Review

Phosphate oxygen isotopes within aquatic ecosystems: Global data synthesis and future research priorities

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HIGHLIGHTS

• Oxygen isotope ratio in dissolved inorganic phosphate a novel stable isotope tracer.
• Theoretical basis for application of this tracer in aquatic ecosystems reviewed.
• Protocols for determining phosphate oxygen isotope ratio summarised.
• Synthesis of global data from marine and freshwater ecosystems reported.
• Priorities for future research in this rapidly evolving field identified.

Abstract

The oxygen isotope ratio of dissolved inorganic phosphate (δ18Op) represents a novel and potentially powerful stable isotope tracer for biogeochemical research. Analysis of δ18Op may offer new insights into the relative importance of different sources of phosphorus within natural ecosystems. Due to the isotope fractionations that occur alongside the metabolism of phosphorus, δ18Op could also be used to better understand the intracellular and extracellular reaction mechanisms that control phosphorus cycling. In this review focussed on aquatic ecosystems, we examine the theoretical basis to using stable oxygen isotopes within phosphorus research. We consider the methodological challenges involved in accurately determining δ18Op, given aquatic matrices in which potential sources of contaminant oxygen are ubiquitous. Finally, we synthesise the existing global data regarding δ18Op in aquatic ecosystems, concluding by identifying four key areas for future development of δ18Op research.

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http://dx.doi.org/10.1016/j.scitotenv.2014.07.057
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1. Introduction

Phosphorus (P) is an essential element for all life, being integral to the structure and function of key biomolecules including DNA, RNA, adenosine triphosphate (ATP) and phospholipids. Under ambient conditions in natural ecosystems, P is tightly cycled within the biosphere and can limit or co-limit primary producer and microbial communities (Ruttenberg, 2003). However, inputs of P to terrestrial and aquatic ecosystems from a range of sources, alongside fluxes of P between these ecosystems, have increased significantly as a result of global population growth and attempts to expand and intensify food production for human society (Filippelli, 2008; Liu et al., 2008). Recent increases in bioavailable P fluxes and concentrations have been linked to undesirable ecosystem changes, including increases in primary production, shifts in community composition, increased frequency of algal blooms and hypoxia, and reduced biodiversity within aquatic ecosystems (Sondergaard and Jeppesen, 2007; Smith and Schindler, 2009).

Inefficient P use and the lack of effective recycling of P within urban and agricultural systems define a 'broken' P biogeochemical cycle, leading to concern over whether human use of P resources is sustainable (Cordell et al., 2009; Childers et al., 2011; Elser and Bennett, 2011). Whilst estimates of mineral P reserves have increased dramatically in recent years (Jasinski, 2012), these estimates remain highly uncertain and future exhaustion of reserves is likely without major advances in P mining and recycling technologies (Vaccari and Strigul, 2011; Seyhan et al., 2012). Securing access to P remains a globally significant issue, given the reliance of agricultural production on inorganic P fertilisers and the current lack of alternative sources for P beyond mineral reserves (Beardsley, 2011).

Despite the importance of P to human society, and the implications for natural ecosystems of perturbations to the P cycle, understanding of the sources and reaction mechanisms controlling biogeochemical cycling of P within ecosystems remains limited (Blake et al., 2005; Slomp, 2011). A critical reason for this is the lack of inherent tracers for analysing the sources and the metabolism of P in natural ecosystems (Karl, 2000). For example, quantifying the importance of different sources of P in aquatic ecosystems has previously relied on mass flux budgets, spatial and temporal analysis of P concentration, export coefficient models and indirect tracers such as boron as a marker for wastewater treatment plant (WWTP) effluent, or microbial source tracking to identify human versus agricultural sources of faecal contamination (Dillon and Kirchner, 1975; Smith et al., 1989; Neal et al., 2000; Jarvie et al., 2002; Scott et al., 2002; Simpson et al., 2002; Holt et al., 2003; Bowes et al., 2008). However, none of these approaches provides an inherent label for P. As a result, none offers a direct means of tracing specific sources or biogeochemical processes relevant to the P cycle.

The stable oxygen isotope ratio within phosphate (hereafter, $\delta^{18}$O$_{P}$) has recently emerged as a novel and potentially powerful inherent tracer for the sources and metabolism of P in natural ecosystems (McLaughlin et al., 2004; Elsbury et al., 2009; Young et al., 2009; Goldhammer et al., 2011a; Li et al., 2011). This review focusses on the use of $\delta^{18}$O$_{P}$ in aquatic ecosystems and particularly within freshwater ecosystems, complimenting a recent review dealing with $\delta^{18}$O$_{P}$ in soil–plant systems (Tamburini et al., 2014). Our objectives are to: i) examine the theoretical basis to the use of $\delta^{18}$O$_{P}$ in P research; ii) consider the methodological challenges involved in the analysis of $\delta^{18}$O$_{P}$ in aquatic ecosystems; and iii) synthesise global data from initial application of $\delta^{18}$O$_{P}$ within aquatic ecosystems. We conclude by identifying future priorities for $\delta^{18}$O$_{P}$ research within freshwater ecosystems, aiming to stimulate further development and application of this technique.

2. Stable isotopes and P biogeochemistry: theoretical background

Phosphorus has three major isotopes, $^{31}$P (stable), $^{32}$P ($t_{1/2} = 14.36$ days) and $^{33}$P ($t_{1/2} = 25.34$ days) (Smith et al., 2011). Biogeochemical cycling of P in aquatic ecosystems has previously been explored using the radioactive isotopes $^{32}$P and $^{33}$P (Benitez-Nelson and Buesseler, 1999; Benitez-Nelson, 2000). However, the use of radioisotopes is constrained by short isotope half-lives, perturbation of experimental systems associated with labelling, or the use of incubations which omits irregular events that occur in natural ecosystems, such as seasonal algal blooms (Levine et al., 1986; Thingstad et al., 1993; Benitez-Nelson, 2000).

Stable isotope ratios have been widely used to understand long-term trends and processes in both local and global biogeochemical cycles (Beveridge and Shackleton, 1994; Newton and Bottrell, 2007) and references therein). Stable isotope analysis requires an element to have a minimum of two naturally-occurring stable isotopes, with changes in the ratio of individual isotopes of an element in a sample, against a known ratio in a reference, providing insight into biogeochemical processes controlling the element cycle. Carbon (C), nitrogen (N) and oxygen (O) are commonly used in stable isotope research on the basis of ratios of $^{13}$C/$^{12}$C, $^{15}$N/$^{14}$N and $^{16}$O/$^{18}$O respectively. However, stable isotope analyses cannot be conducted on the P atom in P-containing compounds, because $^{31}$P is the only stable P isotope.

