1	Variable neuroendocrine responses to ecologically-relevant challenges in sticklebacks
2	
3	
4	
5	
6	
7	Alison M. Bell*
8	Integrative Biology, University of Illinois at Urbana-Champaign
9	505 South Goodwin Ave., Urbana, IL 61801, USA
10	Phone: (217) 265-5469, Fax: (217) 244-4565, Email: alisonmb@life.uiuc.edu
11	
12	Tobias Backström
13	Evolutionary Biology Centre, Dep't of Comparative Physiology, Uppsala University, Sweden
14	
15	Felicity A. Huntingford
16	Environmental and Evolutionary Biology, University of Glasgow, UK
17	
18	Tom G. Pottinger
19	Centre for Ecology and Hydrology, Lancaster, UK
20	
21	Svante Winberg
22	Basic Science and Aquatic Medicine, Norwegian School of Veterinary Sciences, Norway and
23	Evolutionary Biology Centre, Dep't of Comparative Physiology, Uppsala University, Sweden

24	ABSTRACT

25	BELL, A.M., BACKSTRÖM, T.B., HUNTINGFORD, F.A., POTTINGER, T.P., WINBERG, S.
26	Variable neuroendocrine responses to ecologically-relevant challenges in sticklebacks.
27	PHYSIOL BEHAV 00(0) 000-000, 2006. Here, we compare the behavioral, endocrine and
28	neuroendocrine responses of individual sticklebacks exposed to either an unfamiliar conspecific
29	or to a predator. We found that the two stressors elicited a similar hypothalamic-pituitary-
30	interrenal response as assessed by whole-body concentrations of immunoreactive corticosteroids,
31	but produced quite different patterns of change in brain monoamine and monoamine metabolite
32	content as assessed by concentrations of serotonin (5-HT), dopamine (DA), norepinephrine (NE)
33	and the monoamine metabolites 5-hydroxyindole acetic acid (5-HIAA), homovanillic acid
34	(HVA) and 3-4-dihydroxyphenylacetic acid (DOPAC). For example, relative to baseline levels,
35	NE levels were elevated in individuals exposed to a predator but were lower in individuals
36	confronted by a challenging conspecific. Levels of monoamine neurotransmitters in specific
37	regions of the brain showed extremely close links with behavioral characteristics. Frequency of
38	attacking a conspecific and inspecting a predator were both positively correlated with
39	concentrations of NE. However, whereas serotonin was negatively correlated with frequency of
40	attacking a conspecific, it was positively associated with predator inspection. The data indicate
41	that the qualitative and quantitative nature of the neuroendocrine stress response of sticklebacks
42	varies according to the nature of the stressor, and that interindividual variation in behavioural
43	responses to challenge are reflected by neuroendocrine differences.
44	Key words: stickleback, Gasterosteus aculeatus, behavioral syndromes, antipredator
45	behavior, aggression, glucocorticoid, serotonin, stress, coping styles, individual differences
46	Running head: Individual differences in sticklebacks

INTRODUCTION

Both attacking a conspecific and confronting a potential predator are dangerous. In addition to energetic costs [1], aggression can result in injury [2] and exposure to predation risk while fighting [3]. Similarly, an encounter with a potential predator can impose energetic costs of escape [4], injury [5] or even death. Not surprisingly, both confrontation by a challenging conspecific [6-11] and exposure to a predator [12-15] elicit a neuroendocrine stress response.

The neuroendocrine stress response involves a coordinated activation of both the hypothalamic-pituitary-adrenal (or interrenal, in the case of fishes, HPI) axis and the brain monoamine neurotransmitter systems [16]. When a stimulus evokes a stress response, both systems are activated by the same central mechanism, resulting in the elevation of plasma corticosteroids and brain monoaminergic activity. In general, exposure to stressors is associated with increased concentrations of plasma glucocorticoids and increased turnover of 5-HT to 5-hydroxyindoleacetic acid (5-HIAA) [17].

Individual differences in behavior are often related to individual differences along both axes of the stress response [18-22]. With respect to the HPA axis, individual differences in aggressiveness are negatively correlated with concentrations of plasma glucocorticoids in trout [23] and chickens [24]. In humans, individual differences in behaviors that are analogous to risk-taking behaviors and aggression are associated with increased norepinephrine and dopamine activity [25,26]. Finally, aggression and risk-taking behaviors in several species have been linked to serotonin turnover. For example, individual differences in aggression are negatively related to serotonin turnover in monkeys [24,27-29], trout [21] and anolis lizards [30-32]. However, the relationship between 5-HT, stress, the HPI axis and aggression is complex and

depends on the duration of the stressor. For example, in salmonids, 5-HT turnover is usually positively associated with plasma ACTH and cortisol concentrations and negatively associated with aggression. However, long-term stimulation of the serotonergic system has inhibitory (negative) effects on the HPI axis [33] and aggression [17].

In previous work, we have shown that behavioral reactions to predators and competing conspecifics covary at the individual level in threespined sticklebacks (*Gasterosteus aculeatus*) [34-36]. While some individuals are willing to engage in behavior that appears to be dangerous, such as foraging under predation risk or performing predator inspection, other individuals are much more cautious around predators. Individuals that take more risks in this context are also more aggressive toward conspecifics. Covariance among suites of behavioral traits is common [37,38] and in several species the shy-bold continuum and the proactive-reactive axis have been associated with individual differences in stress responsiveness [39]. Therefore it is possible that differences in how individual sticklebacks respond to dangerous situations might be linked with differences in the stress response.

Here, we investigated natural variation in behavioral, glucocorticoid and monoamine responses of individual sticklebacks to two potentially dangerous situations. We wished to establish whether wild-caught animals responding to ecologically-relevant challenges show stress responses that are comparable in nature and extent to those described for laboratory animals, and whether the stress response might be an underlying root of the covariance of behavioral responses in sticklebacks. With this in mind, we exposed individuals to either an unfamiliar conspecific or to a potential predator and recorded their behavior. Although the danger of predation is greater than the danger posed by a territorial intrusion, we hypothesized that both situations would induce a stress response because social stress is one of the most

effective stressors in inducing a high magnitude response in other animals [40]. We sampled individuals at 15, 30 or 60 minutes after exposure to determine the time course of the glucocorticoid and monoaminergic responses to these two threats. This design allowed us not only to follow the neuroendocrine responses to these stressors through time, but also to determine whether individual differences in behavioral responses to these challenges could be related to underlying neuroendocrine physiology.

METHODS

Overview: Individuals were presented with one of two potential threats, either an unfamiliar conspecific or a predator, hereafter referred to as 'conspecific' and 'predator', respectively, and their behavior was recorded. Individuals exposed to the 'conspecific' or the 'predator' were subdivided into three different treatment groups, sacrificed 15, 30 or 60 minutes after exposure to the potentially threatening stimulus. Individuals were randomly assigned to a treatment group prior to observing their behavior. The responses to the stressors were compared across time periods and against a 'baseline control' group, which consisted of individuals sampled directly from an undisturbed stock tank. Each treatment group comprised ten individuals.

