

1 **Migratory carryover effects and endocrinological correlates of reproductive decisions and**
2 **reproductive success in female albatrosses.**

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25 **Abstract**

26 Physiological mechanisms mediating carryover effects, wherein events or activities occurring in
27 one season, habitat, or life-history stage affect important processes in subsequent life-history
28 stages, are largely unknown. The mechanism most commonly invoked to explain carryover
29 effects from migration centres on the acquisition and utilization of resources (e.g. body mass, or
30 individual ‘condition’). However, other mechanisms are plausible, e.g. trade-offs reflecting
31 conflict or incompatibility between physiological regulatory systems required for different
32 activities or life-history stages (e.g. migration vs. reproduction). Here we show that in female
33 black-browed albatrosses (*Thalassarche melanophris*) the decision to reproduce or to defer
34 reproduction, made prior to their arrival at breeding colonies after long-distance migration, is
35 associated with condition related (body mass, hematocrit, hemoglobin concentrations) and
36 hormonal (progesterone, testosterone, estrogen-dependent yolk precursors) traits. In contrast,
37 reproductive success showed little association with condition traits but showed significant
38 associations with the steroidogenic processes underlying follicle development. Specifically,
39 success was determined by reproductive readiness via differences in steroid hormones and
40 hormone-dependent traits. Successful albatrosses were characterised by high progesterone and
41 high estradiol-dependent yolk precursor levels, whereas failed albatrosses had high testosterone
42 and low yolk precursor levels. Results are discussed with reference to migratory carryover
43 effects and how these can differentially affect the physiologies influencing reproductive decisions
44 and reproductive success.

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47 **Keywords:** Trade-offs, vitellogenin, physiological conflict, yolk precursors, reproductive
48 hormones, seabirds.

49 **1. Introduction**

50 An ever-increasing number of studies are documenting the existence of carryover effects in
51 animal populations, wherein events or activities occurring in one season, habitat, or life-history
52 stage affect important processes at subsequent stages (see reviews by [16,25]). The majority of
53 studies examining carryover effects have examined the influence of over-wintering and migratory
54 experiences on aspects of reproduction including breeding decisions, timing of reproduction,
55 reproductive output, and in some cases reproductive success [16]. However, despite the growing
56 number of studies examining these phenomena, our understanding of the physiological and
57 hormonal mechanisms driving carryover effects remains rudimentary.

58 In migratory animals, the mechanism most commonly invoked to explain carryover
59 effects on reproduction centres on the acquisition and utilization of resources, i.e. differences in
60 individual condition. Under such a model, the events and activities occurring at the transition
61 from the migratory non-breeding period in winter to spring breeding can “carry over” to affect
62 patterns of resource acquisition and utilization which directly determine reproductive success
63 [2,16,20,23,24,28]. Some studies show links between individual condition at arrival and aspects
64 of reproduction (laying date and egg size), but individual variation in these traits tends to be high
65 and their direct effects on reproductive success are currently largely speculative (but see [11]).
66 As an alternative hypothesis, there is growing evidence that endocrine processes themselves
67 might mediate carryover effects to influence reproductive decisions. For example, Goutte et al.
68 [13] showed that in an annually-breeding seabird, elevated baseline corticosterone levels during
69 the pre-laying period after migration were associated with a higher probability of deferred
70 breeding, and speculated that elevated corticosterone might inhibit luteinizing hormone (LH) and
71 the downstream secretion of sex-steroids necessary for reproduction. Other studies have shown
72 that non-breeding or deferring birds are fully capable of elevating LH, when presented with a

73 luteinizing hormone releasing hormone (LHRH) challenge, but they do not or cannot sustain
74 gonadal steroid production [5,14,15]. These studies suggest that the metabolic demands of
75 migration might down-regulate the reproductive axis in terms of gonadal steroid production, an
76 effect that might be mediated by, or simply correlated with, elevated corticosterone levels and/or
77 inhibition of LH. Few studies have considered individual variation in circulating gonadal
78 steroids at arrival in migratory birds in relation to subsequent reproduction. Furthermore, the
79 studies cited above only considered breeding decisions as breeding/not breeding (all-or-none
80 decisions), and did not extend the analysis of hormonal patterns at arrival to consider the
81 consequence of the decision to breed in terms of success or failure. Recently, Crossin et al. [8]
82 showed that migratory carryover effects can place constraints on the estrogen-dependent
83 production of yolk precursors by egg-producing females, with negative effects on egg sizes, and
84 that these effects are unrelated to variation in individual condition. However, the functional
85 significance of reduced or constrained sex-steroid production, during the migration-reproduction
86 transition, to reproductive success remains largely unknown. Nevertheless, what is emerging is
87 the recognition that condition (or resource state) at arrival need not be the only mechanism
88 affecting reproductive success. The endocrine processes controlling female reproduction may be
89 directly affected by migratory carryover effects, via constraints on, or incompatibilities between,
90 competing physiological regulatory systems [8,17,37].

