 23 Estonia (Paalits, 1992; Pöldvere and
1054 Paalits, 1998; Uutela, 2008); 24 Swedish Caledonides, Sweden (Moczydłowska, 2002; Vidal and

1055 Moczyłowska, 1996); 25 Moscow Syncline, Russia (Volkova, 1990; Volkova, 1995); 26
1056 Leningrad Oblast, Russia (Volkova, 1993); 27 Digermul Peninsula, Norway (Welsch, 1986); 28
1057 Kolguev Island, Russia (Moczyłowska and Stockfors, 2004); 29 Severnaya Zemlya, Russia – Kara
1058 (Raevskaya and Golubkova, 2006); 30 NE Siberia (Vidal et al., 1995); 31 Jilin, China (Chen et al.,
1059 1985); 32 Hubei, China (Ding et al., 1992; Zang, 1992); 33 Guizhou, China (Yang and Yin, 2001;
1060 Yin et al., 2010); 34 Yunnan, China (Yin, 1990; Zang, 1992); 35 Sichuan, China (Wang and Chen,
1061 1987); 36 Shaanxi, China (Yin, 1987); 37 Grainger County, Tennessee, USA (Clendening and
1062 Wood, 1981); 38 East Svalbard, Norway (Knoll and Swett, 1987); 39 Mackenzie Mountains,
1063 Canada (Baudet et al., 1989)

1064

1065 **Fig. 3. Classes of taxa (in regards to ranges) present in a stratigraphic interval.**

1066 After Foote (2000).

1067

1068 **Fig. 4. Global and regional taxonomic diversity curves of Cambrian acritarchs based on**
1069 **stages.**

1070 A, global species diversity. B, global genus diversity. C, Baltica species diversity (no data available
1071 for the Drumian). D, Gondwana species diversity (no data available for the Fortunian, Stages 2-3).

1072

1073 **Fig. 5. Taxonomic diversity based on biozones.**

1074 A, global species diversity. B, global genus diversity. C, Baltica species diversity (no data available
1075 for the *Parabolina spinulosa* zone). D, Gondwana species diversity (no data available for the
1076 *Trichophycus pedum* zone, Stage 2, the *Schmidtiellus mickwitzi* and *Leptoplastus* zones).

1077

1078 **Fig. 6. Taxonomic changes of global species and genus diversity.**

1079 A, B, species. C, D, genera. A, C, raw data. B, D, rates of taxonomic changes per stage duration

1080 [My].

1081

1082 **Fig. 7. Poly-cohort analyses.**

1083 A, cohort survivorship. B, cohort pre-nascence.

1084

1085 **Fig. 8. Comparison of global species and genus diversities with the distribution of studies and**
1086 **a previously published diversity curve.**

1087

1088 **Fig. 9. Correlation of taxonomic (global species per stage) and monographic (number of**
1089 **references per stage) diversity.**

1090

1091 **Fig. 10. Comparison of global sampled-in-bin genus diversity of acritarchs and marine**
1092 **invertebrates.**

1093 Values for invertebrates taken from the supporting information of Na and Kiessling (2015).

1094

1095 **Fig. 11. Correlation of global sampled-in-bin genus diversity of acritarchs and marine**
1096 **invertebrates.**

1097 Values for invertebrates taken from the supporting information of Na and Kiessling (2015). A,

1098 scatterplot of acritarch vs. marine invertebrate diversity. B, Spearman's r_s calculated with a moving

1099 window for successive intervals, each comprising eight stages; values above 0.75 or below -0.75

1100 (dashed lines) indicate strong correlation. The middle square indicates strong and significant

1101 correlation ($r_s = 0.88, p = 0.004$) for the middle set of eight stages, from Stage 2 to the Jiangshanian

1102 Stage (9).

1103

1104 **Supplementary Material**

1105 **Table S1.** Database of Cambrian acritarch occurrences.

1106 **Table S2.** List of source references, including bibliography, countries and palaeogeographical units
1107 corresponding to study areas.

1108 **Table S3.** List of acritarch genera and species reported from the Cambrian System, with marked
1109 known synonymies and homonymies, including references.

1110 **Table S4.** Global species range charts based on stages, blue = presence reported; green = presence
1111 inferred; white = absence.

1112 **Table S5.** Global species range charts based on biozones, blue = presence reported; green =
1113 presence inferred; white = absence.

1114 **Table S6.** Baltica species range charts based on stages, blue = presence reported; green = presence
1115 inferred; white = absence.

1116 **Table S7.** Baltica species range charts based on biozones, blue = presence reported; green =
1117 presence inferred; white = absence.

1118 **Table S8.** Gondwana species range charts based on stages, blue = presence reported; green =
1119 presence inferred; white = absence.

1120 **Table S9.** Gondwana species range charts based on biozones, blue = presence reported; green =
1121 presence inferred; white = absence.

1122 **Table S10.** Global genus range chart based on stages; blue = presence reported; green = presence
1123 inferred; white = absence.

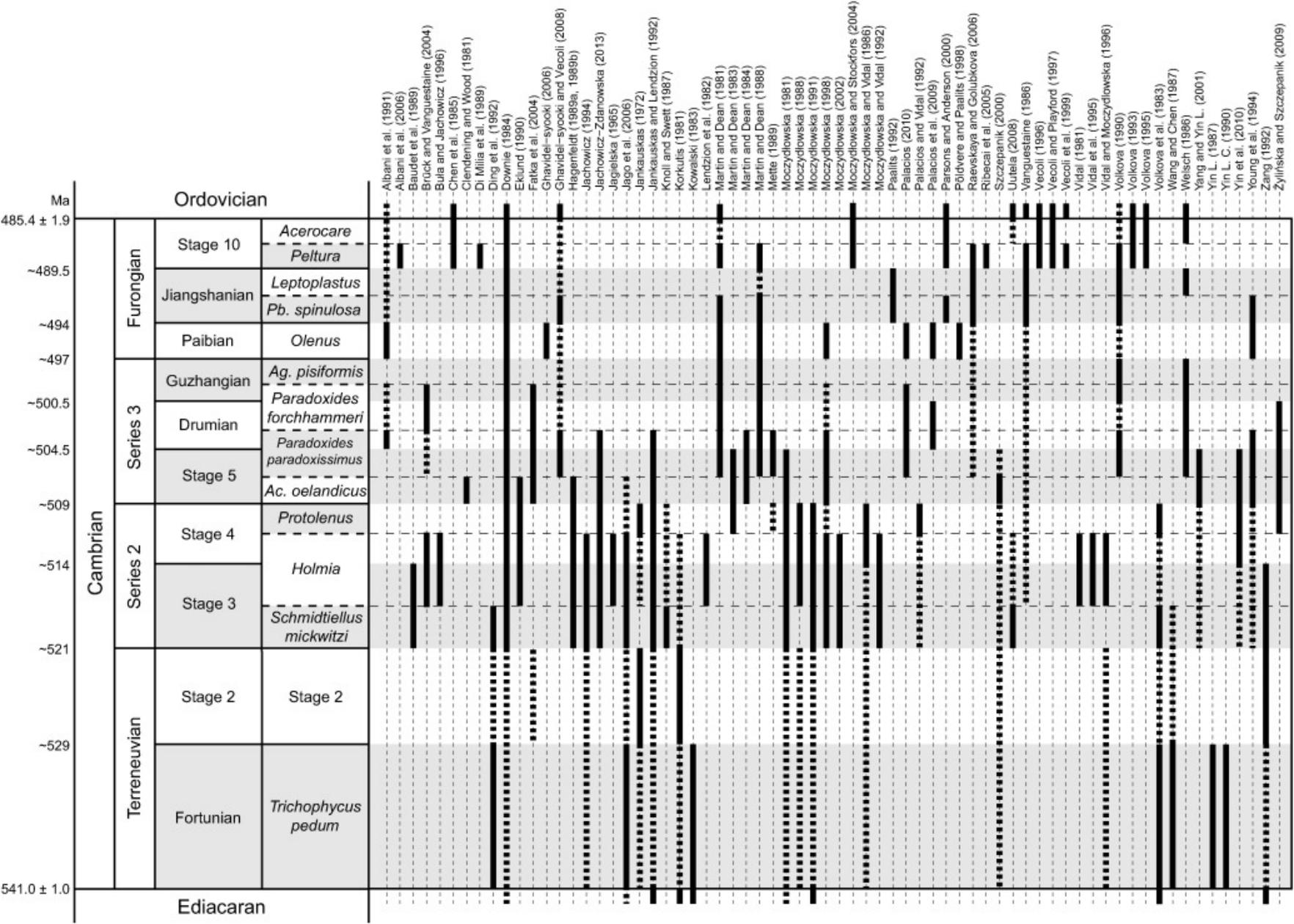
1124 **Table S11.** Global genus range chart based on biozones; blue = presence reported; green = presence
1125 inferred; white = absence.

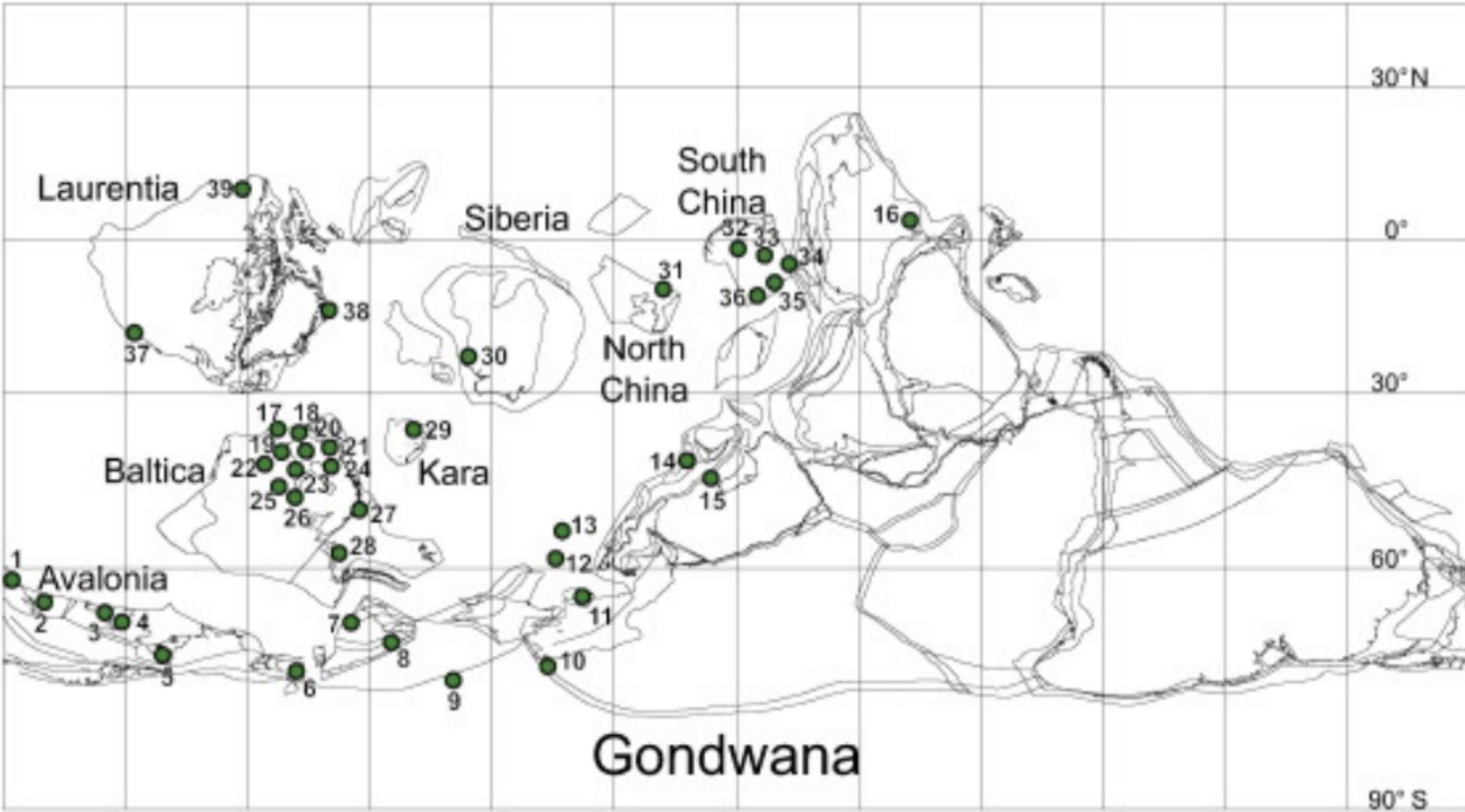
1126 **Table S12.** Values of diversity indices based on stages.

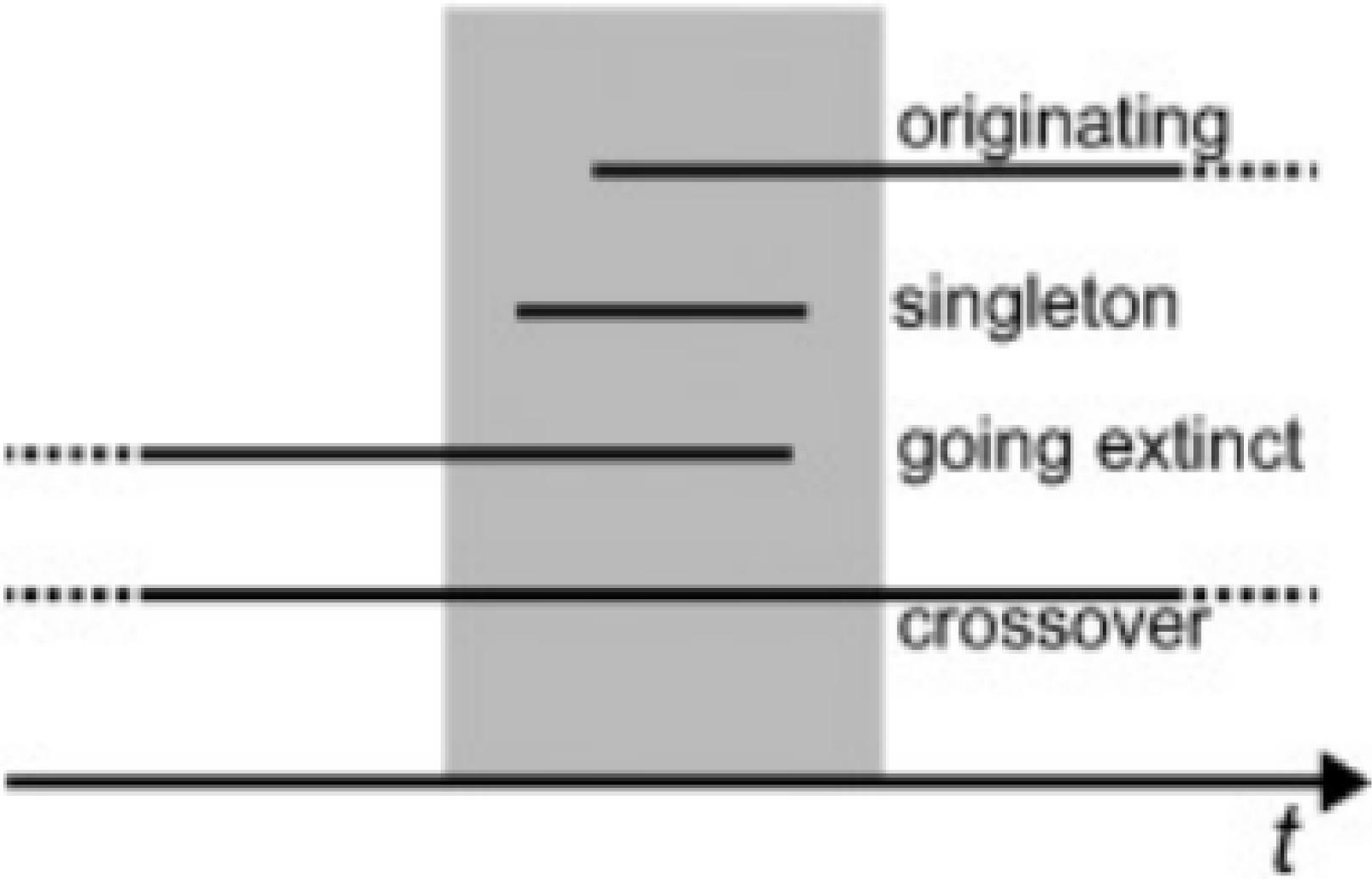
1127 **Table S13.** Values of diversity indices based on biozones.

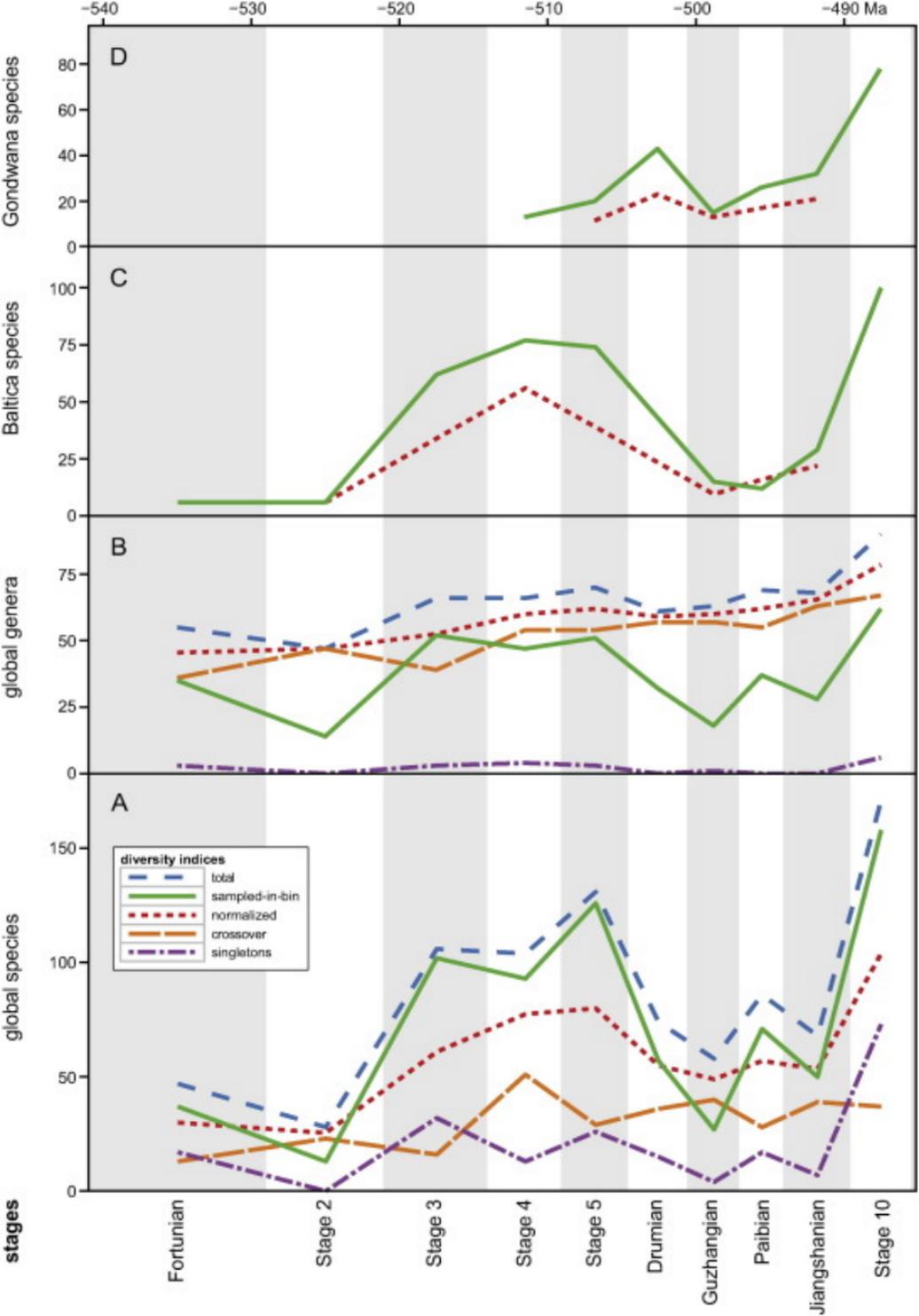
1128 **Table S14.** Spearman's r_s and probability value p for correlations between acritarch diversity
1129 indices and the number of studies per stage, and between global sampled-in-bin genus diversities of
1130 acritarchs and marine invertebrates (values for marine invertebrates were taken from the supporting
1131 information of Na and Kiessling, 2015).

