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| 5      | Connecting the seasonal productivities: migratory movements of   |
| 6      | rhinoceros auklets in the northwestern Pacific   |
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| 24     | ABSTRACT: Spatial and temporal variability in marine biological productivity may   |
| 25     | drive heterogeneity in seasonal resources available for marine animals in temperate  |
| 26     | waters. Migratory seabirds are expected to adjust the annual cycle of breeding activities  |
| 27     | and migratory movements to exploit seasonally available resources efficiently. We  |
| 28     | studied the movement and trophic position of rhinoceros auklets Cerorhinca   |
| 29     | monocerata breeding at Teuri Island, Japan Sea, during the nonbreeding and early   |
| 30     | breeding periods over two years. After breeding, the auklets moved northward from the  |
| 31     | colony to the Sea of Okhotsk, where phytoplankton blooms enhanced biological   |
| 32     | productivity in autumn. The birds then moved southward to the southwestern Japan Sea   |

33 (~1470 km from the colony), where major epipelagic fish and squid concentrations have 34 been reported in winter. Stable isotope analyses suggest that the auklets fed on higher-trophic level prey, including fish and/or squid during the autumn and winter 35 nonbreeding periods. The auklets moved northward and returned to the colony in 36 mid-March. During the early breeding period, the birds foraged close to the colony 37 38 (~380 km) on lower-trophic level prey, including fish and/or krill, which were available during the spring phytoplankton bloom. The timing of the return migration does not 39 40 match with the northward migration of warm-water anchovy, a profitable prey during summer, but may be related to the adjustment of the chick-rearing period to anchovy 41 42 arrival. We suggest that rhinoceros auklets follow spatial and seasonal changes in prey availability by a distinctive 'three-step' migration (first northward, second southward, 43 third northward) in a temperate marine system of northwestern Pacific. 44

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KEY WORDS: Seabird foraging, Light-based geolocation, Productivity, Seasonal
movements, Stable isotope analysis, Japan Sea

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#### 50 INTRODUCTION

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Seabirds are faced with challenges in finding their prey in the spatially and 52 53 temporally variable marine environment (Weimerskirch 2007), under the different foraging constraints that exist during the breeding and nonbreeding periods. During the 54 55 breeding period, parent seabirds are central place foragers that have to commute between their colony and feeding grounds at sea. Their foraging ranges are relatively 56 57 limited especially during early chick-rearing period to provision their offspring (e.g., 58 Charrassin & Bost 2001, Rayner et al. 2012). Despite the constraints on their foraging 59 range, parent seabirds have to feed intensively in order to meet the high energetic demands associated with breeding (Hamer et al. 2001). During the nonbreeding period, 60 61 seabirds can move over larger distances without the constraints of central-place foraging (e.g., Thiebot et al. 2011), but may experience reduced prey availability due to 62 decreased marine productivity outside the summer breeding season and/or possible 63 changes in the accessibility of prey in the water column (e.g., Charrassin & Bost 2001). 64

An increasing number of studies have shown that seabirds, especially proficient-flying procellariiform seabirds, adjust the annual cycle of breeding activities and migratory movements to seasonally available resources (Shaffer et al. 2006, González-Solís et al. 2007, Guilford et al. 2009, Rayner et al. 2011, Carey et al. 2014); though, such adjustments may vary among species of different migration abilities and among different marine regions.

Recent advances in bio-logging technologies, such as light-based geolocators, 7172 have allowed us to examine the year-round movement of individual seabirds over 73 extensive periods (Burger & Shaffer 2008, Wakefield et al. 2009). In addition, stable 74 isotope analysis of animal tissues can be used to examine the trophic position of seabirds during their year-round movement (Hobson & Bond 2012). Based on these 75 techniques, long-distance trans-equatorial migrations have been documented for 76 proficient-flying medium-sized petrels, which presumably allow them to exploit the 77 summer peaks in biological productivities in both northern and southern hemispheres 78 79 successively (Shaffer et al. 2006, González-Solís et al. 2007, Rayner et al. 2011). However, relatively less is known about the movements of seabirds with higher 80 energetic cost of flying, such as auks (Elliott et al. 2013), in relation to seasonal changes 81 82 in regional marine productivities and breeding constraints. So far, all the previous studies on the migratory movements of alcids have been conducted in the North Atlantic, 83 84 highlighting the areas off Newfoundland as important foraging area during autumn and 85 winter (Mosbech et al. 2012, Jessopp et al. 2013, Linnebjerg et al. 2013, McFarlane Tranquilla et al. 2013). In contrast, no such information is yet available for any alcid 86 87 species breeding in the North Pacific. Information on migratory patterns will also be valuable for understanding population trends with respect to marine environmental 88 89 change and for assessing any potential conservation issues (Harris et al. 2013, 90 McFarlane Tranquilla et al. 2013).