91 Procellariiforme seabirds (the albatrosses and petrels) are pelagic species which also make
92 good models for exploring the effects of migratory activity on the hormonal processes underlying
93 reproduction. Like many seabirds, the Procellariiformes have a slow life-history characterized by
94 delayed age of reproduction, a single-egg clutch, prolonged parental care, and longevity, and they
95 make very long, pelagic migrations during a non-breeding period which lasts 6-16 months in
96 *Thalassarche spp.* [9,26]. Among breeding-age albatrosses and petrels, there is typically a high

97 proportion of non-breeding individuals, and among even the most experienced breeders, breeding
98 deferral for one or more years is a common tactic underlying lifetime fitness [27,36]. The annual
99 breeding pattern of black-browed albatrosses (*Thalassarche melanophris*) make this species a
100 particularly tractable model for the study of carryover effects. Each year, black-brows return to
101 breeding colonies from distant foraging areas occupied during the non-breeding period [26], and
102 in the weeks preceding arrival, birds are presumably balancing their own physiological
103 requirements against those needed for the initiation of reproduction. Upon arrival at breeding
104 colonies, they then experience one of three inevitable outcomes: they will defer reproduction
105 entirely until the next season, or they will lay and then either fail or succeed at fledging a chick
106 [26].

107 In this study, we investigate the physiological mechanisms that mediate reproductive
108 decisions (breeding or deferral) and reproductive outcomes (success or failure) in female black-
109 browed albatrosses during the transition between winter migration and spring breeding.
110 Specifically we consider condition or resource related traits (mass, hematology), as well as the
111 steroidogenic hormones and estrogen-dependent yolk precursor pathways underlying
112 reproductive readiness (*sensu* [8]), in mediating decisions and outcomes. We predicted that the
113 decision to lay or to defer would be condition-dependent, and that females deferring reproduction
114 would be in poorer condition (low body mass, low hematocrit [Hct] and hemoglobin [Hb]
115 concentrations) relative to breeding females. However, we also predicted that reproductive
116 success (e.g. fledging of chicks) might reflect differences in “reproductive readiness” upon
117 arrival at the colony, as indicated by the estrogen-dependent production of the yolk precursors
118 vitellogenin [VTG] and yolk-targeted very low density lipoprotein [VLDLy], as well as by
119 patterns of production of the reproductive steroid hormones progesterone [P4] and testosterone
120 [T].

121

122 **2. Materials and methods**

123 *2.1. Study site and field sampling protocol*

124 Fieldwork was conducted during the austral summer beginning in September 2008 at a large
125 long-term demographic study-colony of black-browed albatrosses (Colony J) breeding on Bird
126 Island, South Georgia (54°01'S, 38°02'W). Research was conducted through permits issued by
127 the British Antarctic Survey, and conformed to guidelines established by the Canadian Council
128 on Animal Care (Simon Fraser University Animal Care Permit # 897B-8).

129 Beginning in late September when black-browed albatrosses historically return from sea,
130 we made daily visits to the colony in order to identify newly arrived females with previous
131 breeding experience. All females had first bred at least six years previously, and were
132 identifiable by unique leg rings. We then captured the newly arrived, previously unsighted
133 females (N=33) at nest sites and collected 2 ml blood samples from tarsal veins using heparinized
134 syringes fitted with 25G needles. We recorded the time (± 1 sec) from initial approach until the
135 end of blood sampling for each bird. Blood was then transferred to heparinized 2.5 ml Eppendorf
136 vials, centrifuged for 5 min at 10,000 g, and the isolated plasma was transferred to labelled 0.6 ml
137 vials for storage at -20 °C. We recorded body mass (± 10 g), and measured bill length and tarsus
138 length (both ± 1 mm). After sampling, we continued daily (later reduced to weekly) visits to
139 monitor breeding fate, noting dates of laying, hatching, failure (loss of an egg or chick), and
140 fledging.

141

142 *2.2. Blood and plasma analysis*

143 Blood samples were assayed in duplicate for hematocrit and hemoglobin concentrations.
144 Hematocrit (Hct) was measured on fresh whole blood at the time of blood sampling as reported

145 as packed cell volume (%) following centrifugation of whole blood in microhematocrit tubes for
146 5 min at 10,000 g. Hemoglobin (Hb, g dl⁻¹ whole blood) was measured with the
147 cyanomethemoglobin method modified for use with a microplate spectrophotometer, using 5µl
148 whole blood diluted in 1.25 ml Drabkin's reagent (D5941 Sigma-Aldrich Canada, Oakville,
149 Ontario, Canada). Absorbance was measured at 540 nm.

150 Progesterone (P4) and testosterone (T) were assayed in duplicate from each of 33 samples
151 by liquid chromatography-tandem mass spectrometry (LC-MS/MS). Both steroids were assayed
152 in a single injection, starting from a sample volume between 50 and 100 µl. All samples received
153 a deuterated internal standard representing a final concentration of 5 ng ml⁻¹ P4-d9 and 1 ng ml⁻¹
154 T5-d2, and were diluted to 500µl with water. Sample preparation consisted of solid phase
155 extraction over C18, with elution in 1 ml of ethyl acetate. Samples were dried under nitrogen gas
156 and reconstituted in 50% MeOH. Liquid chromatography (Agilent 1200 SL system) used an
157 injection volume of 40 µl, a 100x300 mm Kinetex C18 Column (Phenomenex), and
158 water/methanol as mobile phases. Mass spectrometry (AB Sciex Q-trap 5500) used APCI +ve
159 mode, with the following MRM transitions: Progesterone 315/97; Progesterone-d9 324/100;
160 Testosterone 289/97; Testosterone-d2 291/99.