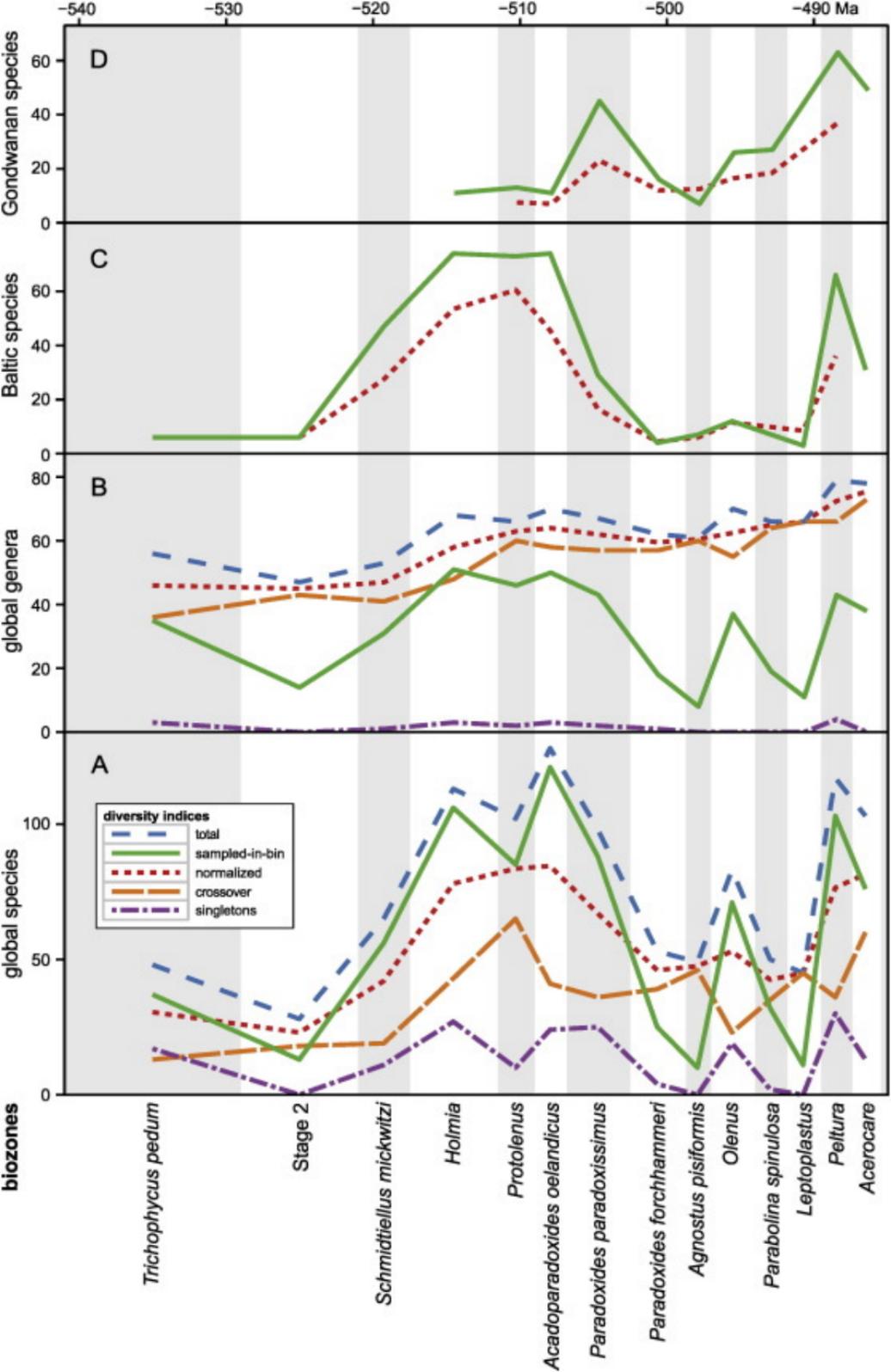
1132 **Table S15.** Comparison of regional species diversities based on biozones; Baltica and Gondwana:
1133 this study, sampled-in-bin diversity; Upper Silesia: Moczyłowska (1998), total? diversity.



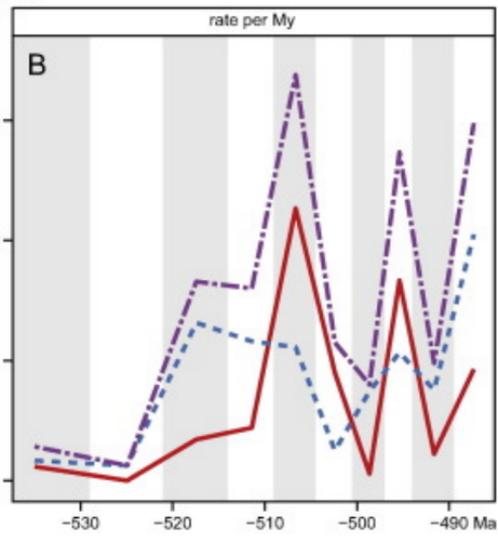
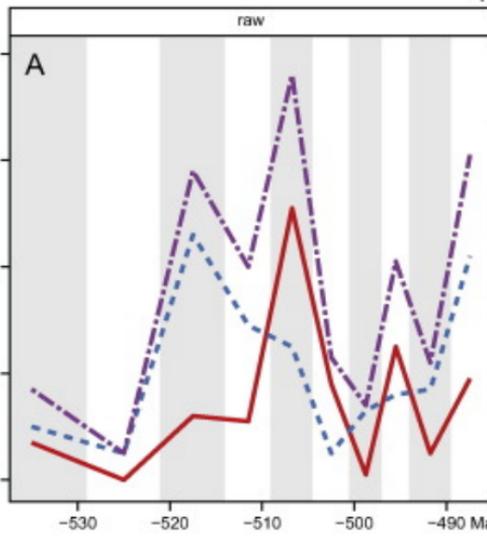




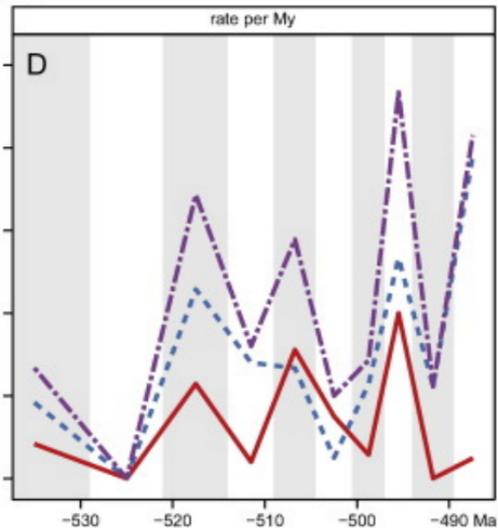
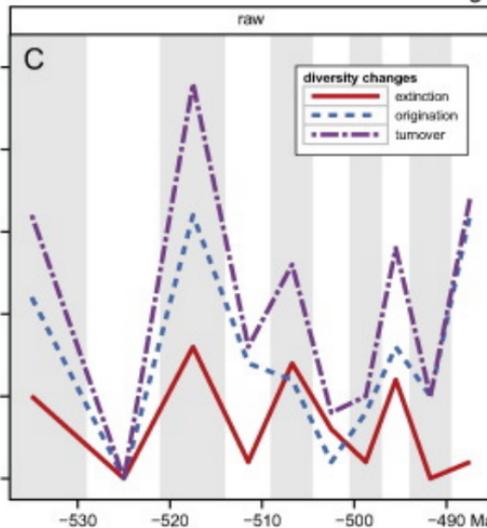