In natural aquatic ecosystems, P is often bound strongly to O in the form of the dissolved inorganic phosphate ion, defined hereafter as P$_{i}$. Therefore, attention has recently focussed on whether the stable isotope composition of P$_{i}$ can be used to gain insight into the biogeochemical cycling of P. Isotope fractionation involves the preferential incorporation of one isotope from a starting material into another during a reaction product. Kinetic fractionation describes a process that preferentially selects one isotope (generally the lighter isotope) due to a faster reaction rate in a unidirectional reaction. In contrast, equilibrium fractionation is a thermodynamic effect in which a system has time to exchange isotopes continuously in order to achieve the lowest energy system, wherein the heavier isotope forms the strongest bond possible (Hoefs, 2008). The P–O bonds in P$_{i}$ are resistant to inorganic hydrolysis under typical temperature and pressure conditions in the Earth’s surface water and groundwater ecosystems (O’Neil et al., 2003). Therefore, without biological mediation, negligible isotope exchange occurs between P$_{i}$ and water within these ecosystems (Tudge, 1960; Blake et al., 2001). Whilst the initial stages of some abiotic reactions, including the sorption of P$_{i}$ to solid-phase iron-oxide or the formation of apatite, may be associated with kinetic fractionation, this does not persist and through time the stable isotope composition of the solid–phase P$_{i}$ approaches that of the aqueous-phase P$_{i}$ (Liang and Blake, 2007; Jaisi et al., 2010, 2011). In contrast, enzyme-catalysed processes cleave P–O bonds leading to kinetic or equilibrium fractionation between the isotopes of P in P$_{i}$ and O in a surrounding fluid, either within a cell or within the extracellular environment (Blake et al., 2005). The fractionation between the stable isotopes of O ($^{16}$O and $^{18}$O) within a
sample is expressed as $\delta^{18}O$, relative to Vienna Standard Mean Ocean Water (VSMOW), defined as $0\%_{\text{VSMOW}}$ on the $\delta^{18}O$ scale (Eq. (1)).

$$
\delta^{18}O_{\text{sample}} = 1000 \left( \frac{\delta^{18}O_{\text{sample}}}{\delta^{18}O_{\text{standard}}} - 1 \right)_{\text{VSMOW}}
$$

(1)

Phosphate oxygen isotopes have previously been used within both the archaeological and environmental communities to determine palaeoclimate conditions at the time of formation of biogenic phosphatic material, such as apatite in fossil bones and teeth, as well as to reconstruct historical salinity and water temperatures from biogenic phosphate derived from fish remains (Longinelli and Nuti, 1973; Bryant et al., 1994; Fricke and O’Neil, 1996; Lécuyer et al., 1996; Stephan, 2000). Recent methodological advances in the analysis of $\delta^{18}O_p$ now provide the opportunity to use $\delta^{18}O_p$ as both a tracer of source and of the extent of metabolism of P in aquatic ecosystems (McLaughlin et al., 2004; Elsbury et al., 2009; Goldhammer et al., 2011b).

3. Isotope effects controlling $\delta^{18}O_p$ during biogeochemical cycling of P

A range of isotope effects associated with metabolic processes have the potential to influence $\delta^{18}O_p$. These effects are summarised in Fig. 1 and described in Sections 3.1 to 3.3.

3.1. Intracellular metabolism of P$_i$

A variety of P-containing compounds are present within aquatic ecosystems, including P$_i$, inorganic polyphosphates and organic compounds such as proteins, sugars and phospholipids (Karl and Tien, 1997). However, P$_i$ is the species preferentially utilised by organisms as it is capable of diffusion across cytoplasmic cell membranes (Björkman and Karl, 1994; Liang and Blake, 2006b). Consequently, P$_i$ is required for a number of intracellular reactions, including cellular signalling through phosphorylation and dephosphorylation, as well as being incorporated within biomass during cell growth (Blake et al., 1997, 2005). The major enzyme that catalyses intracellular P$_i$ cycling is inorganic pyrophosphatase, due to its ability to mediate reversible reactions such as the cleavage of adenosine-5′-triphosphate to adenosine-5′-diphosphate (Cohn, 1958; Blake et al., 2005; Fig. 2). Intracellular reactions catalysed by inorganic pyrophosphatase cause complete and very rapid (in the order of minutes) O isotope exchange between O atoms in P$_i$ and O atoms in surrounding water molecules that donate O atoms to newly produced P$_i$ molecules (Kolodny et al., 1983; Blake et al., 1997). Thermodynamic fractionation characterises the exchange of O between P$_i$ and the surrounding fluid, resulting in temperature-dependent equilibrium between O in P$_i$ and O within intracellular water ($\delta^{18}O_w$) expected to be identical to extracellular $\delta^{18}O_w$ (Fricke et al., 1998).

Release of P from biomass into the extracellular environment is driven by two mechanisms. Firstly, from live biomass as a by-product of uptake and intracellular metabolic reactions, in order to maintain non-toxic intracellular P concentrations (Cooperman et al., 1992). Secondly, following the death and lysis of cells (see Section 3.3). Within environments in which P$_i$ is extensively taken up and recycled through live biomass, an initially distinctive $\delta^{18}O_p$ composition in the extracellular environment, potentially reflecting sources of P, could be overprinted following intracellular, equilibrium fractionation and release of P$_i$ to the extracellular environment. The $\delta^{18}O$ value of a P$_i$ molecule in equilibrium with surrounding water can be predicted, based on the measured $\delta^{18}O_w$ and temperature of the surrounding water, using the empirically-derived Eq. (2) (Longinelli and Nuti, 1973). This is the basis for using historical $\delta^{18}O_p$ in solid-phase samples, such as tooth enamel, as a palaeothermometer, given an ecosystem in equilibrium via intracellular metabolism (Kolodny et al., 1983; Fricke et al., 1998).

$$
T(\degree C) = 111.4 - 4.3 \left( \delta^{18}O_p - \delta^{18}O_w \right)
$$

(2)

where T is the average growth temperature during shell formation for sampled organisms and $\delta^{18}O_p$ and $\delta^{18}O_w$ refer to the isotopic composition of P$_i$ and environmental water (at 25.2 °C) respectively. Whilst predominantly based on biogenic apatite from marine organisms, one freshwater species, Unio pictorum, was included in the data from which Eq. (2) was derived, suggesting no significant difference between seawater and freshwater equilibria equations. Although Eq. (2) has been used widely, more recent development of techniques for purification of P$_i$ from solid materials and for determination of $\delta^{18}O_p$ led Pucéat et al. (2010) to propose a revised phosphate–water fractionation equation based on empirical work (Eq. (3)):

$$
T(\degree C) = 118.7(\pm 4.9) - 4.3(\pm 0.20) \left( \delta^{18}O_p - \delta^{18}O_w \right)
$$

(3)

Based on empirical Eqs. (2) or (3), if both the ambient water temperature and $\delta^{18}O_p$ are determined then the expected equilibrium value for $\delta^{18}O_p$ can be calculated. Comparing the theoretical value for $\delta^{18}O_p$ at

Fig. 1. Schematic diagram describing the major isotope effects that can occur within the intracellular or extracellular environment due to metabolic processes. P$_{org}$ = organic P compounds; P$_i$ = dissolved inorganic phosphate ion; H$_2$O = water molecule; T = temperature.
equilibrium with $\delta^{18}O_p$ observed in an environmental sample can provide insight into the extent to which Pi has been recycled through intracellular metabolic reactions, assuming that $\delta^{18}O_p$ in the sources of P to an ecosystem was originally at disequilibrium.

3.2. Uptake of extracellular Pi

Many biologically-mediated processes associated with isotope effects are kinetically controlled, for example bacterial sulfate reduction and methanogenesis (Nakai and Jensen, 1964; Whiticar, 1999). During bacterial sulfate reduction, bacteria preferentially reduce $^{32}$S- and $^{16}$O-containing sulfate molecules, because these molecules have lower bond energies due to a lower combined mass. Thus, bacteria can metabolise isotopically lighter sulfate at a faster rate, resulting in the formation of isotopically light sulfide relative to the starting sulfate pool, assuming that only partial reduction of the available sulfate occurs (e.g. Habicht and Canfield, 1997). A similar kinetic fractionation may also be imparted on the Pi pool during biological uptake of Pi from the extracellular environment. Initial research reported by Blake et al. (2005) has demonstrated that *Escherichia coli* grown in controlled laboratory conditions preferentially take up $^{31}$P$^{16}$O$_4^-\text{P}$ compared to the isotopically-heavier $^{31}$P$^{18}$O$_4^-\text{P}$, resulting in enrichment of the extracellular environment with $^{31}$P$^{18}$O$_4^-\text{P}$. If this kinetic isotope effect operates more widely in natural ecosystems, the effect would only be observed

![Fig. 2. Reversible reaction of the dephosphorylation of adenosine-5'-triphosphate to adenosine-5'-diphosphate, in which Pi is generated as a by-product (Gajewski et al., 1986).](image)

![Fig. 3. Schematic diagram representing extracellular metabolism of Pi, in which enzymes are released to the extracellular environment beyond the cytoplasmic membrane to metabolise $P_{org}$ and release Pi, which is subsequently transported across the cell membrane. Reactions include: A: Hydrolysis of a 5'-nucleotide molecule through the action of 5'-nucleotidase; and B: Hydrolysis of glucose-6-phosphate through the action of glucose-6-phosphatase. Both reactions yield a Pi molecule as a by-product.](image)
given unidirectional reactions in which the reactants are not fully consumed and where competing fractionations do not over-print that associated with biological uptake of P.