Rhinoceros auklets *Cerorhinca monocerata* are medium-sized alcids (500 - 600 g) that breed in the North Pacific. The largest breeding colony is located at Teuri Island in the northern Japan Sea, where approximately 0.3 million pairs breed annually (Watanuki & Ito 2012). The auklets feed intensively on Japanese anchovy *Engraulis japonicus* close to the breeding colony (~130 km) during chick-rearing period (Kato et al. 2003, Watanuki & Ito 2012), but their movement and diet have been largely

97 unknown outside the breeding period. The objective of this study is therefore to 98 characterize the migratory movements and trophic position of rhinoceros auklets during nonbreeding and early breeding periods, by combining information from light-based 99 100 geolocation and stable isotope analyses. It is hypothesized that the migratory 101 movements of the auklets follow seasonal changes in regional prey availability, given 102 the heterogeneity of seasonal productivity patterns in Japan Sea and the adjacent Sea of Okhotsk (Lee et al. 2009, Radchenco et al. 2010). We expect that rhinoceros auklets 103 104 maintain their fish diet by moving among different marine regions during the nonbreeding period, but they may be confined to feed close to the colony on different 105 106 prey items during the early breeding period.

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#### 109 MATERIALS AND METHODS

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111 • Geolocator deployments. The study was conducted at Teuri Island (44°24' N, 141°17' E), located in the northern Japan Sea (Fig. 1). Rhinoceros auklets first return 112 113 to Teuri Island from March to April and lay a single egg in April and May. The single chick hatches between May and June, and fledges in July (Watanuki & Ito 2012). In 114 115 July 2010, we captured 10 chick-rearing birds from nest burrows, and attached a 116 geolocator (Mk15, 16 x 18 x 6mm, British Antarctic Survey, Cambridge, UK) on the tarsus of each bird using a plastic leg ring. The total mass of geolocators including the 117ring was 3.5 g (0.62 % of the mean body mass of tracked birds). We retrieved 118 geolocators from 8 out of 10 birds and in May 2011. Upon recapture, the 10<sup>th</sup> primary 119 120 feather and 2-3 breast feathers were taken as samples for stable isotope analysis. One additional instrumented bird was recaptured in the breeding season of 2013. We also 121 attached geolocators on 20 birds in July 2011. We retrieved geolocators from 16 out 122 123 of 20 birds and in May-June 2012. Three individuals were tracked during the nonbreeding and early breeding periods of both 2010-11 and 2011-12. All the 124 geolocators that were recovered recorded data successfully, but 5 geolocators from 125 126 the 2011-12 deployments stopped recordings prematurely after 1-3 months. We therefore analyzed 20 complete tracks recorded from 17 birds (10 males and 7 127 females). The sex of the birds was determined on the basis of bill and head 128

measurements (Niizuma et al. 1999). The study birds maintained their body mass between device attachment and retrieval on average (average mass change: +12.5 g, range: +75 g to -45 g), although we do not have comparable data from control birds to examine any device effects.

• Geolocation data analysis. The geolocators record time, light intensity, immersion 133 in seawater and water temperature. The geolocators were programmed to measure 134 light levels at 1 min intervals, and to record the maximum value after each 10 min 135 period. Immersion in seawater was checked every 3 s and the data were compiled as a 136 proportion over each 10 min period. Water temperature was recorded only after 137 continuous immersion for 20 min as the temperature sensors require 10 min to 138 139 stabilize. Accuracy of the temperature recordings was 0.5 °C as indicated by the manufacturer. Light and water temperature data were used to estimate the daily bird 140 141 locations (Yamamoto et al. 2011). Times of sunset and sunrise were estimated from 142 the thresholds in the light curves. Latitude was derived from day length and longitude 143 from time of local midday with respect to Greenwich Mean Time and Julian day, using TransEdit and Locator software (British Antarctic Survey). To improve the 144 145 location estimates from light levels, the daily median of the water temperature records from the geolocators were compared with remotely sensed sea surface temperature 146 (SST) data (8-day composite, 9-km resolution, measured by Aqua-MODIS, 147 148 downloaded from the ocean color web http://oceancolar.gsfc.nasa.gov). We retained the longitude estimates obtained from light-based geolocation, and searched the 149 latitude where SST matches with median water temperature records from geolocators. 150 Since SST varies with latitude (cooler towards higher latitude), this procedure 151 152 enabled us to refine the latitude estimates during the entire period of at-sea trips, including during the equinoxes. Finally, these daily positions were smoothed using a 153 154 theoretical movement model in a Bayesian framework (Thiebot & Pinaud 2010). This 155 movement model calculates the most probable location estimates along the tracks 156 (without deleting any locations) that meet with the following three conditions: 1) no locations occur on land, 2) the start and end locations of the tracks are the breeding 157 158colony, and 3) mean movement speed is 10 km/h (based on flying speeds measured from GPS tracking, Watanuki et al., unpublished data, corrected by the average time 159160 spent flying per day over the nonbreeding period, ~4.5 h, this study), with 5 km /h

allowed for variance of the mean (see Thiebot & Pinaud 2010 and references thereinfor details).