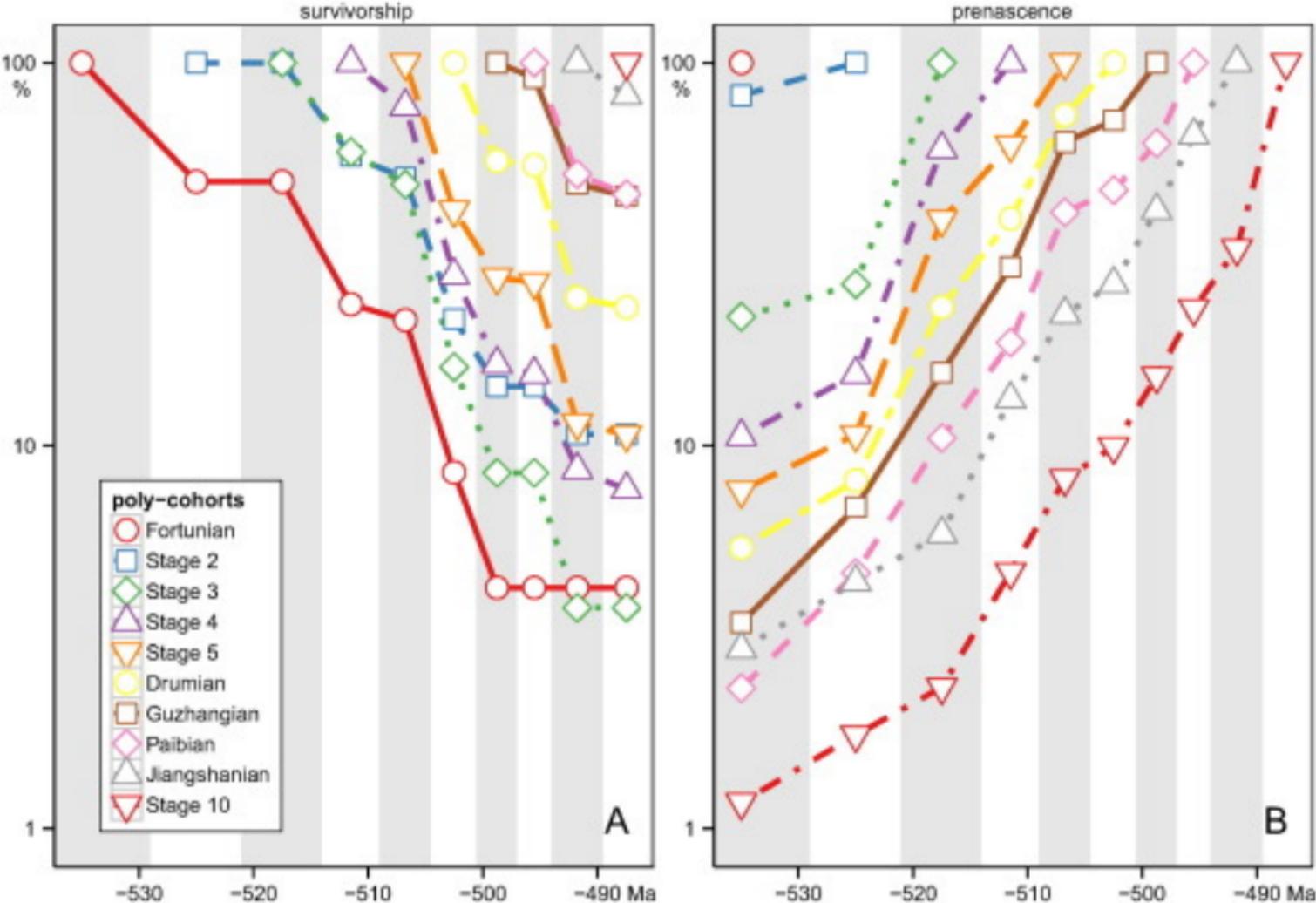


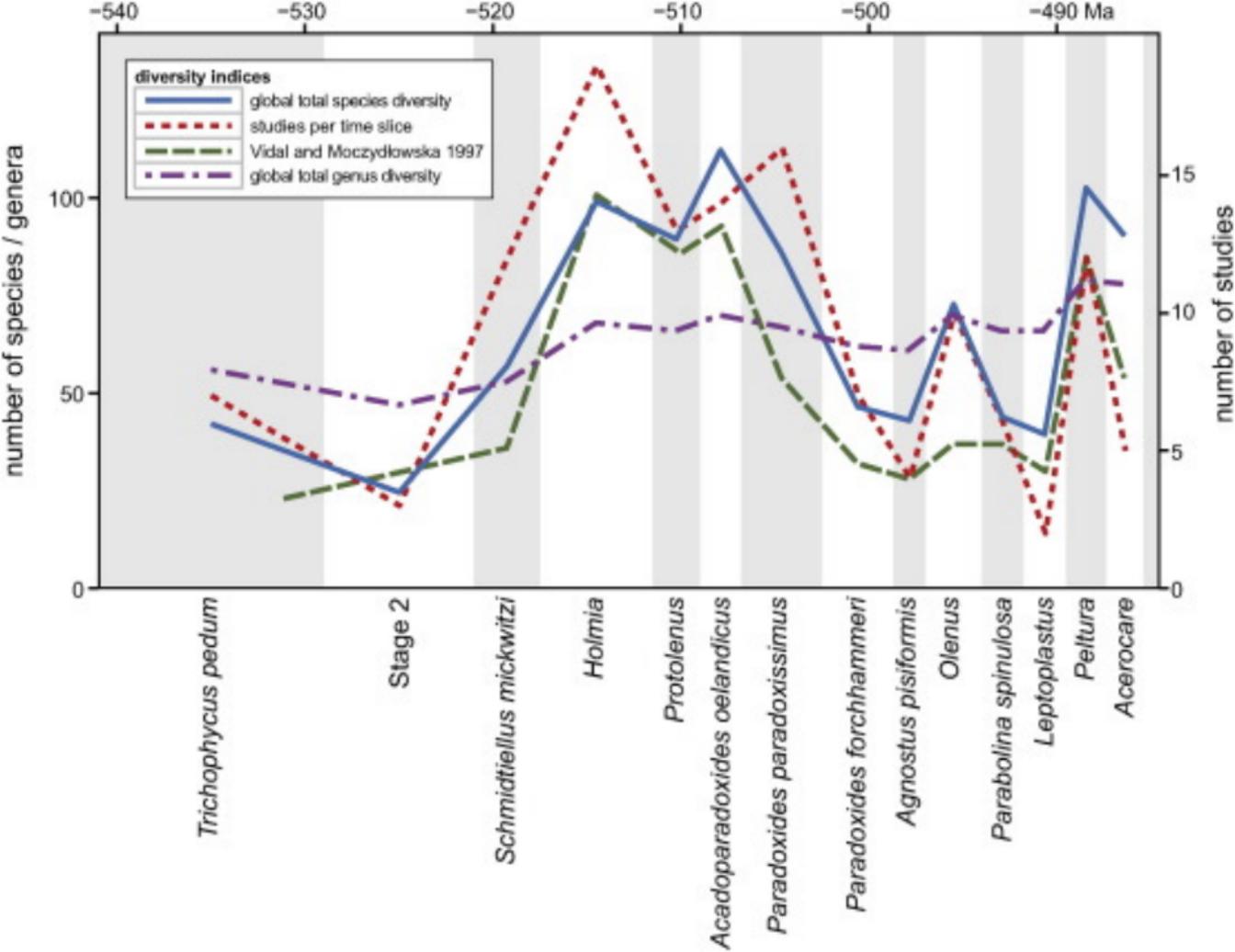
species

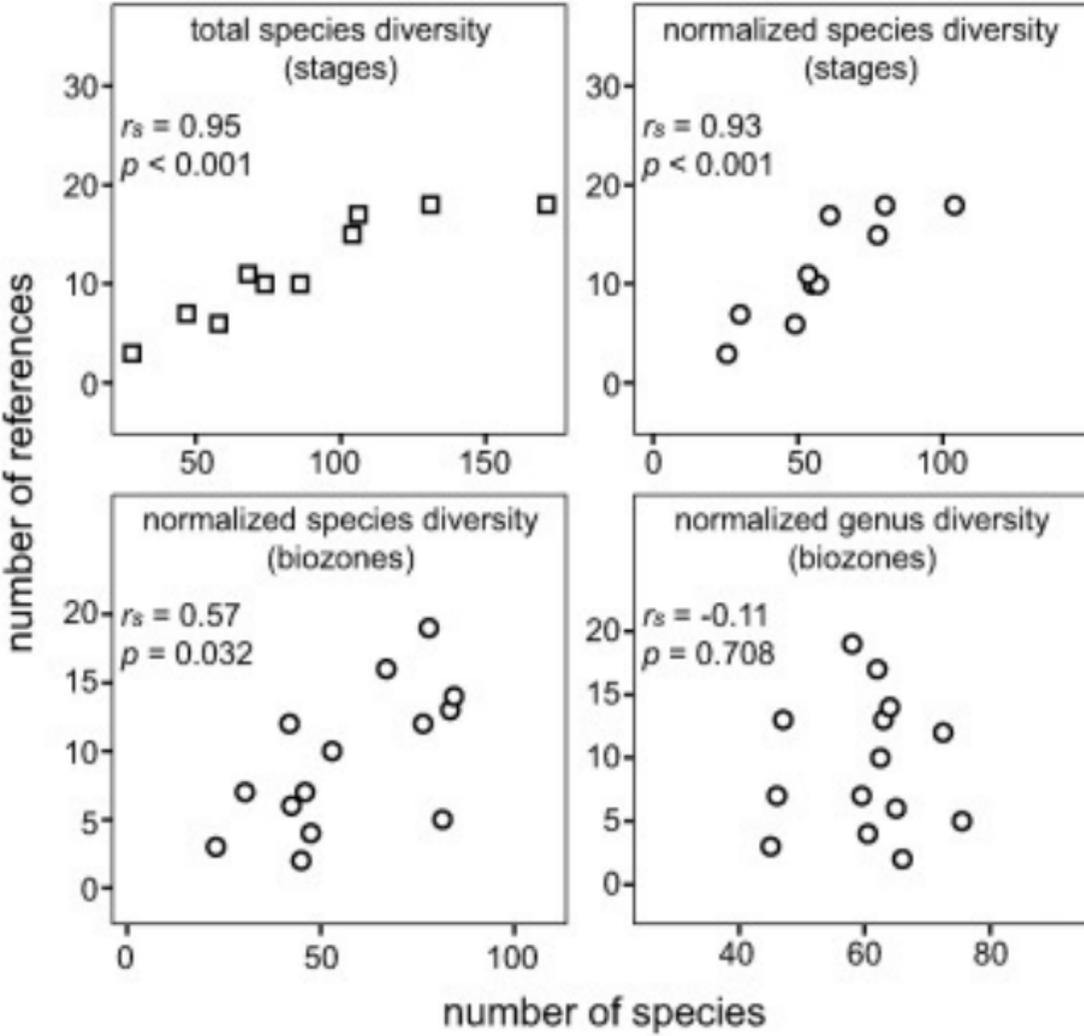


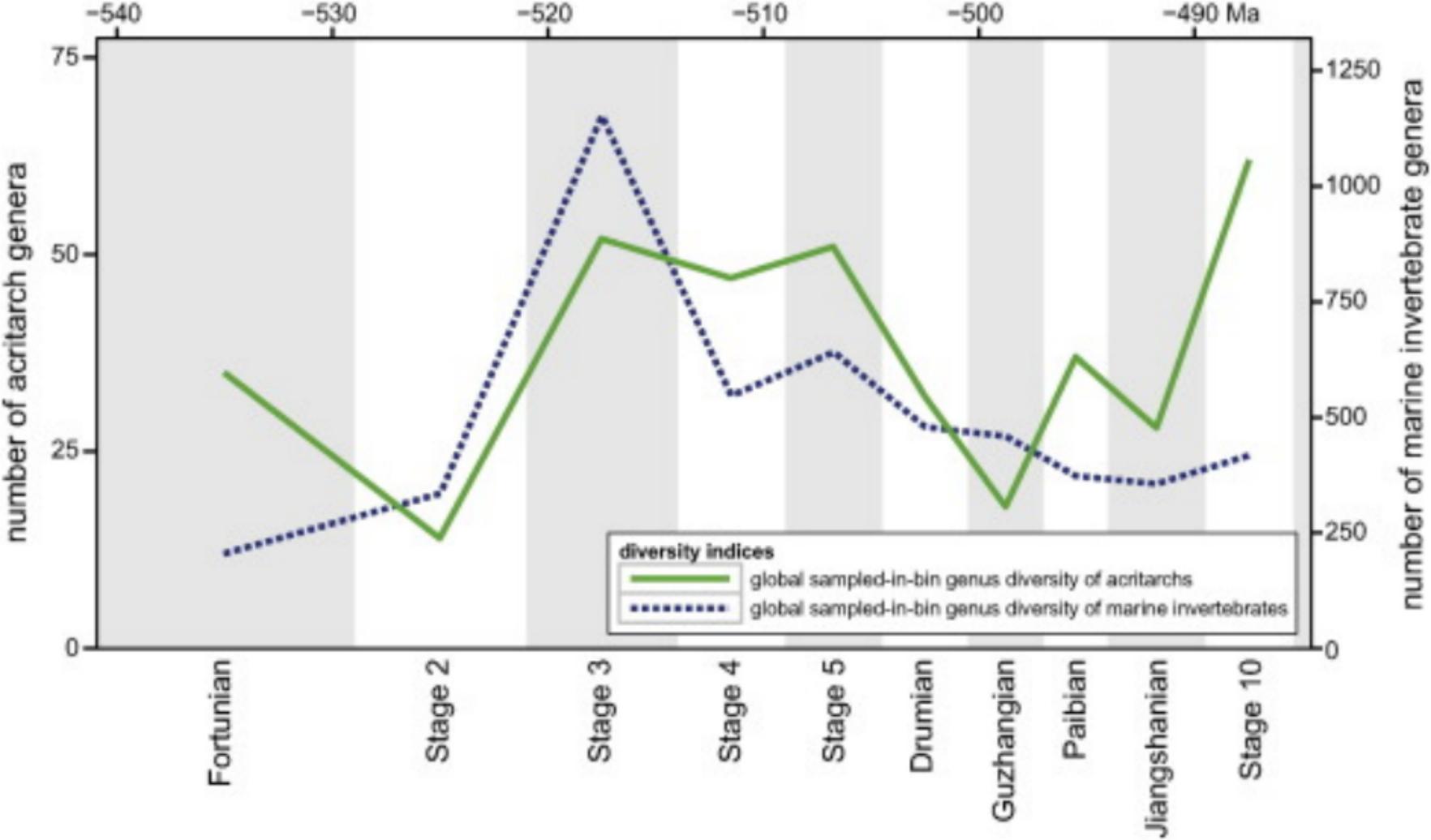
genera











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Comments on systematic palaeontology

Among the so-called acritarchs reported from the Cambrian, the genera *Chuaria* Walcott 1899, *Cymatiosphaera* Wetzel 1933 ex Deflandre 1954, *Dictyotidium* Eisenack 1955, *Leiopsophosphaera* Naumova 1961 ex Naumova 1968, *Polyedryxium* Deunff 1954, *Pterospermella* Eisenack 1972, and *Tasmanites* Newton 1875 are now considered to be (likely) prasinophytes (e.g. Guy-Ohlson 1996, Traverse 2007). At least a part of the rather featureless sphaeromorphs, especially *Leiosphaeridia* Eisenack 1958 and *Protoleiosphaeridium* Timofeev 1959 ex Timofeev 1960, might also belong to this group (comp. e.g. Guy-Ohlson 1996, Strother 1996). *Huroniospora* Barghoorn in Barghoorn & Tyler 1965 and *Myxococcoides* Schopf 1968 have been identified as cyanobacteria (Chroococcales; e.g. Shukla et al. 2006, Schopf 1968). *Caryosphaeroides* Schopf 1968 was described as a chlorococcoid (Chlorophyceae) green alga (Schopf 1968). According to Butterfield (2005), *Germinosphaera* Mikhailova 1986 (considered by him to be a synonym of *Tappania* Yin 1997) belongs to the fungi. *Ceratophyton* Kiryanov in Volkova et al. 1979 has been established to comprise animal remains (Fatka & Konzalová 1995). Wang and Luo (1982) reported the artificial production of structures resembling *Trematosphaeridium* Timofeev 1959 and *Polyporata* Pykhova 1966, among others. They suggested that these genera might – at least in part – represent pseudofossils. Baudet et al. (1989) tentatively identified ?*Hemisphaerium* Hemer & Nygreen 1967 from the lower Cambrian Sekwi Formation of the Canadian Mackenzie Mountains. According to Grenfell (1995), *Hemisphaerium* is a probable synonym of *Brazilea* Tiwari & Navale 1967, which he classified as a zygnetacean spore. From the Sinian (Late Proterozoic-?lower Cambrian) of northern China, Timofeev (1966) described *Pulvinomorpha* Timofeev 1966, considered by Grenfell (1995) as a possible synonym of *Tetraporina* Naumova 1939, another zygnetacean spore. This tentative interpretation would make *Brazilea* (*Hemisphaerium*) and *Tetraporina* (*Pulvinomorpha*) the oldest recorded zygnetaceans by far, with a gap in the record ranging up to the Carboniferous (Grenfell 1995). The genus *Pseudozonosphaeridium* Andreeva 1966 and the species *P. fengxiangense* Zhong Guofang 1978 in particular have been reported from the Proterozoic and the Silurian, but not the Cambrian (Fensome et al. 1990). The existence of *Veryhachium* Deunff 1954 in the Cambrian was rejected by Raevskaya and Servais (2009), but several species reported from the Cambrian have been assigned to this genus and are currently not formally transferred to other genera. Moczydłowska (1991) proposed the transfer of Cambrian forms assigned to *Micrhystridium* Deflandre 1937 (which has received numerous emendations) to the new genera *Asteridium* Moczydłowska 1991 and *Heliosphaeridium* Moczydłowska 1991. Not all concerned species have been transferred so far and not all authors have followed the proposal (e.g. Sarjeant & Stancliffe 1994).

List of acritarch genera and species reported from the Cambrian

names in “ ” have been found to be invalid

« → » indicates current name

“*Abacum*” Fombella 1978

→ *Virgatasporites* Combaz 1967 [acc. to Fensome et al. 1990]

type species: “*Abacum normale*” Fombella 1978

→ *Virgatasporites normalis* (Fombella 1978) Fensome et al. 1990

“*Abacum normale*” Fombella 1978

→ *Virgatasporites normalis* (Fombella 1978) Fensome et al. 1990

Acanthodiacrodium Timofeev 1958 emend. Deflandre & Deflandre-Rigaud 1962 restrict.
Moczydłowska & Stockfors 2004

type species: *Acanthodiacrodium dentiferum* Timofeev 1958

“*Acanthodiacrodium achrasii*” (Martin 1972) Martin & Dean 1988

→ *Actinotodissus achrasii* (Martin 1972) Yin L. 1986

Acanthodiacrodium anceps Timofeev 1959b

Acanthodiacrodium angustum (Downie 1958) Combaz 1967

Acanthodiacrodium complanatum (Deunff 1961a) Vavrdová 1965

Acanthodiacrodium enodum Timofeev 1959b

Acanthodiacrodium golubii Fensome et al. 1990

Acanthodiacrodium cf. *ignoratum* (Deunff 1961a) Downie & Sarjeant 1965

Acanthodiacrodium lanatum Timofeev 1959b ex Downie & Sarjeant 1965

Acanthodiacrodium petrovii Timofeev 1959b

“*Acanthodiacrodium polymorphum*” Timofeev 1959b

→ *Actinotodissus polimorphus* (Timofeev 1959b) Moczydłowska & Stockfors 2004

“*Acanthodiacrodium secundarium*” Timofeev 1959b

→ *Actinotodissus secundarius* (Timofeev 1959b) Moczydłowska & Stockfors 2004

Acanthodiacrodium sinuosum Rasul 1979

Acanthodiacrodium snookense Parsons & Anderson 2000

Acanthodiacrodium cf. *spinum* Rasul 1979

“*Acanthodiacrodium spinutisum*” Timofeev 1959b

- *Actinotodissus spinutisus* (Timofeev 1959b) Moczyłowska & Stockfors 2004
- “*Acanthodiacrodium timofeevii*” Golub & Volkova in Volkova & Golub 1985
- *Acanthodiacrodium golubii* Fensome et al. 1990
- Acanthodiacrodium tricornis* (Timofeev 1959b ex Downie & Sarjeant 1965) Martin 1969
- “*Acanthodiacrodium tuberatum*” (Downie 1958) Martin 1972
- *Goniosphaeridium tuberatum* (Downie 1958) Welsch 1986
- “*Acanthodiacrodium ubuii*” Martin 1969
- *Actinotodissus ubuii* (Martin 1969) Fensome et al. 1990
- “*Acanthodiacrodium uniforme*” Burmann 1968
- [**senior homonym:** *Acanthodiacrodium uniforme* Timofeev 1959b, acc. to Fensome et al. 1990]
- *Actinotodissus burmanniae* (Fensome et al. 1990) Moczyłowska & Stockfors 2004
- [Moczyłowska & Stockfors 2004 gave authorship of this combination as “(Burmann 1968) Fensome et al. 1990 »]
- {*pro parte* in Chen J. et al. 1985} *Actinotodissus achrasii* (Martin 1972) Yin L. 1986 [acc. to Martin & Dean 1988]
- {*pro parte* in Chen J. et al. 1985} *Dasydiacrodium obsonum* Martin in Martin & Dean 1988 [acc. to Moczyłowska & Stockfors 2004]
- Acanthodiacrodium* sp. A Parsons & Anderson 2000
- Acanthodiacrodium* sp. B Parsons & Anderson 2000
- Acanthodiacrodium* sp. I Yin L. in Chen J. et al. 1985

Acrum Fombella 1977

type species: *Acrum novum* Fombella 1977

?*Acrum araxisii* Di Milia 1991

Acrum cylindriferum Downie 1982

Acrum radiale Fombella 1977

Actinotodissus Loeblich & Tappan 1978

type species: *Actinotodissus longitaleosus* Loeblich & Tappan 1978

Actinotodissus achrasii (Martin 1972) Yin L. 1986

Actinotodissus formosus (Górka 1967) Moczyłowska & Stockfors 2004

Actinotodissus polimorphus (Timofeev 1959b) emend. Moczyłowska & Stockfors 2004

Actinotodissus secundarius (Timofeev 1959b) Moczyłowska & Stockfors 2004

Actinotodissus spinutisus (Timofeev 1959b) Moczyłowska & Stockfors 2004

Actinotodissus ubuii (Martin 1969) Fensome et al. 1990

Actinotodissus sp. A Martin 1992

Adara Fombella 1977 emend. Martin in Martin & Dean 1981
type species: *Adara mutatina* Fombella 1977

Adara alea Martin in Martin & Dean 1981

→ {in Albani et al. 1991} *Multiplicisphaeridium ?ancliforme* Fombella 1978 [acc. to Jachowicz-Zdanowska 2013]

→ {in Buła & Jachowicz 1996; Moczyłowska 1998} *Turrisphaeridium semireticulatum* (Timofeev 1959b) Jachowicz-Zdanowska 2013 [acc. to Jachowicz-Zdanowska 2013]

“*Adara denticulata*” Tongiorgi in Bagnoli et al. 1988

→ *Adara alea* Martin in Martin & Dean 1981 [acc. to Vanguetaine & Brück 2008]

Adara matutina Fombella 1977

“*Adara undulata*” Moczyłowska 1998

→ *Adara alea* Martin in Martin & Dean 1981 [acc. to Vanguetaine & Brück 2008]

Adara sp. A Baudet et al. 1989

Alliumella Fanderflit in Umnova & Fanderflit 1971

type species: *Alliumella baltica* Fanderflit in Umnova & Fanderflit 1971

Alliumella baltica Fanderflit in Umnova & Fanderflit 1971

Alliumella n. sp. Vanguetaine 1986

Ammonidium Lister 1970 emend. Sarjeant & Vavrdová 1997

type species: *Ammonidium microcladum* (Downie 1963) Lister 1970 emend. Sarjeant & Vavrdová 1997

Ammonidium bellulum (Moczyłowska 1998) Sarjeant & Stancliffe 2000

Ammonidium notatum (Volkova 1969) Jachowicz-Zdanowska 2013

Ammonidium oligum (Jankauskas in Jankauskas & Posti 1976) Jachowicz-Zdanowska 2013

Annulum Fombella 1978

type species: *Annulum difuminatum* Fombella 1978

“*Annulum squamaceum*” (Volkova 1968) emend. Martin in Martin & Dean 1983
→ *Granomarginata squamacea* Volkova 1968

Aranidium Jankauskas 1975
type species: *Aranidium izhoricum* Jankauskas 1975

Aranidium granulatum Welsch 1986
Aranidium izhoricum Jankauskas 1975
Aranidium aff. *pycnacanthum* Jankauskas 1975
Aranidium sparsum Volkova in Volkova et al. 1979

Arbusculidium Deunff 1968a emend. Welsch 1986
type species: *Arbusculidium destombesii* Deunff 1968a

?*Arbusculidium adminiculum* Di Milia et al. 1989
Arbusculidium destombesii Deunff 1968a
Arbusculidium cf. *mamillosum* Welsch 1986
Arbusculidium ornatum (Combaz 1967) Fensome et al. 1990
Arbusculidium perlongum Di Milia et al. 1989
Arbusculidium polypus Di Milia et al. 1989
“*Arbusculidium rommelaerei*” Martin in Martin & Dean 1981
→ *Ladogella rommelaerei* (Martin in Martin & Dean 1981) Di Milia et al. 1989
Arbusculidium sp. 1 Volkova 1990

Archaeodiscina Naumova 1961 emend. Volkova 1968
type species: *Archaeodiscina granulata* Naumova 1961

Archaeodiscina bicostata Volkova in Volkova et al. 1979
Archaeodiscina umbonulata Volkova 1968
Archaeodiscina multipunctata Jachowicz-Zdanowska 2013
Archaeodiscina sp. A Baudet et al. 1989
?*Archaeodiscina* sp. B Baudet et al. 1989

Archaeosacculina Pykhova 1967
[**junior homonym:** “*Archaeosacculina*” Naumova 1968]
type species: *Archaeosacculina torosa* Pykhova 1967

“*Archaeosacculina atava*” Pykhova 1966
[not validly published acc. to Fensome et al. 1990]

Aryballomorpha Martin and Yin L. 1988
type species: *Aryballomorpha grootaertii* (Martin 1984) emend. Martin and Yin L. 1988

Aryballomorpha albertana Martin 1992

Asperatopsophosphaera Shepeleva 1963 emend Kowalski 1983
type species: *Asperatopsophosphaera bavlensis* Shepeleva 1963 emend. Kowalski 1983

Asperatopsophosphaera bavlensis Shepeleva 1963

“*Asperatopsophosphaera partialis*” Shepeleva 1963
→ *Asperatopsophosphaera bavlensis* Shepeleva 1963 [acc. to Kowalski 1983]

Asperatopsophosphaera rugosa Ding
[cited without complete reference in Ding et al. 1992]

Asperatopsophosphaera umishanensis Xing & Liu 1973

Asteridium Moczydłowska 1991
type species: *Asteridium lanatum* (Volkova 1969) Moczydłowska 1991

Asteridium cerinum (Volkova 1968) Moczydłowska 1991

Asteridium lanatum (Volkova 1969) Moczydłowska 1991

Asteridium ordense (Downie 1982) emend. Vidal & Peel 1993

Asteridium pallidum (Volkova 1968) Moczydłowska 1991

Asteridium pilare Moczydłowska 1998

Asteridium solidum Moczydłowska 1998

Asteridium spinosum (Volkova 1969) Moczydłowska 1991

Asteridium tornatum (Volkova 1968) Moczydłowska 1991

Asteriscus Kowalski 1983

type species: *Asteriscus irregularis* Kowalski 1983

Asteriscus irregularis Kowalski 1983

Auritusphaera Strother 2008

type species: *Auritusphaera bifurcata* Strother 2008

Auritusphaera bifurcata Strother 2008

Baltisphaeridium Eisenack 1958c ex Eisenack 1959a emend. Staplin et al. 1965 emend.
Eisenack 1969 emend. Eiserhardt 1989

type species: *Baltisphaeridium longispinosum* (Eisenack 1931 ex Wetzel 1933) Eisenack
1959a

Baltisphaeridium acerosum Jankauskas & Posti 1976

“*Baltisphaeridium brachyspinosum*” Kiryanov 1974

→ *Ichnosphaera delicata* Jachowicz-Zdanowska 2013

“*Baltisphaeridium capillatum*” (Naumova 1950) Umnova 1975

[**senior homonym:** *Baltisphaeridium capillatum* Jardiné et al. 1974]