Subadult sticklebacks were collected from the River Endrick in January 2004 and brought to the Glasgow University Field Station, Rowardennan, where all of the behavioral observations were carried out. Groups of fish (n=10-40) were maintained in flow-through stock tanks (210 liters) at $9 \pm 2^{\circ}$ C and on a 14L:10D photoperiod. Fish were fed frozen bloodworms *ad libitum* daily except on the day of observation, when they were unfed.

Behavioral observations took place in March and April 2004 in a U-shaped flume with a live pike (*Esox lucius*) in either arm of the flume. Aquaria that were used for behavioral observation ('observation tanks', 44 liters, 61x32x22 cm) were placed inside the flume and next to a window in the flume so that the behavior of the fish could be observed. The window was covered by a blind with a small opening which allowed the observer to see through the window with minimal disturbance to the fish. Each observation tank contained a one-liter glass conical flask, a plastic plant and a length of opaque tube (12 cm diameter, 36 cm tall) that stood vertically on one side of the tank and allowed fish to be introduced into the tank with a minimum of disturbance. Exterior lines on the tanks divided them into 16 equally-sized areas.

Each arm of the flume contained one of two live pike (46, 41cm standard length) and cloth plants which served as hiding places for the pike. The compartments were fitted with a removable opaque cover which created a dark, shaded area for the pike. The pike were caught by hook and line in February 2004 in a small water body near the Glasgow University Field Station (the Duibh Lochan). The two pike were fed dead minnows and dead sticklebacks *ad libitum*.

Procedure:

Fish were removed from the stock tank and placed into a settling tank (49 liters, 61x31x26 cm) for two nights in order to acclimate to the flume. After the acclimation period, sticklebacks were netted from the settling tank and were randomly assigned to one of eight treatments (see below for a description of the different treatments). The stickleback was deposited into the tube in an observation tank. After 15 minutes, the tube was lifted, which allowed the stickleback to swim freely around the tank. After another 15 minutes, the fish was

presented with either an unfamiliar conspecific or a pike, and the behavioral observation began.

Behavioral observations of response to an unfamiliar conspecific and predator were alternated.

Treatments:

Unfamiliar conspecific: We employed a procedure that was designed to simulate a challenge to the resident fish by an intruding conspecific. Sticklebacks at this size and age $(0.373 \pm 0.02 \text{ g})$, approximately 7-8 months of age) are not breeding and so do not defend breeding territories, but they do display aggressive behavior during competition for food and other resources and can be territorial [41]. Therefore we interpret the behavioural response of sticklebacks to the unfamiliar conspecific in this experiment as a response to a potential competitor for food and/or space. It is also worth considering that the sticklebacks' response to a conspecific might also reflect an affiliative motivation because they were held in isolation.

A live conspecific (within 5mm standard length of the resident) was placed into the flask in the observation tank. Seven different conspecifics were used as intruders throughout the experiment. A fish was never used as an intruder more than once consecutively. The flask effectively standardized the behavior of the intruder by minimizing movement. The frequency of attacking the conspecific (biting) was recorded for 15 minutes after the resident first oriented to the conspecific because some individuals were facing away from the flask when the intruder was introduced. Latency to orient to the intruder ranged from 0.4-482.0 seconds (mean= 104.6 ± 24.7 s). This procedure is roughly analogous to studies with trout where a resident is challenged by an intruder [23]. However, an important difference is that in the present case there is no physical contact between the resident and intruder and the intruder cannot escape. We elected to use this procedure to minimize stress to the intruder. After the behavioral observation, the flask

containing the conspecific was removed from the tank and the resident fish was sacrificed according to treatment (15 minutes, 30 minutes or 60 minutes after the behavioral observation was completed).

161

162

163

164

165

166

167

168

169

170

171

172

173

174

175

176

177

178

179

180

181

182

183

Predator: This procedure was designed to simulate a potential predatory threat by a live pike. We lured the pike into a chamber situated next to the observation tank by removing cover over the pike. In general, the pike willingly swam into the chamber, seeking cover. A removable opaque divider was situated between the observation aquarium and the predator chamber. To start the behavioural observation, the divider separating the observation aquarium from the chamber was gently lifted, allowing the stickleback a clear view of the pike on the other side of the glass. The behavior of the individual stickleback was observed for 15 minutes after the divider was removed and the following behaviors were recorded: predator inspection (swimming next to and orienting to the mouth of the pike) and time orienting (body facing toward the pike). Whether the pike moved or oriented to the stickleback during the observation was also recorded. After the behavioral observation, the opaque divider separating the chamber from the observation aquarium was replaced and the fish was sacrificed according to treatment (15 minutes, 30 minutes or 60 minutes after the behavioral observation completed). In order to eliminate any olfactory cues that might affect subsequent behavioral observations, the water in each of the observation tanks was replaced after each behavioral observation.

The two pike used in this study did not differ in behavior and movement of the pike during the observation period did not have a statistically detectable effect on either the behavior or the physiology of the sticklebacks (all P>0.05).

Baseline control: Each day, for ten days, a single stickleback was netted from a stock tank and sacrificed immediately to contribute to a baseline control value for neuroendocrine and

hormonal measurements. These fish were collected at the same time as individuals in the treatment groups to minimize the amount of disturbance in the stock tank.

Settling tank control: At the end of each observation day, 1-2 remaining individuals in the 'settling tank' were quickly netted from the settling tank and sacrificed immediately. This group (n=10) was analyzed for corticosteroids to determine whether transfer and housing in the flume produced a stress response. However, it is important to note that this group does not control for the effect of isolation. We did not detect a difference in whole-body between the settling tank control and the baseline control and therefore did not analyze this treatment group further (Figure 1, $F_{1.18}$ =0.488, P=0.494).

Tissue collection

Fish were quickly killed by decapitation. The head and body were immediately weighed, the brain dissected out within three minutes and mounted in Tissue-Tek (Sakura). The brain and body were immediately frozen on dry ice and stored at -80° C until physiological analyses. A small amount of tissue from the tail fin was placed in 80% ethanol for DNA extraction for sex determination. Tissue was collected between 0800 and 1800 hours. As in [42], we found no evidence for circadian changes in whole-body cortisol (r=0.045, F_{1.58}=0.118, P=0.773).

Steroid determination

Corticosteroids were assessed by measurement of solvent-extractable immunoreactivity in whole-body homogenates. Corticosteroids were extracted from the tissue by homogenization in ethyl acetate (5:1 volume:carcass weight). Recovery of steroids from homogenized tissue was assessed by adding 50µl radio-labelled cortisol tracer to homogenized tissue and equilibrating for

one hour before extractions. Immunoreactive steroids were quantified in 20-100 µl aliquots of ethyl acetate extracts of whole-body homogenates using a validated cortisol radioimmunoassay procedure as described previously [43-46]. We used the rabbit polyclonal antibody to cortisol produced by the IgG Corporation and supplied by Campro Scientific (code IgG-F-2).

A standard curve of 0-800 pg cortisol per tube was used.