161 Quantification of E₂ was not possible at these volumes as sample preparation and optimal
162 LC-MS conditions at sample concentrations (based on pilot analysis of a pooled sample) differed
163 from the conditions for testosterone and progesterone. However, the principal role of E₂
164 secretion is the production of the yolk precursors vitellogenin (VTG) and yolk-targeted very low
165 density lipoprotein (VLDL_y), which are a strong indicator of reproductive readiness [8 and
166 references therein].

167 Plasma samples were assayed in duplicate for vitellogenic zinc (Zn; zinc kit, Wako
168 Chemicals) and total triglycerides (Glycerol Reagents A and B, Sigma), as indices of the yolk

169 precursors VTG and VLDL_y respectively (e.g. [8]). VTG and VLDL_y are the two main yolk
170 precursors in birds and are transported from the liver to the ovary where they are deposited into
171 developing follicles. Of the two precursors, VTG is generally considered the more reliable
172 indicator of follicle development in birds [8]. All assays were measured using a Biotek 340i
173 microplate reader. Using a 19-week domestic laying hen (*Gallus domesticus*) plasma pool, intra-
174 assay coefficients of variation for VTG and for VLDL_y ranged from 5.9-8.2% and 6.6-8.7%,
175 respectively. Inter-assay coefficients of variation were 7.8% for VTG and 6.9% for total VLDL_y.
176

177 2.3. Statistical analyses

178 Analyses were run with either the JMP 8.0 or SAS 9.0 software packages. All variables were
179 tested for normality, as were residuals from plots against predicted values, using Shapiro-Wilk
180 tests. Data transformations were applied when distributions were non-normal. Analysis of
181 covariance (ANCOVA) tests were used to examine breeding-group differences (e.g. successful,
182 failed, and deferring breeders) in VTG, VLDL_y, Hct, Hb, T, P, and body mass. As many studies
183 show links between age and individual quality and breeding output, we included age and a
184 measure of individual quality as a covariates in all analyses. We defined individual quality as the
185 number of chicks fledged (i.e. breeding quality) during the previous 6 year period (i.e. breeding
186 experience). We also examined whether a female bred successfully or not, or deferred, in the
187 previous year alone (i.e. immediate breeding history). However, because age and the two quality
188 measures are co-linear, we used the covariates in separate models exploring each response
189 variable listed above. Tukey-Kramer *post-hoc* tests were run after univariate tests to identify
190 significant contrasts between breeding groups. Correlations among all variables with dates of
191 colony arrival and with individual quality measures were examined through a Pearson's
192 correlation matrix.

193

194 **3. Results**

195 Female black-browed albatrosses in this study returned to the breeding colony on 10-17 October
196 2008, which is consistent with data from previous years (e.g. [26]). Of the 33 females that we
197 sampled, 8 deferred reproduction entirely, 8 laid eggs which either failed to hatch (N=5), or
198 hatched but failed shortly thereafter (N=3), and 17 fledged chicks successfully. Mean dates of
199 egg-laying occurred approximately 15 days after arrival (range 8-18 days), with a median lay-
200 date of 26 October. Based on observed arrival and laying dates, all 24 females that laid were
201 sampled during the 1-2 day period when newly arrived birds reunite with their mates at nest sites
202 before departing for an approximately 10-12 day pre-laying exodus. So on average, all breeding
203 females were sampled approximately 15 days into the rapid yolk deposition phase of egg
204 production when VTG levels are held at maximal levels (in black-browed albatrosses, RYD =
205 20.4 ± 1.1 days; [3]). All 8 deferring females were sampled over the same 7-day period as all the
206 breeding females.

207 No measured variables were significantly correlated with arrival date (body mass, tarsus
208 length, bill length, Hct, Hb, VTG, VLDLy, T, P; all $P > 0.10$, $N \sim 33$ individuals per variable).
209 Furthermore, no variables were related to age (mean age 22 years, range 12-30), or to individual
210 experience when defined as the number of chicks fledged during the previous 6 year period, or
211 whether a female bred in the immediately preceding year (in all models, both $P > 0.10$). In other
212 words, variation in age and breeding experience in the last 6 years did not significantly affect our
213 subsequent analyses and conclusions. Body mass did not correlate with either tarsus or bill
214 length (both $P > 0.20$), and so the analysis of body mass did not necessitate correction for
215 structural differences in body size (e.g. analysis of covariance or principal components analysis).