3.3. Extracellular hydrolysis of organic P compounds

Organic P (Porg) compounds can be used to support metabolism given low bioavailable P concentrations within the extracellular environment. However, Porg compounds are too large for direct diffusion across cytoplasmic cell membranes (Liang and Blake, 2006b). Therefore, Porg must be hydrolysed through enzyme-mediated reactions involving phosphohydrolases that are either attached to the outside of a cell membrane or are secreted by organisms into the extracellular environment (Liang and Blake, 2006b). These extracellular metabolic reactions release P as a by-product (Fig. 3) that can be transported into a cell via diffusion or via ATP-driven pumps or transport proteins (Ammernman, 1991).

During enzyme-catalysed hydrolysis of Porg, P–O bonds are cleaved and O atoms within the original Porg compound are replaced with O atoms from surrounding water molecules in the newly created Pfl. The specific enzyme and associated hydrolysis pathway will influence the extent of O isotope exchange. For hydrolysis catalysed by a phosphomonoesterase, one O atom within the newly produced Pfl will be incorporated from the surrounding water-O and the remaining three O atoms are inherited from the Porg compound. With hydrolysis catalysed by a phosphodiesterase, two O atoms will be incorporated from the surrounding water-O (in a two-stage process involving incorporation of one O atom due to the action of the phosphodiesterase and the second O atom due to phosphomonoesterase-catalysed hydrolysis of the newly produced monoester), with the remaining two O atoms inherited from the Porg compound. Because extracellular hydrolysis of Porg does not result in complete exchange of all four O atoms in Pfl with O atoms in water, inheritance isotope effects occur in which $\delta^{18}O$P of the newly produced Pfl lies between that of the starting Porg compound and $\delta^{18}O_{\text{water}}$ (Blake et al., 1997; Colman et al., 2005). Incorporation of water-O within Pfl during extracellular hydrolysis of Porg is also accompanied by a kinetic fractionation, due to more rapid incorporation of the isotopically lighter $^{16}O$ atom compared to the $^{18}O$ atom from water molecules in the resulting Pfl (Blake et al., 2005). To date however, the magnitude of the fractionation factor associated with this process has only been quantified for a small number of enzyme–substrate combinations (e.g. Liang and Blake, 2006b).

In many aquatic environments, extracellular hydrolysis is significant in the regeneration of Pfl (Ammernman and Azam, 1985; Chrost, 1991; Colman et al., 2005; Goldhammer et al., 2011a,b). The fate of Pfl regenerated from Porg depends on the initial driver for Pfl regeneration from organic matter. Under conditions of Pfl limitation, regenerated Pfl is required to meet the metabolic demand for P among heterotrophic microorganisms. Alternatively, dephosphorylation of particulate or dissolved Porg may be required prior to uptake of C compounds to meet intracellular energy or C requirements among heterotrophic organisms (Colman et al., 2005; Goldhammer et al., 2011a,b). Because C rather than P demand drives dephosphorylation under these conditions, not all Pfl regenerated from Porg is necessarily taken up and recycled extensively through biomass. For example, it has been estimated that in coastal waters only 10–15% of Pfl produced through the action of secreted 5′-nucleotidase was taken up by microorganisms (Ammernman and Azam, 1985). The remaining 85–90% may enter the Pfl pool of the water column and affect the bulk $\delta^{18}O_{\text{water}}$ value of the extracellular environment. Effects on extracellular $\delta^{18}O_{\text{Pfl}}$ due to regeneration of Pfl from Porg that is driven by C-limitation have also been observed in deep-ocean water and in marine sediment porewater (Colman et al., 2005; Goldhammer et al., 2011a,b).

4. Analytical protocols for determining $\delta^{18}O_{Pfl}$

Historically, $\delta^{18}O_{P}$ has been determined through fluorination or bromination of a phosphate precipitate, traditionally bismuth(III) phosphate (BiPO4) although currently silver(I) phosphate (Ag2PO4) (Lécuyer, 2004). Fluorination (Eq. (4)) and bromination (Eq. (5)) halogenate the metal ion, reducing the bromine present and releasing gaseous O2 which can be quantitatively converted into CO2 for analysis (Vennemann et al., 2002; Lécuyer, 2004). However, fluorination (which can also be achieved using BrF5) and bromination reactions can suffer from the use of hazardous materials, poor O2 yields, time intensive preparation (precipitation of BiPO4 requires 6 days of extraction time relative to the 3 days of the Ag2PO4 precipitation method) and the requirement for large sample sizes in both methods (typically 4–5 mg Ag2PO4) (Lécuyer et al., 1993; Vennemann et al., 2002; Lécuyer, 2004; Grau et al., 2005).

$$\text{BiPO}_4 + \frac{8}{3} \text{BrF}_3 \rightarrow \text{BiF}_3 + \text{PF}_3 + \frac{4}{3} \text{Br}_2 + 2\text{O}_2$$ \hspace{1cm} (4)

$$2\text{Ag}_2\text{PO}_4 + \text{Br}_2 \rightarrow 2\text{Ag}_4\text{P}_2\text{O}_7 + \frac{1}{2}\text{O}_2 + 2\text{AgBr}$$ \hspace{1cm} (5)

Isotope ratio mass spectrometry (IRMS) allows for the precise measurement of small mass differences between different isotopes within a compound and, when coupled to a thermal conversion/elemental analyser (TC/EA), the system can carry out relatively rapid on-line measurements of isotope ratios (LaPorte et al., 2009). These systems have been automated, whilst retaining both precision and accuracy, for $\delta^{13}C$, $\delta^{15}N$ and $\delta^{18}O$ solid sample analysis. A further advantage of TC/EA-IRMS is the relatively small sample mass required for analysis (typically 400–500 μg Ag2PO4). To establish similar TC/EA-IRMS protocols for $\delta^{18}O$ measurements in Pfl, aqueous samples containing Pfl must be converted from dissolved to solid form. In this conversion, particular care must be taken to ensure that: i) the original isotope composition of Pfl is retained by using non-fractionating reactions or ensuring reactions go to completion; ii) Pfl is not introduced through hydrolysis within Porg-rich natural samples; and iii) other O-bearing compounds including dissolved organic carbon (DOC), nitrate (NO3$^-$), sulfate (SO4$^{2-}$) and calcium carbonate (CaCO3) are excluded from the final solid product.