“*Baltisphaeridium cerinum*” Volkova 1968

→ *Globosphaeridium cerinum* (Volkova 1968) Moczydłowska 1991

“*Baltisphaeridium ciliosum*” Volkova 1969

→ *Skiagia ciliosa* (Volkova 1969) Downie 1982

Baltisphaeridium citrinum (Downie 1963) Stockmans & Willièrè 1974

“*Baltisphaeridium compressum*” Volkova 1968

→ *Skiagia compressa* (Volkova 1968) Downie 1982

Baltisphaeridium crinitum Martin in Dean & Martin 1978

Baltisphaeridium dictium Wang & Chen Q. 1987

“*Baltisphaeridium dubium*” Volkova 1968

→ “*Lophosphaeridium dubium*” (Volkova 1968) Moczydłowska 1991]

Baltisphaeridium ellipticum Wang et Chen

[cited without complete reference in Wang & Chen Q. 1987]

Baltisphaeridium emeiense Wang et Chen

[cited without complete reference in Wang & Chen Q. 1987]

Baltisphaeridium favosum Xing 1962

“*Baltisphaeridium implicatum*” Fridriksone 1971
→ *Solisphaeridium implicatum* (Fridriksone 1971) Moczydłowska 1998

“*Baltisphaeridium insigne*” (Fridriksone 1971) Volkova 1974
→ *Skiagia insignis* (Fridriksone 1971) Downie 1982

Baltisphaeridium jiulaodongense Wang & Chen Q. 1987

Baltisphaeridium jilinense Fensome et al. 1990

“*Baltisphaeridium latviense*” Volkova 1974
→ *Comasphaeridium latviense* (Volkova 1974) Hagenfeldt 1989a

Baltisphaeridium longispinosum (Eisenack 1931 ex Wetzel 1933) Eisenack 1959a

“*Baltisphaeridium orbiculare*” Volkova 1968
→ *Skiagia orbicularis* (Volkova 1968) Downie 1982

“*Baltisphaeridium ornatum*” Volkova 1968
→ *Skiagia ornata* (Volkova 1968) Downie 1982

“*Baltisphaeridium papillosum*” (Timofeev 1959) ex Volkova 1968
→ *Filisphaeridium papillosum* (Timofeev 1959 ex Volkova 1968) Hu Yunxu 1986

“*Baltisphaeridium pilosiusculum*” Jankauskas in Volkova et al. 1979
→ *Skiagia pilosiuscula* (Jankauskas in Volkova et al. 1979) Jachowicz-Zdanowska 2013

“*Baltisphaeridium primarium*” Jankauskas in Volkova et al. 1979
→ *Goniosphaeridium primarium* (Jankauskas in Volkova et al. 1979) Downie 1982

Baltisphaeridium pseudofaveolatum Fridriksone 1971

Baltisphaeridium cf. *psilatum* Kjellström 1971a

“*Baltisphaeridium pungens*” Timofeev 1959b ex Martin 1969
→ *Polygonium pungens* (Timofeev 1959b ex Martin 1969) Albani 1989

Baltisphaeridium reticulum Wang & Chen Q. 1987

“*Baltisphaeridium robustum*” Yin L. in Chen J. et al. 1985
→ *Baltisphaeridium jilinense* Fensome et al. 1990

Baltisphaeridium simplex Deunff 1961a
[**junior homonym:** “*Baltisphaeridium simplex*” Stockmans & Willièrè 1962]

Baltisphaeridium solidum Xing 1962

“*Baltisphaeridium stipaticum*” Hagenfeldt 1989a
→ *Ichnosphaera stipaticum* (Hagenfeldt 1989a) Jachowicz-Zdanowska 2013

“?*Baltisphaeridium strigosum*” Jankauskas in Jankauskas & Posti 1976
→ *Comasphaeridium strigosum* (Jankauskas in Jankauskas & Posti 1976) Downie 1982

Baltisphaeridium tuberculatum Fridriksone 1971

“*Baltisphaeridium varium*” Volkova 1969
→ *Eklundia varia* (Volkova 1969) Jachowicz-Zdanowska 2013

Baltisphaeridium sp. 1 Volkova 1980

Baltisphaeridium sp. 1 Volkova et al. 1983

Baltisphaeridium sp. 1 Volkova et al. 1983

Baltisphaeridium sp. 2 Volkova 1980

Baltisphaeridium sp. 2 Volkova et al. 1983

→ *Goniosphaeridium volkovae* Hagenfeldt 1989a [acc. to Jachowicz-Zdanowska 2013]

Bavlinella Shepeleva 1962

[acc. to Muir 1977 “[...] objects like *Bavlinella* are not fossils. They are organic residua from framboids.”]

type species: *Bavlinella faveolata* Shepeleva 1962

Bavlinella faveolata Shepeleva 1962

“*Bothroligtriletes*” Timofeev 1958

[not validly published acc. to Fensome et al. 1990]

variant: “*Bothroligotriletum*”

no type species

“*Bothroligotriletes exasperatus*” Timofeev 1959b

Brevitrichoides Jankauskas 1980

type species: *Brevitrichoides bashkirus* Jankauskas 1980

Brevitrichoides crassus (Xing & Liu 1978) Zang 1992

Brochos

[cited without reference for the genus in Volkova 1995]

type species: ?

Brochos eisenackii Vanderflit et Mikhailova

[cited without complete reference in Volkova 1995]

Bubomorpha Yin L. in Chen J. et al. 1985

type species: *Bubomorpha hunjiangensis* Yin L. in Chen J. et al. 1985

Bubomorpha balangensis Yin L. et al. 2010

Bubomorpha hunjiangensis Yin L. in Chen J. et al. 1985

Buchinia Volkova 1990

type species: *Buchinia variabilis* Volkova 1990

Buchinia variabilis Volkova 1990

Buedingiisphaeridium Schaarschmidt 1963 emend. Lister 1970 emend. Sarjeant & Stancliffe 1994

type species: *Buedingiisphaeridium permicum* Schaarschmidt 1963

Buedingiisphaeridium tremadocum Rasul 1979

Caldariola Molyneux in Molyneux & Rushton 1988

type species: *Caldariola glabra* (Martin 1972) Molyneux in Molyneux & Rushton 1988

Caldariola glabra (Martin 1972) Molyneux in Molyneux & Rushton 1988

Calyxiella Golub & Volkova in Volkova & Golub 1985

type species: *Calyxiella izhoriensis* Golub & Volkova in Volkova & Golub 1985

Calyxiella izhoriensis Golub & Volkova in Volkova & Golub 1985

Caryosphaeroides Schopf 1968

[Chlorococcales]

type species: *Caryosphaeroides pristina* Schopf 1968

Caryosphaeroides pristina Schopf 1968

Celtiberium Fombella 1977 emend. Sarjeant & Stancliffe 2000

type species: *Celtiberium geminum* Fombella 1977

Celtiberium cf. *clarum* Fombella 1978

→ {in Vanguetaine & Van Looy 1983} *Turrisphaeridium semireticulatum* (Timofeev 1959b)
Jachowicz-Zdanowska 2013

Celtiberium dedalinum Fombella 1978

Celtiberium geminum Fombella 1977

Celtiberium cf. *geminum* Fombella 1977

→ {in Vanguetaine & Van Looy 1983} *Turrisphaeridium semireticulatum* (Timofeev 1959b)
Jachowicz-Zdanowska 2013

?*Celtiberium papillatum* Moczyłowska 1998

Celtiberium robustum Hagenfeldt 1989b

Celtiberium n. sp. A Palacios 2010

Celtiberium sp. A Downie 1982

Celtiberium sp. B Downie 1982

Celtiberium sp. B Knoll & Swett 1987

Celtiberium sp. indet. B Jago et al. 2006

Ceratophyton Kiryanov in Volkova et al. 1979

[SCF]

type species: *C. vernicosum* Kiryanov in Volkova et al. 1979

Chuar Walcott 1899 emend. Vidal & Ford 1985

type species: *Chuar* *circularis* Walcott 1899 emend. Vidal & Ford 1985

Chuar *circularis* Walcott 1899 emend. Vidal & Ford 1985

Comasphaeridium Staplin et al. 1965 emend. Sarjeant & Stancliffe 1994

type species: *Comasphaeridium cometes* (Valensi 1949) Staplin et al. 1965

Comasphaeridium agglutinatum Moczyłowska 1988

Comasphaeridium annulare (Wang 1985) Yao et al. 2005

“*Comasphaeridium brachyspinosum*” (Kiryanov 1974) Moczyłowska & Vidal 1988

→ *Ichnosphaera delicata* Jachowicz-Zdanowska 2013

“*Comasphaeridium caesariatum*” Wood & Clendening 1982

[**senior homonym:** *Comasphaeridium caesariatum* Wicander 1974]
→ *Comasphaeridium woodii* Fensome et al. 1990

?*Comasphaeridium densispinosum* Vidal in Vidal & Peel 1993

“*Comasphaeridium filiforme*” Fombella 1979
[nomen nudum, acc. to Fensome et al. 1990]

Comasphaeridium formosum Moczydłowska 1988

Comasphaeridium francinae Jachowicz-Zdanowska 2013

Comasphaeridium gogense (Downie 1982) Sarjeant & Stancliffe 1994

Comasphaeridium latviense (Volkova 1974) Hagenfeldt 1989a

Comasphaeridium longispinosum Hagenfeldt 1989b
[**junior homonym:** “*Comasphaeridium longispinosum*” Vidal in Vidal & Peel 1993]

“*Comasphaeridium longispinosum*” Vidal in Vidal & Peel 1993
[**senior homonym:** *Comasphaeridium longispinosum* Hagenfeldt 1989b]

Comasphaeridium mackenzianum Baudet et al. 1989

Comasphaeridium molliculum Moczydłowska & Vidal 1988

“*Comasphaeridium piliferum*” Fombella 1979
[nomen nudum, acc. to Fensome et al. 1990]
→ *Parmasphaeridium implicatum* (Fridriksone 1971) Jachowicz-Zdanowska 2013

Comasphaeridium silesiense Moczydłowska 1998

Comasphaeridium soniae Jachowicz-Zdanowska 2013

Comasphaeridium spinosum Jachowicz-Zdanowska 2013

Comasphaeridium strigosum (Jankauskas in Jankauskas & Posti 1976) Downie 1982
→ {in Szczepanik 2000} *Comasphaeridium silesiense* Moczydłowska 1998 [acc. to Jachowicz-Zdanowska 2013]

Comasphaeridium velvetum Moczydłowska 1988

Comasphaeridium vozmedianum Fombella 1978 ex Jachowicz-Zdanowska 2013

Comasphaeridium woodii Fensome et al. 1990

Comasphaeridium n. sp. 3 Moczydłowska & Vidal 1986

Comasphaeridium n. sp. 4 Moczydłowska & Vidal 1986

Comasphaeridium n. sp. A Palacios 2010
→ *Comasphaeridium francinae* Jachowicz-Zdanowska 2013

Comasphaeridium n. sp. Vidal 1981

Comasphaeridium sp. 2 Moczydłowska & Vidal 1986

Comasphaeridium sp. A Downie 1982

Comasphaeridium sp. A Palacios et al. 2009

Comasphaeridium sp. B Palacios et al. 2009

Comasphaeridium sp. indet. A Jago et al. 2006

Corollasphaeridium Martin in Dean & Martin 1982 emend. Yin L. 1986
type species: *Corollasphaeridium wilcoxianum* Martin in Dean & Martin 1982

Corollasphaeridium wilcoxianum Martin in Dean & Martin 1982

Corrugasphaera Fensome et al. 1990
type species: *Corrugasphaera terranovana* (Martin in Martin & Dean 1988) Fensome et al. 1990

Corrugasphaera terranovana (Martin in Martin & Dean 1988) Fensome et al. 1990

Coryphidium Vavrdová 1972
type species: *Coryphidium bohemicum* Vavrdová 1972

Coryphidium sichuanense Wang & Chen Q. 1987

Cristallinium Vanguetaine 1978
type species: *Cristallinium cambriense* (Slavíková 1968) Vanguetaine 1978

Cristallinium aciculatum Tongiorgi in Bagnoli et al. 1988

Cristallinium baculatum Volkova 1990

Cristallinium cambriense (Slavíková 1968) Vanguetaine 1978

Cristallinium compactum Jachowicz-Zdanowska 2013

?*Cristallinium delicatum* Yin L. in Chen J. et al. 1985

Cristallinium dubium Volkova 1990

Cristallinium locale Volkova 1990

“*Cristallinium ovillense*” (Cramer & Díez 1972) Fensome et al. 1990
→ *Cristallinium cambriense* (Slavíková 1968) Vanguetaine 1978 [acc. to Moczydłowska 1998]

“*Cristallinium ovillense*” (Cramer & Díez 1972) Martin in Martin & Dean 1981
→ *Retisphaeridium ovillense* (Cramer & Díez 1972) Vanguetaine 2002

Cristallinium pilosum Golub & Volkova in Volkova & Golub 1985

Cristallinium cf. randomense Martin in Martin & Dean 1981 emend. Martin in Martin & Dean 1988

Cymatiogalea Deunff 1961a emend. Deunff 1964
type species: *Cymatiogalea margaritata* Deunff 1961a

Cymatiogalea aspergillum Martin in Martin & Dean 1988
Cymatiogalea bellicosa Deunff 1961a
Cymatiogalea bouvardii Martin 1972
→ {in Martin & Dean 1981} *Stelliferidium* sp. A Parsons & Anderson 2000
“*Cymatiogalea columellifera*” (Deunff 1961a) Eisenack et al. 1973
→ *Cymatiogalea cuvillieri* (Deunff 1961a) Deunff 1964 [acc. to Rasul 1974]
Cymatiogalea cristata (Downie 1958) Rauscher 1973
Cymatiogalea cuvillieri (Deunff 1961a) Deunff 1964
“*Cymatiogalea cylindrata*” Rasul 1974
→ *Stelliferidium cylindratum* (Rasul 1974) Elaouad-Debbaj 1988
Cymatiogalea dentalea Paalits 1992b
Cymatiogalea fascicularis Yin L. in Chen J. et al. 1985
Cymatiogalea fimbriata Volkova 1990
Cymatiogalea geometrica Di Milia et al. 1989
Cymatiogalea gorkae Rauscher 1973
Cymatiogalea hunjiangensis Yin L. in Chen J. et al. 1985 emend Yin L. 1986
Cymatiogalea membranispina Deunff 1961a
?*Cymatiogalea membranula* Martin in Dean & Martin 1978
Cymatiogalea multarea (Deunff 1961a) Eisenack et al. 1973
Cymatiogalea parviveia Di Milia 1991
Cymatiogalea regularis Yin L. in Chen J. et al. 1985 emend. Yin L. 1986
Cymatiogalea velifera (Downie 1958) Martin 1969
Cymatiogalea virgulta Martin in Martin & Dean 1988
Cymatiogalea wironica Paalits 1992b
Cymatiogalea sp. 1 Paalits 1992b
Cymatiogalea sp. 2 Paalits 1992b
Cymatiogalea sp. 3 Paalits 1992b

Cymatiosphaera Wetzel 1933 ex Deflandre 1954
type species: *Cymatiosphaera radiata* Wetzel 1933

Cymatiosphaera baarstadi Eklund 1990

“*Cymatiosphaera capsulara*” Jankauskas in Jankauskas & Posti 1976
→ *Retisphaeridium capsulatum* (Jankauskas in Jankauskas & Posti 1976) Vanguestaine in Brück & Vanguestaine 2005

Cymatiosphaera crameri Slavíková 1968

Cymatiosphaera cristata Jankauskas 1976

“*Cymatiosphaera favosa*” Jankauskas 1976
→ *Cristallinium cambriense* (Slavíková 1968) Vanguestaine 1978

Cymatiosphaera gotlandica Hagenfeldt 1989a

“*Cymatiosphaera luminosa*” Fombella 1978
→ *Duplisphaera luminosa* (Fombella 1978) Moczydłowska 1998

“?*Cymatiosphaera membranacea*” Kiryanov 1974
→ *Fimbriaglomerella membranacea* Kiryanov 1974 ex Moczydłowska & Vidal 1988

“*Cymatiosphaera minuta*” Jankauskas in Volkova et al. 1979
→ *Fimbriaglomerella minuta* (Jankauskas in Volkova et al. 1979) Moczydłowska & Vidal 1988

Cymatiosphaera cf. *mirabilis* Deunff 1959

Cymatiosphaera cf. *nebulosa* (Deunff 1954b) Deflandre 1954

“*Cymatiosphaera nerisica*” Jankauskas 1976
→ *Cristallinium cambriense* (Slavíková 1968) Vanguestaine 1978 [acc. to Welsch 1986]

“*Cymatiosphaera ovillensis*” Cramer & Díez 1972
→ *Cristallinium cambriense* (Slavíková 1968) Vanguestaine 1978

Cymatiosphaera aff. *pavimenta* (Deflandre 1945) Deflandre 1954

“*Cymatiosphaera postii*” Jankauskas in Volkova et al. 1979
→ *Retisphaeridium postae* (Jankauskas in Volkova et al. 1979) Vanguestaine in Brück & Vanguestaine 2004

“*Cymatiosphaera pusilla*” Moczydłowska 1998
→ *Retisphaeridium pusillum* (Moczydłowska 1998) Vanguestaine in Brück & Vanguestaine 2005

Cymatiosphaera sp. 1 Hagenfeldt 1989b

Cymatiosphaera sp. 1 Volkova et al. 1979

Cymatiosphaera sp. 2 Hagenfeldt 1989a

Cymatiosphaera sp. A Moczydłowska & Stockfors 2004

Cymatiosphaera sp. A Moczydłowska 1998

Cymatiosphaera sp. A Vanguestaine 1978
[dated 1977 in Baudet et al. 1989]

Cymatiosphaera sp. A Vanguestaine 1986

Dactylofusa Brito & Santos 1965 emend Combaz et al. 1967 emend Cramer 1970
type species: *Dactylofusa maranhensis* Brito & Santos 1965

“*Dactylofusa simplex*” Combaz 1967
→ *Leiofusa simplex* (Combaz 1967) Martin 1975
“*Dactylofusa squama*” (Deunff 1961a) Rauscher 1973
→ *Poikilofusa squama* (Deunff 1961a) Martin 1973

Dasydiacrodium Timofeev 1959b ex Deflandre & Deflandre-Rigaud 1962 emend.
Moczyłowska & Stockfors 2004
type species: *Dasydiacrodium eichwaldii* Timofeev 1959b

Dasydiacrodium cf. *angulare* Timofeev 1959b ex Downie & Sarjeant 1965
“*Dasydiacrodium caudatum*” Vanguetstaine 1973b
→ *Ninadiacrodium caudatum* (Vanguetstaine 1973b) Raevskaya & Servais 2009
Dasydiacrodium obsonum Martin in Martin & Dean 1988
Dasydiacrodium palmatilobum Timofeev 1959b ex Downie & Sarjeant 1965
?*Dasydiacrodium setuensis* Paalits 1992a
“*Dasydiacrodium tricornis*” Timofeev 1959b ex Downie & Sarjeant 1965
→ *Acanthodiacrodium tricornis* (Timofeev 1959b ex Downie & Sarjeant 1965) Martin 1969
Dasydiacrodium tumidum (Deunff 1961a) Tongiorgi in Bagnoli et al. 1988
Dasydiacrodium veryhachioides Di Milia et al. 1989
Dasydiacrodium sp. A Moczyłowska & Stockfors 2004
Dasydiacrodium sp. B Moczyłowska & Stockfors 2004

Deunffia Downie 1960 emend. Thusu 1973
type species: *Deunffia monospinosa* (Deunff 1951) Downie 1960

“*Deunffia dentifera*” Volkova 1969
→ *Volkovia dentifera* (Volkova 1969) Downie 1982

“***Dichasphaira***” Vanguetstaine 1973a
[not validly published, acc. to Fensome et al. 1990]
type species: ?