216 Supporting our first prediction, body mass at colony arrival differed among females with
217 differing breeding outcomes (whole model, $F_{2,30}=4.7$, $P=0.011$), independently of age effects
218 ($P=0.26$). Successful females weighed 3.83 ± 0.07 kg, which was significantly heavier (*post hoc*
219 contrast, $P=0.006$) than deferring females which weighed 3.53 ± 0.10 kg. Failed breeders were
220 intermediate in mass, and weighed 3.62 ± 0.10 kg (Fig. 1A) which was not significantly different
221 from either successful ($P=0.150$) or deferring ($P=0.289$) females. Therefore, as predicted,
222 deferring birds were significantly lighter than successfully breeding birds. When we pooled
223 failed and successful birds and compared them to deferring birds, the breeding birds were still
224 significantly heavier than deferring birds (3.79 ± 0.06 kg vs. 3.52 ± 0.09 kg; $F_{2,30}=4.4$, $P=0.016$).
225 Hct, but not Hb, also supported our first prediction (Fig. 1B-C), and Hct levels followed the same
226 trend as body mass. Hct was significantly higher in successful ($41.7 \pm 0.8\%$; $P=0.004$) and failed
227 ($41.3 \pm 1.2\%$; $P=0.033$) breeders than in deferring breeders ($37.8 \pm 1.0\%$) ($F_{2,30}=5.18$, $P=0.012$).
228 Hb did not differ significantly between breeding categories ($F_{2,30}=0.739$, $P=0.486$).

229 Also supporting our second prediction, differences in reproductive readiness, as measured
230 by differences in sex steroid hormone and yolk precursor levels (Fig. 1D-G), varied in relation to
231 reproductive status independently of age and breeding quality effects. Differences in P4 (Fig.
232 1D) showed that successful ($P=0.009$) and failed ($P=0.030$) females had higher (up-regulated)
233 concentrations than deferring females (successful = 1.71 ± 0.38 ng ml⁻¹, failed = 1.41 ± 0.57 ng
234 ml⁻¹, deferring = 0.34 ± 0.53 ng ml⁻¹; $F_{2,26}=4.10$, $P=0.028$). T was significantly higher in failed
235 females (2.79 ± 0.50 ng ml⁻¹) relative to both successful ($P=0.003$) and deferring ($P=0.016$)
236 females (successful = 1.20 ± 0.28 ng ml⁻¹, deferring = 0.68 ± 0.39 ng ml⁻¹) ($F_{2,27}=5.62$, $P=0.009$;
237 Fig. 1E). VTG varied significantly ($F_{2,21}=7.33$, $P=0.004$; Fig. 1G), with higher concentrations in
238 breeding females (2.91 ± 0.34 µg Zn ml⁻¹) than in both deferring (0.74 ± 0.42 µg Zn ml⁻¹, $P=0.0$

239 12) and failed ($1.21 \pm 0.46 \mu\text{g Zn ml}^{-1}$, $P=0.032$) females. VLDLy concentrations however did
240 not vary significantly among deferred, failed and successful females ($F_{2,22}=2.05$, $P=0.15$; Fig.
241 1F).

242 Among the breeding birds, T and P4 were related to VTG concentrations but in opposite
243 directions: P4 was positively related with VTG in a natural log manner ($r=0.64$, $P=0.001$, $N=14$,
244 Fig. 2A), while T was negatively related to VTG ($r=-0.56$, $P=0.007$, $N=16$, Fig. 2B). T and P4
245 were not significantly correlated in the breeding birds ($r=-0.22$, $P=0.32$, $N=24$), or in all breeding
246 and deferring birds collectively ($r=-0.15$, $P=0.42$, $N=33$).

247 Using regression analysis, date of breeding failure (known for 7 of the 8 failed females)
248 was significantly related to pre-breeding T, VTG, and VLDLy concentrations ($N=7$ females, all
249 $P<0.017$, Fig. 3A-C). T tended to be higher in females which failed shortly after laying relative
250 those failing later. Similarly, VTG and VLDLy were generally lower in females failing early,
251 and higher values were associated with later failure. In contrast, pre-breeding body mass was not
252 significantly related to date of breeding failure ($P=0.128$, Fig. 3D). Multiple logistic regression
253 analysis revealed a significant role of VTG in predicting breeding success or failure ($\chi^2=4.98$,
254 $P=0.026$), but the influence of body mass was not significant ($\chi^2=1.13$, $P=0.287$). In other words,
255 resource-independent VTG effects were more strongly predictive of breeding success than
256 resource-dependent mass effects.

257

258 **4. Discussion**

259 In this study we show that after long-distance migrations to a breeding colony, reproductive
260 decisions (breeding or deferring) and reproductive outcome (successful or failed) in female
261 black-browed albatrosses are associated with marked differences in patterns of circulating

262 gonadal steroids (P4, T) and steroid-dependent traits (yolk precursors). Previous studies of
263 black-browed albatrosses have shown that all breeding-age females generally return to natal
264 colonies at the start of the breeding season, whether they breed or not, and that plasma LH levels
265 are elevated upon arrival [18]. It is therefore parsimonious to assume that day-length drives the
266 general timing of their return to colonies, indicating a general seasonality of migratory behaviour,
267 and that this is sufficient to “switch on” the reproductive axis at the level of the hypothalamus
268 and pituitary as in most temperate-breeding birds [10]. However, our data clearly show that
269 female black-browed albatrosses are not all equally prepared for reproduction upon arrival in
270 terms of “downstream” components of the reproductive axis. As we predicted, deferring females
271 had low plasma progesterone (P4), and consequently low testosterone (T) and low estrogen-
272 dependent vitellogenin (VTG) levels, which suggests that the decision to defer reproduction by
273 these annually breeding birds was made prior to arrival perhaps due to “stressful” metabolic
274 demands incurred at sea during the over-winter or migratory phase, i.e. carryover effects. Also as
275 predicted, successful females had high plasma P4 and high VTG, but low T, which indicates a
276 responsiveness of the ovary to stimulation by pituitary LH. The low plasma T in these
277 successfully breeding birds is presumably due to rapid conversion (aromatisation) to E₂ to
278 support vitellogenesis, as indicated by the high VTG levels that we measured. The failed
279 breeders however are especially interesting as they had high P4 and high T, which like the
280 successful breeders indicates gonadal sensitivity to LH secretion, but these females had low VTG
281 which suggests a possible disruption of aromatisation (as per [12]) in the E₂-mediated
282 vitellogenic pathway. Ultimately, and rather amazingly, VTG in pre-laying females predicted not
283 only their breeding success or failure, but it also the length of time from their arrival at the colony
284 to breeding failure (Fig. 3B). This suggests very strongly that hormonal differences among
285 individuals at colony arrival, during the transition from migratory to reproductive states, affect