Bismuth(III) phosphate (BiPO4) is a hygroscopic material, with full rehydration following dehydration possible after 15 min in air. The BiPO4 precipitate requires significant preparation before isotopic analysis to remove adsorbed water which can otherwise increase O yields by up to 12.5% in $\delta^{18}O$ analysis (Karhu and Epstein, 1986), with $\delta^{18}O$ enrichment possible when drying at temperatures greater than 200 °C (Mooney-Slater, 1962; Shemesh and Kolodny, 1988). Such constraints limit the utility of BiPO4, leading to consideration of alternative solid-phase compounds for analysis of $\delta^{18}O_{Pfl}$. Silver(I) phosphate (Ag2PO4) has been identified as a suitable alternative, due to its weakly hygroscopic nature, stability, low solubility and relatively short preparation time (Lécuyer, 2004; McLaughlin et al., 2004; Tamburini et al., 2010). Firsching (1961) was among the first to precipitate Ag2PO4 in order to gravimetrically determine the phosphate concentration of a homogeneous solution. Whilst silicates and multivalent ions were observed to interfere with Ag2PO4 precipitation, ions with lower valences did not significantly interfere (e.g. NO3$^-$, ammonium (NH4$^+$) and SO4$^{2-}$). However, Ag2PO4 has only more recently become a viable basis for analysis of $\delta^{18}O_{P}$ in aqueous environmental samples, following advances in extraction protocols that enable the precipitation of a sufficient mass of Ag2PO4 for analysis from complex matrices that often contain low concentrations of Pfl (Colman, 2002; McLaughlin et al., 2004).

4.1. Pfl extraction protocols for aqueous matrices

The major published protocols for extraction of Pfl via precipitation of Ag2PO4 are summarised in Fig. 4. None of the protocols in Fig. 4 currently represents a definitive approach for all aquatic matrices. Development
and evaluation of extraction protocols are required across the range of aquatic samples in which $\delta^{18}$Op research is undertaken, to ensure that accurate determination of $\delta^{18}$Op is achieved within these complex matrices.

There are commonalities across these protocols, particularly across procedures 1–4: i) concentration of P$_i$ through co-precipitation of P$_i$ with brucite following the magnesium-induced co-precipitation (MagIC) method of Karl and Tien (1992); ii) redissolution of the brucite precipitate in an acid matrix; iii) removal of other potential sources of O using anion exchange resins and/or sequential precipitations; iv) removal of cations that have the potential to interfere with Ag$_3$PO$_4$ precipitation, using a cation exchange resin; iv) precipitation of Ag$_3$PO$_4$. The method of Gruau et al. (2005) in Fig. 4 was developed for freshwaters and includes an initial dissolved organic matter (DOM) removal step using activated carbon. This protocol is based on the BiPO$_4$ method of Longinelli et al. (1976), with the benefit of a weakly hygroscopic end precipitate (Ag$_3$PO$_4$) relative to BiPO$_4$.

Final precipitation of Ag$_3$PO$_4$ is accomplished using either ‘slow’ or ‘fast’ approaches. Both precipitation methods involve the addition of silver nitrate (AgNO$_3$), ammonium hydroxide (NH$_4$OH) and ammonium nitrate (NH$_4$NO$_3$) to form a near-neutral pH solution. Whilst ‘slow’ precipitation is achieved by holding a solution at ~50 °C over several days to allow large Ag$_3$PO$_4$ crystals to grow, ‘fast’ precipitation can achieve full precipitation in a matter of minutes (Dettman et al., 2001; Tamburini et al., 2010). The ‘fast’ precipitation also has the added advantage of 100% Ag$_3$PO$_4$ yields independent of sample sizes, whereas lower recovery has been observed using the ‘slow’ precipitation, particularly for smaller P$_i$ masses (Dettman et al., 2001). However, the isotopic difference in Ag$_3$PO$_4$ generated by the two methods has been found to be within expected interlaboratory variation (Dettman et al., 2001).

Some research has applied the McLaughlin et al. (2004) protocol that was originally developed for marine systems to freshwater matrices without modification, whilst other research has adapted one of the major Ag$_3$PO$_4$ precipitation protocols to include clean-up stages for samples containing high concentrations of particulate organic matter and DOM. Particulate organic matter in environmental samples is typically removed through filtration at a maximum filter pore size of 0.45 μm (Elsbury et al., 2009; Li et al., 2011). In organic-rich sample matrices, including many freshwaters, inefficient removal of DOM could significantly influence measured $\delta^{18}$Op, because DOM can consist of up to 45% O by weight and has been shown to persist until the precipitation
of Ag₃PO₄ (Ma et al., 2001; Lécuyer, 2004; McLaughlin et al., 2004; Zohar et al., 2010). For accurate determination of ₁⁸O in Ag₃PO₄, all O-containing contaminants, especially Ag₃PO₄ precipitated from Ag₃PO₄, must be removed from the Ag₃PO₄ precipitate prior to TC/EA-IRMS. This remains a significant challenge for research seeking to apply ₁⁸O to complex freshwater matrices. Removal of DOM from samples has been attempted through adsorption of organic compounds to activated carbon (Gruau et al., 2005), washing (Liang and Blake, 2006a) or through the use of resins to specifically remove DOM (Tamburini et al., 2010), repetition of precipitation steps such as MgCl₂ in order to isolate P from a matrix of potential contaminants (Goldhammer et al., 2011b), and pH-specific precipitations to remove fulvic and/or humic acids (Zohar et al., 2010).

Beyond a source of contaminant O, DOM includes P-org compounds that may undergo acid hydrolysis during the protocols described in Fig. 4, for example during the dissolution of brucite. Hydrolysis of P-org that is co-precipitated with brucite can yield Pₗ that is subsequently washed from the Ag₃PO₄ precipitate with hydrogen peroxide to oxidise organic compounds (Tamburini et al., 2010; Zohar et al., 2010), using resins to specifically remove DOM (Tamburini et al., 2010), repetition of precipitation steps such as MgCl₂ in order to isolate P from a matrix of potential contaminants (Goldhammer et al., 2011b), and pH-specific precipitations to remove fulvic and/or humic acids (Zohar et al., 2010).

A major analytical challenge remains the lack of internationally certified Ag₃PO₄ standards, despite the long-term use of Ag₃PO₄ in the determination of ₁⁸O in archaeological and palaeoclimate studies (Stephan, 2000; Vennemann et al., 2002, Halas et al., 2011). Using materials other than Ag₃PO₄ as calibration standards, such as benzoic acids and barium sulfates (LaPorte et al., 2009; Halas et al., 2011), the precision for isotopic analysis is generally quoted as better than ±0.3‰VSMOW (1 standard deviation) (Lécuyer et al., 2007; LaPorte et al., 2009; Halas et al., 2011).

4.2. Determination of ₁⁸O through EA-IRMS

On-line EA-IRMS has been utilised for analysis of Ag₃PO₄. Dried samples (typically 400–500 μg) are introduced in silver capsules as the traditional and cheaper tin alternatives can lead to significant deterioration of chromatography results (Lécuyer et al., 2007). Conversion of Ag₃PO₄ to CO for analysis is achieved through pyrolysis at 1270 to 1450 °C in the presence of a carbon source to aid full conversion, typically nickelised graphite and/or glassy carbon (Vennemann et al., 2002; Lécuyer et al., 2007; Halas et al., 2011). Water vapour is removed through a water trap and CO is separated from other gaseous impurities through gas chromatography using purge-and-trap technology or a packed GC column (Meier-Augenstein, 1999; Fourel et al., 2011). Helium is used as a carrier gas to transfer the sample gas into a mass spectrometer via a continuous flow mode. Mass signals of 28 (¹²C¹⁶O) and 30 (¹²C¹⁸O;¹⁴C¹⁶O;¹³C¹⁷O) are integrated and compared to those in an independently introduced pulse of pure CO reference gas to calculate the ¹⁸O/¹⁶O ratio. Subsequently, these ratios are calibrated to the VSMOW scale in per mill notation (%VSMOW) using international reference materials, typically benzoic acids and barium sulfates (LaPorte et al., 2009; Halas et al., 2011).

A number of these uncertified compounds have been distributed to other laboratories (Excess c.Acetic Acid) when an 8-fold volume excess of acid was used. Mean values given, error bars show ±1 standard deviation (n = 3 for each acid system; Davies et al., 2014).
and initial inter-laboratory comparison undertaken (Lécuyer et al., 2007; Halas et al., 2011). However, finite availability and the lack of a quality control network in production represent significant challenges for broader use of these materials.