“*Dichasphaera vorax*” Vanguetaine 1973a
[not validly published, acc. to Fensome et al. 1990]

Dichotisphaera Turner 1984
type species: *Dichotisphaera caradocensis* Turner 1984

Dichotisphaera gregalis (Hagenfeldt 1989a) Vanguetaine 1991

Dicrodiacrodium Burmann 1968
type species: *Dicrodiacrodium ancoriforme* Burmann 1968

Dicrodiacrodium ramusculosum (Combaz 1967) Volkova 1990

Dictyotidium Eisenack 1955 emend. Staplin 1961
type species: *Dictyotidium dictyotum* (Eisenack 1938a) Eisenack 1955

Dictyotidium birvetense Paškevičienė in Volkova et al. 1979

“*Dictyotidium cambriense*” Slavíková 1968
→ *Cristallinium cambriense* (Slavíková 1968) Vanguetaine 1978

Dictyotidium hasletianum Vanguetaine 1974

“*Dictyotidium microreticulatum*” Hagenfeldt 1989b
[**senior homonym:** *Dictyotidium microreticulatum* Jiabo 1978 (= Song et al. 1978)]

Dictyotidium perforatum Vidal in Vidal & Peel 1993

Dictyotidium priscum Kiryanov & Volkova in Volkova et al. 1979

Dictyotidium sp. 1 Hagenfeldt 1989b

Dictyotidium sp. 1 Volkova 1980

Dictyotidium sp. 1 Volkova et al. 1979

Dominopolia Kiryanov 1974
type species: *Dominopolia longispinosa* Kiryanov 1974

Dominopolia lata Kiryanov 1974

Dominopolia longispinosa Kiryanov 1974

Duplisphaera Moczydłowska 1998

type species: *Duplisphaera luminosa* (Fombella 1978) Moczydłowska 1998

Duplisphaera luminosa (Fombella 1978) Moczydłowska 1998

Eklundia Jachowicz-Zdanowska 2013

type species: *Eklundia campanula* (Eklund 1990) Jachowicz-Zdanowska 2013

Eklundia campanula (Eklund 1990) Jachowicz-Zdanowska 2013

Eklundia florentinata Jachowicz-Zdanowska 2013

Eklundia pusilla Jachowicz-Zdanowska 2013

Eklundia varia (Volkova 1969) Jachowicz-Zdanowska 2013

Elektoriskos Loeblich 1970

type species: *Elektoriskos aurora* Loeblich 1970

“*Elektoriskos cerinus*” (Volkova 1968) Vanguetaine 1978

→ *Globosphaeridium cerinum* (Volkova 1968) Moczydłowska 1991

“*Elektoriskos flexuosus*” Eklund 1990

→ *Ichnosphaera flexuosa* (Eklund 1990) Jachowicz-Zdanowska 2013

→ {in Brück & Vanguetaine 2004} *Ichnosphaera robusta* Jachowicz-Zdanowska 2013

Elektoriskos sp. A Vidal & Peel 1993

Elenia Volkova 1984

type species: *Elenia armillata* Fanderflit in Umnova & Fanderflit 1971 ex Volkova 1984

Elenia armillata Fanderflit in Umnova & Fanderflit 1971 ex Volkova 1984

Eliasum Fombella 1977

type species: *Eliasum llaniscum* Fombella 1977

Eliasum asturicum Fombella 1977

Eliasum hutchinsonii Martin in Martin & Dean 1984

Eliasum jennessii Martin in Martin & Dean 1984

Eliasum llaniscum Fombella 1977

“*Eliasum microgranulatum*” Hagenfeldt 1989b

→ *Eliasum llaniscum* Fombella 1977 [acc. to Vanguetaine & Brück 2008]

Eliasum pisciforme Fombella 1977

Eliasum pisiformis

[cited without reference in Palacios 2008, 2010; presumably a misspelling of *Eliasum pisciforme* Fombella 1977]

Eliasum n. sp. A Palacios 2010

Eliasum sp. 1 Volkova 1990

Eliasum sp. A Żylińska & Szczepanik 2009

Eomicrhystridium Deflandre 1968

type species: *Eomicrhystridium barghoornii* Deflandre 1968

Estiastra Eisenack 1959a emend. Sarjeant & Stancliffe 1994

type species: *Estiastra magna* Eisenack 1959a

Estiastra minima Volkova 1969

“***Eupoikilofusa***” Cramer 1970

→ *Dactylofusa* Brito & Santos 1965 [acc. to Fensome et al. 1990]

type species: “*Eupoikilofusa striatifera*” (Cramer 1964a) Cramer 1970

→ *Dactylofusa striatifera* (Cramer 1964a) Fensome et al. 1990

“*Eupoikilofusa squama*” (Deunff 1961a) Eisenack et al. 1979

→ *Poikilofusa squama* (Deunff 1961a) Martin 1973

Evittia Brito 1967a emend. Lister 1970

type species: *Evittia sommeri* Brito 1967a

Evittia irregulare Downie 1982

Filisphaeridium Staplin et al. 1965 emend. Sarjeant & Stancliffe 1994 (emend. rejected by Mullins 2001)

type species: *Filisphaeridium setasessitante* (Jansonius 1962) Staplin et al. 1965

Filisphaeridium hirtum (Timofeev 1959b ex Umnova 1975) Sarjeant & Stancliffe 1994

Filisphaeridium papillosum (Timofeev 1959b ex Volkova 1968) Hu Yunxu 1986

Filisphaeridium sp. A Jago et al. 2006

Fimbriaglomerella Loeblich & Drugg 1968

type species: *Fimbriaglomerella divisa* Loeblich & Drugg 1968

Fimbriaglomerella gothlandica

[cited without reference in Jago et al. 2006]

Fimbriaglomerella membranacea Kiryanov 1974 ex Moczydłowska & Vidal 1988

Fimbriaglomerella minuta (Jankauskas in Volkova et al. 1979) Moczydłowska & Vidal 1988

Germinosphaera Mikhailova 1986

[fungi]

type species: *Germinosphaera bispinosa* Mikhailova 1986

Globosphaeridium Moczydłowska 1991

type species: *Globosphaeridium cerinum* (Volkova 1968) Moczydłowska 1991

Globosphaeridium arenulum Jachowicz-Zdanowska 2013

Globosphaeridium cerinum (Volkova 1968) Moczydłowska 1991

Globus Vidal in Moczydłowska & Vidal 1988

type species: *Globus gossipinus* Vidal in Moczydłowska & Vidal 1988

Globus gossipinus Vidal in Moczydłowska & Vidal 1988

Gloeocapsomorpha Zalessky 1917 emend. Foster et al. 1989
[incertae sedis]
type species: *Gloeocapsomorpha prisca* Zalessky 1917

Goniomorpha Yin L. 1986
type species: *Goniomorpha rara* Yin L. 1986

Goniosphaeridium Eisenack 1969 emend. Kjellström 1971a emend. Turner 1984
type species: *Goniosphaeridium polygonale* (Eisenack 1931 ex Eisenack 1938a) Eisenack 1969

“*Goniosphaeridium* aff. *akrochordum*” (Rasul 1979) Dean & Martin 1982
[misspelled as “*akrochodermum*” in Dean & Martin 1982 and Martin 1992]
→ {in Dean & Martin 1982; *pro parte* in Martin 1992} *Polygonium martiniae* Moczydłowska & Crimes 1995 [acc. to Moczydłowska & Stockfors 2004]
→ {*pro parte* in Martin 1992} *Solisphaeridium akrochordum* (Rasul 1979) Moczydłowska & Stockfors 2004

Goniosphaeridium canningia (Combaz & Peniguel 1972) Playford & Martin 1984

“*Goniosphaeridium* aff. *dentatum*” (Timofeev 1959b ex Konzalová-Mazancová 1969) Rauscher 1973
→ *Polygonium pungens* (Timofeev 1959b ex Martin 1969) Albani 1989 [acc. to Moczydłowska & Stockfors 2004]

“*Goniosphaeridium implicatum*” (Fridriksone 1971) Downie 1982
→ *Parmasphaeridium implicatum* (Fridriksone 1971) Jachowicz-Zdanowska 2013

Goniosphaeridium primarium (Jankauskas in Volkova et al. 1979) Downie 1982

Goniosphaeridium rasulii Welsch 1986

“*Goniosphaeridium regulare*” Yin L. in Chen J. et al. 1985
→ *Stellechinatum uncinatum* (Downie 1958) Molyneux 1987 [acc. to Yin L. 1986]

Goniosphaeridium tener Timofeev 1959b ex Elaouad-Debbaj 1988

Goniosphaeridium tuberatum (Downie 1958) Welsch 1986
[comb. not validly published in Wolf 1980]

“*Goniosphaeridium uncinatum*” (Downie 1958) Kjellström 1971b
→ *Stellechinatum uncinatum* (Downie 1958) Molyneux 1987

“*Goniosphaeridium varium*” (Volkova 1969) Downie 1982
→ *Eklundia varia* (Volkova 1969) Jachowicz-Zdanowska 2013

Goniosphaeridium volkovae Hagenfeldt 1989a

Goniosphaeridium sp. A Downie 1982

Goniosphaeridium sp. A Vidal & Peel 1993

Goniosphaeridium sp. B Downie 1982

Gorgonisphaeridium Staplin et al. 1965 emend. Kiryanov 1978

type species: *Gorgonisphaeridium winslowiae* Staplin et al. 1965

?*Gorgonisphaeridium* sp. A Wood & Clendening 1982

?*Gorgonisphaeridium* sp. B Wood & Clendening 1982

?*Gorgonisphaeridium* sp. C Wood & Clendening 1982

?*Gorgonisphaeridium* sp. D Wood & Clendening 1982

?*Gorgonisphaeridium* sp. E Wood & Clendening 1982

?*Gorgonisphaeridium* sp. F Wood & Clendening 1982

Granomarginata Naumova 1961

type species: *Granomarginata prima* Naumova 1961

Granomarginata parva Jachowicz-Zdanowska 2013

Granomarginata prima Naumova 1961

Granomarginata squamacea Volkova 1968

Granomarginata vulgaris Naumova 1968

Granularia Kowalski 1983

type species: *Granularia saccoformis* Kowalski 1983

Granularia saccoformis Kowalski 1983

Heliosphaeridium Moczyłowska 1991

type species: *Heliosphaeridium dissimulare* (Volkova 1969) Moczyłowska 1991

Heliosphaeridium ampliatum (Wang 1985) Yao et al. 2005

“*Heliosphaeridium bellulum*” Moczyłowska 1998

→ *Ammonidium bellulum* (Moczyłowska 1998) Sarjeant & Stancliffe 2000
Heliosphaeridium coniferum (Downie 1982) Moczyłowska 1991
Heliosphaeridium dissimulare (Volkova 1969) Moczyłowska 1991
Heliosphaeridium exile Moczyłowska 1998
Heliosphaeridium lanceolatum (Vanguetaine 1974) Moczyłowska 1998
“?*Heliosphaeridium llynense*” Martin in Young et al. 1994
→ *Multiplicisphaeridium llynense* (Martin in Young et al. 1994) Jachowicz-Zdanowska 2013
Heliosphaeridium longum (Moczyłowska 1988) Moczyłowska 1998
Heliosphaeridium lubomlense (Kiryanov 1974) Moczyłowska 1991
Heliosphaeridium nodosum Moczyłowska 1998
“*Heliosphaeridium notatum*” (Volkova 1969) Moczyłowska 1991
→ *Ammonidium notatum* (Volkova 1969) Jachowicz-Zdanowska 2013
Heliosphaeridium obscurum (Volkova 1969) Moczyłowska 1991
“*Heliosphaeridium oligum*” (Jankauskas in Jankauskas & Posti 1976) Moczyłowska 1998
→ *Ammonidium oligum* (Jankauskas in Jankauskas & Posti 1976) Jachowicz-Zdanowska 2013
Heliosphaeridium radzyncicum (Volkova in Volkova et al. 1979) Moczyłowska 1991
Heliosphaeridium serridentatum Moczyłowska 1998
Heliosphaeridium n. sp. B Moczyłowska & Crimes 1995

Hemisphaerium Hemer & Nygreen 1967

→ *Brazilea* Tiwari & Navale 1967 [acc. to Grenfell 1995; Zygnemataceae]
type species: *Hemisphaerium inornatum* Hemer & Nygreen 1967

Hubeisphaera Xing & Liu 1978

type species: *Hubeisphaera radiata* Xing & Liu 1978

Hubeisphaera radiata Xing & Liu 1978

Huroniospora Barghoorn in Barghoorn & Tyler 1965

type species: *Huroniospora microreticulata* Barghoorn in Barghoorn & Tyler 1965

Huroniospora microreticulata Barghoorn in Barghoorn & Tyler 1965

Ichnosphaera Jachowicz-Zdanowska 2013

type species: *Ichnosphaera flexuosa* (Eklund 1990) Jachowicz-Zdanowska 2013

Ichnosphaera aranea Jachowicz-Zdanowska 2013

Ichnosphaera delicata Jachowicz-Zdanowska 2013

Ichnosphaera flexuosa (Eklund 1990) Jachowicz-Zdanowska 2013

Ichnosphaera robusta Jachowicz-Zdanowska 2013

Ichnosphaera stipatica (Hagenfeldt 1989a) Jachowicz-Zdanowska 2013

Impluviculus Loeblich & Tappan 1969 emend. Martin 1975

type species: *Impluviculus milonii* (Deunff 1968b) Loeblich & Tappan 1969

?*Impluviculus bibulbulus* Parsons & Anderson 2000

Impluviculus cleae (Martin 1972) Martin 1975

Impluviculus lenticularis Martin 1975

Impluviculus milonii (Deunff 1968b) Loeblich & Tappan 1969

Impluviculus multiangularis (Umnova in Umnova & Fanderflit 1971) Volkova 1990

Impluviculus stellaris Martin 1975

Impluviculus villosiusculus Volkova 1990

Impluviculus sp. A Martin & Dean 1988

?*Impluviculus* sp. A Parsons & Anderson 2000

Izhoria Golub & Volkova in Volkova & Golub 1985

type species: *Izhoria angulata* Golub & Volkova in Volkova & Golub 1985

Izhoria angulata Golub & Volkova in Volkova & Golub 1985

“*Kildinella*” Shepeleva & Timofeev 1963 ex Timofeev 1966

→ *Leiosphaeridia* Eisenack 1958a [acc. to Volkova 1968 and Lindgren 1982]

type species: “*Kildinella hyperboreica*” Timofeev 1966

→ *Leiosphaeridia hyperboreica* (Timofeev 1966) Fensome et al. 1990

“*Kildinosphaera*” Vidal in Vidal & Siedlecka 1983

→ *Valeria* Jankauskas 1982 [acc. to Fensome et al. 1990]

type species: “*Kildinosphaera chagrinata*” Vidal in Vidal & Siedlecka 1983
→ *Valeria sinica* (Timofeev 1966) Fensome et al. 1990

“*Kildinosphaera chagrinata*” Vidal in Vidal & Siedlecka 1983
→ *Valeria sinica* (Timofeev 1966) Fensome et al. 1990

“*Kildinosphaera granulata*” Vidal in Vidal & Siedlecka 1983
→ *Megasacculina atava* Naumova 1961

Ladogella Golub & Volkova in Volkova & Golub 1985

type species: *Ladogella rotundiformis* Golub & Volkova in Volkova & Golub 1985

?*Ladogella aries* Di Milia et al. 1989

Ladogella filifera Di Milia et al. 1989

?*Ladogella intermedia* Parsons & Anderson 2000

Ladogella rommelaerei (Martin in Martin & Dean 1981) Di Milia et al. 1989

Ladogella rotundiformis Golub & Volkova in Volkova & Golub 1985

Ladogella saharica Vecoli & Playford 1997

Ladogella volkovae Di Milia et al. 1989

Ladogella sp. A Parsons & Anderson 2000

Lechistania Jachowicz-Zdanowska 2013

type species: *Lechistania magna* Jachowicz-Zdanowska 2013

Lechistania magna Jachowicz-Zdanowska 2013

Leiofusa Eisenack 1938a emend. Eisenack 1965a emend. Combaz et al. 1967 emend. Cramer 1970

type species: *Leiofusa fusiformis* Eisenack 1934 ex Eisenack 1938a

Leiofusa cf. *bicornuta* Xing & Liu 1973

Leiofusa cf. *gravida* Pittau 1985

“*Leiofusa pristina*” Potter in Baudelot & Géry 1979

[nomen nudum, acc. to Fensome et al. 1990; Baudelot & Géry 1979 gave authorship as “Potter 1974”, referring to an unpublished thesis]

Leiofusa simplex (Combaz 1967) Martin 1975

“*Leiofusa squama*” Deunff 1961a
→ *Poikilofusa squama* (Deunff 1961a) Martin 1973
Leiofusa stoumonensis Vanguetaine 1973b

“***Leioligotriletes***” Timofeev 1958
[not validly published acc. to Fensome et al. 1990]
variant: “*Leioligotriletum*”
type species: “*Leioligotriletes crassus*” (Naumova 1949) Timofeev 1958 [not validly published, acc. to Fensome et al. 1990]

“*Leioligotriletes crassus*” (Naumova 1949) Timofeev 1958
[not validly published, acc. to Fensome et al. 1990]

Leiomarginata Naumova 1961
type species: *Leiomarginata simplex* Naumova 1961

Leiomarginata simplex Naumova 1961

“***Leiominuscula***” Naumova 1961
→ *Leiosphaeridia* Eisenack 1958a [acc. to Cramer & Díez 1979]
type species: “*Leiominuscula minuta*” Naumova 1961
→ *Leiosphaeridia riphiana* Fensome et al. 1990

“*Leiominuscula minuta*” Naumova 1961
→ *Leiosphaeridia riphiana* Fensome et al. 1990

“***Leiopsophosphaera***” Naumova 1961 ex Naumova 1968
→ *Leiosphaeridia* Eisenack 1958a [acc. to Volkova 1968 and Lindgren 1982]
type species: “*Leiopsophosphaera convexiplicata*” Naumova 1968