286 not only breeding decisions but also the degree of reproductive readiness with consequences for
287 reproductive success. Furthermore, these effects influence processes at the *downstream* end of
288 the reproductive axis, at the level of ovary steroidogenesis and E₂-dependent regulation of hepatic
289 vitellogenesis.

290 Collectively, our data suggest that reproductive decisions (breeding or deferral) and
291 reproductive success are influenced by two very different mechanistic pathways in female black-
292 browed albatrosses. In addition to hormonal differences, the decision to defer reproduction was
293 associated with more traditional measures of individual condition, characterized by lower body
294 mass and lower hematocrit levels at arrival relative to breeding individuals. This is consistent
295 with previous studies of breeding deferral in seabirds (blue petrels [7]; gulls [19]; shearwaters
296 [21]), and with the idea that the initiation of breeding attempts is resource- or condition-
297 dependent. Furthermore, the low P4 and T levels in deferring birds is a hormonal profile that has
298 been linked to poor condition in seabirds [13,14,15]. These finding support our first prediction:
299 breeding deferral is characterized by low body mass, low blood hematocrit, and a generally
300 compromised condition upon arrival at the breeding colony after overwinter migration. In
301 contrast, and supporting our second prediction, reproductive success in albatrosses that initiated a
302 breeding attempt was not related to body mass or to hematocrit (i.e. to resource state or
303 condition) but showed a significant relationship with the potentially “resource-independent”
304 endocrine processes underlying follicle development [8,17,37].

305 It is important to note that, contrary to many studies, female age and breeding experience
306 were never significant effects in our models. However, this is perhaps not surprising as all but
307 three of the birds we sampled (30 of 33) were experienced breeders between 17-25 years of age
308 and within the range of peak breeding performance of black-browed albatrosses [1]. The
309 significance of this is that we have found hormonal differences in female albatrosses at the peak

310 of breeding performance which were predictive of reproductive success, failure, and deferral, and
311 which were unrelated to senescence or inexperience.

312

313 *4.1. Resource-dependent breeding decisions*

314 Female black-browed albatrosses showed considerable inter-individual variation in body mass
315 upon arrival at the breeding colony, the extremes of which differed by as much as 1 kg (range
316 3.25-4.25 kg, N=33). The initial breeding decision (breed/defer) was very strongly associated
317 with body mass, independent of age or quality, and the simplest explanation for this pattern is
318 that returning females can assess their post-migratory physiological state, and if some relative
319 threshold of condition has not been attained they will forgo reproduction thereby minimizing
320 fitness costs (as per life-history theory, [29]). This might be mediated in part by the effect of
321 body condition on hormonal processes which affect HPG functioning [13,14,15], and in our study
322 P4 and T were not up-regulated in deferring females to a degree that would support gamete
323 production. Previous studies of carryover effects in migratory animals, including seabirds,
324 provide some evidence for this by arguing that the environmental conditions experienced during
325 the non-breeding period can influence resource state at the onset of reproduction ([16] and
326 references therein). The low body masses and hematocrit levels of deferring females suggest that
327 these albatrosses had very different or difficult experiences during the nonbreeding period (e.g.
328 different foraging opportunities, feeding successes, distributions, migration routes or
329 oceanographic conditions) relative to successful females, and that deferring females were less
330 able to cope with the physical demands of migration, perhaps via reduced hematocrit-dependent
331 oxygen transport capacities. However, direct observation of migratory behaviour via telemetry is
332 needed to confirm these links. Nevertheless, as in other avian species [31,32], the deferring
333 albatrosses were in a poorer post-migratory / pre-breeding condition independent of arrival date,

334 age, or recent breeding history, and the low steroid hormone levels (P4 and T) and near-basal
335 yolk precursor levels suggests that they had ‘decided’ before their arrival at the breeding colony
336 to forgo steroidogenesis and thus egg formation. We therefore propose that a migratory
337 carryover effect(s) constraining resource acquisition prior to arrival is the most parsimonious
338 explanation for breeding deferral.

