



Article (refereed) - postprint

Maberly, S.C.; Berthelot, S.A.; Stott, A.W.; Gontero, B. 2015. Adaptation by macrophytes to inorganic carbon down a river with naturally variable concentrations of CO2.

Crown Copyright © 2014 Published by Elsevier GmbH.

This version available http://nora.nerc.ac.uk/508868/

NERC has developed NORA to enable users to access research outputs wholly or partially funded by NERC. Copyright and other rights for material on this site are retained by the rights owners. Users should read the terms and conditions of use of this material at http://nora.nerc.ac.uk/policies.html#access

NOTICE: this is the author's version of a work that was accepted for publication in *Journal of Plant Physiology*. Changes resulting from the publishing process, such as peer review, editing, corrections, structural formatting, and other quality control mechanisms may not be reflected in this document. Changes may have been made to this work since it was submitted for publication. A definitive version was subsequently published in *Journal of Plant Physiology* (2015), 172. 120-127. 10.1016/i.jplph.2014.07.025

www.elsevier.com/

Contact CEH NORA team at noraceh@ceh.ac.uk

The NERC and CEH trademarks and logos ('the Trademarks') are registered trademarks of NERC in the UK and other countries, and may not be used without the prior written consent of the Trademark owner.

The productivity and ecological distribution of freshwater plants can be controlled by the availability of inorganic carbon in water despite the existence of different mechanisms to ameliorate this, such as the ability to use bicarbonate. Here we took advantage of a short, natural gradient of CO₂ concentration, against a background of very high and relatively constant concentration of bicarbonate, in a spring-fed river, to study the effect of variable concentration of CO₂ on the ability of freshwater plants to use bicarbonate. Plants close to the source, where the concentration of CO₂ was up to 24-times air equilibrium, were dominated by Berula erecta. pH-drift results and discrimination against ¹³C were consistent with this and the other species being restricted to CO₂ and unable to use the high concentration of bicarbonate. There was some indication from stable ¹³C data that *B. erecta* may have had access to atmospheric CO₂ at low water levels. In contrast, species downstream, where concentrations of CO₂ were only about 5-times air-equilibrium were almost exclusively able to use bicarbonate, based on pH-drift results. Discrimination against ¹³C was also consistent with bicarbonate being the main source of inorganic carbon for photosynthesis in these species. There was, therefore, a transect downstream from the source of increasing ability to use bicarbonate that closely matched the decreasing concentration of CO₂. This was produced largely by altered species composition, but partly by phenotypic changes in individual species.

1	Adaptation by macrophytes to inorganic carbon down a river with naturally variable
2	concentrations of CO ₂
3	
4	
5	Maberly S.C. ^{1,2} , Berthelot S.A. ^{1,3} , Stott A.W. ² , Gontero B. ¹
6	
7	
8	¹ Aix-Marseille Université CNRS, BIP UMR 7281, 31 Chemin Joseph Aiguier, 13402
9	Marseille Cedex 20, France
10	² Centre for Ecology & Hydrology, Lancaster Environment Centre, Library Avenue, Bailrigg,
11	Lancaster LA1 4AP UK
12	³ Present address: Universitat Konstanz, Universitatstrasse 10, 78464, Konstanz, Germany
13	
14	
15	
16	
17	To whom correspondence should be addressed:
18	Stephen C. Maberly
19	Lake Ecosystems Group, Centre for Ecology & Hydrology, Lancaster Environment Centre,
20	Library Avenue, Bailrigg, Lancaster LA1 4AP UK
21	email: scm@ceh.ac.uk
22	tel: +44 1524595851

	L - 4	
-	∖bst	racı

23

24

25

26

27

28

29

30

31

32

33

34

35

36

37

38

39

40

41

The productivity and ecological distribution of freshwater plants can be controlled by the availability of inorganic carbon in water despite the existence of different mechanisms to ameliorate this, such as the ability to use bicarbonate. Here we took advantage of a short, natural gradient of CO₂ concentration, against a background of very high and relatively constant concentration of bicarbonate, in a spring-fed river, to study the effect of variable concentration of CO₂ on the ability of freshwater plants to use bicarbonate. Plants close to the source, where the concentration of CO₂ was up to 24-times air equilibrium, were dominated by Berula erecta. pH-drift results and discrimination against ¹³C were consistent with this and the other species being restricted to CO₂ and unable to use the high concentration of bicarbonate. There was some indication from stable ¹³C data that *B. erecta* may have had access to atmospheric CO₂ at low water levels. In contrast, species downstream, where concentrations of CO₂ were only about 5-times air-equilibrium were almost exclusively able to use bicarbonate, based on pH-drift results. Discrimination against ¹³C was also consistent with bicarbonate being the main source of inorganic carbon for photosynthesis in these species. There was, therefore, a transect downstream from the source of increasing ability to use bicarbonate that closely matched the decreasing concentration of CO₂. This was produced largely by altered species composition, but partly by phenotypic changes in individual species.

42

43

44

Keywords: bicarbonate, Fontaine de Vaucluse, photosynthesis, river Sorgue, stable carbon isotope

Introduction

The function and biodiversity of inland waters is controlled by interactions between the physical and chemical environment and the physiological and biochemical acclimation and adaptation of the organisms present as well as their short-term behavioural responses. High biological demand for resources, created by high potential metabolic rates or high biomass density, or both, can exceed the rate of re-supply and so alter the chemical and physical conditions in inland waters. Conversely, uncoupling of biogeochemical cycles of production and decomposition can generate a high supply of resource when the demand is low.

The primary producers at the base of the aquatic food web can be planktonic, epiphytic or benthic. In rivers, benthic macrophytes, Plantae that are mainly derived from land plants that have re-invaded the water, are often dominant because the rapid water transit time can prevents the formation of large populations of phytoplankton, especially in upstream regions where nutrient concentrations can be low (Reynolds and Descy, 1996). Macrophytes can survive high flushing rates or low nutrient resources in the water, as in many upland rivers, because most have roots, anchoring them to the substrate and allowing them to access nutrients in the substrate.