“*Leiopsophosphaera indefinita*” Pichova
[cited as “*Leiopsophosphaera indefinitus* Pichova” without complete reference in Pykhova 1967]

“*Leiopsophosphaera infriata*” Shepeleva
[cited without complete reference in Ding et al. 1992]
[possible homonym: “*Leiopsophosphaera infriata* (Andr.) Sin et Liu (comb. nov.)” in Xing &

Liu 1978, wherein the basionym was not properly referenced; Fensome et al. 1990 also reported citations of “*Leiopsophosphaera infriata*” without complete references and authorship given as: “(Andreeva)” in Xing et al. 1979; “(Timofeev)” in Xing & Liu 1980]

“*Leiopsophosphaera pseudozonalis*” Wang et Chen
[cited without complete reference in Wang & Chen Q. 1987]

“*Leiopsophosphaera rotunda*” Pykhova 1967
[not validly published, acc. to Fensome et al. 1990]

“*Leiopsophosphaera solida*” (Liu et Sin) Xing & Liu 1978
[the basionym was not properly referenced in Xing & Liu 1978, most likely
Protoleiosphaeridium solidum Liu & Xing in Xing & Liu 1973]

Leiosphaeridia Eisenack 1958a emend. Downie & Sarjeant 1963 emend. Turner 1984
type species: *Leiosphaeridia baltica* Eisenack 1958a

Leiosphaeridia annulata Yin L. in Chen J. et al. 1985

Leiosphaeridia asperata (Naumova 1950) Lindgren 1982

“*Leiosphaeridia atava*” (Naumova 1961) Cramer & Díez 1979
→ *Megasacculina atava* Naumova 1961

Leiosphaeridia bicrura Jankauskas in Jankauskas & Posti 1976

Leiosphaeridia cerebriformis Volkova 1969

Leiosphaeridia crassa (Naumova 1949) Jankauskas in Jankauskas et al. 1989
[**junior homonym:** “*Leiosphaeridia crassa*” (Pykhova 1973) Fensome et al. 1990]

Leiosphaeridia dehisca Paškevičienė in Volkova et al. 1979

“*Leiosphaeridia densa*” (Timofeev 1966) Pyatiletov 1978
[**junior homonym:** *Leiosphaeridia densa* (Maithy 1975) Nautiyal 1983]
→ *Protoleiosphaeridium laccatum* (Timofeev 1966) Fensome et al. 1990 [acc. to Vidal 1974]

Leiosphaeridia desmosa Yin L. in Chen J. et al. 1985

“*Leiosphaeridia fragile*” Downie 1982
→ *Saharidia fragilis* (Downie 1958) Combaz 1967

Leiosphaeridia fumiana Vanguetstaine 1974

“*Leiosphaeridia gregalis*” Hagenfeldt 1989a
→ *Dichotisphaera gregalis* (Hagenfeldt 1989a) Vanguetstaine 1991

Leiosphaeridia macrostomata Yin L. in Chen J. et al. 1985

Leiosphaeridia minutissima (Naumova 1949) Jankauskas in Jankauskas et al. 1989

“*Leiosphaeridia papillata*” (Staplin 1961) Downie & Sarjeant 1965
→ *Lophosphaeridium papillatum* (Staplin 1961) Martin 1969

Leiosphaeridia pylomifera Paškevičienė in Volkova et al. 1979

Leiosphaeridia subgranulata Kiryanov 1974
Leiosphaeridia tenuissima Eisenack 1958b
Leiosphaeridia ternata (Timofeev 1966) Mikhailova & Jankauskas in Jankauskas et al. 1989
Leiosphaeridia tubulosa Eisenack 1963
Leiosphaeridia sp. 1 Cramer & Díez 1972
Leiosphaeridia sp. 1 Eklund 1990
Leiosphaeridia sp. 1 Moczydłowska & Vidal 1986
Leiosphaeridia sp. 1 Volkova et al. 1979
Leiosphaeridia sp. 2 Moczydłowska & Vidal 1986
Leiosphaeridia sp. A Parsons & Anderson 2000

Leiovalia Eisenack 1965a ex Górka 1969
type species: *Leiovalia ovalis* (Eisenack 1938a) Górka 1969

Leiovalia tenera Kiryanov 1974

Liepaina Jankauskas and Volkova in Volkova et al. 1979
type species: *Liepaina plana* Jankauskas and Volkova in Volkova et al. 1979

Liepaina plana Jankauskas and Volkova in Volkova et al. 1979

Liepaina rigida Eklund 1990

?*Liepaina* n. sp. Vidal 1981

→ *Turrisphaeridium semireticulatum* (Timofeev 1959b) Jachowicz-Zdanowska 2013

Lophodiacrodium Timofeev 1958 emend. Deflandre & Deflandre-Rigaud 1962
type species: *Lophodiacrodium obtusum* Timofeev 1958

Lophodiacrodium valdaicum (Timofeev 1959b) Downie & Sarjeant 1965

“*Lopholigotriletum*” Timofeev 1959b
[not validly published, acc. to Fensome et al. 1990]
no type species

“*Lopholigotritetum spathaeforme*” Timofeev 1959b
→ *Lophosphaeridium spathaeforme* Timofeev 1959b ex Hu Yunxu 1986

Lophomiuscula Naumova 1961
type species: *Lophomiuscula prima* Naumova 1961

Lophomiuscula acietata Sun in Sun & Zhai 1989
[complete reference only available in Chinese]

Lophomiuscula crassa Xing & Liu 1978

Lophomiuscula prima Naumova 1961

Lophopsophosphaera Naumova
[cited without complete reference in Pykhova 1967; most likely a misspelling of
Leiopsophosphaera Naumova 1961]
type species: ?

Lophosphaeridium Timofeev 1959b ex Downie 1963 emend. Lister 1970
type species: *Lophosphaeridium rarum* Timofeev 1959b ex Downie 1963

Lophosphaeridium acietatum Xing & Liu 1978

Lophosphaeridium bacilliferum Vanguetaine 1974

Lophosphaeridium bottnicum Hagenfeldt 1989b

“*Lophosphaeridium citrinum*” Downie 1963

→ *Baltisphaeridium citrinum* (Downie 1963) Stockmans & Willièrè 1974

“*Lophosphaeridium dubium*” (Volkova 1968) Moczydłowska 1991

[**senior hyponym:** *Lophosphaeridium dubium* Jankauskas & Vaitekunene 1972, acc. to
Molyneux et al. 1996]

Lophosphaeridium kryptoradiatum Vanguetaine 1974

Lophosphaeridium latviense (Volkova 1974) Moczydłowska 1998

Lophosphaeridium orbiculatum Xing 1962

Lophosphaeridium papillatum (Staplin 1961) Martin 1969

Lophosphaeridium spathaeforme Timofeev 1959b ex Hu Yunxu 1986

Lophosphaeridium tentativum Volkova 1968

Lophosphaeridium truncatum Volkova 1969

Lophosphaeridium tynnii Hagenfeldt 1989b

Lophosphaeridium variabile Volkova 1974

Lophosphaeridium yichangense Xing & Liu 1978

Lophosphaeridium sp. A Potter 1974

Lusatia Burmann 1970 emend. Sarjeant & Vavrdová 1997

type species: *Lusatia dendroidea* Burmann 1970 emend. Albani et al. 2007

Lusatia dendroidea Burmann 1970 emend. Albani et al. 2007

“*Lusatia triangularis*” (Umnova 1975) Volkova 1990

→ *Lusatia dendroidea* Burmann 1970 emend. Albani et al. 2007

Macroptycha Timofeev 1973a ex Timofeev et al. 1976

type species: *Macroptycha uniplicata* Timofeev in Timofeev et al. 1976

Macroptycha uniplicata Timofeev in Timofeev et al. 1976

Margominuscula Naumova 1961

type species: *Margominuscula rugosa* Naumova 1961

Margominuscula rugosa Naumova 1961

Medousapalla Wood & Clendening 1982

type species: *Medousapalla choanoklosma* Wood & Clendening 1982

Medousapalla choanoklosma Wood & Clendening 1982

Megasacculina Naumova 1961

type species: *Megasacculina atava* Naumova 1961

Megasacculina atava Naumova 1961

Micrhystridium Deflandre 1937 emend. Staplin 1961 emend. Downie & Sarjeant 1963
emend. Staplin et al. 1965 emend. Sarjeant 1967 emend. Lister 1970 emend. Sarjeant &
Stancliffe 1994
type species: *Micrhystridium inconspicuum* (Deflandre 1935) Deflandre 1937

Micrhystridium ampliatum Wang 1985

[**junior homonym:** “*Micrhystridium ampliatum*” Wicander & Playford 1985, acc. to Fensome
et al. 1990]

→ *Heliosphaeridium ampliatum* (Wang 1985) Yao et al. 2005

Micrhystridium breviacanthum Slavíková 1968

Micrhystridium brevicornum Jankauskas 1976

→ {in Welsch 1986} *Heliosphaeridium serridentatum* Moczydłowska 1998

Micrhystridium cerinum

[cited without reference in Yang & Yin L. 2001]

Micrhystridium confusum (Jankauskas 1975) Fensome et al. 1990

[acc. to Fensome et al. 1990, this combination was not validly published in Pozaryski et al.
1981]

“*Micrhystridium coniferum*” Downie 1982

→ *Heliosphaeridium coniferum* (Downie 1982) Moczydłowska 1991

Micrhystridium cf. *coronatum* Stockmans & Willièrè 1963

“*Micrhystridium dissimulare*” Volkova 1969

→ *Heliosphaeridium dissimulare* (Volkova 1969) Moczydłowska 1991

Micrhystridium dubium

[cited without reference in Yang & Yin L. 2001]

Micrhystridium echinulatum Luo & Wang in Wang & Luo 1984

Micrhystridium ellaense Downie 1982

“*Micrhystridium ellipticum*” Downie 1982

→ *Heliosphaeridium obscurum* (Volkova 1969) Moczydłowska 1991

Micrhystridium flexispinosum Downie 1982

“*Micrhystridium gogense*” Downie 1982

→ *Comasphaeridium gogense* (Downie 1982) Sarjeant & Stancliffe 1994

Micrhystridium guizhouense Fensome et al. 1990

Micrhystridium henryi Paris & Deunff 1970

“*Micrhystridium lanatum*” Volkova 1969

→ *Asteridium lanatum* (Volkova 1969) Moczydłowska 1991

“*Micrhystridium lanceolatum*” Vanguetstaine 1974

[**junior homonym:** “*Micrhystridium lanceolatum*” Yin L. 1985, acc. to Palacios & Vidal 1992]

→ *Heliosphaeridium lanceolatum* (Vanguetstaine 1974) Moczydłowska 1998

“*Micrhystridium lanceolatum*” Yin L. 1985

[**senior homonym:** “*Micrhystridium lanceolatum*” Vanguetstaine 1974, **now** *Asteridium
lanceolatum* (Vanguetstaine 1974) Moczydłowska 1998, acc. to Palacios & Vidal 1992]

- Micrhystridium longispiniferum* Wang & Luo 1984
- “*Micrhystridium longum*” Moczydłowska 1988
→ *Heliosphaeridium longum* (Moczydłowska 1988) Moczydłowska 1998
- “*Micrhystridium lubomlense*” Kiryanov 1974
→ *Heliosphaeridium lubomlense* (Kiryanov 1974) Moczydłowska 1991
- Micrhystridium maidingense* Wang & Chen Q. 1987
- Micrhystridium minimum* Xing
[cited without complete reference in Yin C. & Gao 1996]
- Micrhystridium minutum* Downie 1982
[**junior homonym:** “*Micrhystridium minutum*” Luo & Wang in Wang & Luo 1984, acc. to Fensome et al. 1990]
- “*Micrhystridium minutum*” Luo & Wang in Wang & Luo 1984
[**senior homonym:** *Micrhystridium minutum* Downie 1982, acc. to Fensome et al. 1990]
→ *Micrhystridium guizhouense* Fensome et al. 1990
- Micrhystridium ningqiangense* Xing
[cited without complete reference in Yin C. & Gao 1996]
- “*Micrhystridium notatum*” Volkova 1968
→ *Ammonidium notatum* (Volkova 1969) Jachowicz-Zdanowska 2013
- Micrhystridium obscurum* Volkova 1969
→ *Heliosphaeridium obscurum* (Volkova 1969) Moczydłowska 1991
- “*Micrhystridium oligum*” Jankauskas in Jankauskas & Posti 1976
→ *Ammonidium oligum* (Jankauskas in Jankauskas & Posti 1976) Jachowicz-Zdanowska 2013
- “*Micrhystridium ordense*” Downie 1982
→ *Asteridium ordense* (Downie 1982) Vidal & Peel 1993
- “*Micrhystridium pallidum*” Volkova 1968
→ *Asteridium pallidum* (Volkova 1968) Moczydłowska 1991
- ?*Micrhystridium profusum* Wicander 1974
- Micrhystridium pseudozonale* Wang & Chen Q. 1987
- “*Micrhystridium radzyncicum*” Volkova in Volkova et al. 1979
→ *Heliosphaeridium radzyncicum* (Volkova in Volkova et al. 1979) Moczydłowska 1991
- Micrhystridium regulare* Yin L. 1987
- Micrhystridium resistens* Timofeev 1959b ex Deflandre & Deflandre-Rigaud 1965
- Micrhystridium semiapertum* Welsch 1986
- Micrhystridium setulerum* Wang et al. 1987
[cited as “in press” in Wang & Chen Q. 1987]
- Micrhystridium shinetonense* Downie 1958
- “*Micrhystridium sinuosum*” Potter in Baudelot & Géry 1979
[nomen nudum, acc. to Fensome et al. 1990; Baudelot & Géry 1979 gave authorship as “Potter 1974”, referring to an unpublished thesis]
- “*Micrhystridium spinosum*” Volkova 1969

→ *Asteridium spinosum* (Volkova 1969) Moczydłowska 1998

Micrhystridium stellatum Deflandre 1945

“*Micrhystridium tornatum*” Volkova 1968

→ *Asteridium tornatum* (Volkova 1968) Moczydłowska 1991

Micrhystridium varium

[cited without reference in Yang & Yin L. 2001]

Micrhystridium villosum Kiryanov 1974

Micrhystridium n. sp. Vidal 1981

Micrhystridium sp. 1 Cramer & Díez 1972

Micrhystridium sp. 1 Hagenfeldt 1989b

Micrhystridium sp. 1 Paalits 2005

Micrhystridium sp. 2 Hagenfeldt 1989b

Micrhystridium sp. 2 Paalits 2005

Micrhystridium sp. A Albani et al. 1991

Micrhystridium sp. A Knoll & Swett 1987

Micrhystridium sp. A Parsons & Anderson 2000

Micrhystridium sp. B Albani et al. 1991

Micrhystridium sp. B Potter 1974

Microconcentrica Naumova 1961 ex Naumova 1968

type species: *Microconcentrica atava* Naumova 1961 ex Naumova 1968

Microconcentrica atava Naumova 1961 ex Naumova 1968

Microconcentrica induplicata Liu & Xing in Xing & Liu 1973

Monocrodium Pittau 1985

type species: *Monocrodium mediterraneum* Pittau 1985

Monotrematosphaeridium Sin in Xing & Liu 1978

[authorship in Xing & Liu 1978 was given as “Sin 1962 (MS)”, presumably referencing an unpublished manuscript or Xing 1962]

type species: *Monotrematosphaeridium asperum* Xing & Liu 1978

“*Monotrematosphaeridium quadratum*” Sin 1962

[probably not validly published]

Monotrematosphaeridium simplex Wang et Chen

[cited without complete reference in Wang & Chen Q. 1987]

Multiplicisphaeridium Staplin 1961 emend. Staplin et al. 1965 emend. Eisenack 1969 emend. Lister 1970 emend. Eiserhardt 1992 emend. Sarjeant & Vavrdová 1997
type species: *Multiplicisphaeridium ramispinosum* Staplin 1961

Multiplicisphaeridium ?ancliforme Fombella 1978

“*Multiplicisphaeridium campanulum*” Eklund 1990

→ *Eklundia campanula* (Eklund 1990) Jachowicz-Zdanowska 2013

Multiplicisphaeridium cervinacornuum Welsch 1986

Multiplicisphaeridium chakor Vanguetaine & Van Looy 1983

Multiplicisphaeridium constipatum Eklund 1990

Multiplicisphaeridium dactylum Vidal in Moczydłowska & Vidal 1988

“*Multiplicisphaeridium dendroideum*” (Jankauskas 1976) Jankauskas & Kiryanov in Volkova et al. 1979

[**senior homonyms:** *Multiplicisphaeridium dendroidium* Morbey 1975; *Multiplicisphaeridium dendroideum* (Burmans 1970) Eisenack et al. 1976, acc. to Fensome et al. 1990]

→ *Multiplicisphaeridium xianum* Fombella 1977 [acc. to Moczydłowska 1998]

Multiplicisphaeridium cf. *eodigitatum* Fombella 1978

Multiplicisphaeridium cf. *eopiriferum* Fombella 1978

“?*Multiplicisphaeridium* cf. *furcatum*” (Deunff 1961a) Eisenack et al. 1973

→ *Stelliferidium* cf. *furcatum* (Deunff 1961a) emend. Deunff et al. 1974

Multiplicisphaeridium jaroslavicum Volkova 1990

“*Multiplicisphaeridium lancarae*” Cramer & Díez 1972

→ *Timofeevia lancariae* (Cramer & Díez 1972) Vanguetaine 1978

Multiplicisphaeridium llynense (Martin in Young et al. 1994) Jachowicz-Zdanowska 2013

Multiplicisphaeridium martae Cramer & Díez 1972

Multiplicisphaeridium ramosum Moczydłowska 1998

“*Multiplicisphaeridium raquelinae*” Cramer & Díez 1972

→ *Timofeevia raquelinae* (Cramer & Díez 1972) Cramer & Díez 1979

Multiplicisphaeridium parvum (Hagenfeldt 1989b) Moczydłowska 1998

Multiplicisphaeridium sosnowiecense Moczydłowska 1998

Multiplicisphaeridium varietatis Moczydłowska 1998

“*Multiplicisphaeridium vilnense*” (Jankauskas 1976) Jankauskas in Volkova et al. 1979

→ *Timofeevia lancariae* (Cramer & Díez 1972) Vanguetaine 1978

Multiplicisphaeridium waltonii Downie 1982
Multiplicisphaeridium xianum Fombella 1977
Multiplicisphaeridium yankauskasii Fensome et al. 1990
Multiplicisphaeridium sp. 1 Volkova 1990

Myxococcoides Schopf 1968
type species: *Myxococcoides minor* Schopf 1968

Myxococcoides inornata Schopf 1968
Myxococcoides minor Schopf 1968
Myxococcoides staphylidion Lo 1980

Nanocyclopia Loeblich & Wicander 1976
type species: *Nanocyclopia aspratilis* Loeblich & Wicander 1976