339

340 *4.2. Reproductive readiness and breeding success*

341 Once a female albatross decides to provision an egg and that egg is laid, she will then either fail
342 or succeed at producing a chick. Among the female albatrosses that initiated a breeding attempt,
343 reproductive success was not related to measures of resource state or physiological condition -
344 neither body mass nor hematocrit differed significantly between failed and successful females
345 upon their arrival at the colony (and reproductive success was independent of age and
346 experience). In contrast, success was strongly related to the steroidogenic processes underlying
347 follicle development and yolk precursor production. In fact, multiple logistic regression
348 indicated a significant role of VTG, but not body mass, in reproductive success. This strongly
349 suggests that the reproductive failure of female black-browed albatrosses in this study was most
350 likely the result of a constraint on the endocrine pathways regulating normal yolk precursor
351 production.

352 So how might variation in pre-breeding VTG concentrations affect breeding success or
353 failure weeks to months after laying? The answer may lie in the dynamics of egg production.
354 During the course of normal follicle development, testosterone synthesized in ovarian follicle
355 cells is converted to 17 β -estradiol which is then secreted to general circulation to stimulate the
356 production of yolk precursors VTG and VLDL_y by the liver (reviewed by [35]). VTG and

357 VLDL_y are the principal sources of protein and lipid for developing embryos, so it might be
358 expected that a constraint on either precursor would have deleterious effects on embryonic
359 development and egg/chick survival. In fact, we show that failed females had significantly lower
360 VTG concentrations than successful females, and that VTG more strongly predicted
361 success/failure than body mass (Fig. 1G). Furthermore, when we examined VTG and VLDL_y
362 levels in failed females (Fig. 3), we found that low levels were related to failure shortly after
363 laying, whereas higher levels delayed egg/chick mortality by some weeks/months. Overall, failed
364 females had a mean VTG value which was nearly identical to that of the deferring, non-breeding
365 females which did not produce eggs, and closer to those of females sampled 6 months after the
366 egg-producing stage of the annual cycle just prior to winter out-migrations (mean 0.17 $\mu\text{g Zn ml}^{-1}$,
367 G.T. Crossin, unpublished data). In contrast, pre-breeding body mass was not significantly
368 related to the date of failure (Fig. 3). Some caution may be warranted when interpreting the
369 effect of VTG on breeding failure as the number of failed albatrosses in this study was somewhat
370 low (N=8). Nevertheless, even with a low sample size, we were still able to resolve significantly
371 lower VTG concentrations in failed versus successful breeders; generally, low samples sizes
372 increase the risk of a Type II statistical error, but this is not the case here.

373 The consequences of low circulating maternal VTG concentrations for egg and offspring
374 quality have been demonstrated in experimental studies with female birds. Experimental
375 reduction in circulating vitellogenin decreased yolk size and quality [33,34], with deleterious
376 effects on offspring quality and survival [8,30]. In the absence of any age or breeding experience
377 effects, we therefore propose that the provisioning of constituents to the yolks of failed albatross
378 eggs was probably insufficient to sustain normal embryonic development, which as indicated in
379 Fig. 3 could result in either relatively rapid or delayed mortality. The mechanism(s) responsible

380 for this constraint in post-migratory females is not known, but might involve a glucocorticoid-
381 mediated carryover effect or inhibition of reproductive physiology [4,8].

382 Although we did not directly measure plasma E_2 , variation in P4 and T provide some
383 indication why VTG levels were so low in post-migratory female albatrosses which subsequently
384 failed in their breeding attempt. P4 is the common precursor for T and E_2 , and after a series of
385 enzymatic reactions P4 is converted to T, which can then be aromatized to E_2 . Elevated levels of
386 both P4 and E_2 are required for normal reproductive function in females, mediating oviduct
387 development, yolk precursor production (e.g. P4 is synergistic with E_2 binding on vitellogenin
388 gene II, [6]), follicle development, and reproductive behaviours. This is supported by the strong,
389 positive relationship we observed between P4 and E_2 -dependent plasma VTG levels (Fig. 2A),
390 which we argue is indicative of greater “reproductive readiness” *sensu* [8]. In contrast, females
391 which failed had significantly higher T levels and lower VTG levels than successful birds (Fig.
392 2B). This suggests a potential stress-related inhibition of P_{450} aromatase which prevented
393 conversion of ovarian T to E_2 in these birds (as occurs in the brain nuclei of various bird species
394 [4,12]), and which effectively constrained E_2 -dependent vitellogenesis. This supports the idea of
395 reduced reproductive readiness in females with high T. It is important to reiterate that the
396 variables we examined were measured during the pre-breeding period, weeks to months before
397 ultimate reproductive outcome. Although we found no significant associations between pre-
398 breeding body mass and reproductive success, this does not discount the possibility of a resource
399 link to reproductive success later during the incubation or chick rearing stage. In fact, one may
400 very well assume that resource-dependent and resource-independent factors work synergistically
401 to affect ultimate reproductive fate (but as we have indicated, the focus of this present study was
402 on the pre-breeding physiology and condition of female albatrosses). Ultimately, we have shown
403 that at this early stage of the breeding season, immediately following overwinter migration, there

404 was a very clear and significant effect of reproductive physiology on breeding success,
405 independent of resource effects, which we attribute to a migratory carryover effects on
406 reproductive readiness operating downstream in the reproductive axis.