Linearity effects, referring to a change in the reported isotope ratio due to a change in sample mass introduced to the EA-IRMS and thus the peak size of the CO ion in the mass spectrometer (Brand, 2004), can also affect determination of δ18O. If this effect is consistent and can be quantified then the measured isotope ratios can be corrected (Brand, 2004). One alternative solution to address linearity effects is to match CO ion peak heights across a run for both samples and standards by using a narrow sample mass range. However this is made more complicated by variable Ag3PO4 pyrolysis efficiency and CO yield between samples, which can cause variations in peak heights. Difficulties have been found in ensuring complete conversion of Ag3PO4 to CO in the TC/EA, with Ag3PO4 requiring pyrolysis in the hot spot of a furnace (LaPorte et al., 2009). Whilst many compounds can achieve complete conversion easily (especially at temperatures exceeding 1400 °C), thereby ensuring accurate analysis of the bulk δ18O, incomplete conversion of other matrices, such as carbonates, has been implicated in isotope fractionation (Koziet, 1997; Kornexl et al., 1999). If conversion to CO is not complete then the reaction order of the O atoms in a compound becomes significant, with those being most strongly bound to the P atom being under-represented in the CO product. If this fractionation can be shown to affect all samples equally then the results can be properly calibrated. However, if the kinetic fractionation is not equivalent across all samples due to variable conversion to CO, then correction becomes problematic. This highlights the need to ensure the pyrolysis of Ag3PO4 is efficient and standardised across a sample run, which requires precise pyrolysis of samples within the hot spot of a furnace.

5. Synthesis of global δ18O data from aquatic ecosystems

Recent research has highlighted the potential to use δ18O as both a tracer of P sources and as a dynamic tracer of metabolic processes affecting P cycling in aquatic ecosystems. The characterisation of significant sources of P within an ecosystem can be achieved using δ18O, assuming:

i) δ18O for major sources of P is constrained;
ii) Individual sources of P possess distinct δ18O signatures;
iii) δ18O for P sources is not equal to the theoretical equilibrium;
iv) δ18O signatures for P sources are maintained and not rapidly over-printed by fractionation associated with metabolic processes.

Given these pre-requisites, if δ18O within an ecosystem tends towards the value of a particular source, inferences regarding the dominant P sources within ecosystems can be made. However, under conditions in which iv) above is not met and δ18O tends towards the theoretical equilibrium (Eqs. (2) or (3)), δ18O can then be used to probe the extent of intracellular metabolism of P. Insights into other metabolic processes, including biological P uptake and extracellular hydrolysis of Porg, may be gained through the kinetic and inheritance isotope effects associated with these processes. Table 1 synthesises published δ18O data currently available across a range of aquatic environments.

Table 1: Synthesis of global δ18O data derived from a range of aquatic ecosystems.

<table>
<thead>
<tr>
<th>Type</th>
<th>Setting</th>
<th>Min δ18O/‰</th>
<th>Max δ18O/‰</th>
<th>Av δ18O/‰</th>
<th>1σ Number of samples</th>
<th>Geographical locations</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Marine</td>
<td>Coastal surface waters</td>
<td>15.9</td>
<td>22.3</td>
<td>20.0</td>
<td>1.34</td>
<td>141 Monterey Bay, California; Long Island Sound, USA</td>
<td>McLaughlin et al. (2004); Colman et al. (2005); McLaughlin et al. (2006a)</td>
</tr>
<tr>
<td></td>
<td>Open ocean surface (~200 m depth)</td>
<td>13.8</td>
<td>23.7</td>
<td>18.5</td>
<td>3.27</td>
<td>17 Monterey Bay, California; Sargasso Sea</td>
<td>McLaughlin et al. (2004); McLaughlin et al. (2013)</td>
</tr>
<tr>
<td></td>
<td>Deep ocean (~200 m depth)</td>
<td>17.4</td>
<td>25.4</td>
<td>22.6</td>
<td>2.08</td>
<td>31 North Atlantic and Pacific Ocean tropical gyres; Sargasso Sea</td>
<td>McLaughlin et al. (2004); Colman et al. (2005); McLaughlin et al. (2013)</td>
</tr>
<tr>
<td>Estuarine</td>
<td></td>
<td>7.8</td>
<td>20.3</td>
<td>15.9</td>
<td>2.78</td>
<td>54 North San Francisco Bay California; Elk Slough, California;</td>
<td>McLaughlin et al. (2004); McLaughlin et al. (2006c); McLaughlin et al. (2006a)</td>
</tr>
<tr>
<td>Freshwater</td>
<td>River (main)</td>
<td>8.6</td>
<td>15.2</td>
<td>12.4</td>
<td>1.46</td>
<td>32 San Joaquin River and Lake Tahoe California; Lake Erie, USA</td>
<td>Elsbury et al. (2009); McLaughlin et al. (2006a); Young et al. (2009)</td>
</tr>
<tr>
<td></td>
<td>River (tributaries)</td>
<td>9.2</td>
<td>17.2</td>
<td>12.5</td>
<td>2.03</td>
<td>32 Elk Slough, San Joaquin River and Lake Tahoe California; Lake Erie, USA</td>
<td>McLaughlin et al. (2006a); Young et al. (2009)</td>
</tr>
<tr>
<td>Groundwater</td>
<td></td>
<td>15.1</td>
<td>22.4</td>
<td>18.6</td>
<td>2.13</td>
<td>9 Elk Slough and San Joaquin River, California</td>
<td>Blake et al. (2001); McLaughlin et al. (2006a); Young et al. (2009)</td>
</tr>
<tr>
<td>Lake (surface water – 1 m)</td>
<td></td>
<td>10</td>
<td>17.1</td>
<td>14.1</td>
<td>1.57</td>
<td>34 Lake Erie, USA</td>
<td>Elsbury et al. (2009)</td>
</tr>
<tr>
<td>Lake (5.5–22 m)</td>
<td></td>
<td>8.4</td>
<td>15.8</td>
<td>13.6</td>
<td>1.58</td>
<td>25 Lake Erie, USA</td>
<td>Elsbury et al. (2009)</td>
</tr>
<tr>
<td>Lake (58 m)</td>
<td></td>
<td>–</td>
<td>–</td>
<td>12.7</td>
<td>–</td>
<td>1 Lake Erie, USA</td>
<td>Elsbury et al. (2009)</td>
</tr>
<tr>
<td>WWTP final effluent</td>
<td></td>
<td>8.4</td>
<td>18.4</td>
<td>12.9</td>
<td>4.02</td>
<td>14 Brittany, France;</td>
<td>Gruau et al. (2005); Young et al. (2009); McLaughlin et al. (2006c)</td>
</tr>
<tr>
<td>Constructed wetlands (canals, inlet, interior and outlet)</td>
<td></td>
<td>20.0</td>
<td>25.5</td>
<td>21.5</td>
<td>1.32</td>
<td>25 Florida Everglades, USA</td>
<td>Li et al. (2011)</td>
</tr>
<tr>
<td>Marsh</td>
<td>Marine porewater</td>
<td>21.6</td>
<td>25.1</td>
<td>23.5</td>
<td>1.61</td>
<td>4 Florida Everglades, USA</td>
<td>Li et al. (2011)</td>
</tr>
<tr>
<td>Sediments</td>
<td>Marine porewater</td>
<td>12.8</td>
<td>26.6</td>
<td>22.2</td>
<td>2.59</td>
<td>119 Benguela Upwelling, South Atlantic</td>
<td>Goldhammer et al. (2011a)</td>
</tr>
</tbody>
</table>
ecosystems. Below, we consider the ways in which recent research has begun to explore the use of δ^{18}O, in aquatic ecosystems, highlighting the insights into P biogeochemistry that can be derived using this stable isotope technique.