Nanocyclopia sp. 1 Uutela 2008

Navifusa Combaz et al. 1967 ex Eisenack 1976
type species: *Navifusa navis* (Eisenack 1938b) Eisenack 1976

Nellia Golub & Volkova in Volkova & Golub 1985
type species: *Nellia longiuscula* Golub & Volkova in Volkova & Golub 1985

Nellia acifera (Umnova in Umnova & Fanderflit 1971) Volkova 1990
? *Nellia longispinata* Parsons & Anderson 2000
Nellia longiuscula Golub & Volkova in Volkova & Golub 1985
Nellia magna Volkova 1990
Nellia sukatschevii (Timofeev 1959b) Volkova 1990

Neoveryhachium Cramer 1970
type species: *Neoveryhachium carminae* (Cramer 1964b) Cramer 1970

Ninadiacrodium Raevskaya & Servais 2009

type species: *Ninadiacrodium dumontii* (Vanguetaine 1973b) Raevskaya & Servais 2009

Ninadiacrodium caudatum (Vanguetaine 1973b) Raevskaya & Servais 2009

Ninadiacrodium dumontii (Vanguetaine 1973b) Raevskaya & Servais 2009

Nodosus Hagenfeldt 1989b

type species: *Nodosus irregulare* Hagenfeldt 1989b

Nodosus irregulare Hagenfeldt 1989b

Nucellophaeridium Timofeev 1966

type species: *Nucellophaeridium deunffii* Timofeev 1966

Nucellophaeridium sp. A Martin & Dean 1983

“*Ocridoligo-triletes*” Timofeev 1958

variant: “*Ocridoligo-triletum*”

→ *Trachytriletes* Naumova 1939 ex Naumova 1949 [acc. to Fensome et al. 1990]

type species: *O. kryshstofovichii* (Naumova 1949) Timofeev 1958 [comb. illegitimate acc. to Fensome et al. 1990]

→ *Trachytriletes kryshstofovichii* Naumova 1949

“*Ocridoligo-triletum kryshstofovichii*” (Naumova 1949) Timofeev 1958 [comb. illegitimate acc. to Fensome et al. 1990]

→ *Trachytriletes kryshstofovichii* Naumova 1949

Octaedryxium Rudavskaya 1973a

or *Octoedryxium* Rudavskaya 1973b [uncertain seniority acc. to Fensome et al. 1990]

type species: *Octaedryxium truncatum* Rudavskaya 1973a or Rudavskaya 1973b

Octaedryxium minutum Wang & Chen Q. 1987

Octaedryxium simplex Wang & Chen Q. 1987

Ooidium Timofeev 1957 emend. Norris & Sarjeant 1965
type species: *Ooidium rossicum* Timofeev 1957

?*Ooidium clavigerum* Parsons & Anderson 2000

“*Ooidium* aff. *revinium*” Vanguetaine 1973b

→ *Trunculumarium* aff. *revinium* (Vanguetaine 1973b) Loeblich & Tappan 1976

Ooidium rossicum Timofeev 1957

Ooidium timofeevii Loeblich 1970

Ooidium sp. A Vanguetaine 1986

?*Ooidium* sp. A Martin & Dean 1981

Orthosphaeridium Eisenack 1968 emend. Kjellström 1971a emend. Turner 1984
type species: *Orthosphaeridium rectangulare* (Eisenack 1963) Eisenack 1968

?*Orthosphaeridium extensum* Parsons & Anderson 2000

→ {*pro parte* in Parsons & Anderson 2000} *Lusatia dendroidea* Burmann 1970 emend. Albani et al. 2007

“?*Orthosphaeridium triangulare*” (Umnova 1975) Parsons & Anderson 2000

→ *Lusatia dendroidea* Burmann 1970 emend. Albani et al. 2007

Orygmatosphaeridium Timofeev 1959b
type species: *Orygmatosphaeridium ruminatum* Timofeev 1959b

Orygmatosphaeridium holtedahlii (Timofeev 1966) Fensome et al. 1990

“*Orygmatosphaeridium semireticulatum*” Andreeva 1966

→ *Orygmatosphaeridium holtedahlii* (Timofeev 1966) Fensome et al. 1990

“**Ovulum**” Jankauskas 1975

[not validly published, acc. to Fensome et al. 1990]

→ *Revinotesta* Vanguetaine 1974 [acc. to Cramer & Díez 1979]

type species: *Ovulum saccatum* Jankauskas 1975

→ *Revinotesta saccata* (Jankauskas 1975) ex Fensome et al. 1990

“*Ovulum lanceolatum*” Jankauskas 1975

→ *Revinotesta lanceolata* Jankauskas 1975 ex Fensome et al. 1990

“*Ovulum saccatum*” Jankauskas 1975

→ *Revinotesta saccata* Jankauskas 1975 ex Fensome et al. 1990

Paracymatiosphaera Wang 1985

type species: *Paracymatiosphaera regularis* Wang 1985

Paracymatiosphaera annularis Wang 1985

Paracymatiosphaera hunnanensis Wang 1985

Paracymatiosphaera irregularis Wang 1985

Paracymatiosphaera regularis Wang 1985

Paracymatiosphaera spinosa Yin C. in Yin C. & Liu 1988

Parmasphaeridium Jachowicz-Zdanowska 2013

type species: *Parmasphaeridium implicatum* (Fridriksone 1971) Jachowicz-Zdanowska 2013

Parmasphaeridium implicatum (Fridriksone 1971) Jachowicz-Zdanowska 2013

Parmasphaeridium robustispinosum Jachowicz-Zdanowska 2013

Peramorpha Martin in Martin & Dean 1983

type species: *Peramorpha manuelsenensis* Martin in Martin & Dean 1983

Peramorpha manuelsenensis Martin in Martin & Dean 1983

Petaloferidium Jacobson 1978

type species: *Petaloferidium stigii* Jacobson 1978

Petaloferidium lacrimiferum Palacios et al. 2009

Peteinosphaeridium Staplin et al. 1965

type species: *Peteinosphaeridium bergstroemii* Staplin et al. 1965

Phenacoon Vecoli & Playford 1997

type species: *Phenacoon imperfectum* Vecoli & Playford 1997

Phenacoon imperfectum Vecoli & Playford 1997

Pirea Vavrdová 1972

type species: *Pirea dubia* Vavrdová 1972

Pirea orbicularis Volkova 1990

“*Plicatosphaera*” Potter in Baudelot & Géry 1979

[nomen nudum, acc. to Fensome et al. 1990]

type species: “*Plicatosphaera elementaria*” Potter in Baudelot & Géry 1979 [nomen nudum acc. to Fensome et al. 1990]

“*Plicatosphaera elementaria*” Potter in Baudelot & Géry 1979

[nomen nudum, acc. to Fensome et al. 1990; Baudelot & Géry 1979 gave authorship as “Potter 1974”, referring to an unpublished thesis]

Poikilofusa Staplin et al. 1965

type species: *Poikilofusa spinata* Staplin et al. 1965

Poikilofusa chalaza Rasul 1979

Poikilofusa squama (Deunff 1961a) Martin 1972

Polyedryxium Deunff 1954a emend. Deunff 1971

type species: *Polyedryxium deflandrei* Deunff 1954a ex Deunff 1961b

“*Polyedryxium sarjeantii*” Slavíková 1968

[not validly published, acc. to Fensome et al. 1990]

Polygonium Vavrdová 1966 restrict. Le Hérisse 1989 emend. Sarjeant & Stancliffe 1994
emend. Moczyłowska & Stockfors 2004
type species: *Polygonium gracile* Vavrdová 1966

“*Polygonium dentatum*” (Timofeev 1959b ex Konzalová-Mazancová 1969) Albani 1989
→ *Polygonium pungens* (Timofeev 1959b ex Martin 1969) Albani 1989 [acc. to Moczyłowska
& Stockfors 2004]

Polygonium gracile Vavrdová 1966 emend. Jacobson & Achab 1985 emend. Sarjeant & Stancliffe
1996

Polygonium martinae Moczyłowska & Crimes 1995

Polygonium minimum Timofeev 1959b ex Volkova 1990

Polygonium pellicidum (Timofeev 1959b ex Tynni 1975) Volkova 1990 emend. Moczyłowska &
Stockfors 2004

Polygonium pungens (Timofeev 1959b ex Martin 1969) Albani 1989
[this combination was not validly published by Baudelot & Bessiere 1977, acc. to Fensome et
al. 1990]

Polygonium sexradiatum (Timofeev 1959b) Volkova 1990

“*Polygonium varium*” (Volkova 1969) Sarjeant & Stancliffe 1994
[Sarjeant & Stancliffe 1994 tentatively assigned this species to *Polygonium*; Moczyłowska
1998 marked this as a new combination]
→ *Eklundia varia* (Volkova 1969) Jachowicz-Zdanowska 2013

Polygonium sp. A Vanguetaine 1973a

“***Polyporata***” Pykhova 1966
[nomen nudum, acc. to Fensome et al. 1990]
[**homonym:** *Polyporata* Xing & Liu 1973]
no type species

“*Polyporata nidia*” Pykhova 1966
[not validly published, acc. to Fensome et al. 1990]

“*Polyporata verrucosa*” Pykhova 1966
[not validly published, acc. to Fensome et al. 1990]

Priscogalea Deunff 1961a emend. Rasul 1974
type species: *Priscogalea barbara* Deunff 1961a

“*Priscogalea cortinula*” Deunff 1961a
→ *Stelliferidium cortinulum* (Deunff 1961a) emend. Deunff et al. 1974

“*Priscogalea cuvillieri*” Deunff 1961a
→ *Cymatiogalea cuvillieri* (Deunff 1961a) Deunff 1964
“*Priscogalea gautieri*” Martin 1972
→ *Stelliferidium gautieri* (Martin 1972) Pittau 1985
“*Priscogalea multarea*” Deunff 1961a
→ *Cymatiogalea multarea* (Deunff 1961a) Eisenack et al. 1973
Priscogalea primordialis Yin L. in Chen J. et al. 1985 emend. Yin L. 1986
Priscogalea promptusa Yin L. in Chen J. et al. 1985
?*Priscogalea* sp. A Vanguetaine 1986

Priscotheca Deunff 1961a
type species: *Priscotheca raia* Deunff 1961a

Priscotheca notata Volkova 1988

Protoarchaeosacculina Naumova 1961
type species: *Protoarchaeosacculina atava* Naumova 1961

Protoleiosphaeridium Timofeev 1959b ex Timofeev 1960
type species: *Protoleiosphaeridium conglutinatum* Timofeev 1959b ex Timofeev 1960

Protoleiosphaeridium conglutinatum Timofeev 1959b ex Timofeev 1960

Protoleiosphaeridium crassum Timofeev 1963

“*Protoleiosphaeridium densum*” (Timofeev 1966) Yin L. 1979
→ *Protoleiosphaeridium laccatum* (Timofeev 1966) Fensome et al. 1990

Protoleiosphaeridium laccatum (Timofeev 1966) Fensome et al. 1990

Protoleiosphaeridium minutissimum Timofeev
[cited without complete reference in Jagielska 1965]

Protoleiosphaeridium papyraceum (Timofeev 1966) Zang 1992

Protoleiosphaeridium rugulosum (Fombella 1978) Fensome et al. 1990

Protoleiosphaeridium sp. forme A Potter 1974

Protolophosphaeridium Timofeev 1963
type species: *Protolophosphaeridium faetonum* Timofeev 1963

Protolophosphaeridium crispum Timofeev
[cited without complete reference in Jagielska 1965]

Protolophosphaeridium grumosum Timofeev
[cited without complete reference in Jagielska 1965]

Protomyceterosphaeridium Timofeev 1963
type species: *Protomyceterosphaeridium marmoratum* Timofeev 1963

Protomycterosphaeridium marmoratum Timofeev 1963

“*Protosphaeridium*” Timofeev 1966
→ *Protoleiosphaeridium* Timofeev 1959b ex Timofeev 1960 [acc. to Loeblich & Tappan 1976]
type species: “*Protosphaeridium densum*” Timofeev 1966
→ *Protoleiosphaeridium laccatum* (Timofeev 1966) Fensome et al. 1990

“*Protosphaeridium flexuosum*” Timofeev 1966
→ *Orygmatosphaeridium holtedahlii* (Timofeev 1966) Fensome et al. 1990

Prototrachysphaeridium Timofeev 1963
type species: *Prototrachysphaeridium staplinii* Timofeev 1963

Prototrachysphaeridium nevelense Timofeev
[cited without complete reference in Jagielska 1965]

Pseudotasmanites Kiryanov 1974
type species: *Pseudotasmanites parvus* Kiryanov 1974

Pseudotasmanites parvus Kiryanov 1974

Pseudozonosphaera Xing & Liu 1973
type species: *Pseudozonosphaera verrucosa* Xing & Liu 1973

Pseudozonosphaera asperella Xing & Liu 1978

[authorship in Xing & Liu 1978 was given as “Sin et Liu (MS)”, presumably referencing an unpublished manuscript]

Pseudozonosphaera sinica Xing & Liu 1973

Pseudozonosphaera verrucosa Xing & Liu 1973

Pseudozonosphaeridium Andreeva 1966

type species: *Pseudozonosphaeridium populosum* Andreeva 1966

Pseudozonosphaeridium fengxiangense Zhong Guofang 1978

[not reported from the Cambrian, but from the Proterozoic and the Silurian]

Psophosphaera Naumova 1939 ex Ishchenko 1952 emend. Potonié 1958

[gymnospermous palynomorphs]

type species: ?

Psophosphaera obscura Pykhova 1967

Pterospermella Eisenack 1972

type species: *Pterospermella aureolata* (Cookson & Eisenack 1958) Eisenack 1972

Pterospermella gigantea Jachowicz-Zdanowska 2013

Pterospermella inordinata Jachowicz-Zdanowska 2013

Pterospermella solida (Volkova 1969) Volkova in Volkova et al. 1979

Pterospermella velata Moczyłowska 1988

Pterospermella vitalis Jankauskas in Volkova et al. 1979

Pterospermella vitrea (Volkova 1974) Fensome et al. 1990

“*Pterospermella wolynica*” Kiryanov

[cited in Vidal 1981 as “*P. wolynica* Kiryanov”, “*P.*” implicitly representing *Pterospermella* in the context, but it should be *Pterospermopsimorpha*]

→ *Pterospermopsimorpha wolynica* Kiryanov 1974

Pterospermella n. sp. 1 Moczyłowska & Vidal 1986

Pterospermella sp. A Palacios & Vidal 1992

Pterospermella sp. B Moczyłowska & Stockfors 2004

Pterospermopsimorpha Timofeev 1966

type species: *Pterospermopsimorpha pileiformis* Timofeev 1966

Pterospermopsimorpha rugulosa Jachowicz-Zdanowska 2013

Pterospermopsimorpha wolynica Kiryanov 1974

Pulvinosphaeridium Eisenack 1954a

type species: *Pulvinosphaeridium pulvinellum* Eisenack 1954a

Pulvinosphaeridium antiquum Paškevičienė 1980

Pulvinosphaeridium delicatum Yin L. in Chen J. et al. 1985

Raphesphaera Volkova 1990

type species: *Raphesphaera spinulifera* Volkova 1990

Raphesphaera obsoleta Volkova 1990

Raphesphaera spinulifera Volkova 1990

→ *Vulcanisphaera spinulifera* (Volkova 1990) Parsons & Anderson 2000

Raphesphaera striatula Volkova 1993a

“*Raphesphaera turbata*” (Martin in Martin & Dean 1981) Volkova 1990

→ *Vulcanisphaera turbata* Martin in Martin & Dean 1981

Retisphaeridium Staplin et al. 1965

type species: *Retisphaeridium dichamerum* Staplin et al. 1965

Retisphaeridium brayense (Gardiner & Vanguetaine 1971) Moczydłowska & Crimes 1995

Retisphaeridium capsulatum (Jankauskas in Jankauskas & Posti 1976) Vanguetaine in Brück & Vanguetaine 2005

Retisphaeridium densum Paškevičienė in Volkova et al. 1979

Retisphaeridium dichamerum Staplin et al. 1965

“*Retisphaeridium howellii*” Martin in Martin & Dean 1983

→ *Retisphaeridium postae* (Jankauskas in Volkova et al. 1979) Vanguetaine in Brück & Vanguetaine 2004 [acc. to Vanguetaine in Brück & Vanguetaine 2004]

Retisphaeridium lechistanium Jachowicz-Zdanowska 2013

Retisphaeridium ovillense (Cramer & Díez 1972) Vanguetaine 2002

Retisphaeridium postae (Jankauskas in Volkova et al. 1979) Vanguetaine in Brück & Vanguetaine 2004

Retisphaeridium pusillum (Moczydłowska 1998) Vanguetaine in Brück & Vanguetaine 2005

Revinotesta Vanguetaine 1974 emend. Moczydłowska 1998
type species: *Revinotesta microspinosa* Vanguetaine 1974

Revinotesta izhorica (Jankauskas 1975) Moczydłowska 1998

“*Revinotesta laevigata*” Vanguetaine in Baudelot & Géry 1979
[nomen nudum, acc. to Fensome et al. 1990; Baudelot & Géry 1979 gave authorship as
“Vanguetaine 1973”, referring to an unpublished thesis (Vanguetaine 1973a)]

Revinotesta lanceolata Jankauskas 1975 ex Fensome et al. 1990

Revinotesta microspinosa Vanguetaine 1974

Revinotesta ordensis Downie 1982

Revinotesta saccata Jankauskas 1975 ex Fensome et al. 1990

Revinotesta n. sp. Vanguetaine 1986

“*Rugasphaera*” Martin in Martin & Dean 1988
[**senior homonym:** *Rugasphaera* Jiabo 1978 (= Song et al. 1978), acc. to Fensome et al. 1990]
→ *Corrugasphaera* Fensome et al. 1990
type species: “*Rugasphaera terranova*” Martin in Martin & Dean 1988
→ *Corrugasphaera terranova* Fensome et al. 1990

“*Rugasphaera terranova*” Martin in Martin & Dean 1988
→ *Corrugasphaera terranova* Fensome et al. 1990

Sagatum Vavrdová & Bek 2001
type species: *Sagatum priscum* (Kiryanov & Volkova in Volkova et al. 1979) Vavrdová & Bek 2001

Sagatum priscum (Kiryanov & Volkova in Volkova et al. 1979) Vavrdová & Bek 2001

Saharidia Combaz 1967

type species: *Saharidia downiei* Combaz 1967

Saharidia fragilis (Downie 1958) Combaz 1967

Saharidia sp. A Parsons & Anderson 2000

Schizodiacrodium Burmann 1968

type species: *Schizodiacrodium ramiferum* Burmann 1968

Schizodiacrodium armatum Volkova 1990

Schizodiacrodium brevicrinatum Golub & Volkova in Volkova & Golub 1985

Schizodiacrodium digermulense (Welsch 1986) Parsons & Anderson 2000

Schizodiacrodium fibrosum Golub & Volkova in Volkova & Golub 1985

Sinianella Yin L. 1980 emend. Zang in Zang & Walter 1992

type species: *Sinianella uniplicata* Yin L. 1980 emend. Zang in Zang & Walter 1992