However, freshwater macrophytes require high concentrations of CO₂ to saturate photosynthesis because of a high transport resistance across relatively large external boundary layers (Black et al., 1981, Maberly and Madsen, 1998). Photosynthesis can be limited further by intermittent depletion of CO₂ produced when rates of photosynthetic demand exceed rates of resupply and by the generation of high concentrations of oxygen that can promote photorespiration (Maberly and Madsen, 2002, Pedersen et al., 2013). However, aquatic photoautotrophs possess avoidance, exploitation and amelioration strategies to minimize the effects of potential carbon limitation (Klavsen et al., 2011). Avoidance strategies involve living in environments where the concentration of CO₂ is naturally high, for example as a result of heterotrophic decomposition of organic matter (Maberly, 1985). Exploitation strategies include gaining access to more reliable sources of

CO₂ such as the atmosphere (Maberly and Madsen, 2002). Just over half of the freshwater macrophytes tested so far have amelioration strategies based on biophysical or biochemical carbon concentrating mechanisms (CCMs) to minimise carbon limitation photorespiration (Maberly and Madsen, 2002). The most widespread CCM is the use of bicarbonate as an alternative source of carbon. For biochemical details see also (Kroth, 2015) and Sage ans Stata (2015). Because of physico-chemical equilibria, bicarbonate is less susceptible than CO₂ to depletion during removal of inorganic carbon by photosynthesis and is present at concentrations that exceed CO₂ at sites where the pH exceeds the pH representing the first pK value of the carbonate system at approximately pH 6.3.

Although the use of bicarbonate is widespread, it is not universal, probably because there are costs involved in its use. First, bicarbonate uptake involves moving bicarbonate against its electrochemical potential gradient and across an inherently impermeable plasmalemma and is therefore an active process requiring expenditure of ATP. In situations where light energy is low, species therefore tend to lack the ability to use bicarbonate. One example is the predominance of species at depth that lack an ability to use bicarbonate such as sublittoral marine red macroalgae (Maberly, 1990) and in freshwater, aquatic bryophytes (Krause-Jensen and Sand-Jensen, 1998). Secondly, there are further energetic costs associated with the production and maintenance of the proteins involved in bicarbonate use (Jones, 2005). Thirdly, there is also a 'permeability cost' involved in using bicarbonate: species that use bicarbonate have a lower affinity for CO₂ than species that are restricted to CO₂ (Maberly and Madsen, 1998). This is probably caused by a high internal resistance to CO₂ uptake (Madsen and Maberly, 2003) that minimises the diffusive loss of carbon actively pumped into the photosynthesising tissue, but consequently reduces rates of photosynthesis at limiting concentrations of CO₂.

Comparisons of the ability of freshwater macrophytes to use bicarbonate or operate a CCM are usually based on a comparison of plants collected from sites with very different physical and chemical characteristics. An alternative comparative approach compares the

physiology of plants or algae at sites with locally high concentrations of CO2 with those at nearby sites with lower concentrations of CO₂. An example in the terrestrial environment includes studies at natural CO₂ vents associated with volcanic activity (Korner and Miglietta, 1994). An example from the marine environment is the use of geothermal vents to study the effects of high CO₂ and low pH as a surrogate for future ocean acidification (Hall-Spencer et al., 2008). This approach does not appear to have been used for freshwater macrophytes at a specific site despite the high variability in CO₂ concentrations in different surface waters and the frequently high CO₂ concentrations found in groundwater and groundwater-fed streams (Sand-Jensen and Staehr, 2012, Stets et al., 2009), although comparisons have been made over wider spatial areas e.g. (Demars and Tremolieres, 2009). The aim of the work reported here was to assess the ability of river macrophytes to use bicarbonate at different sites down a natural gradient of high CO₂ concentration from the groundwater-fed source of a river to lower concentrations downstream. The comparison is therefore made on plants experiencing very similar background physico-chemical conditions apart from CO₂. Furthermore, since this CO₂ gradient is likely to have been present for thousands of years, it allows adaptive responses to varying CO₂ to be distinguished from acclimative responses to short-term variability.

117

100

101

102

103

104

105

106

107

108

109

110

111

112

113

114

115

Materials and methods

118

143

144

119	La Sorgue at Fontaine de Vaucluse
120	Fontaine de Vaucluse, the source of the River Sorgue, is located in a Karst landscape in
121	Provence, in south-east France (Fig. 1). It is the largest spring in France, and one of the
122	largest in the world with a mean discharge of about 23 m ³ s ⁻¹ and minimum and maximum
123	discharges of 4 and over 100 m ³ s ⁻¹ respectively (Bonacci, 2007). The recharge area
124	comprises 1115 km² of Lower Carboniferous limestone, 1500 m thick, on largely uncultivated
125	land dominated by Mediterranean forest and 'garrigue'- dry limestone scrubland (Blavoux et
126	al., 1992) which on average receives an annual rainfall of 1096 mm (Bonacci, 2007). The
127	outflow has a very high concentration of bicarbonate of about 4.25 mmol L ⁻¹ (Emblanch et
128	al., 2003) with calcium as the dominant cation with a concentration of around 2 mmol L ⁻¹
129	(Garry et al., 2008). The geometric mean pH of 7.25, calculated from the frequent outflow
130	data over a year presented in (Emblanch, Zuppi, 2003), was used to estimate an average
131	CO ₂ concentration of about 0.6 mmol L ⁻¹ assuming a water temperature of 12°C and an ionic
132	strength of 0.0065 mol L ⁻¹ based on ionic composition from adjacent cave water given in
133	(Garry, Blondel, 2008) which had a similar bicarbonate concentration to the outflow.
134	
135	Field methods
136	Samples were collected on 12/09/2013, between 11:00 and 16:00, down a transect from
137	close to the source of the spring at Fontaine de Vaucluse to about 20 km downstream on
138	one of the many distributaries in this region (Fig. 1). On this date, the discharge was
139	relatively low at about 9 m ³ s ⁻¹ (REALPACA, 2013) which is slightly less than half the annual
140	mean. At each site, water was collected in a large bucket and water temperature was
141	measured immediately and the concentration of CO ₂ measured with a Vaisala non-
142	dispersive infrared gas analyser protected for use in water with a waterproof membrane as