We quantified cortisol in whole-body homogenates rather than plasma because successful extraction of the brain for monoamine analyses required that it be dissected out and frozen as soon as possible, which precluded rapid blood sampling from the body. The whole-body homogenate method measures cortisol in multiple body compartments. Therefore in addition to measuring plasma concentrations of cortisol, this method also detects cortisol derivatives in the liver and gall bladder that might have cross-reacted with the antibody [47], This does not detract from the ability of this method to detect the onset of a stress response, because corticosteroids are synthesized de novo and not stored prior to release. This method has been employed previously to monitor the stress response in fish from which, because of their small size, blood samples could not be obtained, including juvenile trout [48], zebra fish [49] and sticklebacks [46]. Simultaneous measurement of plasma cortisol and whole-body cortisol in fish exposed to acute and chronic stressors has confirmed that the method is appropriate for detecting stress-induced changes in HPI activity [48]. Hereafter we refer to concentrations of corticosteroids we measured on whole body preps as ng/g of 'whole-body cortisol'.

Analysis of brain monoamines

Brains were sectioned in a frozen state on a cryostat and mounted on glass slides.

Sections of 300 µm thickness were cut in the coronal plane. Brain-punch microdissection was

performed as described by [30]. The hypothalamus, telencephalon and region posterior to the hypothalamus ('reticular formation') were identified for punching.

Punches from each of these three regions were collected and homogenized in 50µl ice-cold 4% perchloric acid containing 40 ng/ml DHBA (dihydroxybenzamine) as internal standard, using an MSE 100-W ultrasonic disintegrator. Samples were then centrifuged at 13000rpm for 10 minutes at 4°C and the supernatants were analyzed for serotonin (5-HT), dopamine (DA) and norepinephrine (NE) and their metabolites 5-hydroxyindoleacetic acid (5-HIAA), 3,4-dihydroxyphenylacetic acid (DOPAC) and homovanillic acid (HVA) using high performance liquid chromatography with electrochemical detection [50] immediately, or stored at -80°C for no more than two days prior to analysis. Pellets were stored at -80°C for subsequent analysis of protein content in an Eppendorf Biophotometer by a pre-made program measuring absorbance at 280nm. The monoamines and monoamine metabolites were quantified using standard solutions and corrected for recovery of the internal standard using HPLC software (CSW, DataApex Ltd., the Czech republic). The concentration of monoamines and monoamine metabolites is expressed as ng per mg protein.

We did not detect strong differences between brain regions in concentrations of brain monoamines: the only effect that we detected was that levels of DA ($F_{2,81}$ =3.36, P=0.04), 5-HIAA ($F_{2,81}$ =4.57, P=0.013) and 5-HT ($F_{2,81}$ =5.21, P=0.007) were significantly lower in the reticular formation in the 'predator' treatment (Table 1). Therefore we summed the concentration of each monoamine across regions and focused our subsequent analysis of treatment differences on the whole-brain values. However, the failure to detect strong region-specific differences should not be overinterpreted because we did not have the resolution to detect fine-scale

differences. Other studies have found region-specific differences in monoamine turnover during aggression [32].

A decrease in the concentration of a monoamine neurotransmitter could reflect a reduction in the release of the neurotransmitter (decrease in activity) or an increase in turnover to its metabolite (increase in activity). Therefore, it is preferable to use the ratio of the parent neurotransmitter to its metabolite (5-HIAA:5-HT, DOPAC:DA AND HVA:DA) as an index of neurotransmitter activity. However, we were unable to quantify the NE metabolite, 3-methoxy-4-hydroxyphenylglycol (MHPG) in any of the samples as a consequence of non-identified interfering peaks. In addition, in some samples the monoamines (especially 5-HIAA and 5-HT) became degraded during the sampling procedure, resulting in our failure to detect 5-HIAA. This was particularly a problem for the 'conspecific' treatments (Table 1). Samples with undetectable levels of a monoamine were omitted from that analysis.

Here, we report data on the concentration of both the parent monoamine and metabolite, and we focus our interpretation on differences between treatment groups, rather than on the functional significance of absolute levels.

Determining genetic sex

DNA was extracted from each fin clip and genetic sex was determined by genotyping each individual for a male-specific genetic marker validated for sticklebacks [51].

Data analysis

We compared the behavioral and physiological responses of sticklebacks to an unfamiliar conspecific and a predator across time using general linear models except when data were non-

normal. We tested for the effects of sex, body size, time and treatment on each of the dependent variables (behavior, whole-body cortisol and brain monoamines in the different regions). We did not detect sex differences in behavior, whole-body cortisol or brain monoamines and therefore did not analyze this factor further (all P>0.4). The least-squares difference post-hoc test was used to test for differences between groups, except when the distribution was non-normal, in which case we tested for differences between treatments using the nonparametric Mann-Whitney U test.

Pearson correlations were used to test for statistically significant relationships between variables when the data were normally distributed; otherwise, Spearman rank correlation statistics were computed. Because the same behavioral data was used to test for associations with brain monoamine concentrations, we used the sequential Bonferroni procedure to correct for multiple tests. Briefly, for each brain region within a treatment group, we replaced the correlation statistics with their corresponding P-values and then ranked them from smallest to largest. Results that were significant (P<0.05) after the sequential Bonferroni procedure are reported [52]. All tests were two-tailed.

All of the procedures were carried out according to institutional guidelines and in accordance with the U.K. Animals (Scientific Procedures) Act of 1986.

RESULTS

Behavioural and physiological responses to an unfamiliar conspecific

Presentation of an unfamiliar conspecific elicited a behavioral response; on average, individuals approached the intruder 8 times and attacked 11 times within the observation period. However, individuals differed in their behavioral reaction to the simulated intrusion; while one individual attacked the conspecific over 40 times, other individuals spent most of their time

hiding, and scarcely left the refuge. Body size explained some of this individual variation; bigger fish were more aggressive toward their size-matched opponents (number of attacks: r=0.433, P=0.024, n=27). All of the fish oriented to and approached the conspecific and one-half of the fish attacked it at least once.

Interaction with the unfamiliar conspecific quickly produced a glucocorticoid response (Figure 1). Whole-body cortisol levels were highest 15 minutes after the simulated intrusion and then returned to baseline levels by 30 minutes.

The serotonergic system was quickly suppressed in response to the presence of the unfamiliar conspecific, as indicated by reduced whole-brain levels of 5-HT (Figure 2A, Table 1).

Dopamine turnover to DOPAC was elevated 60 minutes following the aggressive interaction (Figure 2C and 2D), while levels of norepinephrine were consistently low (Figure 2F).

Individual differences in concentrations of brain monoamines were related to differences among individuals in aggressiveness. Individuals with lower hypothalamic 5HT were more aggressive (r=-0.806, P=0.016, n=8, Figure 3A), while norepinephrine (r=0.883, P=0.020, n=6, Figure 3B) and DOPAC (r=0.815, P=0.048, n=6, Figure 3C) were positively associated with aggressiveness.