407 We have thus far used the words “resource-dependence” and “condition-dependence”
408 synonymously, but there exists the possibility that reproductive success might be resource-
409 dependent but not condition-dependent. For example, we have shown that there is a relative
410 resource or condition threshold below which breeding does not occur, and here both terms are
411 generally synonymous with “body mass”. However, once this body mass threshold was met and
412 egg production proceeded in the breeding albatrosses, then a possible distinction between
413 “resource” and “condition” emerged. Our data show that both failed and successful breeders
414 were similar in mass, and thus in similar condition. However, the resources available to these
415 birds to allocate towards reproduction (e.g., steroidogenesis, vitellogenesis, etc.) might still be
416 subject to limitation. Protein limitation is thus one possible explanation for the presumed
417 inhibition of P450 aromatase activity and the low VTG levels that distinguished failed from
418 successful breeders. As such, it is possible that breeding failure is resource- but not condition-
419 dependent.

420

421 **5. Conclusion**

422 We present data which suggest that migratory carryover effects can impact two very different
423 aspects of reproductive effort. In the short term, carryover effects constraining the acquisition of
424 resources and body condition during the pre-breeding period mediated the trade-off between
425 current and future reproductive investment (e.g. breeding decision), via hormonal mediators.
426 However, for females deciding to initiate reproduction, the steroidogenic processes underlying
427 yolk precursor production become more directly involved in ultimate breeding success, which in

428 this case involved a constraint on the E₂-mediated yolk-precursor production pathway
429 independent from any condition-related effect. As few studies reporting carryover effects on
430 reproductive success show strong or consistent relationships with measures of pre-breeding
431 resource allocation or condition in females [8,16,22,28], it is important to consider a much
432 broader set of potential mechanisms [17,35], especially as virtually none of the studies reviewed
433 by Harrison et al. [16] show definitive links between resource state and improved breeding
434 success in females. Having controlled for age and experience related effects, the pleiotropic
435 interactions and incompatibilities among endocrine and other physiological regulatory systems
436 like those described in this study are the most likely candidates underlying reproductive success.

437

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- 534

535 **Figure Legends**

536 Fig. 1: Body mass (A), hematological (B-C), hormonal (D-E), and yolk precursor (F-G) profiles
537 in pre-breeding female black-browed albatrosses (*Thalassarche melanophris*) upon arrival at a
538 breeding colony at Bird Island, South Georgia. Birds are grouped according to breeding
539 outcome, and bars represent least square means \pm SEM. Numbers at the base of bars indicate
540 sample sizes. Differing letters indicate statistical significance when $\alpha=0.05$. Note that when
541 failed and successful birds are pooled in Panel A, the deferring birds are significantly lighter
542 (see Results section for details). Abbreviations: Hct- hematocrit; Hb- hemoglobin; P4-
543 progesterone; T- testosterone; VLDLy- yolk-targeted very low density lipoprotein; VTG-
544 vitellogenin.

545
546 Fig. 2: Relationship between plasma vitellogenin and progesterone (A) and testosterone (B) in
547 pre-breeding female black-browed albatrosses (*Thalassarche melanophris*) which eventually
548 laid eggs. Open circles indicate failed breeders and closed circles are successful breeders.
549 Lines are natural log fits. See Figure 1 for key to abbreviations.

550
551 Fig. 3: Relationships of testosterone (A), vitellogenin (B), very low density lipoprotein (C), and
552 body mass (D) with date of breeding failure in female black-browed albatrosses. Lines are
553 natural log fits. See Figure 1 for key to abbreviations.