described by (Johnson et al., 2010). Water was stored in two completely-filled 50 mL Falcon

polypropylene centrifuge tubes and kept in the dark in a cool box for analysis in the

laboratory on the next day. For stable carbon isotope measurement, 10 mL of la	ake or stream
water was injected via a syringe into evacuated (< 5 Pa) 12 mL exetainer (Labo	o) containing
0.15 mL degassed concentrated phosphoric acid (Maberly et al., 2013, Waldron	n et al.,
2007). Samples were collected in duplicate and kept inverted prior to analyses.	At each site,
macrophytes were collected by hand or with a drag rake and placed, moist, in p	olythene
bags and stored in a cool box overnight.	

Laboratory analyses

In the laboratory, macrophyte material for stable carbon isotope measurement was cleaned carefully and dried at 80°C for 24 hours and stored in aluminium foil before analysis. The pH of water from the river was measured with a combination pH-electrode (PHEL-GS2, Labbox) and meter (pH201, Hanna Instruments). Gran titration was used to measure alkalinity by duplicate titration of 15 mL samples with approximately 0.17 mol L⁻¹ HCl that had been standardised against 1 mol L⁻¹ NaHCO₃ (Mackereth et al., 1978). Inorganic carbon concentrations were calculated from measured temperature, pH and alkalinity following (Maberly, 1996). Calcite saturation was calculated as in (Maberly, 1996) using the equations of (Jacobson and Langmuir, 1974) to estimate the temperature-dependent calcite solubility product.

pH-drift experiments

Macrophytes and benthic filamentous algae were identified, cleaned carefully in tap water using a soft brush and rinsed in 1 mol L⁻¹ NaHCO₃ for at least 20 minutes. This concentration was chosen because it has been frequently used and while high enough to allow sufficient bicarbonate, should it be present, it is not so high as to require extremely high pH values to be generated to deplete inorganic carbon. Three species from each site were placed in 13 mL of fresh 1 mmol L⁻¹ NaHCO₃ in 15 mL Falcon tubes that contained about 2 mL of air. The

tubes were capped and placed horizontally in an incubator (Innova 4230, New Brunswick
Scientific) at 17°C under continuous illumination from fluorescent tubes producing about 170
μmol m ⁻² s ⁻¹ photosynthetically active radiation (Macam Q201, Macam Photometrics,
Livingstone, Scotland). pH was measured with the combination pH-electrode after 24 hours
and roughly after every 12 hours until a maximum pH had been reached. Alkalinity was
measured at the end of the experiment on duplicate aliquots by Gran titration.

Stable isotope analysis

For the analysis of inorganic carbon δ^{13} C, four mL of helium (99.999%) was injected into the headspace of each exetainer to overpressurise. After shaking and 30 minutes equilibration a 40 µL gas sample was removed for analysis. δ^{13} C of the CO₂ was measured on an Isoprime Ltd Tracegas Preconcentrator coupled to an Isoprime Ltd Isoprime isotope ratio mass spectrometer. Pulses of known reference CO₂ and blanks were run prior to each batch.

Small amounts of the plant were re-dried at 105°C and aliquots sealed into 6 x 5 mm tin capsules and loaded into an autosampler (Eurovector Elemental Analyser, Eurovector, Milano Italy) coupled in-line to a stable isotope ratio mass spectrometer (Isoprime, Manchester, UK). Each sample was combusted at 1020°C with a pulse of oxygen and products were carried by a flow of helium through a reduction reactor containing copper wire at 650°C and dried with magnesium perchlorate. N₂ and CO₂ were separated by a packed GC column and delivered, via an 'open-split' to the isotope ratio mass spectrometer. Values were compared to pulses of CO₂ reference gas and to a solid working standard of known isotopic composition. Stable isotope methods for inorganic and organic C were accredited to UKAS ISO17025.

The δ^{13} C values of CO₂ and bicarbonate were calculated from the measured δ^{13} C of inorganic carbon and carbonate speciation using the temperature-dependent equations in (Mook et al., 1974) following (Maberly et al., 1992). Discrimination of plants against these

potential carbon sources was calculated as in (Maberly, Raven, 1992) and against atmospheric CO₂ with an assumed value of -8‰ (Verburg, 2007).

Results

197

198

199

200

201

202

203

204

205

206

207

208

209

210

211

212

213

214

215

216

217

218

219

220

221

222

223

The water temperature at the five sites varied by 2°C between 13.8°C at site A and 15.8°C at site E (data not shown). Over the first 8 km downstream from the source, represented by the first four sites, there was a large increase in pH from 7.35 to 8.05 (Fig. 2A). This was linked to a decrease in concentration of CO₂ from about 0.46 to 0.09 mmol L⁻¹ (24- to 5-times air-equilibrium; Fig. 2B). The CO₂ concentration declined with distance downstream over the first four sites, covering a distance of 6.6 km and a reduction in altitude of 15 m (i.e. a slope of 2.27 m km⁻¹), at about $\log_e 0.188$ mmol L⁻¹ km⁻¹; R² = 0.96. Even at the fifth site, 20 km downstream from the source, the concentration of CO2 was about five times the airequilibrium concentration. These CO₂ concentrations were calculated from pH, alkalinity and temperature, but direct measurements in the field with the non-dispersive infra-red sensor gave very similar values and a comparison of the two methods across the five sites gave an R² of 0.94, but these data from the sensor are not used further. The alkalinity was high at all the sites and in contrast to the concentration of CO₂, its value only changed slightly from 4.84 to 4.73 mequiv L⁻¹ down the transect. The concentration of bicarbonate ions was correspondingly high and rather constant, varying between 4.83 and 4.68 mmol L⁻¹. The concentration of carbonate was only 0.005 mmol L⁻¹ at the source, but because of increasing pH, the concentration had increased to 0.028 mmol L⁻¹ (about 4.8-fold increase) at the fifth site. Assuming a calcium concentration of 2 mmol L⁻¹, calcite was about 1.4-times oversaturated at the source increasing to about 7-times over-saturated at the two lower sites (saturation index for calcite 0.15 to 0.85) and this may have been one of the reasons for the slightly lower alkalinity at the lower sites.