Behavioural and physiological responses to a predator

When presented with the pike, most individuals inspected the predator at least once and oriented to it more than nine times. As in the 'conspecific' treatment, individuals differed in their behavior: some individuals inspected the pike as many as seven times during the 15-minute observation period, while others spent the entire observation period hiding in the refuge.

Exposure to the predator elicited a significant glucocorticoid response within 15 minutes which reached a maximum 60 minutes after exposure to the predator (Figure 1). Concentrations of DOPAC fell at 60 minutes (Figure 2D) while concentrations of HVA increased at 15 minutes (Figure 2E), indicating that predator-induced stress stimulated the rapid turnover of DA to HVA.

Activity under predation risk and predator inspection behavior (both of which potentially involve a risk of predation) were positively associated with neurotransmitter concentrations. For example, individuals with greater levels of NE engaged in riskier behavior (r=0.766, P=0.027, n=8, Figure 4A). Serotonin turnover was also associated with predator inspection behavior: the number of predator inspections was significantly positively correlated with hypothalamic serotonin (r=0.928, P=0.003, n=7, Figure 4B) and negatively correlated with whole-brain serotonergic activity (r=-0.669, P=0.049, n=9, Figure 4C).

Comparing responses to the conspecific and predator

Both confrontation by a conspecific and exposure to a predator elicited a cortisol response, but the time course of the cortisol response differed between treatments (Figure 1), as evidenced by the significant interaction between time and treatment ($F_{2,58}$ =5.5, P=0.006). Moreover, the magnitude (average across the three time periods) of the cortisol response was greater to the predator compared to a conspecific (Conspecific: 47 ± -4.97 ng/g, P=0.002).

Relative to the conspecific treatment, NE (Figure 2F) and to a lesser extent, DA (Figure 2C) were higher in the predator treatments.

DISCUSSION

In this experiment, we tested the hypothesis that both the HPI axis and brain monoaminergic systems are activated in response to fighting with an unfamiliar conspecific and exposure to a predator. While other studies have found links between these systems in laboratory animals, the results from this study extends these findings to wild-caught animals that were confronted by ecologically relevant challenges [28,53]. We found that both stressors elicited a similar HPI response, but produced very different patterns of change in monoamine content.

Our design permitted us to determine the time course of the neuroendocrine response to these stressors and to ascertain whether individual differences in behavioral responses to the stressors were related to underlying physiology. We showed that not only do these challenges elicit a neuroendocrine response, but that different behavioral responses of individuals were related to their particular neuroendocrine profiles.

The cortisol response to a conspecific and predator were broadly similar, but exposure to a predator was more stressful

During the present study, both confrontation with an unfamiliar conspecific and exposure to a predator resulted in activation of the HPI axis and significant alterations in the levels of brain monoamines in sticklebacks. These results are consistent with other studies which have shown that both confrontation by a challenging conspecific [10,23] and exposure to a predator [54] elicit a neuroendocrine stress response in fishes.

In the present study both exposure to a conspecific or to a predator resulted in highly significant increases in whole-body cortisol concentrations within 15 minutes relative to controls. In the conspecific-exposed group, whole-body cortisol levels were statistically indistinguishable from control fish after 30 minutes and remained so at 60 minutes. In contrast, whole-body cortisol concentrations in the predator-exposed group remained highly elevated after 60 minutes, significantly exceeding levels attained after 15 minutes. We interpret these data to indicate that the magnitude of the initial response to both stressors was similar, resulting in similar whole-body cortisol concentrations at 15 minutes, but that the HPI axis in the predator-exposed fish remained active for longer, resulting in a greater accumulation of whole-body cortisol with time. The overall significant difference in total cortisol between the two treatment groups detected across all time points indicates a quantitative difference in the response of the fish to the two stressors.

Other studies have found evidence for a more rapid recovery to baseline cortisol levels following less threatening situations compared to more threatening situations [55]. A longer-lasting cortisol response to threat of predation as compared to other stressors has been documented in stonechats [56] and rodents [57,58]. Therefore in this experiment, we hypothesize that the different time course of the cortisol response to a competitor versus to a predator is related to the perceived magnitude of the two different challenges. Sticklebacks are social fish, and frequently interact with other sticklebacks in shoals. Because encounters with conspecifics are frequent, natural selection might have favored individuals which do not mount a severe stress response to frequent interactions with conspecifics, and should favor individuals which recover quickly from fights. In contrast, encounters with predators are less frequent and more threatening

than encounters with conspecifics, so selection might have favored individuals with a greater and longer-lasting stress response.

The levels of whole-body cortisol detected in unstressed sticklebacks during the present study were similar to those previously reported for this species (2 – 8 ng g⁻¹; [46]) and levels detected in the stressed fish in the present study, although slightly higher, were also broadly consistent with previous observations (50 ng g⁻¹;[46]). The difference in magnitude of whole-body cortisol levels between this and previous studies may be related to the nature of the stressor.

Links between stress-induced blood cortisol levels and behavioral traits have been shown in fish [10,23], mammals [59] and reptiles [9]. However, while exposure to both stressors elicited a behavioral and whole-body cortisol response in the treatment groups, we did not detect a relationship at the individual level between concentrations of whole-body cortisol and behavior. It is possible that our method might not have had the resolution to detect fine-scale individual differences.

We did not detect any sex differences in whole-body cortisol. The stress response in vertebrates, including fish [60], is modulated by gonadal steroids with androgens suppressing and estrogens enhancing corticosteroid responsiveness [61]. However, the fish employed in this study were not reproductively active and it is therefore unsurprising that no sex-dependent differences in stress response were observed.

The monoamine responses to a conspecific and a predator were qualitatively different

Whereas the cortisol response was broadly similar across stressors, the monoamines showed a differential response across the two stressors, some being suppressed in response to a conspecific but elevated in response to the predator.

For example, relative to the control group, concentrations of NE were consistently *higher* in the 'predator' treatments, and *lower* in the 'conspecific' treatments. Without data on the NE metabolite, MHPG, we cannot distinguish if reduced concentrations reflect a reduction in NE release (decrease in NE activity) or an increased turnover to MHPG (increase in NE activity). However, at an individual level we found that NE was consistently associated with risk-taking behaviors in both kinds of situations: NE was positively correlated with aggressive behaviors as well as predator inspection behaviors. These positive correlations suggest that more bold or aggressive individuals were more 'aroused', active or uninhibited, results which are consistent with other studies showing positive relationships between NE activity and behavioral impulsivity in monkeys [28] and sensation seeking in humans [62]. The fact that serotonin and NE had opposite relationships with risk-taking behaviors in this experiment is consistent with the observation that 5-HT and catecholamines can have antagonistic effects on behavior [17].

Associations between serotonin, risk-taking behaviors and aggression

In agreement with other studies which have shown that risk-taking behaviors are negatively associated with brain serotonergic activity [24,27-29], we found that risk-taking behaviors performed while under predation risk (e.g. inspection) were negatively correlated with serotonin turnover to 5-HIAA (Figure 4C).