554

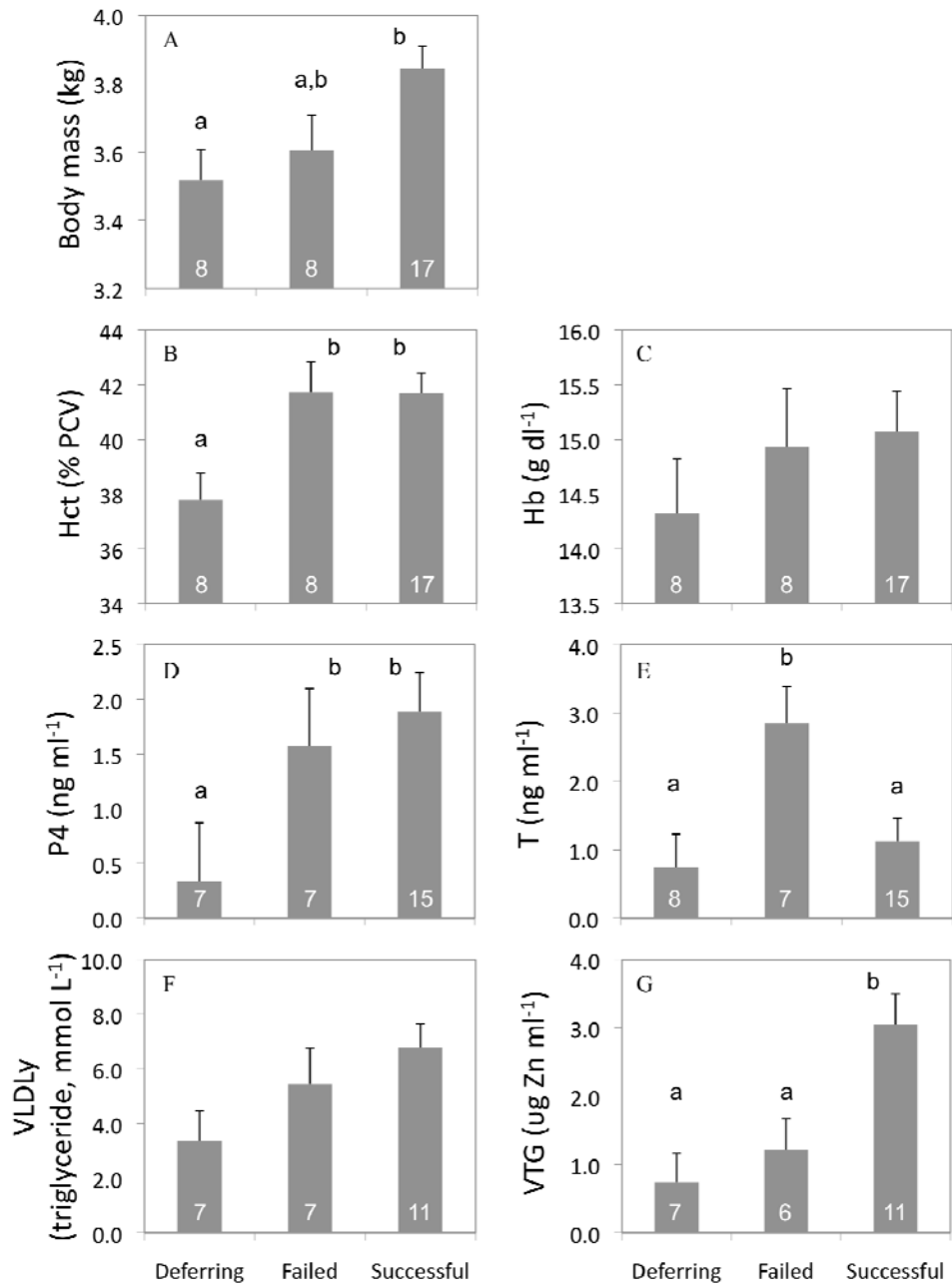


Fig. 1

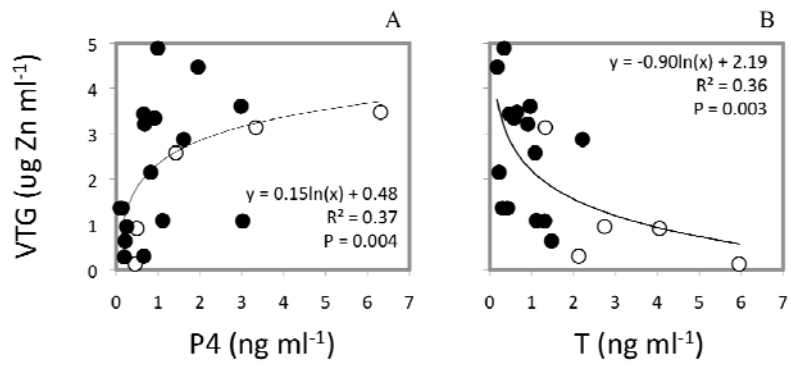


Fig. 2

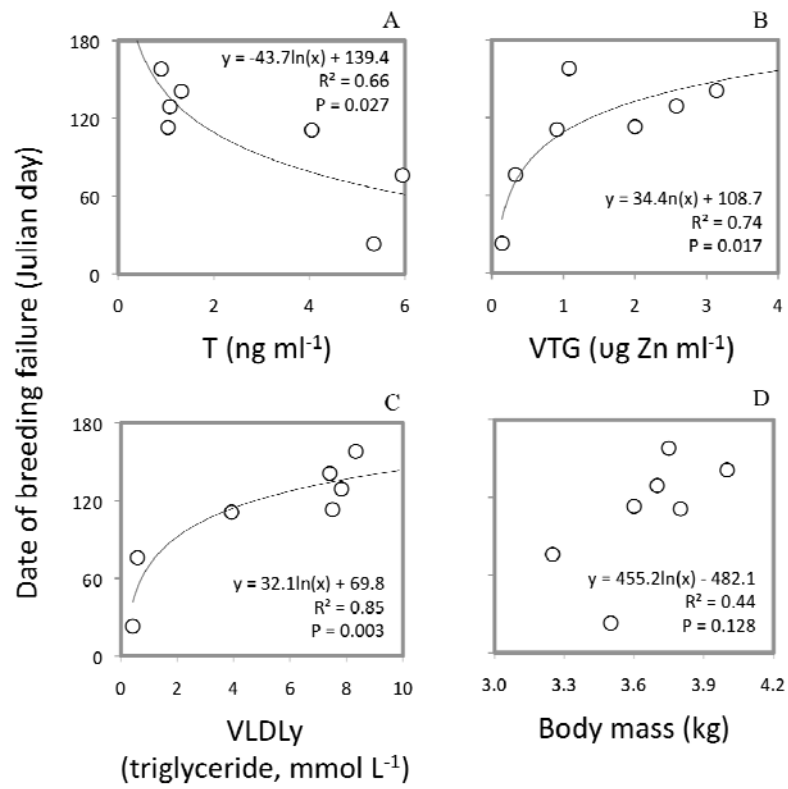


Fig. 3