Nine species or genera of macrophytes and filamentous macroalgae were found at the five sites (Table 1). At the top of the transect, the vegetation was dominated by large underwater populations of *Berula erecta* (Fig. 3) along with the freshwater moss *Fontinalis*

antipyretica. At sites D and E with the lowest concentrations of CO₂ these were largely replaced by *Potamogeton nodosus*, *Stuckenia pectinata* (previously *Potamogeton pectinatus*) *Ceratophyllum demersum* and *Ranunculus flutians* (Table 1).

All the species of macrophytes at the three sites nearest the source (sites A to C) had final CO₂ concentrations at the end of the pH-drift experiment greater than 1 µmol L⁻¹, raised the pH to less than 9.1 and had C_T/Alk quotients close to 1 (Table 2). The data are consistent with these species being restricted to CO₂ as a source of inorganic carbon for photosynthesis. The same was true for *B. erecta* at site D, but all the other species at this site showed evidence of bicarbonate use with pH values over 10.0 and final CO₂ concentrations that were between 20- and 150-times below the CO₂ compensation concentrations generated by the species restricted to CO₂. This was also the case for the species from the fifth site (site E). One species, *Ceratophyllum demersum*, however, showed signs of deterioration during the drift experiment and the final alkalinity at the end of the experiment was over twice the starting alkalinity; pH-drift data from this species are not used further.

The δ^{13} C value of the inorganic carbon was relatively constant: it was most depleted at the source at -11.6% and varied between -10.5 and -10.9% at the four other sites- very similar to values reported in (Emblanch, Zuppi, 2003). Plant δ^{13} C values were more negative (depleted) at sites with high concentrations of CO₂ (Fig. 4A). Similarly plant δ^{13} C values were more negative for plants that were unable to use bicarbonate and hence had higher final CO₂ concentrations at the end of pH-drift experiments (Fig. 4B).The δ^{13} C values for *Berula erecta* tended to be less negative than other plants that were restricted to CO₂.

Discrimination, Δ , was calculated for the different plant species for different putative sources of inorganic carbon (Fig. 5). The wholly submerged species that were restricted to CO_2 had an average discrimination value of 26.4 % (SD = 1.4 %). If *B. erecta* was restricted to CO_2 in water its average discrimination value would be 15.5 % (SD = 3.0 %). If instead its

organic carbon was produced solely from atmospheric CO ₂ , the average discrimination value
would be 28.6 $\%$ (SD = 3.2 $\%$). For species with the ability to use bicarbonate, two potential
forms of inorganic carbon are available. If these species were using CO ₂ , discrimination
against CO_2 would be only 11.2 % (SD = 2.7 %) but their discrimination against bicarbonate
would be 21.6 ‰ (SD = 2.7 ‰).
There was a strong linear relationship between the final CO ₂ in pH-drift experiments and
the CO ₂ concentration at the collection site (Fig. 6). The correlation was even stronger if B.
erecta from site 4 was excluded from the data set ($R^2 = 0.99$; data not shown).

Discussion

260

261

262

263

264

265

266

267

268

269

270

271

272

273

274

275

276

277

278

279

280

281

282

283

284

285

Rivers generally have high concentrations of CO₂ because they receive large amounts of terrestrial organic carbon that has been degraded to inorganic carbon in the soil and have short residence times that limit evasion to the atmosphere (Cole et al., 2007, Raymond et al., 2013, Sand-Jensen and Staehr, 2012) although some rivers can be undersaturated with CO₂ during periods of high productivity (Neal et al., 1998). Groundwater-fed rivers often have particularly high concentrations of CO₂ at source because of sub-surface respiration and restricted exchange with the atmosphere (Cole, Prairie, 2007). In the case of the Fontaine de Vaucluse, data in (Emblanch, Zuppi, 2003) were used to calculate a mean concentration of CO₂ of 0.61 mmol L⁻¹ over a year at the source. On our sampling day, 0.3 km downstream from the source, the concentration was about 0.46 mmol CO₂ L⁻¹, corresponding to the mean in a compilation from groundwaters from New Zealand and Denmark (Sand-Jensen and Staehr, 2012) and similar to the maximum values reported in high alkalinity rivers in eastern France (Massabuau and Fritz, 1984). In limestone regions, these high concentrations of CO₂ are associated with high concentrations of bicarbonate, but high CO₂ associated with groundwater inputs can also be found in non-limestone regions where alkalinity is relatively low (Demars and Thiebaut, 2008).

Turbulent flow, especially in streams and rivers with steep slopes, leads to rapid reduction in the concentration of CO₂ by evasion to the atmosphere and net photosynthetic uptake causes the concentration to decline further (Butman and Raymond, 2011, Sand-Jensen and Staehr, 2012). This reduction in concentration of CO₂ was also found here; concentrations declined 2-fold over 3.7 km.