Our results support the view that 5-HT has an inhibitory effect on aggressive behavior [16,54]. We found a negative relationship at the individual level between concentrations of 5-HT and aggressive behavior, and that confrontation by an unfamiliar conspecific resulted in lower 5-HT. Other studies have shown that winners of agonistic interactions have up-regulated brain 5-HT activity [21,30-32]. One possible explanation for this different pattern is that in our experiment, there was no physical contact between the resident and the intruder because the intruders were confined to a flask. As a result, the resident fish were unable to complete their attacks and therefore might not be analogous to the winners in the forementioned studies. We remain provisional in our interpretation of these results because 5-HIAA was degraded in many of the samples in the 'conspecific' treatments, preventing us from calculating serotonin turnover in those treatments. However, it is worth noting that while more aggressive behaviors were negatively associated with serotonin (Figure 3A), risk-taking behavior under predation risk showed the opposite pattern – it was *positively* correlated with 5HT (Figure 4B), and negatively associated with serotonin turnover to 5-HIAA (Figure 4C).

Overall, these data provide evidence that the response of fish to stressors is not identical regardless of the nature of the challenge, but rather that the response varies according to the magnitude, frequency and predictability of the stressor, as is the case for other vertebrates [56,63]. Further studies on individual variation in responses to different stressors would benefit from repeated sampling of the same physiological measures on the same individuals. While it is currently a challenge to measure brain monoamines noninvasively, noninvasive methods for measuring glucocorticoids in fish [64] are a promising alternative. In addition, the roles played by upstream elements of the stress response such as corticotropin releasing hormone (CRH) and variation in the binding characteristics of corticosteroid receptors and corticotropin binding

proteins should also be investigated [65]. Given that other studies have shown that interindividual differences in stress responsiveness have a high heritable component [66], further investigation will provide insight into the mechanisms that have produced adaptive, heritable behavioral variation in sticklebacks in diverse ecological settings.

ACKNOWLEDGEMENTS

We thank Stuart Wilson, David Alvarez, Susie Coyle for help with the fish, Andy Young for catching the pike, Per-Ove Thörnqvist and Joachim Schjolden for technical assistance with HPLC and brain punches, Kim Pulman for technical help with the cortisol RIA. Funding was provided by an NSF International Postdoctoral Research Fellowship to AMB. AMB was supported by a fellowship from the American Association of University Women during preparation of the manuscript. Work in the Winberg lab was supported by the Swedish Research Council for Environment, Agricultural Sciences and Spatial Planning (FORMAS).

REFERENCES

- 1. Thorpe, K.E.; A.C. Taylor; F.A. Huntingford. How costly is fighting? Physiological effects of sustained exercise and fighting in swimming crabs, *Necora puber*. Animal Behaviour. 1995, 50: 1657-1666.
- Neat, F.C.; A.C. Taylor; F.A. Huntingford. Proximate costs of fighting in male cichlid fish: the role of injuries and energy metabolism. Animal Behaviour. 1998, 55: 875-882.

- 475 3. Diaz-Uriarte, R. Anti-predator behaviour changes following an aggressive encounter in
- 476 the lizard *Tropidurus hispidus*. Proceedings of the Royal Society of London Series B-
- 477 Biological Sciences. 1999, 266(1437): 2457-2464.
- 478 4. Ydenberg, R.C.; L.M. Dill. The economics of fleeing from predators. Advances in the
- 479 Study of Behavior. 1986, 16: 229-249.
- 480 5. Lin, Z.H.; Y.F. Qu; X. Ji. Energetic and locomotor costs of tail loss in the Chinese skink,
- 481 Eumeces chinensis. Comparative Biochemistry and Physiology a-Molecular &
- 482 Integrative Physiology. 2006, 143(4): 508-513.
- 483 6. Elofsson, U.O.E.; L. Mayer; B. Damsgard; S. Winberg. Intermale competition in sexually
- mature arctic charr: Effects on brain monoamines, endocrine stress responses, sex
- hormone levels, and behavior. General and Comparative Endocrinology. 2000, 118(3):
- 486 450-460.
- Fox, H.E.; S.A. White; M.H.F. Kao; R.D. Fernald. Stress and dominance in a social fish.
- 488 Journal of Neuroscience. 1997, 17(16): 6463-6469.
- 489 8. Winberg, S.; O. Lepage. Elevation of brain 5-HT activity, POMC expression, and plasma
- 490 cortisol in socially subordinate rainbow trout. American Journal of Physiology-
- Regulatory Integrative and Comparative Physiology. 1998, 43(3): R645-R654.
- 492 9. Summers, C.H.; M.J. Watt; T.L. Ling; G.L. Forster; R.E. Carpenter; W.J. Korzan; J.L.
- Lukkes; O. Overli. Glucocorticoid interaction with aggression in non-mammalian
- vertebrates: Reciprocal action. European Journal of Pharmacology. 2005, 526(1-3): 21-
- 495 35.

- 496 10. Sloman, K.A.; N.B. Metcalfe; A.C. Taylor; K.M. Gilmour. Plasma cortisol
- concentrations before and after social stress in rainbow trout and brown trout.
- 498 Physiological and Biochemical Zoology. 2001, 74: 383-389.
- 499 11. Knapp, R.; M.C. Moore. Hormonal responses to aggression vary in different types of
- agonistic encounters in male tree lizards. Hormones and Behavior. 1995, 29: 85-105.
- 501 12. Cockrem, J.F.; B. Silverin. Sight of a predator can stimulate a corticosterone response in
- the great tit. General and Comparative Endocrinology. 2002, 125: 248-255.
- 503 13. Scheuerlein, A.; T.J. Van't Hof; E. Gwinner. Predators as stressors? Physiological and
- reproductive consequences of predation risk in tropical stonechats. Proceedings of the
- Royal Society Biological Sciences Series B. 2001, 268: 1575-1582.
- 506 14. Eilam, D.; T. Dayan; S. Ben-Eliyahu; I. Schulman; G. Shefer; C.A. Hendries. Differential
- behavioural and hormonal responses of voles and spiny mice to owl calls. Animal
- 508 Behaviour. 1999, 58: 1085-1093.
- 509 15. Winberg, S.; G.E. Nilsson. Roles of brain monoamine neurotransmitters in agonistic
- behaviour and stress reactions, with particular reference to fish. Comparative
- Biochemistry and Physiology C Comparative Pharmacology and Toxicology. 1993,
- 512 106(3): 597-614.
- 513 16. Winberg, S.; A. Nilsson; P. Hylland; V. Soderstrom; G.E. Nilsson. Serotonin as a
- regulator of hypothalamic-pituitary-interrenal activity in teleost fish. Neuroscience
- 515 Letters. 1997, 230(2): 113-116.
- 516 17. Johnsson, J.I.; S. Winberg; K.A. Sloman. Social interactions. In: K.A. Sloman, R.W.
- Wilson, and S. Balshine, Behavior: Interactions with fish physiology. A volume of Fish
- Physiology, Editor^Editors. 2006, Elsevier.