Attendance at the colony was assessed from the immersion records from 163 geolocators. We defined colony attendance as periods in the data when the geolocator 164 was continuously dry for >5 hours across midnight, because most birds that are not at 165 166 the colony rest on the sea surface during night (Kato et al. 2003). The nonbreeding 167 period was defined as the period from the last attendance at the colony, defined using 168 activity data, after breeding (late July) until the first attendance at the colony in the following year (mid March). Similarly, the early breeding period was defined as the 169 170 period from the first attendance at the colony to the end of April, as geolocators were mostly retrieved from birds in early May, when birds were normally in the incubation 171period (Watanuki & Ito 2012). 172

• Stable isotope analysis of feathers. We analyzed stable isotopes of feathers grown 173 during the nonbreeding period. Although accurate molt cycles are not known for 174 rhinoceros auklets, the 10<sup>th</sup> primary and breast feathers presumably grow during the 175 pre-basic molt (August - October) and pre-alternate molt (February - March), 176 respectively (Pyle 2009, Sorensen et al. 2010). Feathers were stored in a freezer 177 (-20°C) until laboratory analyses. Nitrogen and carbon stable isotope ratios ( $\delta^{15}$ N and 178  $\delta^{13}$ C) were measured at Meijo University (Nagoya, Japan) for 2010-11 samples and at 179 180 Port and Airport Research Institute (Yokosuka, Japan) for 2011-12 samples. At Meijo University, feathers were washed using 0.25 mol/L sodium hydroxide aqueous 181 solution and distilled water, dried in a oven (60°C) for 24 hours, placed in liquid 182 nitrogen for 1 minute, then homogenized using a sample crasher (TK-AM5, TITEC). 183 At Port and Airport Research Institute, feathers were washed using 0.25 mol/L 184 sodium hydroxide aqueous solution and distilled water, freeze-dried for 36-48 hours, 185 cut as small as possible, then homogenized using a mortar.  $\delta^{15}N$  and  $\delta^{13}C$  were 186 measured using a SerCon ANCA-GSL, Hydra 20-20MASS spectrometer (SerCon 187 Ltd.) (Meijo University) or a Delta Plus Advantage mass spectrometer (Thermo 188 Electron) coupled with an elemental analyzer (Flash EA 1112, Thermo Electron) 189 (Port and Airport Research Institute). Stable isotope ratios are expressed in  $\delta$  notation 190 as the deviation from standard (air) in parts per thousand (‰) according to the 191 following equation:  $\delta^{15}N$  or  $\delta^{13}C = [R_{sample} / R_{standard} - 1] \times 1000$ , where  $R = {}^{15}N/{}^{14}N$ 192

or  ${}^{13}C/{}^{12}C$ . Replicate measurements of internal laboratory standards indicated that the measurement precision was  $\pm 0.16$  ‰ and  $\pm 0.12$  ‰ for  $\delta^{15}N$  (SD) and  $\pm 0.16$  ‰ and  $\pm 0.11$  ‰ for  $\delta^{13}C$  (SD) in the laboratories at Meijo University and Port and Airport Research Institute, respectively.

197 We need to take into account the tissue-specific discrimination factors of isotopic values, to compare isotopic values of different tissues of consumers with that 198 of prey. We used the discrimination factors as follows, based on a study of common 199 murres Uria aalge (Becker et al. 2007): 3.7 % and 3.6 % for  $\delta^{15}$ N and 1.9 % and 200 1.0 % for  $\delta^{13}C$ , for primary and breast feathers. We compared the  $\delta^{15}N$  and  $\delta^{13}C$ 201 values of feathers with those of egg yolk and blood plasma obtained during breeding 202 203 season as well as potential prey species (Euphausiids: Thysanoessa inermis; 0+ and 204 >1+ sandlance Ammodytes personatus; Japan sea greenling Pleurogrammus azonus; Japanese anchovy; Squid Coleoides; juvenile chum salmon Oncorhynchus keta), 205 206 reported in Ito et al. (2009, 2012). Ito et al. (2012) used the following discrimination factors, 3.4 % and 2.8 % for  $\delta^{15}$ N and 0.1 % and -1.3 % for  $\delta^{13}$ C, for egg yolk 207 and blood plasma, respectively, based on other avian studies (egg yolk) and their own 208 measurements from rhinoceros auklet chicks (blood plasma). 209

210 • **Primary productivity.** To characterize the seasonal patterns in primary productivity, monthly average values of sea surface chlorophyll a concentrations were 211 212 calculated from January 2003 to December 2012 for an area in the Sea of Okhotsk 213 (area A) and two areas in the Japan Sea (areas B and C; Fig. S1). The areas were 214 chosen on the basis of 50 % kernel density boundaries of bird locations during the nonbreeding (areas A and C) and early breeding (area B) periods. We used the Aqua 215 MODIS level 3 monthly standard mapped image 9-km resolution chlorophyll a data 216 that were downloaded from the ocean color web. 217

Statistics. Maximum distance reached from the colony by the birds was compared between years or sexes with one-way ANOVA. Two males and one female had repeated measurements (i.e. tracked during both 2010-11 and 2011-12). We show here the statistics that include the six tracks from these three birds as independent.
 Statistical results were similar, even if we included only one (the track of either 2010-11 or 2011-12) of the two tracks for these three birds in the analyses. Minitab software (Minitab Inc., Pennsylvania, USA) was used for statistical analyses. Kernel

densities for geolocation data were calculated using the ArcGIS Spatial Analyst Density tool (ESRI, California, USA) with a cell size of 0.2 degree and a search radius of 2 degrees. Means ( $\pm$  SE) are shown.