5.1. Marine ecosystems

Marine upwelling zones, in which nutrient-rich bottom water is driven towards the surface, are areas of enhanced primary production and potentially rapid P turnover (Pennington and Chavez, 2000). Research in upwelling zones has reported δ^{18}Op values that approach equilibrium with water-O, indicating substantial intracellular metabolism of P in these coastal ecosystems (Colman et al., 2005; McLaughlin et al., 2006b). However, a lack of complete equilibrium between δ^{18}Op and water-O in upwelling zones suggests biological turnover of P, relative to inputs from terrestrial and deep-water sources can be lower in coastal upwelling zones compared to the open ocean (McLaughlin et al., 2006b).

In the surface water of the open ocean, P concentrations in the photic zone can be extremely low (e.g. 0.2–1 nM in the Sargasso Sea), often limiting or co-limiting primary production (Wu et al., 2000). Under these conditions, δ^{18}Op values that tend towards temperature-dependent equilibrium with water-O have been reported, indicating extensive cycling of P through biomass (Colman et al., 2005). Insights into temporal variations in P sources and the extent of biological turnover of P within marine ecosystems can also be gained through δ^{18}Op. For example, in Monterey Bay, California, McLaughlin et al. (2006b) observed δ^{18}Op values in the upper water column (<200 m) that were significantly influenced by source contributions from terrestrial freshwater and deep water (500–800 m). However, during the spring–summer periods of upwelling, δ^{18}Op in the upper water column was isotopically heavier relative to non-upwelling autumn/winter periods, and more closely reflected the δ^{18}Op value expected at theoretical equilibrium. The most likely explanation for this trend is enhanced intracellular cycling of P by phytoplankton relative to P, inputs during periods of upwelling. Upwelling periods provide nutrients to the surface zone that, combined with a greater intensity of sunlight in the spring–summer months, can enhance productivity and consequently biological turnover of P (McLaughlin et al., 2006b). Extensive extracellular–intracellular–extracellular cycling of P during the upwelling periods would lead to an over-printing of source δ^{18}Op signatures and a shift of δ^{18}Op, in the extracellular environment towards equilibrium values. However, the data reported by McLaughlin et al. (2006b) suggest that the extent of extracellular–intracellular–extracellular was still not sufficient, even in the spring–summer period, to establish complete equilibrium between δ^{18}Op and water-O.

The preferential remineralisation of P over C from dissolved organic compounds has been observed in marine ecosystems, particularly in the euphotic zone (Clark et al., 1998; Aminot and Kerouel, 2004). This selective remineralisation of Porg is caused by the presence of extracellular enzymes and their action on Porg substrates, meaning that the turnover rate of Porg molecules in this environment is dependent on the specific enzymes and Porg compounds present. For example, it has been shown that the flux of dissolved ATP through an oligotrophic ocean ecosystem can be up to fivefold greater than for the bulk Porg pool (Bjorkman and Karl, 2005). McLaughlin et al. (2013), in research conducted in the Sargasso Sea, used δ^{18}Op to investigate the role of Porg hydrolysis in P-cycling in marine ecosystems. Based on the fractionation imparted on δ^{18}Op, when Porg is regenerated from a specific Porg molecule following a specific hydrolysis pathway, McLaughlin et al. used Eq. (6) to estimate the fraction of Porg in samples that had been regenerated from Porg:

$$ \%P_{\text{Porg remineralisation}} = \frac{\delta^{18}O_{\text{sample}} - \delta^{18}O_{\text{org}}}{\delta^{18}O_{\text{org rem}} - \delta^{18}O_{\text{org}}} \times 100\% $$

(6)

where δ^{18}O_{rem} is the calculated value of δ^{18}Op based on a particular combination of P substrate and hydrolysis pathway; δ^{18}O_{org} is the expected equilibrium value of δ^{18}Op and δ^{18}O_{sample} is the measured δ^{18}Op in a sample. McLaughlin et al. concluded that 5–80% of Porg present in the upper 200 m of an oligotrophic marine ecosystem was the product of Porg remineralisation, dependent on which enzyme/substrate system was used in Eq. (6). These data, combined with expressed enzyme activity and low P concentration, suggest that Porg regeneration from Porg was coupled with uptake by organisms to support metabolism. The proportion of Porg thought to derive from Porg remineralisation was significantly reduced (12–35%) at a monitoring station closest to the shoreline, suggesting increased P availability in this region and thus reduced requirement for Porg mineralisation. Despite the limited number of samples and large contrasts between differing enzyme/substrate systems, this research highlights the potential for using δ^{18}Op to gain insights into P cycling within the marine ecosystem, without the need to directly determine the concentrations of or fluxes affecting Porg and Porg pools.

Colman et al. (2005) also demonstrated that δ^{18}Op was close to or at theoretical equilibrium in deep water samples from both the Atlantic (900–4000 m) and Pacific (300–3000 m) oceans. These data are consistent with intracellular equilibration between δ^{18}Op and water-O in surface waters, followed by advection of Porg to depth. However, small offsets (−1.5‰ vs. SMOW) from the expected theoretical equilibrium in Porg were observed from deep-ocean samples collected below the thermocline were used by Colman et al. to infer that significant Porg mineralisation occurred in this zone, imparting a disequilibrium fractionation on δ^{18}Op. The disequilibrium effects were not fully overprinted by equilibrium fractionations, supporting estimates that 80–95% of the remineralised Porg was not utilised by the surrounding microorganisms in these deep-ocean waters to meet their P requirements, because metabolism in these zones is hypothesised to be limited by C and energy rather than by P. However, the size of the δ^{18}Op offset from equilibrium was smaller than that expected on the basis of Porg regeneration from Porg through the action of phosphohydrolases, suggesting partial biological turnover of Porg in these deep ocean environments (Colman et al., 2005).

5.2. Estuarine ecosystems

Source δ^{18}Op signatures have been shown to be retained within some estuaries (e.g. McLaughlin et al., 2006a). Source signals representing inputs derived from surrounding land (e.g. chemical fertilisers associated with runoff from agricultural land) and from other aquatic ecosystems (e.g. groundwater and surface water tributaries) are thought to dominate the bulk δ^{18}Op composition in many estuarine ecosystems, due to the short residence time of P compounds in tidal estuaries (McLaughlin et al., 2006a). The limited contact time with biomass in these ecosystems constrains the opportunity for metabolism of Porg, increasing the potential for source δ^{18}Op signatures to be retained. McLaughlin et al. (2006a) demonstrated that towards the mouth of Elkhorn Slough in California, δ^{18}Op was dominated by that of oceanic-derived Porg (~20‰ vs. SMOW) due to tidal flushing of the ecosystem in this area. Similar δ^{18}Op values were also found upstream in the estuary. However, these were attributed to chemical fertilisers, entering the estuarine ecosystem through runoff from agricultural fields, that have an isotopic composition similar to that of the marine ore from which the fertilisers were formed. One mid-estuary site had a significantly lower δ^{18}Op composition. This observation, combined with the use of other natural tracers, suggested that groundwater discharge significantly affected bulk δ^{18}Op at this sampling location. δ^{18}Op can also provide insights into the presence of temporal and spatial patterns in Porg sources to estuaries. For example, it was observed that δ^{18}Op composition throughout the main estuary channel was not affected by either tidal or seasonal variations (McLaughlin et al., 2006a). However, a nearby harbour, close to the mouth of the estuary, did show evidence of tidal patterns through δ^{18}Op with high tides associated with lower δ^{18}Op values higher and closer to that of oceanic-derived Porg and low tides associated with lower δ^{18}Op values towards those observed upstream in the estuary.
5.3. Sediments within aquatic ecosystems