- 519 18. Campbell, T.; S. Lin; C. DeVries; K. Lambert. Coping strategies in male and female rats
- exposed to multiple stressors. Physiology and Behavior. 2003, 78: 495-504.
- 521 19. Ebner, K.; C.T. Wotjak; R. Landgraf; M. Engelmann. Neuroendocrine and behavioral
- response to social confrontation: residents versus intruders, active versus passive coping
- 523 styles. Hormones and Behavior. 2005, 47: 14-21.
- 524 20. Clement, T.S.; V. Parikh; M. Schrumph; R.D. Fernald. Behavioral coping strategies in a
- 525 cichlid fish: the role of social status and acute stress response in direct and displaced
- aggression. Hormones and Behavior. 2005, 47: 336-342.
- 527 21. Øverli, O.; C. Sorensen; G.E. Nilsson. Behavioral indictors of stress-coping style in
- rainbow trout: do males and females react differently to novelty? Physiology and
- 529 Behavior. 2006, 87: 506-512.
- 530 22. Schjolden, J.; A. Stoskhus; S. Winberg. Does individual variation in stress responses and
- agonistic behavior reflect divergent stress coping strategies in juvenile rainbow trout?
- Physiological and Biochemical Zoology. 2005, 78: 715-723.
- 533 23. Øverli, O.; S. Winberg; T.G. Pottinger. Behavioral and neuroendocrine correlates of
- selection for stress responsiveness in rainbow trout a review. Integrative & Comparative
- 535 Biology. 2005, 45(3): 463-474.
- van Hierden, Y.M.; S.M. Korte; E.W. Ruesink; C.G. van Reenen; B. Engel; G.A.H.
- Korte-Bouws; J.M. Koolhaas; H.J. Blokhuis. Adrenocortical reactivity and central
- serotonin and dopamine turnover in young chicks from a high and low feather-pecking
- line of laying hens. Physiology & Behavior. 2002, 75(5): 653-659.

- 540 25. Ballenger, J.C.; R.M. Post; D.C. Jimerson; C.R. Lake; D. Murphy; M. Zuckerman; C.
- Cronin. Biochemical correlates of personality traits in normals an exploratory study.
- Personality and Individual Differences. 1983, 4(6): 615-625.
- 543 26. Depue, R.A. Neurobiological factors in personality and depression. European Journal of
- Personality. 1995, 9(5): 413-439.
- 545 27. Ferrari, P.F.; P. Palanza; S. Parmigiani; R.M.M. de Almeida; K.A. Miczek. Serotonin and
- aggressive behavior in rodents and nonhuman primates: Predispositions and plasticity.
- 547 European Journal of Pharmacology. 2005, 526(1-3): 259-273.
- 548 28. Fairbanks, L.A.; M.B. Fontenot; J.E. Phillips-Conroy; C.J. Jolly; J.R. Kaplan; J.J. Mann.
- CSF monoamines, age and impulsivity in wild grivet monkeys (*Cercopithecus aethiops*
- *aethiops*). Brain Behavior and Evolution. 1999, 53(5-6): 305-312.
- 551 29. Mehlman, P.T.; J.D. Higley; I. Faucher; A.A. Lilly; D.M. Taub; J. Vickers; S.J. Suomi;
- M. Linnoila. Low Csf 5-Hiaa concentrations and severe aggression and impaired impulse
- control in nonhuman primates. American Journal of Psychiatry. 1994, 151(10): 1485-
- 554 1491.
- 555 30. Korzan, W.J.; T.R. Summers; C.H. Summers. Monoaminergic activities of limbic regions
- are elevated during aggression: Influence of sympathetic social signaling. Brain
- 557 Research. 2000, 870(1-2): 170-178.
- 558 31. Summers, C.H.; T.R. Summers; M.C. Moore; W.J. Korzan; S.K. Woodley; P.J. Ronan; E.
- Hoglund; M.J. Watt; N. Greenberg. Temporal patterns of limbic monoamine and plasma
- corticosterone response during social stress. Neuroscience. 2003, 116: 553-563.
- 561 32. Summers, C.H.; W.J. Korzan; J.L. Lukkes; M.J. Watt; G.L. Forster; O. Øverli; E.
- Höglund; E.T. Larson; P.J. Ronan; J.M. Matter; T.R. Summers; K.J. Renner; N.

- Greenberg. Does serotonin influence aggression? Comparing regional activity before and
- during social interaction. Physiol Biochem Zool. 2005, 78: 679-694.
- 565 33. Lepage, O.; O. Tottmar; S. Winberg. Elevated dietary intake of l-tryptophan counteracts
- the stress-induced elevation of plasma cortisol in rainbow trout. Journal of Experimental
- 567 Biology. 2002, 205: 3679-3687.
- 568 34. Bell, A.M.; J.A. Stamps. The development of behavioural differences between
- individuals and populations of stickleback. Animal Behaviour. 2004, 68: 1339-1348.
- 570 35. Bell, A.M. Differences between individuals and populations of threespined stickleback.
- Journal of Evolutionary Biology. 2005, 18: 464-473.
- 572 36. Huntingford, F.A. The relationship between inter- and intra-specific aggression. Animal
- 573 Behaviour. 1976, 24: 485-497.
- 574 37. Sih, A.; A.M. Bell; J.C. Johnson. Behavioral syndromes: an ecological and evolutionary
- overview. Trends in Ecology & Evolution. 2004, 19(7): 372-378.
- 576 38. Sih, A.; A.M. Bell; J.C. Johnson; R. Ziemba. Behavioral syndromes: an integrative
- overview. Quarterly Review of Biology. 2004, 79: 241-277.
- 578 39. Koolhaas, J.M.; S.M. Korte; S.F. De Boer; B.J. Van Der Vegt; C.G. Van Reenen; H.
- Hopster; I.C. De Jong; M.A.W. Ruis; H.J. Blokhuis. Coping styles in animals: Current
- status in behavior and stress-physiology. Neuroscience & Biobehavioral Reviews. 1999,
- 581 23(7): 925-935.
- 582 40. Tamashiro, K.L.K.; M.M.N. Nguyen; R.R. Sakai. Social stress: From rodents to primates.
- 583 FRONTIERS IN NEUROENDOCRINOLOGY. 2005, 26: 27-40.
- 584 41. Bakker, T.C.M. Aggressiveness in sticklebacks (Gasterosteus aculeatus) a behavior-
- 585 genetic study. Behaviour. 1986, 98(1-4): 1-144.