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#### 230 **RESULTS**

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#### 232 Seasonal movement patterns

After the breeding season, tracked birds departed the colony on 23<sup>rd</sup> July in both 233 vears ( $\pm$  0.7 and  $\pm$  1.4 days for 2010-11 and 2011-12, respectively), indicated by the 234immersion records of geolocators. In the first phase of migration, the birds moved 235northward from the colony to the Sea of Okhotsk, which was consistent during both 236years (Fig. 2, Fig. 3). The birds were distributed mainly at the east or south of Sakhalin 237Island until September, and then moved southward to the northeastern or southeastern 238 shore of Hokkaido Island in October (Fig. 4). In the second phase, the birds moved 239 southward across the Japan Sea, during November - January. Most birds (7 of 9 birds 240 and 8 of 11 birds in 2010-11 and 2011-12, respectively) reached the sea around the 241 Korean Peninsula and Tsushima Strait (Fig. 3, Fig. 4). The remaining four birds (5 242 tracks) appeared to spend winter farther east in Japan Sea, with a mean westernmost 243 longitude of 135.6 °E. From February, the birds undertook their third movement phase 244 by migrating northward again along the coast of Japan, and returned to the colony on 245 $11^{\text{th}}$  and  $18^{\text{th}}$  March (± 2.3 and ± 1.1 days) in the 2010-11 and 2011-12 seasons, 246 247 respectively (Fig. 3). Overall, kernel density of locations identified the Sea of Okhotsk and the sea around the Korean Peninsula and Tsushima Strait as key areas during the 248 249 nonbreeding period in both 2010-11 and 2011-12 (Fig. 4).

The average maximum distance reached from the colony by the auklets during the nonbreeding period did not differ between the two years ( $1549 \pm 77$  km vs.  $1403 \pm 91$ km for 9 and 11 tracks in 2010-11 and 2011-12, respectively;  $F_{1,18} = 1.43$ , p = 0.25) nor between sexes ( $1481 \pm 96$  km vs.  $1460 \pm 83$  km for 8 and 12 tracks from females and males, respectively;  $F_{1,18} = 0.03$ , p = 0.87).

After the initial arrival to the colony, the birds made foraging trips during pre-laying and the early incubation periods, until the end of the geolocator records.

- During this early breeding period, the birds stayed relatively close to the colony (Fig.4); half of the daily at-sea locations were within  $157 \pm 10$  km and  $237 \pm 21$  km from the colony, for 2010-11 and 2011-12, respectively. The average maximum distance reached from the colony by the auklets during the early breeding period did not differ between the two years ( $376 \pm 24$  km vs.  $386 \pm 19$  km for 9 and 11 tracks in 2010-11 and 2011-12, respectively;  $F_{1,18} = 0.1$ , p = 0.75) nor between sexes ( $407 \pm 29$  km vs.  $365 \pm 14$  km for 8 and 12 tracks from females and males;  $F_{1,18} = 2.12$ , p = 0.16).
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#### Activity and water temperature records

During the nonbreeding period, the leg-mounted geolocators remained dry for 4.42  $\pm$  0.42 h per day (n = 20 tracks). There were no apparent seasonal patterns in the time spent dry, and no clear evidence of flightless periods (Fig. 2). During the early breeding period, the daily time spent dry increased as the birds attended the colony at night or throughout a day for incubation (Fig. 2).

Water temperature recorded by the geolocators showed large seasonal changes in both years (Fig. 5). Water temperature experienced by the birds remained around 11 -14 °C from October to late February, but dropped to 4 - 6 °C in early March associated with a northward migration of auklets to the breeding colony.

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#### 276 Isotopic value of feathers

277Mean measured isotopic values (without adjustment for discrimination factors) were 15.6  $\pm$  0.3  $\,\%\,$  and 15.1  $\pm$  0.1  $\,\%\,$  for  $\delta^{15}N$  and -19.8  $\pm$  0.2  $\,\%\,$  and -18.8  $\pm$  0.1  $\,\%\,$ 278 for  $\delta^{13}C$  in primary feathers, versus 13.5 ± 0.2 ‰ and 14.1 ± 0.1 ‰ for  $\delta^{15}N$  and 279  $-19.5 \pm 0.2$  ‰ and  $-18.1 \pm 0.2$  ‰ for  $\delta^{13}$ C in breast feathers, in 2010-11 (n = 8) and 280 2011-12 (n = 16) respectively (Fig. 6). The  $\delta^{15}$ N values of primary and breast feathers 281 adjusted for discrimination factors (3.7 ‰ and 3.6 ‰ for primary and breast feathers) 282 were higher than those of egg yolk and blood plasma (adjusted for discrimination 283 factors) during the prelaying, incubation and chick-rearing periods (Fig. 6). The  $\delta^{13}$ C 284 285 values of primary feathers were lower than those of breast feathers, egg yolk and blood plasma (all values adjusted for discrimination factors) (Fig. 6). 286

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#### 288 Marine primary productivity

Chlorophyll a concentration was highest in April over the annual cycle in each area of A-C, and the peak values were higher in areas A and B (Sea of Okhotsk and northern Japan Sea) compared to area C (southwestern Japan Sea) (Fig. 7). A second, clear peak in chlorophyll a concentration was observed in autumn (September – November) in area A only.