Sinianella uniplicata Yin L. 1980 emend. Zang in Zang & Walter 1992

Skiagia Downie 1982 emend. Moczydłowska 1991

type species: *Skiagia scottica* Downie 1982

Skiagia brachyspinosa (Kiryanov 1974) Jachowicz-Zdanowska 2013

“*Skiagia brevispinosa*” Downie 1982

→ *Skiagia brachyspinosa* (Kiryanov 1974) Jachowicz-Zdanowska 2013

Skiagia ciliosa (Volkova 1969) Downie 1982

Skiagia compressa (Volkova 1968) Downie 1982

Skiagia insignis (Fridriksone 1971) Downie 1982

Skiagia orbicularis (Volkova 1968) Downie 1982

Skiagia ornata (Volkova 1968) Downie 1982

Skiagia pilosiuscula (Jankauskas in Volkova et al. 1979) Jachowicz-Zdanowska 2013

Skiagia pura Moczydłowska 1988

Skiagia scottica Downie 1982

Skiagia sp. A Knoll & Swett 1987

Solisphaeridium Staplin et al. 1965 emend. Sarjeant 1968 emend. Moczyłowska 1998
type species: *Solisphaeridium stimuliferum* (Deflandre 1938) Staplin et al. 1965 [Fensome
et al. 1990 gave authorship for this comb. as “(Deflandre 1938) Pocock 1972”]

Solisphaeridium akrochordum (Rasul 1979) Moczyłowska & Stockfors 2004

“*Solisphaeridium baltoscandium*” Eklund 1990 emend. Moczyłowska 1998
→ *Goniosphaeridium volkovae* Hagenfeldt 1989a

Solisphaeridium bimodulentum Moczyłowska 1998

Solisphaeridium chinese Moczyłowska & Stockfors 2004

Solisphaeridium cylindratum Moczyłowska 1998

Solisphaeridium elegans Moczyłowska 1998

Solisphaeridium flexipilosum Slavíková 1968 emend. Moczyłowska 1998

“*Solisphaeridium implicatum*” (Fridriksone 1971) Moczyłowska 1998
→ *Parmasphaeridium implicatum* (Fridriksone 1971) Jachowicz-Zdanowska 2013

Solisphaeridium lucidum (Deunff 1959) Turner 1985

Solisphaeridium multiflexipilosum Slavíková 1968 emend. Moczyłowska 1998

Solisphaeridium sp. A Moczyłowska 1998

“***Sphaerocongregus***” Moorman 1974

→ *Bavlinella* Shepeleva 1962 [acc. to Vidal 1976]

type species: “*Sphaerocongregus variabilis*” Moorman 1974

→ *Bavlinella faveolata* Shepeleva 1962 [acc. to Vidal 1976]

“*Sphaerocongregus variabilis*” Moorman 1974

→ *Bavlinella faveolata* Shepeleva 1962 [acc. to Vidal 1976]

Spumosopsophosphaera Naumova

[cited without complete reference in Pykhova 1967]

?= *Spumososphaera* Naumova [cited without complete reference in Naumova 1968, acc. to
Fensome et al. 1990]

type species: ?

Stellechinatum Turner 1984

type species: *Stellechinatum celestum* (Martin 1969) Turner 1984

Stellechinatum uncinatum (Downie 1958) Molyneux 1987

Stelliferidium Deunff et al. 1974

type species: *Stelliferidium striatulum* (Vavrdová 1966) Deunff et al. 1974

?*Stelliferidium anomalum* Di Milia et al. 1989

Stelliferidium cortinulamorphum Paalits 1995

Stelliferidium cortinulum (Deunff 1961a) emend. Deunff et al. 1974

Stelliferidium cylindratum (Rasul 1974) Elaouad-Debbaj 1988

Stelliferidium distinctum (Rasul 1974) Pittau 1985

?*Stelliferidium* cf. *furcatum* (Deunff 1961a) emend. Deunff et al. 1974

Stelliferidium gautieri (Martin 1972) Pittau 1985

Stelliferidium magnum Palacios et al. 2009

Stelliferidium pingiculum Martin in Martin & Dean 1988

Stelliferidium aff. *pseudoornatum* Pittau 1985

Stelliferidium robustum Moczydłowska 1998

Stelliferidium striatulum Vavrdová 1966) Deunff et al. 1974

Stelliferidium sp. 1 Volkova 1990

Stelliferidium sp. A Di Milia 1991

Stelliferidium sp. A Parsons & Anderson 2000

Stelliferidium sp. A Parsons & Anderson 2000

Stelliferidium sp. A Parsons & Anderson 2000

Stelliferidium sp. B Di Milia 1991

Stelliferidium sp. B Parsons & Anderson 2000

Stelliferidium sp. C Di Milia 1991

Stellinium Jardiné et al. 1972

type species: *Stellinium octoaster* (Staplin 1961) Jardiné et al. 1972

?*Stellinium* n. sp. Vidal 1981

Stictosphaeridium Timofeev 1962 ex Timofeev 1966
type species: *Stictosphaeridium sinapticuliferum* Timofeev 1966

“*Stictosphaeridium brayense*” Gardiner & Vanguetaine 1971
→ *Retisphaeridium brayense* (Gardiner & Vanguetaine 1971) Moczyłowska & Crimes 1995

“*Stictosphaeridium implexum*” Timofeev 1966
→ *Protoleiosphaeridium laccatum* (Timofeev 1966) Fensome et al. 1990

Stictosphaeridium sinapticuliferum Timofeev 1966

Striatotheca Burmann 1970
type species: *Striatotheca principalis* Burmann 1970

?*Striatotheca oculifera* Volkova 1990

?*Striatotheca randomensis* Parsons & Anderson 2000

Synplassosphaeridium Timofeev 1959b ex Timofeev 1969
type species: *Synplassosphaeridium tumidulum* Timofeev 1959b

Synplassosphaeridium cambriense Slavíková 1968 ex Fensome et al. 1990

Synsphaeridium Eisenack 1965b
[**junior homonym:** “*Synsphaeridium*” Timofeev 1966, acc. to Fensome et al. 1990]
type species: *Synsphaeridium gotlandicum* Eisenack 1965b

“*Synsphaeridium conglutinatum*” (Timofeev 1959b ex Timofeev 1960) Timofeev 1969
→ *Protoleiosphaeridium conglutinatum* Timofeev 1959b ex Timofeev 1960

Synsphaeridium cf. *paulum* Potter in Baudelot & Géry 1979
[nomen nudum, acc. to Fensome et al. 1990; Baudelot & Géry 1979 gave authorship as “Potter 1974”, referring to an unpublished thesis]

Synsphaeridium switjasium Kiryanov 1974

Synsphaeridium sp. 1 Cramer & Díez 1972

Synsphaeridium sp. 1 Young et al. 1994

Tasmanites Newton 1875

type species: *Tasmanites punctatus* Newton 1875

Tasmanites bobrowskae Ważyńska 1967

Tasmanites convolutus Volkova & Piskun 1985

[cited without complete reference in Jankauskas & Lendzion 1992 as “*Tasmanites convolutus* Volkova et Pisk.”; in Jankauskas 2002 as “*T. convolutum*” (p. 81), “*Tasmanites convolutus*” (p. 83), “*Tasmanites convolutus* Volkova et Piskun, 1985” (p. 110); in Vetluzhskikh et al. 2009 as “*Tasmanites convolutus* Volk. et Pisk”]

Tasmanites piritaensis Posti & Jankauskas in Jankauskas & Posti 1976

Tasmanites tenellus Volkova 1968

“*Tasmanites variabilis*” Volkova 1968

→ *Tasmanites bobrowskae* Ważyńska 1967

Tasmanites volkova Kiryanov 1974

Tasmanites sp. A Vanguetaine 1986

Tawuia Hofmann in Hofmann & Aitken 1979

type species: *Tawuia dalensis* Hofmann Hofmann & Aitken 1979

Tectitheca Burmann 1968

type species: *Tectitheca valida* Burmann 1968

Tectitheca cf. *filigera* Burmann 1968

?*Tectitheca multispinula* Yin L. in Chen J. et al. 1985 emend. Yin L. 1986

Teohipolia Kiryanov in Volkova et al. 1979

type species: *Teohipolia lacerata* Kiryanov in Volkova et al. 1979

Teohipolia lacerata Kiryanov in Volkova et al. 1979

Thymadora Clendening & Wood 1981

type species: *Thymadora kerka* Clendening & Wood 1981

Thymadora kerka Clendening & Wood 1981

Timofeevia Vanguetstaine 1978

type species: *Timofeevia lancariae* (Cramer & Díez 1972) Vanguetstaine 1978

Timofeevia estonica Volkova 1990

Timofeevia ianischewskyi (Timofeev 1959b ex Vavrdová 1976) Volkova 1990

Timofeevia lancariae (Cramer & Díez 1972) Vanguetstaine 1978

Timofeevia manata Albani et al. 1991

Timofeevia microrotis Martin in Martin & Dean 1981

Timofeevia parva Hagenfeldt 1989b

Timofeevia pentagonalis (Vanguetstaine 1974) Vanguetstaine 1978

Timofeevia phosphoritica Vanguetstaine 1978

Timofeevia raquelinae (Cramer & Díez 1972) Cramer & Díez 1979

Timofeevia tacheddirtensis Vanguetstaine & Van Looy 1983

Timofeevia n. sp. A Palacios 2010

Timofeevia n. sp. B Palacios 2010

Timofeevia sp. 1 Paalits 2005

Timofeevia sp. 1 Volkova 1990

Timofeevia sp. 1 Volkova 1993b

Timofeevia sp. 1 Volkova 1995

Timofeevia sp. A Raevskaya & Golubkova 2006

Timofeevia sp. A Vanguetstaine 1986

Trachydiacrodium Timofeev 1959b emend. Deflandre & Deflandre-Rigaud 1962

type species: *Trachydiacrodium productum* Timofeev 1959b

Trachydiacrodium coarctatum Timofeev 1959b

Trachyoligotriletes Timofeev 1958 ex Timofeev 1960

[variant: *Trachyoligotriletum*]

type species: *Trachyoligotriletes nevelensis* Timofeev 1958

Trachyoligotriletes incrassatus (Naumova 1949) Timofeev 1960

Trachyoligotriletes laminaritus Timofeev 1958 ex Timofeev 1960

Trachyoligothrites obsuletes (Naumova 1949) Timofeev 1960

Trachyoligothrites planus Timofeev 1958

Trachysphaeridium Timofeev 1959b ex Timofeev 1966

[**junior synonym:** *Menneria* Lopukhin 1971, acc. to Vidal 1974]

type species: *Trachysphaeridium attenuatum* Timofeev 1959b

“*Trachysphaeridium cultum*” (Andreeva 1966) Xing in Xing & Liu 1973

→ *Leiosphaeridia culta* (Andreeva 1966) Paškevičienė 1980 [acc. to Fensome et al. 1990]

Trachysphaeridium hyalinum Xing & Liu 1973

Trachysphaeridium laminaritum Timofeev 1966

Trachysphaeridium leve (Lopukhin 1971) Vidal 1974

Trachysphaeridium minor Liu & Xing in Xing & Liu 1973

Trachysphaeridium rude Xing & Liu 1978

Trachysphaeridium rugosum Xing in Xing & Liu 1973

Trachysphaeridium simplex Xing 1962

“*Trachysphaeridium stipticum*” Sin et Liu

[cited without complete reference in Ding et al. 1992, presumably referring to Xing & Liu 1973. In that publication, authorship is given as “Sin (in litt.)”, apparently referring to an unpublished document. Since no type specimen was designated by Xing & Liu 1973, the species was not validly published]

Trachysphaeridium timofeevii Vidal 1976

Trachytriletes Naumova 1939

type species: *Trachytriletes kryshtofovichii* Naumova 1949

Trachytriletes kryshtofovichii Naumova 1949

Trachytriletes timofeevii Fensome et al. 1990

Trematosphaeridium Timofeev 1959b

type species: *Trematosphaeridium decoratum* Timofeev 1959b

“*Trematosphaeridium holtedahlii*” Timofeev 1966

→ *Orygmatosphaeridium holtedahlii* (Timofeev 1966) Fensome et al. 1990

Trematosphaeridium minutum Xing & Liu 1978

Trematosphaeridium simplex Wang et Chen

[cited without complete reference in Wang & Chen Q. 1987; possible homonym of “*Trematosphaeridium simplex*” Pykhova 1973, which was not validly published acc. to Fensome et al. 1990]

Trichosphaeridium Timofeev 1966

type species: *Trichosphaeridium annolovaense* Timofeev 1966

Trichosphaeridium annolovaense Timofeev 1966

Trichosphaeridium hirtum Timofeev 1959b ex Fensome et al. 1990

→ *Filisphaeridium hirtum* (Timofeev 1959b ex Umnova 1975) Sarjeant & Stancliffe 1994

Trunculumarium Loeblich & Tappan 1976

type species: *Trunculumarium revinium* (Vanguetaine 1973b) Loeblich & Tappan 1976

Trunculumarium revinium (Vanguetaine 1973b) Loeblich & Tappan 1976

Tubulosphaera Palacios in Palacios & Moczyłowska 1998

type species: *Tubulosphaera perfecta* Palacios in Palacios & Moczyłowska 1998

Tubulosphaera n. sp. A Palacios 2010

Turrisphaeridium (Timofeev 1959b) Jachowicz-Zdanowska 2013

type species: *Turrisphaeridium semireticulatum* (Timofeev 1959b) Jachowicz-Zdanowska 2013

Turrisphaeridium semireticulatum (Timofeev 1959b) Jachowicz-Zdanowska 2013

Turrisphaeridium turgidum Jachowicz-Zdanowska 2013

“*Uniporata*” Naumova in Pykhova 1969

[not validly published, acc. to Fensome et al. 1990]

type species: “*Uniporata typica*” Naumova in Pykhova 1969 [nomen nudum, acc. to Fensome et al. 1990]

“*Uniporata nidia*” Pykhova 1966
[not validly published, acc. to Fensome et al. 1990]

“*Uniporata torosa*” Pykhova 1966
[not validly published, acc. to Fensome et al. 1990]

Valeria Jankauskas 1982
type species: *Valeria lophostriata* (Jankauskas 1979b) Jankauskas 1982

Valeria granulata (Vidal in Vidal & Siedlecka 1983) Fensome et al. 1990

Valeria sinica (Timofeev 1966) Fensome et al. 1990

Veryhachium Deunff 1954c emend. Downie & Sarjeant 1963 emend. Turner 1984 emend.
Sarjeant & Stancliffe 1994
type species: *Veryhachium trisulcum* Deunff 1951 ex Deunff 1959

“*Veryhachium dumontii*” Vanguetaine 1973b
→ *Ninadiacrodium dumontii* (Vanguetaine 1973b) Raevskaya & Servais 2009

Veryhachium incus Paalits 1992a

Veryhachium cf. *minutum* Downie 1958

Veryhachium mutabile Di Milia et al. 1989

“*Veryhachium primaevum*” Deunff 1967
[nomen nudum, acc. to Fensome et al. 1990]

Veryhachium cf. *rhomboideum* Downie 1959

“*Veryhachium setuensis*” Paalits 1992a
→ ?*Dasydiacrodium setuensis* Paalits 1992a

“*Veryhachium* cf. *trapezionarion*” Loeblich 1970
→ *Veryhachium* cf. *rhomboideum* Downie 1959 [acc. to Turner 1984]

Veryhachium trisentium Zang in Gravestock et al. 2001

Veryhachium sp. A Martin in Martin & Dean 1981

Veryhachium sp. A Parsons & Anderson 2000

Virgatasporites Combaz 1967
type species: *Virgatasporites rudii* Combaz 1967

Virgatasporites normale (Fombella 1978) Fensome et al. 1990

Virgatasporites rudii Combaz 1967

Vogtlandia Burmann 1970 emend. Sarjeant & Vavrdová 1997
type species: *Vogtlandia ramificata* Burmann 1970

Vogtlandia carvinacornua (Welsch 1986) Sarjeant & Vavrdová 1997

Vogtlandia notabilis Volkova 1990

Vogtlandia petropolitana (German 1974) Volkova 1990

Vogtlandia simplex Moczydłowska 1998

Volkovia Downie 1982
type species: *Volkovia dentifera* (Volkova 1969) Downie 1982

Volkovia conifera (Jankauskas 1975) Downie 1982

Volkovia dentifera (Volkova 1969) Downie 1982

Volkovia flagellata (Jankauskas 1975) Downie 1982

Vulcanisphaera Deunff 1961a emend. Rasul 1976
type species: *Vulcanisphaera africana* Deunff 1961a

Vulcanisphaera africana Deunff 1961a

Vulcanisphaera britannica Rasul 1976

Vulcanisphaera capillata Jardiné et al. 1974

“*Vulcanisphaera cirrita*” Rasul 1976

→ *Vulcanisphaera africana* Deunff 1961a [acc. to Elaouad-Debbaj 1988]

Vulcanisphaera ferosa Fombella 1977

Vulcanisphaera frequens Górka 1967

Vulcanisphaera lanugo Martin in Martin & Dean 1988

Vulcanisphaera nebulosa Deunff 1961a

Vulcanisphaera spinulifera (Volkova 1990) Parsons & Anderson 2000

“*Vulcanisphaera tuberata*” (Downie 1958) Eisenack et al. 1973

→ *Goniosphaeridium tuberatum* (Downie 1958) Welsch 1986

Vulcanisphaera turbata Martin in Martin & Dean 1981

Vulcanisphaera sp. 1 Volkova 1980

?*Vulcanisphaera* sp. 1 Paalits 2005

Yurtusia Dong et al. 2009

type species: *Yurtusia uniformis* Dong et al. 2009

Yurtusia uniformis Dong et al. 2009

“**Zonosphaeridium**” Timofeev 1959b

→ *Tasmanites* Newton 1875 [acc. to Timofeev 1966]

no type species

“*Zonosphaeridium ovillensis*” Cramer & Díez 1972

→ *Retisphaeridium ovillense* (Cramer & Díez 1972) Vanguetstaine 2002

Acritarch gen. et sp. nov. Martin & Dean 1984

→ *Comasphaeridium francinae* Jachowicz-Zdanowska 2013

Acritarch gen. et sp. nov. Martin & Dean 1988

Acritarch gen. et sp. nov. Young et al. 1994

Acritarch sp. 1 Hagenfeldt 1989b

Acritarch sp. Wood & Clendening 1982

Gen. et sp. ind. A Di Milia 1991

Gen. et sp. ind. B Di Milia 1991

Gen. et sp. indet. 1 Paalits 2005

Gen. et sp. indet. 2 Paalits 2005

Gen. et sp. indet. C Jago et al. 2006

Gen. et sp. indet. Żylińska & Szczepanik 2009

Genus et species ind. A Di Milia et al. 1989

Genus et species ind. B Di Milia et al. 1989

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