δ18O has also been used as a tracer of particulate inorganic P sources and sinks within aquatic ecosystems (Markel et al., 1994; Jaisi and Blake, 2010). The first major lacustrine site for this application was Lake Kinneret, situated in the Dead Sea Rift Valley (Markel et al., 1994). Markel et al. (1994) showed that δ18O, in sediment-bound P, varied significantly with sediment grain size, with clay-bound P demonstrating significant enrichment of δ18O compared to silt or sand-bound P. These δ18O signatures enabled an internal P-cycling model of Lake Kinneret to be developed, which included major processes such as the sedimentation of both detrital and authigenic apatite and CCP (Ca₅(PO₄)₃(OH), as well as the dissolution of these materials. In conjunction with other internal P-cycling studies of Lake Kinneret, δ18O was used as a tool to elucidate processes such as the release of P through dissolution of authigenic apatite and to support the quantitative analyses of sediment–lake interactions found in other studies. Further work has also shown that δ18O may act as a tracer of P source and as a paleotemperature proxy (particularly in detrital and authigenic P phases) in other ecosystems, such as the sediments of continental margins (Jaisi and Blake, 2010).

More recent methodological advances that enable accurate measurements of δ18O in samples containing a small mass of P have underpinned studies of δ18O within the porewaters of deep marine sediments (Goldhammer et al., 2011a,b). These studies demonstrate that δ18O in extracted porewater, P concentration profiles and two-endmember isotope mixing models can be used to determine whether marine sediments are dominated by P release (regeneration of P2 during microbial respiration of organic matter and the desorption of P from mineral phases), P burial from sources in the overlying water column, or P movement through advection or diffusion within the sediment column (Goldhammer et al., 2011a,b). In particular, it was shown that marine sediment porewaters demonstrate significant disequilibrium in δ18O, reflecting the dominance of disequilibrium effects associated with P2 regeneration from organic matter (Goldhammer et al., 2011a).

However, it was noted that simple isotope mass balance models can overlook processes which could be significant within these environments and thus improvements need to be made for future analyses, including the construction of more complex and realistic isotope models that allow for the incorporation of physical mixing processes, such as diffusion and advection.

6. Freshwater ecosystems

Beyond the relatively intensive study of Lake Erie reported by Elsberry et al. (2009), the catalogue of δ18O data for freshwater ecosystems remains more constrained than for marine and terrestrial ecosystems. However, a limited number of initial studies have explored the use of δ18O to assess sources and metabolism of P in freshwater ecosystems.

6.1. Application of δ18O to identify sources of P in freshwater ecosystems

A range of potential sources of P to freshwater ecosystems may be distinguished on the basis of δ18O, as long as the source isotope signature is not rapidly over-printed by isotopic fractionations associated with metabolism (e.g. Young et al., 2009; Li et al., 2011). However, this emerging dataset also shows potential for significant overlap in δ18O between a number of individual sources. For example, whilst δ18O may be used to distinguish P derived from fertilisers compared to WWTP effluents, no significant differences were found between δ18O from other potential sources, such as vegetation and detergents (Young et al., 2009). Table 2 synthesizes currently available data regarding δ18O in potential sources of P to freshwaters, with average δ18O in these sources ranging from 13.2 to 21.5‰. However, note that for many of these sources, the size of the available dataset is currently small and geographically constrained.

Gruau et al. (2005) compared δ18O across three commonly used inorganic P fertilisers and effluent samples from three WWTPs in France. The origin of the phosphatic rock used in the production of the fertilisers was found to be significant in the resulting fertiliser δ18O (19.6–23.1‰VSMOW), which was consistent with the typical ranges for the two largest phosphatic rock suppliers to France–Morocco (18.5‰VSMOW to 20.5‰VSMOW) and Florida (17.2‰VSMOW to 23.2‰VSMOW) respectively. Laboratory dissolution experiments demonstrated no significant fractionation of δ18O, suggesting that fertiliser δ18O could be preserved in runoff from agricultural land (Gruau et al., 2005). Phosphate in WWTP effluents from the study by Gruau et al. was assumed to originate from two main sources: human waste, including faeces, urine and waste food disposal (30–50%) and phosphate-based detergents (50–70%). For these WWTP effluents, δ18O values fell in the narrow range of 16.6 to 18.1‰VSMOW, whilst δ18O for a phosphate builder used in detergents fell within the same range (17.9‰VSMOW), indicating possible maintenance of source δ18O composition in the final WWTP effluent. However, the δ18O composition of WWTP effluents could also reflect microbial metabolism of Porg, as reported values also fall in the δ18O range expected following fractionation due to the degradation of Porg associated with microbial metabolism (Blake et al., 1997). The difference between δ18O ranges for fertiliser- and WWTP-derived phosphate (<2‰VSMOW) reported by Gruau et al. was statistically significant. However, given the relatively small absolute difference, and the variability in δ18O of the two sources (ranges for both sources were ≥1.5‰VSMOW), these authors concluded that δ18O may not be suitable for determining the sources of anthropogenic P.

However, the δ18O of other potentially significant sources of P to freshwaters has not been thoroughly characterised, including soil leachate, waste from aquaculture and, in particular, groundwater and septic tank discharge. Where sources such as inorganic fertiliser or WWTP effluent have begun to be characterised, available data tend to focus on specific geographical regions such as California (USA) (Young et al., 2009), the Great Lakes (USA) (Elsberry et al., 2009; Young et al., 2009).

### Table 2

<table>
<thead>
<tr>
<th>Source</th>
<th>Min δ18O (‰)</th>
<th>Max δ18O (‰)</th>
<th>Nδ18O (‰)</th>
<th>1σ</th>
<th>Number of samples</th>
<th>Geographical location</th>
</tr>
</thead>
<tbody>
<tr>
<td>Chemical fertilisers</td>
<td>15.5</td>
<td>25.3</td>
<td>22.1</td>
<td>2.32</td>
<td>33</td>
<td>–</td>
</tr>
<tr>
<td>Fertiliser ore and processing</td>
<td>18.2</td>
<td>21.6</td>
<td>20</td>
<td>1.43</td>
<td>5</td>
<td>Israel</td>
</tr>
<tr>
<td>Aerosols and dust</td>
<td>14.2</td>
<td>24.9</td>
<td>20.1</td>
<td>2.17</td>
<td>17</td>
<td>Israel</td>
</tr>
<tr>
<td>WWTP final effluent and processing</td>
<td>8.4</td>
<td>18.4</td>
<td>13.5</td>
<td>3.5</td>
<td>18</td>
<td>USA; France</td>
</tr>
<tr>
<td>Detergent</td>
<td>13.3</td>
<td>18.6</td>
<td>16.8</td>
<td>1.83</td>
<td>7</td>
<td></td>
</tr>
<tr>
<td>Toothpaste</td>
<td>–</td>
<td>–</td>
<td>17.7</td>
<td>–</td>
<td>1</td>
<td>–</td>
</tr>
<tr>
<td>Animal faeces</td>
<td>15.7</td>
<td>23.1</td>
<td>20</td>
<td>1.82</td>
<td>11</td>
<td>South Pacific; California, USA</td>
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<tr>
<td>Soil leachate</td>
<td>17.9</td>
<td>19.1</td>
<td>18.5</td>
<td>0.474</td>
<td>5</td>
<td>Israel</td>
</tr>
<tr>
<td>Compost</td>
<td>23.3</td>
<td>27</td>
<td>25.2</td>
<td>2.62</td>
<td>2</td>
<td>California, USA</td>
</tr>
<tr>
<td>Vegetation leachate (Dead, Live and Decayed)</td>
<td>14.2</td>
<td>23.1</td>
<td>16.8</td>
<td>2</td>
<td>27</td>
<td>California, USA</td>
</tr>
</tbody>
</table>