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#### 296 **DISCUSSION**

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298 Our results, together with previous results from the chick-rearing period (Kato et al. 2003), suggest that, interannually, rhinoceros auklets from the largest colony in the 299 northwestern Pacific mainly use three marine regions year-round: the Sea of Okhotsk in 300 301 autumn, the southwestern Japan Sea in winter, and the northern Japan Sea during the 302 breeding season in spring and summer. The auklets consistently used these areas over two successive years. The Sea of Okhotsk, northern Japan Sea and southwestern Japan 303 Sea are highly productive areas (Lee et al. 2009, Radchenko et al. 2010) among which 304 the seasonal patterns of biological productivity differ considerably (Fig. 7). We 305 306 hypothesize that the auklets are able to connect such seasonal peaks in productivity by a 307 distinctive 'three-step' migration pattern.

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#### 309 Migratory movements and regional prey availability

Rhinoceros auklets feed mainly on warm-water Japanese anchovy during the 310 311 chick-rearing period (Watanuki & Ito 2012), and change their foraging locations from south to north of the colony during May - July, presumably following the northward 312 313 migration of anchovy (Deguchi et al. 2010). Our results showed that the auklets 314 continued to move northward into the Sea of Okhotsk after the breeding season, which 315 is in accordance with previous ship-based surveys that reported the concentrations of 316 auklets along Soya Strait and in the Sea of Okhotsk in late July – late August (Shuntov 317 2000, Deguchi et al. 2010). The auklets moved within the Sea of Okhotsk until October (Fig. 3). The Sea of Okhotsk is one of the southernmost seasonal sea ice zones in the 318 319 northern hemisphere, and supports high biological productivity associated with spring 320 ice-edge blooms as well as autumn blooms (Mustapha et al. 2009, Radchenko et al.

321 2010). High summer as well as autumn primary productivity was measured by satellite 322 (area A in Fig. 7) as well as from ship-based observations in this region, associated with the seasonal intrusion of the Tsushima Warm Current and the East Sakhalin Current 323 324 along the northern coast of Hokkaido Island (Mustapha et al. 2009). The high biological productivity in summer and autumn attracts forage fish, including Japanese anchovy (at 325 326 shallow depths 0 - 60 m; Nagasawa et al. 1998), and their predators to the Sea of Okhotsk (Radchenko et al. 2010, Sakurai et al. 2013). For example, nonbreeding 327 328 flesh-footed shearwaters Puffinus carneipes from breeding colonies in New Zealand and Australia move into the Sea of Okhotsk in August and September, presumably mostly 329 330 feeding on fish (Rayner et al. 2011) and other southern hemisphere migrants such as 331 sooty and short-tailed shearwaters (Puffinus griseus and P. tenuirostris) have been observed through July and August (Shuntov 2000, Sakurai et al. 2013, Carey et al. 332 2014). Similarly, black-tailed gulls Larus crassirostris that breed in a colony in the 333 northern Japan Sea move into the Sea of Okhotsk in August (Kazama et al. 2013). Such 334 enhanced marine food web might also attract rhinoceros auklets in autumn. 335

From November to December, the auklets moved to the southwest region of the 336 Japan Sea, and 75 % of the tracks reached as far as the sea around the Korean Peninsula. 337 They stayed in this region until mid February before migrating back to their breeding 338 colony in mid March. The southwest region of the Japan Sea is known to be productive 339 340 due to coastal upwelling (Lee et al. 2009). The region is a major fishing ground for 341 warm-water pelagic fish such as Japanese anchovy, Chub and Jack mackerels (Scomber japonicus and Trachurus japonicus) (Kim et al. 2008). In particular, Japanese anchovy 342343 are found distributed around the Korean Peninsula in winter, and are available at shallow depths (5 - 60 m; Iversen et al. 1993). This region is also known as a major 344 345 autumn spawning ground for Japanese common squid Todarodes pacificus, so juvenile squid would be available for auklets in the water column during winter (Kidokoro et al. 346 2010). 347

The three-step migration, or three-phased migratory pattern (first northward, second southward, third northward) will increase the total distance travelled during the nonbreeding period, compared with the direct migration from the colony to the southern wintering area. The costs of travelling by flight are expected to be high in alcids that have high wing loading such as rhinoceros auklets (Elliott et al. 2013). The auklets presumably gain sufficient energy in autumn in the Sea of Okhotsk, which outweighs the energy costs of moving any extra distances to southern wintering areas. The birds appeared to experience a similar temperature range (12 - 14 °C) between October and late February (Fig. 5) due to southward seasonal movement, and relatively constant sea temperature may be favorable for thermoregulation.