- 586 42. Audet, C.; G.J. Fitzgerald; H. Guderley. Photoperiod effects on plasma cortisol levels in
- 587 Gasterosteus aculeatus. General and Comparative Endocrinology. 1986, 61(1): 76-81.
- 588 43. Pottinger, T.G.; T.A. Moran; J.A.W. Morgan. Primary and secondary indices of stress in
- the progeny of rainbow trout (*Oncorhynchus mykiss*) selected for high and low
- responsiveness to stress. Journal of Fish Biology. 1994, 44(1): 149-163.
- 591 44. Øverli, O.; T.G. Pottinger; T.R. Carrick; E. Øverli; S. Winberg. Brain monoaminergic
- activity in rainbow trout selected for high and low stress responsiveness. Brain Behavior
- 593 and Evolution. 2001, 57(4): 214-224.
- 594 45. Øverli, O.; T.G. Pottinger; T.R. Carrick; E. Øverli; S. Winberg. Differences in behaviour
- between rainbow trout selected for high- and low-stress responsiveness. Journal of
- 596 Experimental Biology. 2002, 205(3): 391-395.
- 597 46. Pottinger, T.G.; T.R. Carrick; W.E. Yeomans. The three-spined stickleback as an
- environmental sentinel: effects of stressors on whole-body physiological indices. Journal
- 599 of Fish Biology. 2002, 61(1): 207-229.
- 600 47. Pottinger, T.G.; T.A. Moran; P.A. Cranwell. The biliary accumulation of corticosteroids
- in rainbow trout, *Oncorhynchus mykiss*: a stable indicator of chronic stress. Fish
- Physiology and Biochemistry. 1992, 10: 55-66.
- 603 48. Pottinger, T.G.; E.M. Musowe. The corticosteroidogenic response of brown and rainbow
- trout alevins and fry during a 'critical' period. General and Comparative Endocrinology.
- 605 1994, 95: 350-362.
- 606 49. Ramsay, J.M.; G.W. Feist; Z.M. Varga; M. Westerfield; M.L. Kent; C.B. Schreck.
- Whole-body cortisol is an indicator of crowding stress in adult zebrafish, *Danio rerio*.
- 608 Aquaculture. 2006, 258(1-4): 565-574.

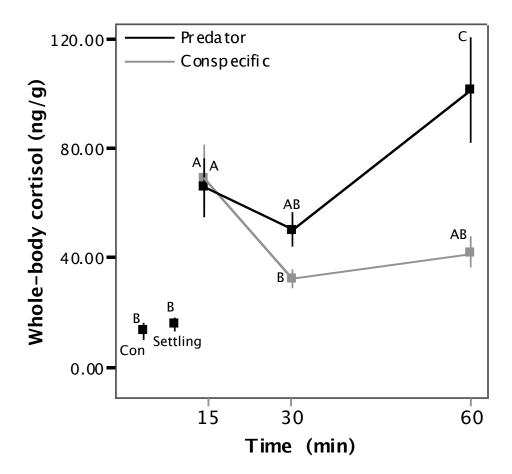
- 609 50. Øverli, O.; C.A. Harris; S. Winberg. Short-term effects of fights for social dominance and
- the establishment of dominant-subordinate relationships on brain monoamines and
- cortisol in rainbow trout. Brain Behavior and Evolution. 1999, 54: 263-275.
- 612 51. Peichel, C.; J. Ross; C. Matson; M. Dickson; J. Grimwood; J. Schmutz; R. Myers; S.
- Mori; D. Schluter; D. Kingsley. The master sex-determination locus in threespine
- sticklebacks in on a nascent Y chromosome. Current Biology. 2004, 14: 1416-1424.
- 615 52. Rice, W.R. Analyzing tables of statistical tests. Evolution. 1989, 43: 223-225.
- 616 53. Kaplan, J.R.; J. Phillips-Conroy; M.B. Fontenot; C.J. Jolly; L.A. Fairbanks; J.J. Mann.
- 617 Cerebrospinal fluid monoaminergic metabolites differ in wild anubis and hybrid (*Anubis*
- *hamadryas*) baboons: Possible relationships to life history and behavior.
- Neuropsychopharmacology. 1999, 20(6): 517-524.
- 620 54. Winberg, S.; A.A. Myrberg; G.E. Nilsson. Predator exposure alters brain serotonin
- metabolism in bicolour damselfish. Neuroreport. 1993, 4(4): 399-402.
- 622 55. Djordjevic, J.; G. Cvijic; V. Davidovic. Different activation of ACTH and corticosterone
- release in response to various stressors in rats. Physiological Research. 2003, 52: 67-72.
- 624 56. Canoine, V.; T.J. Hayden; K. Rowe; W. Goymann. The stress response of european
- stonechats depends on the type of stressor. Behaviour. 2002, 139: 1303-1311.
- 626 57. Blanchard, R.J.; J.N. Nikulina; R.S. Sakai; C. McKittrick; B. McEwen; D.C. Blanchard.
- Behavioral and endocrine change following chronic predatory stress. Physiology &
- 628 Behavior. 1998, 63: 561-569.
- 629 58. Adamec, R.E.; T. Shallow. Lasting effects on rodent anxiety of a single exposure to a cat.
- 630 Physiology and Behavior. 1993, 54: 101-109.

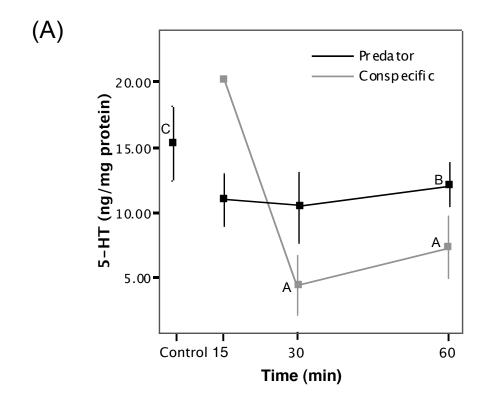
- 631 59. Steimer, T.; P. Driscoll. Divergent stress responses and coping styles in
- psychogenetically selected roman high-(RHA) and low-(RLA) avoidance rats:
- Behavioural, neuroendocrine and developmental aspects. Stress. 2003, 6: 87-100.
- 634 60. Pottinger, T.G.; P.H.M. Balm; A.D. Pickering. Sexual maturity modifies the
- responsiveness of the pituitary-interrenal axis to stress in male rainbow trout. General and
- 636 Comparative Endocrinology. 1995, 98: 311-320.
- 637 61. Pottinger, T.G.; T.R. Carrick; S.E. Hughes; P.H.M. Balm. Testosterone, 11-
- ketotestosterone and estradiol-17_ modify baseline and stress-induced interrenal and
- corticotropic activity in trout. General and Comparative Endocrinology. 1996, 104: 284-
- 640 295.
- 641 62. Gerra, G.; P. Avanzini; A. Zaimovic; R. Sartori; C. Bocchi; M. Timpano; U. Zambelli; R.
- Delsignore; F. Gardini; E. Talarico; F. Brambilla. Neurotransmitters, neuroendocrine
- correlates of sensation-seeking temperament in normal humans. Neuropsychobiology.
- 644 1999, 39(4): 207-213.
- 645 63. Silverin, B. Behavioural and hormonal responses of the pied flycatcher to environmental
- stressors. Animal Behaviour. 1998, 55: 1411-1420.
- 647 64. Scott, A.P.; M. Pinillos; T. Ellis. Why measure steroids in fish plasma when you can
- measure them in water? in *Perspectives in Comparative Endocrinology: Unity and*
- 649 Diversity. 14th International Congress of Comparative Endocrinology. 2001. Sorrento,
- 650 Italy: Monduzzi Editore S.p.A. -Medimond Inc.: Italy.
- 651 65. Sapolsky, R.M.; L.M. Romero; A.U. Munck. How do glucocorticoids influence stress
- responses? Integrating permissive, suppressive, stimulatory and preparative actions.
- Endocrine Reviews. 2000, 21(1): 55-89.