The species of macrophytes found in this study are typical of European rivers, especially those with a groundwater influence. For example, all the macrophytes found here were recorded in a survey of macrophytes from groundwater-fed regions of the rivers Rhine, Rhône and Moosach, (Demars and Tremolieres, 2009) apart from *P. nodosus*. Furthermore,

286

287

288

289

290

291

292

293

294

295

296

297

298

299

300

301

302

303

304

305

306

307

308

309

310

311

312

(Demars and Tremolieres, 2009) showed that plant distribution at different sites was related more to the concentration of CO₂ than to other chemical variables such as the concentration of ammonium or phosphate. We show here that, as previously assumed based on literature information, (Demars and Tremolieres, 2009), this pattern is caused by the differential ability of the different species to utilize bicarbonate as a source of inorganic carbon in addition to CO₂. At the first three sites below the spring down to about 3 km, the concentrations of CO₂ exceeded 10-times air-equilibrium (0.46 to 0.22 mmol L⁻¹) and all the species found were unable to use bicarbonate. The amphibious macrophyte Berula erecta was dominant at these sites and this species is known to be restricted to CO₂ (Sand-Jensen et al., 1992) and this was confirmed here. Based on our measurements of CO₂ concentration and the kinetic response of photosynthesis to CO₂ in (Sand-Jensen, Pedersen, 1992), we estimate that B. erecta would be between 75% and 100% saturated with CO₂ at these sites. At site D, where only small amounts of B. erecta were found, the CO₂ concentration of 0.09 mmol L⁻¹ would only support about 20% of the CO₂ saturated rates. The absence of species able to use bicarbonate at the upper sites, despite substantial concentrations of bicarbonate, exceeding 4.8 mmol L⁻¹, is consistent with substantial costs linked to the reduced affinity for CO₂ (Maberly and Madsen, 1998), and increased costs of running the process and producing and maintaining the extra machinery required (Jones, 2005). Conversely, at the lowermost sites, all species found and tested, had the ability to use bicarbonate. Here, CO₂ concentrations were only about 4.8-times greater than air-equilibrium which is likely to limit rates of photosynthesis if CO₂ was the only source of inorganic carbon (Sand-Jensen and Frost-Christensen, 1999).

The stable carbon isotope data can be used to give some indication of the source of inorganic carbon for the different species of macrophytes at the different sites if putative source values are known (Osmond et al., 1981). pH-drift experiments suggested that *F. antipyretica* and *Vaucheria* sp. were restricted to CO₂. Discrimination against CO₂ was about 25 to 28 % which is consistent with typical C3 photosynthesis with little diffusion limitation

313

314

315

316

317

318

319

320

321

322

323

324

325

326

327

328

329

330

331

332

333

334

335

336

337

338

339

(Lloyd and Farquhar, 1994). Although *B. erecta* was also limited to CO₂, discrimination against this carbon source was only 12 to 19 ‰ which could indicate either diffusion limitation or that *B. erecta* may have taken up CO₂ from the atmosphere at very low water level even though the benefit is likely to be small at these very high CO₂ concentrations in the river as suggested by work on another amphibious species (Madsen and Breinholt, 1995). The discrimination values for species shown to use bicarbonate in the drift experiments were very low (8 to 16 ‰) against CO₂ but much more typical if expressed against bicarbonate (18 to 26 ‰) which may indicate that this was the dominant source of inorganic carbon even at moderate concentrations of CO₂.

B. erecta was found at four of the five sites, growing in CO₂ concentrations from 0.46 to 0.09 mmol L⁻¹. Although we do not have information on seasonal changes in CO₂ at the sites, the downstream gradient is likely to be relatively stable because the annual variation of discharge is relatively low and because the data in (Emblanch, Zuppi, 2003) show no significant correlation between CO_2 concentration at the source and discharge (r = -0.039). There was, however, some slight evidence for change in the CO₂ response of the different populations. In pH-drift experiments, the B. erecta population at the site nearest the spring had a final CO_2 concentration of about 19 $\mu mol \ L^{-1}$ whereas the three downstream populations sampled had final CO₂ concentrations of between 2 and 4 μmol L⁻¹. In contrast, the final CO₂ concentration of the aquatic moss, *F. antipyretica*, that was found at the three upper sites, was invariable at 2 to 3 µmol L⁻¹. More detailed work will be needed to determine whether phenotypic or genotypic change is involved in the putative differences in the B. erecta populations. Genotypic change is possible given the potentially large number of generations that could have occurred at this site over thousands of years. However, a study of soil algae at two high CO₂ (aerial) springs, found little evidence for genetic adaptation to high CO₂ concentrations (Collins and Bell, 2006). The ability of plants such as *B. erecta* to grow well in rivers is strongly linked to the high concentrations of CO₂ that can sometimes be found there (Sand-Jensen, Pedersen, 1992) and also explains why these species are

relatively rare in lakes which tend to have much lower CO₂ concentrations during most of the year (Baattrup-Pedersen et al., 2013). Although bicarbonate users can down-regulate their ability to use bicarbonate depending on inorganic carbon availability (Madsen et al., 1996) this does not appear to have allowed them to compete with *B. erecta* when concentrations of CO₂ were high at upstream sites.

Decomposition processes in the recharge zone can produce high concentrations of CO₂ in groundwater-fed rivers. This, coupled with a gradient of CO₂ loss downstream, produces a natural experiment to test the effect of elevated CO₂ on the biodiversity and physiology of aquatic plants in rivers which helps to explain the distribution of macrophytes in rivers.

Acknowledgements

SCM carried out this work during a visiting scholarship funded by Aix-Marseille Université. SAB was funded by a DAAD Erasmus fellowship. We thank Richard Lansdown and David John for confirming the identifications of some of the macrophytes and filamentous algae, respectively, Helen Grant for making the plant stable isotope measurements and Martin Rouen and David Aspinall for constructing the CO₂ sensor. We thank the referees for their comments on an earlier version of the manuscript.