358 The three-step migration is also observed in little auks Alle alle breeding in East Greenland (Mosbech et al. 2012). The little auk used an autumn staging area in the 359 360 Greenland Sea at the northeast of the colony, and then performed a long southerly migration (~2000 km) to winter off Newfoundland. Other alcids show diverse migratory 361 362 patterns even within a species. For example, common murres in a colony at Svalbard migrate southward (~1000 km from the colony) to the Barents Sea and adjacent areas 363 364 (Fort et al. 2013), but the same species from a colony in central Norway migrate northward to the Barents Sea (Lorentsen et al. 2012). Atlantic puffins Fratercula 365 arctica from the Isle of May off east Scotland stay within the North Sea or move north 366 to the northeast Atlantic (Harris et al. 2013), but the same species from southwest 367 to 368 Ireland make long-distance east-west movements Newfoundland in August-September (2537 km on average; Jessopp et al. 2013). Thick-billed murres Uria 369 lomvia from 5 colonies spanning eastern Canadian Coast from the high Arctic to 370 371 Newfoundland show variable degrees of southward movement (700 - 3500 km) to 372 common wintering areas in the Labrador Sea and the seas off Newfoundland 373 (McFarlane Tranquilla et al. 2013). We suggest that the migratory patterns of alcids are highly flexible depending upon the spatial and seasonal patterns of prey availability 374 375 relative to their breeding location and timing. However, the maximum migration distances of alcids from the colony (~3500 km) are relatively limited, compared to the 376 377 distances traveled by proficient-flying procellariiform seabirds (~12000 km from the colony; Guilford et al. 2009, Yamamoto et al. 2010), suggesting that the high energetic 378 379 costs of flight would constrain the ability of alcids to exploit seasonally available 380 resources through migration.

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#### 2 **Trophic position during the nonbreeding period**

383 The nitrogen and carbon isotope signatures of the 10<sup>th</sup> primary feathers and breast 384 feathers of auklets presumably reflect the diet during autumn and winter, respectively

(see 'Stable isotope analysis of feathers' in Materials and Methods). The relatively high 385  $\delta^{15}$ N values of both primary and beast feathers (adjusted for discrimination factors), 386 compared with  $\delta^{15}$ N values of auklet blood samples during the breeding period, suggest 387 that the auklets fed on higher trophic level prey, including fish and/or squid, rather than 388 zooplankton (Fig. 6). This contrasts with isotopic studies of other alcids such as Atlantic 389 puffins or thick-billed murres, which showed seasonal shifts to low  $\delta^{15}N$  values, 390 suggesting the dependence on lower trophic level zooplankton during the nonbreeding 391 392 period (Hedd et al. 2010, Hobson & Bond 2012).

We have to note that the relatively low  $\delta^{13}$ C values in both primary and breast 393 394 feathers (adjusted for discrimination factors) may reflect the carbon source for primary producers, varying with habitat (nearshore/benthic vs. offshore/pelagic) and/or season 395(summer vs. autumn and winter). Information on regional differences in measured  $\delta^{15}N$ 396 and  $\delta^{13}C$  values of particulate organic matter (POM) are not available in Japan Sea or 397 the Sea of Okhotsk. Nearshore/benthic marine habitat is known to show higher  $\delta^{13}C$ 398 values than offshore/pelagic habitat in general (Graham et al. 2011). However, 399 rhinoceros auklets fed on pelagic fishes and yet showed higher  $\delta^{13}$ C values (adjusted for 400 discrimination factors) during the breeding period than during the nonbreeding period 401 (Fig. 6). In northern Japan Sea, the  $\delta^{13}$ C values of sinking particles were higher in early 402 summer bloom period than the rest of the year (range: -25 - -23 %; Nakanishi & 403 Minagawa 2003), which may partly explain the lower adjusted  $\delta^{13}$ C values observed 404 during the nonbreeding period. 405

Estimation of prey species based on  $\delta^{15}N$  and  $\delta^{13}C$  values are difficult without 406 more data on isotopic baseline and potential prey species in both the Sea of Okhotsk and 407 southwestern Japan Sea. Nagasawa (1998) suggests that rhinoceros auklets would be a 408 major predator of juvenile chum salmon migrating along the coasts of Hokkaido during 409 the summer and autumn season, although the nitrogen isotopic value of juvenile chum 410 salmon (collected during chick-rearing season at colony) was not as high as that of the 411 primary feathers (Fig. 6). Ogi (1980) found juvenile Atka mackerel Pleurogrammus 412 monopterigius in the stomach contents of thick-billed murres, horned Fratercula 413 corniculata and tufted Fratercula cirrhata puffins caught entangled in the Sea of 414 Okhotsk in summer. Juvenile chum salmon and Atka mackerel would also be potential 415 prey for rhinoceros auklets, but isotopic data are lacking in the Sea of Okhotsk. 416

Japanese anchovy sampled in southwestern Japan Sea in summer had  $\delta^{15}N$  and  $\delta^{13}C$ values of 9.73 – 10.16 ‰ and -17.75 – -17.44 ‰, respectively (Tanaka et al. 2008). The  $\delta^{15}N$  values of breast feathers of auklets (adjusted for discrimination factors) were similar to that of anchovy, but the adjusted  $\delta^{13}C$  values were 1.3 – 3.0 ‰ lower than that of anchovy collected in southwestern Japan Sea.