1 Water soluble P fraction.
Evidence of metabolism of P in freshwater ecosystems from $\delta^{18}O_p$ analysis

Evidence of temperature-dependent equilibrium fractionation between $\delta^{18}O_p$ and $\delta^{18}O_w$ has also been used to infer full or partial metabolism of P in some freshwater ecosystems (Blake et al., 2001; Elsbury et al., 2009). For example, Blake et al. (2001) analysed $\delta^{18}O_p$ within groundwater samples from a shallow P-rich aquifer in Cape Cod, Massachusetts, USA. Significant inputs of P to groundwater resulted from sewage contamination, causing P concentrations to rise to 30–108 µmol/L. Full equilibration between $\delta^{18}O_p$ and $\delta^{18}O_w$ in the surrounding groundwater was not achieved, probably due to low DOC concentrations meaning that elevated P concentrations were above metabolic requirements. However, a strong positive correlation was found between $\delta^{18}O_p$ and $\delta^{18}O_w$, suggesting partial metabolism of P and that $\delta^{18}O_p$ could be used to provide insights into metabolism of P in groundwater.

The first major study of the application of $\delta^{18}O_p$ in the water column of a lacustrine setting was conducted in Lake Erie, USA and emphasises the potential to use $\delta^{18}O_p$ within freshwater P cycling studies to identify trends that cannot be detected through analysis of concentration alone (Elsbury et al., 2009). Despite strict regulations on point sources of P since the 1970s and no observed increase in inflow to the lake, P concentration in the Central Basin of Lake Erie has increased since 1990 (Elsbury et al., 2009). As changes in inflow P fluxes could not explain the observed increase in P concentration, analysis of $\delta^{18}O_p$ was initially used in an attempt to identify additional sources of P.

Weighted average riverine $\delta^{18}O_p$ feeding the lake was considerably depleted in $\delta^{18}O_p$ compared to that of the theoretical equilibrium $\delta^{18}O_{water}$. Whereas the $\delta^{18}O_p$ values observed in Lake Erie were generally enriched in $\delta^{18}O_p$ compared to those of the seven sampled tributaries, particularly in the Central Basin. Any samples with $\delta^{18}O_p$ between that of the theoretical equilibrium and the weighted riverine signal (+11‰ VSMOW) could potentially be attributed to incomplete equilibration of the O within riverine phosphate with water-O in the lake though metabolism of P. Consequently, Elsbury et al. (2009) concluded that samples with $\delta^{18}O_p$ values outside of the potential mixing region must be derived from an additional (unconstrained) source of P. Potential sources included smaller tributaries that were not sampled in the study; however this was deemed improbable given the large P fluxes required to achieve the observed concentration increase in the Central Basin. Alternatively, disequilibrium isotopic effects could be operating in the lake, leading to an increase of $\delta^{18}O_p$ in lake P though processes associated with a kinetic fractionation (Blake et al., 2005). However, the most likely explanation for the observed increase in P concentration was concluded to be end-member mixing of riverine P with an isotopically enriched source(s) of P (+17‰ VSMOW), which was suggested by Elsbury et al. to be associated with remineralisation of $\delta^{18}O$-enriched $P_{org}$ within the bed sediments, followed by release of P from the lake sediments to the water column.

7. Concluding remarks and priorities for future research

The use of $\delta^{18}O_p$ in research examining P cycling in natural ecosystems is at an embryonic stage, particularly with respect to freshwater ecosystems. This stable isotope tracer has the potential to offer new insights into both the relative importance of different sources of P to ecosystems, and the extent to which P from individual sources is linked to metabolic activity within ecosystems. These insights would have important implications for understanding the reaction mechanisms controlling P biogeochemistry in nature, and for the design and targeting of future policies and practices to deliver more sustainable stewardship of P. Below, we conclude by identifying four key areas for development of $\delta^{18}O_p$ research that would enable the potential of this isotope tracer to be more fully realised:

- Develop robust analytical protocols for $\delta^{18}O_p$ in freshwater matrices. The primary challenge here is to avoid errors in the determination of $\delta^{18}O_p$...
due to incorporation of contaminant O in a precipitate that is subsequently pyrolysed. In freshwater matrices this involves excluding O derived from DOC compounds and from oxygenates other than P, from the final precipitate. However, it also requires removal of Porg compounds from an aquatic matrix at the appropriate stage of an analytical protocol to ensure that hydrolysis of Porg and production of contaminant P is avoided. Finally, despite recent inter-laboratory comparison of the isotope composition of various sources of Ag5P04, the need for an internationally certified Ag5P04 standard has yet to be addressed.

- Expand the global library of δ18O within sources of P to freshwaters. Proper characterisation of δ18O in P sources provides the basis for using this stable isotope tracer as a source apportionment tool. Initial data suggests that distinguishing certain sources on the basis of δ18O may be possible, whilst the isotopic composition of other sources of P may overlap. However, the global database of δ18O in sources of P remains limited. The priorities are: i) to characterise sources that are not currently included in the global database, for example P within potable water supplies and within septic tank discharges; ii) to extend the geographical extent to which individual sources are characterised, for example covering regions in which inorganic P fertiliser is derived from differing rock P reserves, or regions in which differing δ18O may vary. These may include rivers and estuaries, within which δ18O may vary significantly in the fluvial effluent of WWTPs and in the estuarine plume. Further research may also focus on the temporal variation in δ18O within sources of P, for example associated with seasonal or diurnal cycles in the fluvial effluent from WWTPs.iii) to constrain the temporal variation in δ18O within key sources of P, for example associated with seasonal or diurnal cycles in the fluvial effluent from WWTPs.

- Focus on the potential of δ18O to provide insight into biogeochemical processes controlling P cycling. δ18O will not act as a conservative tracer for the source of P where metabolic reactions significantly influence the P cycle within an ecosystem. Whilst clearly a constraint on source apportionment studies, the isotope fractionations that occur alongside metabolic processes provide the opportunity to use δ18O to explore the reaction mechanisms controlling P cycling. Specifically, the balance between equilibrium and disequilibrium fractionations can provide insight into the relative importance of intracellular metabolism of P versus the extracellular regeneration of P from Porg. This requires further research to determine and interpret variations in δ18Op both through time (for example, from inter-annual or seasonal cycles to event-driven variations within lake or river ecosystems) and through space (for example, tracking downstream changes in δ18Op associated with point or diffuse inputs of P to aquatic ecosystems). Further research could also focus on analysis of δ18O within the phosphate moieties of Porg to better understand the cycling of Porg within aquatic ecosystems.

- Couple δ18O with parallel techniques to develop an integrated isotope framework across C, N and P cycles. Whilst analysis of δ18O provides a tool that can be used as a research tool in aquatic ecosystems, parallel stable isotope fractionations have a longer history of application for N (δ15N/14N in ammonium and δ15N/14N in nitrate) and for C (δ13C/12C). Coupling analysis of δ18O with δ13C of organic matter can potentially provide new insights into process interactions between C, N and P biogeochemistry. A coupled stable isotope framework could draw on natural abundance approaches or, ultimately, on isotopic labelling studies based on production of labelled δ18O to complement isotopically labelled sources of N and C. Finally, by coupling stable isotope and radiocarbon techniques, processes that govern the cycling of P within aquatic ecosystems across both the short and long term could be more readily constrained.

Acknowledgements

This work was supported by a Natural Environment Research Council CASE studentship (NE/K501801/1) and by funding from the British Geological Survey to CD. DCG publishes with the permission of the Executive Director, British Geological Survey (NERC). We are grateful to two anonymous reviewers for their comments that helped to significantly improve the manuscript.

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