358	Figure legends
359	Fig. 1. Location of the sampling site in France (A), showing the catchment area (dashed
360	line), the source (cross) and the downstream flow to the River Rhône (B) and the five sample
361	sites A to E (C).
362	Fig. 2. Changes in carbonate chemistry on the River Sorgue with distance downstream from
363	the source at Fontaine de Vaucluse. pH (A); alkalinity (open circles) and concentration of
364	bicarbonate (closed circles; B); concentration of carbon dioxide measured directly (open
365	circles) and calculated from pH, alkalinity and temperature (closed circles) with air-
366	equilibrium indicated by a dashed line (C) and concentration of carbonate (D).
367	Fig. 3. The River Sorgue downstream from site A. Showing general appearance of the river
368	(A) and the dominance of Berula erecta (B).
369	Fig. 4. Plant δ^{13} C values from the five sites for <i>Berula erecta</i> (open triangles), submerged
370	$\text{CO}_2\text{-users}$ (open circles), and bicarbonate users (closed circles). A Plant $\delta^{13}\text{C}$ as a function
371	of site CO_2 concentration; B Plant $\delta^{13}\text{C}$ as function of final CO_2 concentration in pH-drift
372	experiments. The regression lines relate to all the combined data. Note the Log scale for
373	concentration of CO ₂ .
374	Fig. 5. Calculated discrimination, Δ , against air, (assumed -8%; grey bars), carbon dioxide
375	(white bars) and bicarbonate (black bars) for the different species on the River Sorgue below
376	Fontaine de Vaucluse. Species are ordered by increasing ability to deplete inorganic carbon
377	in pH-drift experiments. The asterisk indicates the likely source of inorganic carbon based on
378	pH-drift experiments.
379	Fig. 6. Correlation between average final concentration of CO ₂ at the end of a pH-drift
380	experiment and the concentration of ${\rm CO}_2$ at the sites from which the samples were collected.
381	

383	References
384	Baattrup-Pedersen A, Madsen TV, Riis T, Cavalli G. Photosynthetic performance of submerged
385	macrophytes from lowland stream and lake habitats with contrasting CO ₂ availability. New
386	Phytologist. 2013;198:1135-42.
387	Black MA, Maberly SC, Spence DHN. Resistances to carbon-dioxide fixation in four submerged
388	freshwater macrophytes. New Phytologist. 1981;89:557-68.
389	Blavoux B, Mudry J, Puig JM. The karst system of the Fontaine de Vaucluse (Southeastern France).
390	Environmental Geology and Water Sciences. 1992;19:215-25.
391	Bonacci O. Analysis of long-term (1878-2004) mean annual discharges of the karst spring Fontaine de
392	Vaucluse (France). Acta Carsologica. 2007;36:151-56.
393	Butman D, Raymond PA. Significant efflux of carbon dioxide from streams and rivers in the United
394	States. Nature Geoscience. 2011;4:839-42.
395	Cole JJ, Prairie YT, Caraco NF, McDowell WH, Tranvik LJ, Striegl RG, et al. Plumbing the global carbon
396	cycle: Integrating inland waters into the terrestrial carbon budget. Ecosystems. 2007;10:171-84.
397	Collins S, Bell G. Evolution of natural algal populations at elevated CO ₂ . Ecology Letters. 2006;9:129-
398	35.
399	Demars BOL, Thiebaut G. Distribution of aquatic plants in the Northern Vosges rivers: implications
400	for biomonitoring and conservation. Aquatic Conservation-Marine and Freshwater Ecosystems.
401	2008;18:619-32.
402	Demars BOL, Tremolieres M. Aquatic macrophytes as bioindicators of carbon dioxide in groundwater
403	fed rivers. Science of the Total Environment. 2009;407:4752-63.
404	Emblanch C, Zuppi GM, Mudry J, Blavoux B, Batiot C. Carbon 13 of TDIC to quantify the role of the
405	unsaturated zone: the example of the Vaucluse karst systems (Southeastern France). Journal of
406	Hydrology. 2003;279:262-74.

407	Garry B, Blondel T, Emblanch C, Sudre C, Bilgot S, Cavaillou A, et al. Contribution of artificial galleries
408	to the knowledge of karstic system behaviour in addition to natural cavern data. International
409	Journal of Speleology. 2008;37:75-82.
410	Hall-Spencer JM, Rodolfo-Metalpa R, Martin S, Ransome E, Fine M, Turner SM, et al. Volcanic carbon
411	dioxide vents show ecosystem effects of ocean acidification. Nature. 2008;454:96-99.
412	Jacobson RL, Langmuir D. Dissociation constants of calcite and CaHCO ³⁺ from 0 to 50°C. Geochimica
413	et Cosmochimica Acta. 1974;38:301–18.
414	Johnson MS, Billett MF, Dinsmore KJ, Wallin M, Dyson KE, Jassal RS. Direct and continuous
415	measurement of dissolved carbon dioxide in freshwater aquatic systems-method and
416	applications. Ecohydrology. 2010;3:68-78.
417	Jones J. The metabolic cost of bicarbonate use in the submerged plant <i>Elodea nuttallii</i> . Aquatic
418	Botany. 2005;83:71-81.
419	Klavsen SK, Madsen TV, Maberly SC. Crassulacean acid metabolism in the context of other carbon-
420	concentrating mechanisms in freshwater plants: a review. Photosynthesis Research.
421	2011;109:269-79.
422	Korner C, Miglietta F. Long-term effects of naturally elevated CO ₂ on mediterranean grassland and
423	forest trees. Oecologia. 1994;99:343-51.
424	Krause-Jensen D, Sand-Jensen K. Light attenuation and photosynthesis of aquatic plant communities.
425	Limnology and Oceanography. 1998;43:396-407.
426	Kroth P. The Biodiversity of Carbon Assimilation. J Plant Physiol 2015; 172: pp-pp
427	Lloyd J, Farquhar GD. C13 discriminationi during CO ₂ assimilation by the terrestrial biosphere.
428	Oecologia. 1994;99:201-15.
429	Maberly SC. Photosynthesis by Fontinalis antipyretica .2. Assessment of environmental-factors
430	limiting photosynthesis and production. New Phytologist. 1985;100:141-55.
431	Maberly SC. Exogenous sources of inorganic carbon for photosynthesis by marine macroalgae.
432	Journal of Phycology. 1990;26:439-49.