To compare the isotopic values of feathers with that of prey, we used the discrimination factors of feathers obtained from a closely-related species (common murre, Becker et al. 2007) but not from rhinoceros auklets. The discrimination factors of feathers can be variable among different seabird species or different studies on the same species, especially for  $\delta^{13}$ C (Becker et al. 2007), and this might have been another confounding factor.

428

#### 429 Adjustment of breeding timing and location

Rhinoceros auklets return to their breeding colony in mid-March. Thereafter, 430 431 they forage around the breeding colony during the prelaying, incubation (Fig. 4), and 432 chick-rearing periods (Kato et al. 2003, Watanuki et al. 2009). The auklets experienced the lowest sea temperatures (4-6 °C) during the early breeding period compared with 433 the rest of the year (11-18 °C), based on the water temperature records from the 434 geolocators (Fig. 5). Therefore, the thermoregulatory energy costs may be highest 435 during the early breeding period, as the lower end of thermoneutral zone of the auklets 436 has been estimated to be 15 °C based on body mass (Shirai et al. 2013). The auklets 437 often attended to the nest burrow during nighttime, and this might improve 438 thermoregulatory efficiency in the early breeding period. Still, it remains unclear as to 439 440 why auklets choose to return in March to breed in the northern Japan Sea, given the relatively high thermoregulatory costs. 441

One explanation could be the availability of euphausiids around the breeding colony in early spring. In the northern Japan Sea, euphausiids *Thysanoessa longipes* and *T. inermis* come to the surface to spawn when spring phytoplankton blooms occur (area B in Fig. 7) and when sea surface temperatures are 4-8 °C (Hanamura et al. 1989). The auklets feed on euphausiids and low trophic level juvenile fish in early spring based on the egg yolk isotope values (Fig. 6). The carotenoid-rich euphausiids may be suitable prey for females during egg production (Ito et al. 2009), and could be an important

449 factor determining the timing of auklets' migration to the northern Japan Sea. Another, 450 not mutually exclusive, explanation could be that the auklets need to adjust their chick-rearing period to the arrival of migrating warm water forage fish in early summer 451 (Watanuki et al. 2009). Japanese anchovy, a profitable prey during the chick-rearing 452 period, migrates to the sea around Teuri Island when warm waters (> 13 °C) from the 453 454 Tsushima Warm Current intrude into the foraging area in late May and mid June (Watanuki et al. 2009). If the auklets are to adjust chick-rearing to the warm water 455 456 arrival, they need to be back in the colony and have laid their eggs by mid April to early May, even though the sea temperature are still low at this time, given that incubation 457 458 takes 45 days on average (Gaston & Jones 1998).

Proficient-flying procellariiform seabirds can move long distances and shift their 459 foraging ranges seasonally, even while attending the colony. For example, streaked 460 shearwaters Calonectris leucomelas from the colony on the Pacific coast of Japan, shift 461 their foraging area from south to the north of the colony in April – July, while attending 462 the colony (Yamamoto et al. 2011). They fly up to a mean distance of 600 km from the 463 colony, presumably following the northward migration of Japanese anchovy 464 (Yamamoto et al. 2011). Rhinoceros auklets shift their foraging area seasonally during 465 466 the early to late breeding period (Deguchi et al. 2010, this study), but the range of shift is much smaller, reflecting higher flight costs (up to a median distance of 240 km from 467 468 the colony, see results). We suggest that the adjustment of breeding timing and location 469 are an important factor affecting the migratory patterns, especially in a species with a short foraging range during the breeding period. In fact, the seasonal mismatch between 470 471 breeding timing and anchovy migration has important fitness consequences in rhinoceros auklets (Watanuki et al. 2009). 472

473

#### 474 **Conservation implications**

The breeding population of rhinoceros auklets at Teuri Island is the largest in the northwestern Pacific, comprising more than 90 % of the total population in Asia (Gaston & Jones 1998). High concentrations of the nonbreeding auklets in the Sea of Okhotsk in autumn and in the southwestern Japan Sea in winter (Fig. 4) indicate that these two regions offer critical nonbreeding habitat for the population of Teuri Island. We suggest that the auklets are susceptible to 1) ocean warming and climatic regime

shifts, and 2) oil spill threats, which have been observed in both the Sea of Okhotsk and
southwestern Japan Sea.

Significant warming and reductions in winter sea-ice production have been 483 observed in the Sea of Okhotsk, and this may negatively influence the biological 484 productivity of these regions (Radchenko et al. 2010, and references therein). In the 485 486 northern part of the Sea of Okhotsk, increased sea surface temperatures negatively affected the breeding performance of planktivorous alcids, but the opposite was the case 487 488 for piscivorous alcids (Kitaysky & Golubova 2000). Climatic regime shifts are suggested to occur in the Japan Sea, and may have influenced the stock size of 489 490 epipelagic fish including Japanese anchovy (Kim et al. 2008) and the migration patterns of Japanese common squid (Kidokoro et al. 2010). 491

Oil platform development is planned for the Sea of Okhotsk, east of Sakhalin 492 Island (Sakurai et al. 2013), where auklets concentrate in autumn. Therefore if oil spills 493 occur in this region in autumn, it would have severe implications for the global 494 population of auklets. Also, in the southwestern Japan Sea, oil spills from tankers 495 passing through the Tsushima Strait may also affect the population if the spill happens 496 in winter. Indeed, 1326 and 482 oiled carcasses of rhinoceros auklets were recovered 497 498 from the shore of the southwestern Japan Sea, during oil spills from unknown sources in January 1986 (Sato 1999) and from a tanker in January 1997 (Fries et al. 1998), 499 500 respectively. Therefore, the information presented here is crucial for identifying key 501 areas of global significance for seabird conservation.

502

#### 503 CONCLUSION

Our results suggest that rhinoceros auklets followed spatial and seasonal changes 504 505 in prey availability by a distinctive 'three-step' migration after breeding; moving 506 northward to the Sea of Okhotsk in autumn, southward to the southwestern Japan Sea in 507 winter, and then returning northward to the breeding colony in the northern Japan Sea in early spring. The auklets appeared to continue feeding on higher trophic level prey 508 509 including fish and/or squid during the autumn and winter nonbreeding periods, but switched to lower trophic level prey in early spring when they were confined to forage 510close to the colony. The accessibility of foraging habitats with different seasonal 511 512 productivity patterns would be important for nonbreeding seabirds that have high

- 513 energetic constraints on migratory movements.
- 514

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685 Figures



# Figure 1

Fig. 1. *Cerorhinca monocerata.* Oceanographic features of study area. Warm and cold
currents displayed in pink and blue, respectively. TWC and ESC indicate the Tsushima
Warm Current and the East Sakhalin Current, mentioned in the text. Teuri Island
(breeding colony) is marked with a star.



Figure 2

Fig. 2. *Cerorhinca monocerata*. An example of time-series data obtained by geolocators for one bird. Latitude and longitude were estimated using light level and water temperature, and were smoothed using a movement model (see Materials and Methods for details). Water temperature recorded by the geolocator and time spent dry per day are also shown. Horizontal arrows indicate the early breeding period (after the first return to the breeding colony).







704

Figure 3 (continued)

Fig. 3. *Cerorhinca monocerata*. Monthly locations of nine rhinoceros auklets during the
nonbreeding and early breeding periods in August 2010 – April 2011. Each colour
shows locations from different individuals. Teuri Island is marked with a star.
Satellite-derived monthly-averaged sea surface temperature contours are also shown.



Figure 4

Fig. 4. Cerorhinca monocerata. Kernel density distribution of auklet locations during the nonbreeding (NB) and early breeding (EB) periods of first and second study year (2010-11 and 2011-12). Kernel densities indicate 25 %, 50 %, and 75 % from darker to lighter colours. Teuri Island is marked with a star. 



Figure 5

Fig. 5. *Cerorhinca monocerata*. Seasonal patterns in water temperature recorded by the geolocators during a) August 2010 – April 2011 and b) August 2011 – April 2012 (means  $\pm$  SE; n = 9 and 11 birds, respectively).



## Figure 6

724 725

Fig. 6. Cerorhinca monocerata.  $\delta^{15}N$  and  $\delta^{13}C$  values of primary and body feathers of 726 rhinoceros auklets collected in 2010-11 and 2011-12, before (black closed squares) and 727 after (grey closed squares) applying the adjustment of discrimination factors (indicated 728 by grey arrows).  $\delta^{15}$ N and  $\delta^{13}$ C values of auklet egg yolks and blood plasma collected 729 during the incubation (INCU) and chick-rearing (CR) periods (open squares; adjusted 730 for discrimination factors), and those of prey species (open circles, with names in 731italics) are also shown. The prey samples were obtained at the colony during the 732 chick-rearing periods in 2004 and 2005 (Ito et al. 2009). Means ± SE are shown. 733 Isotopic values of prey, the egg and blood plasma of auklets (open symbols) are based 734 on Ito et al. (2009, 2012). 735



# Figure 7

Fig. 7. Cerorhinca monocerata. Mean monthly-average chlorophyll a concentrations of 739 three different foraging areas (defined by 50 % kernel boundary of auklet locations) in 740the Sea of Okhotsk (Area A) and the northern and southwestern Japan Sea (Areas B and 741742 C) over 10 years (2003-2012). Mean values are shown with a thick line, and values of Mean  $\pm 1$  SD are shown with dotted lines, to show inter-annual variability. The arrows 743 below each graph show the approximate period when the auklets stayed in each of Area 744 A, B, and C. Please refer to Fig. S1 for the locations of the Areas A-C. Note the 745 746occurrence of autumn bloom in Area A. 747

748 **Electronic Supplement** 





## Figure S1

Fig. S1. Kernel density distribution of auklet locations during the nonbreeding period of the first and second study year combined (2010-11 and 2011-12 combined). Kernel densities indicate 25 %, 50 %, and 75 % from darker to lighter colours. Chlorophyll a concentrations were calculated for Area A (red), Area B (black) and Area C (blue). The areas were chosen on the basis of 50 % kernel density boundaries during nonbreeding (Areas A and C) and early breeding (Area B: see Fig 4) periods. Teuri Island is marked with a star.