433	Maberly SC. Diel, episodic and seasonal changes in pH and concentrations of inorganic carbon in a
434	productive lake. Freshwater Biology. 1996;35:579-98.
435	Maberly SC, Barker PA, Stott AW, De Ville MM. Catchment productivity controls CO ₂ emissions from
436	lakes. Nature Climate Change. 2013;3:391-94.
437	Maberly SC, Madsen TV. Affinity for CO ₂ in relation to the ability of freshwater macrophytes to use
438	HCO ₃ . Functional Ecology. 1998;12:99-106.
439	Maberly SC, Madsen TV. Freshwater angiosperm carbon concentrating mechanisms: processes and
440	patterns. Functional Plant Biology. 2002;29:393-405.
441	Maberly SC, Raven JA, Johnston AM. Discrimination between ¹² C and ¹³ C by marine plants.
442	Oecologia. 1992;91:481-92.
443	Mackereth FJH, Heron J, Talling JF. Water analysis: some revised methods for limnologists.
444	Freshwater Biological Association Scientific Publication. 1978:1-120.
445	Madsen TV, Breinholt M. Effects of air contact on growth, inorganic carbon sources, and nitrogen
446	uptake by an amphibioius freshwater macrophyte. Plant Physiology. 1995;107:149-54.
447	Madsen TV, Maberly SC. High internal resistance to CO ₂ uptake by submerged macrophytes that use
448	HCO ₃ -: measurements in air, nitrogen and helium. Photosynthesis Research. 2003;77:183-90.
449	Madsen TV, Maberly SC, Bowes G. Photosynthetic acclimation of submersed angiosperms to CO ₂ and
450	HCO ₃ . Aquatic Botany. 1996;53:15-30.
451	Massabuau J-C, Fritz B. Respiratory gas content (O ₂ and CO ₂) and ionic composition of river water in
452	the plain of Alsace (eastern France). In: Eriksson E, editor. Hydrochemical balances of
453	freshwater systems. Uppsala: IAHS Press; 1984. p. 107-15.
454	Mook WG, Bommerso.Jc, Staverma.Wh. Carbon Isotope Fractionation between Dissolved
455	Bicarbonate and Gaseous Carbon-Dioxide. Earth and Planetary Science Letters. 1974;22:169-76.
456	Neal C, House WA, Jarvie HP, Eatherall A. The significance of dissolved carbon dioxide in major
457	lowland rivers entering the North Sea. Science of the Total Environment. 1998;210:187-203.

458	Osmond CB, Valaane N, Haslam SM, Uotila P, Roksandic Z. Comparisons of delta-C-13 values in
459	leaves of aquatic macrophytes from different habitats in Britain and Finland - some implications
460	for photosynthetic processes in aquatic plants. Oecologia. 1981;50:117-24.
461	Pedersen O, Colmer TD, Sand-Jensen K. Underwater photosynthesis of submerged plants - recent
462	advances and methods. Frontiers in Plant Science. 2013;4.
463	Raymond PA, Hartmann J, Lauerwald R, Sobek S, McDonald C, Hoover M, et al.Global carbon dioxide
464	emissions from inland waters. Nature. 2013;503:355-59.
465	REALPACA. Bulletin de situation hydrologique. REALPACA (http://www.pacadeveloppement-
466	durablegouvfr/IMG/pdf/Bulletin_hydro_paca_Septembre_2013_cle53be71pdf). 2013;N180:11
467	Reynolds CS, Descy JP. The production, biomass and structure of phytoplankton in large rivers.
468	Archiv fuer Hydrobiologie Supplement. 1996;113:161-87.
469	Sage RF, Stata M. Photosynthetic Diversity meets Biodiversity: The C ₄ plant Example. J Plant Physiol
470	2015: 172: pp-pp.
471	Sand-Jensen K, Frost-Christensen H. Plant growth and photosynthesis in the transition zone between
472	land and stream. Aquatic Botany. 1999;63:23-35.
473	Sand-Jensen K, Pedersen MF, Nielsen SL. Photosynthetic use of inorganic carbon among primary and
474	secondary water plants in streams. Freshwater Biology. 1992;27:283-93.
475	Sand-Jensen K, Staehr PA. CO ₂ dynamics along Danish lowland streams: water-air gradients, piston
476	velocities and evasion rates. Biogeochemistry. 2012;111:615-28.
477	Stets EG, Striegl RG, Aiken GR, Rosenberry DO, Winter TC. Hydrologic support of carbon dioxide flux
478	revealed by whole-lake carbon budgets. Journal of Geophysical Research-Biogeosciences.
479	2009;114.
480	Verburg P. The need to correct for the Suess effect in the application of delta C-13 in sediment of
481	autotrophic Lake Tanganyika, as a productivity proxy in the Anthropocene. Journal of
482	Paleolimnology. 2007;37:591-602.

483	Waldron S, Scott EM, Soulsby C. Stable isotope analysis reveals lower-order river dissolved inorganic
484	carbon pools are highly dynamic. Environmental Science & Technology. 2007;41:6156-62.

Table 1.

Species composition at the five sites on the River Sorgue (see Fig. 1 for location).

Species	Α	В	С	D	Е
Berula erecta (Huds.) Coville					
Fontinalis antipyretica Hedw.					
Vaucheria sp. DC.					
Lemna trisulca L.					
Stuckenia pectinata (L.) Böerner					
Cladophora sp. Kütz.					
Ceratophyllum demersum L.					
Potamogeton nodosus Poir.					
Ranunculus fluitans L.					

Table 2. Mean carbonate equilibria from triplicate pH-drift experiments. Values are the maximal pH, the minimal concentration of total inorganic carbon (C_T) and CO_2 and the quotient of C_T /alkalinity.

			Ст	CO ₂	
Site	Species	рН	(mmol L ⁻¹)	(mmol ^{-L-1})	C _T /Alk
Α	Berula erecta	7.61	0.348	1.98E-02	1.06
Α	Fontinalis antipyretica	9.02	0.960	2.16E-03	0.95
В	Berula erecta	8.55	0.324	2.20E-03	0.99
В	Fontinalis antipyretica	8.91	0.971	2.81E-03	0.96
В	Vaucheria sp.	8.54	1.174	8.16E-03	0.99
С	Berula erecta	8.34	0.328	3.57E-03	1.00
C	Fontinalis antipyretica	9.05	0.957	1.99E-03	0.95
D	Berula erecta	8.29	0.328	4.03E-03	1.00
D	Cladophora sp.	10.73	0.458	6.33E-06	0.42
D	Lemna trisulca	10.31	0.734	4.61E-05	0.61
D	Stuckenia pectinata	10.43	0.339	1.62E-05	0.49
E	Ceratophyllum demersum	9.72	2.266	9.10E-04	0.81
Е	Potamogeton nodosus	10.27	0.805	6.99E-05	0.60
Е	Ranunculus fluitans	10.45	0.294	1.19E-05	0.48

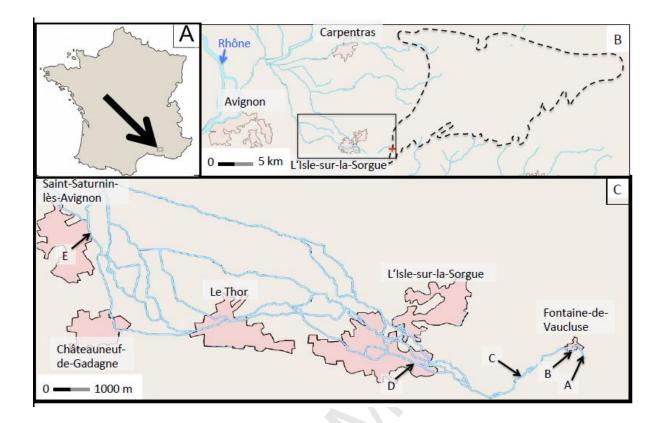


Fig. 1.

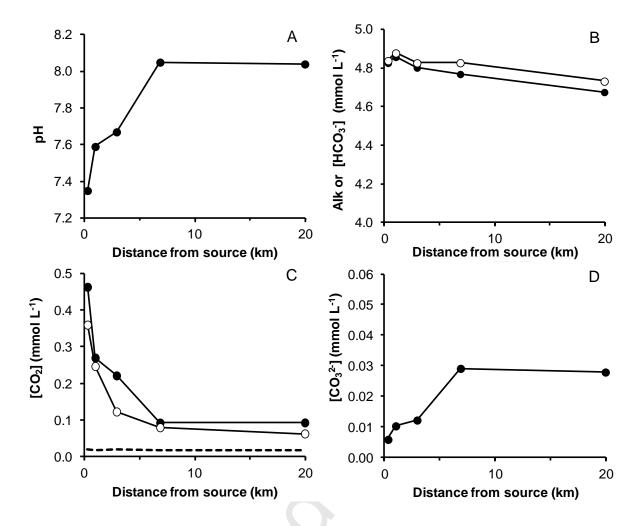


Fig. 2



Fig. 3

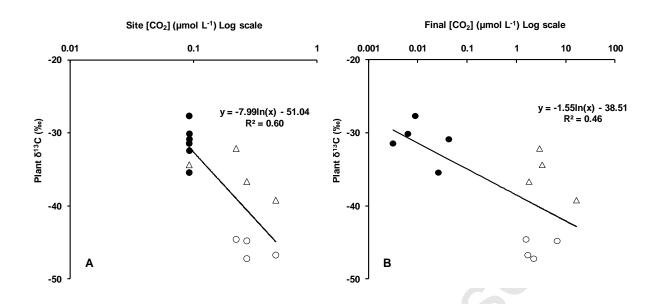


Fig. 4

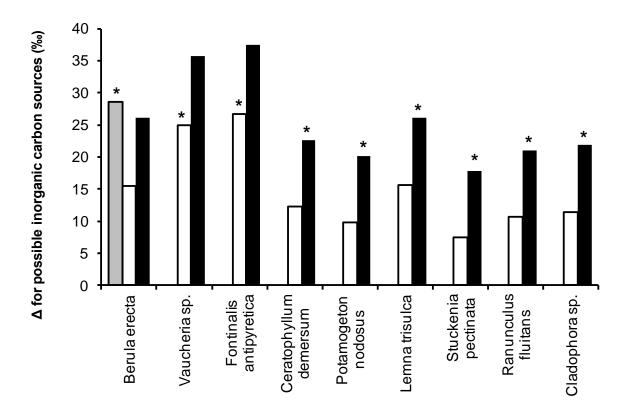


Fig. 5

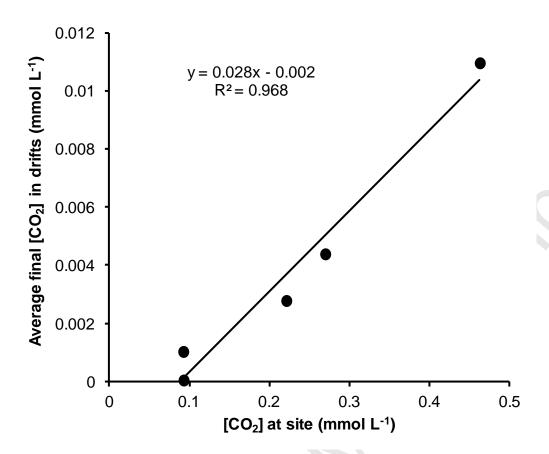


Fig. 6