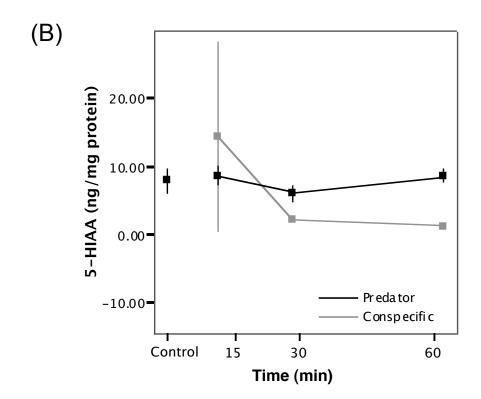
654 66. Pottinger, T.G.; T.R. Carrick. Modification of the plasma cortisol response to stress in rainbow trout by selective breeding. General and Comparative Endocrinology. 1999, 116(1): 122-132.

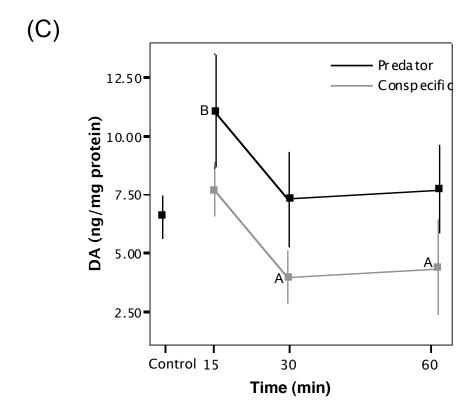
659 Figure legends 660 661 Figure 1. Whole-body cortisol in the different treatments. Statistically similar means share the 662 same letter. 663 Figure 2. Whole-brain concentrations of brain monoamines in different treatments. Statistically 664 665 similar means share the same letter. (A) 5-HT; (B) 5-HIAA; (C) DA; (D) DOPAC; (E) HVA; (F) 666 NE. 667 668 Figure 3. Correlations between monoamine concentrations and aggressive behavior (attacks). (A) 669 Hypothalamic 5-HT 60 minutes after a fight; (B) NE in reticular formation 15 minutes after a 670 fight; (C) Telencephalic DOPAC 30 minutes after a fight. 671 672 Figure 4. Correlations between monoamine concentrations and behavior under predation risk. 673 (A) Telencephalic NE 60 minutes after exposure and time orienting to the predator; (B) 674 Hypothalamic 5-HT 60 minutes after exposure and predator inspections; (C) Whole-brain 5-675 HIAA:5-HT ratio 15 minutes after exposure and predator inspections.

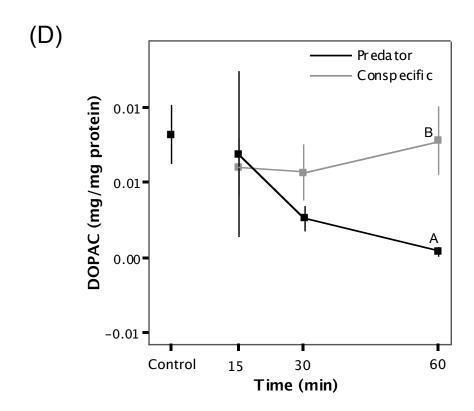
Figure 1

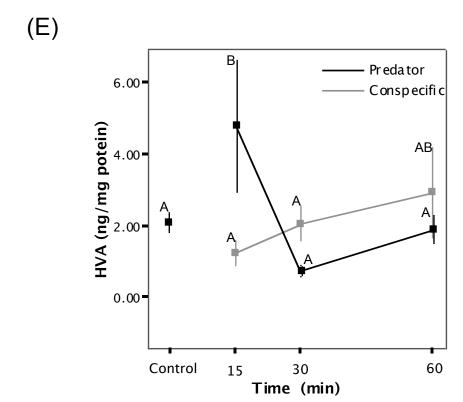




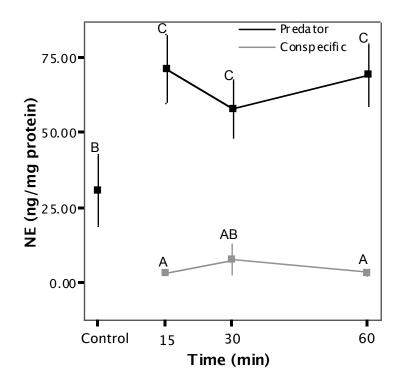


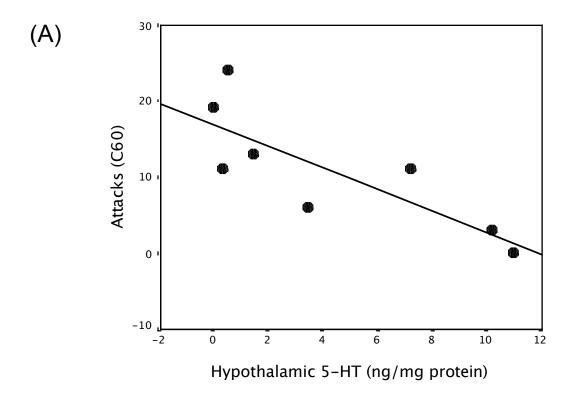


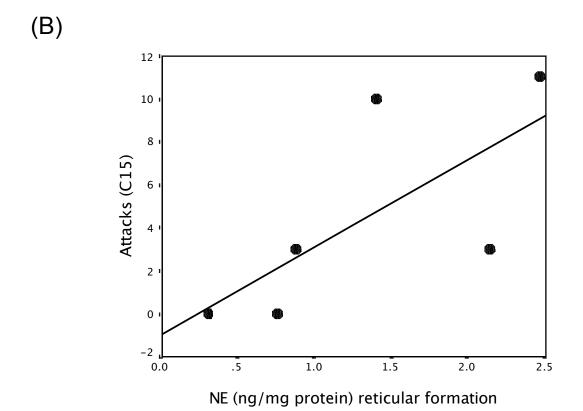




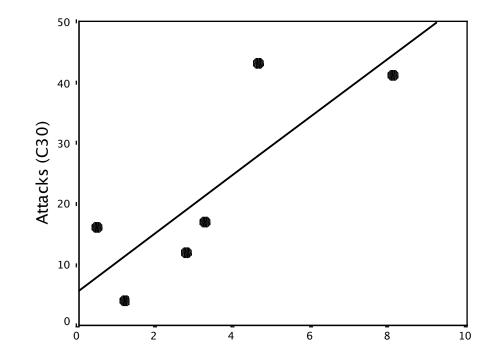






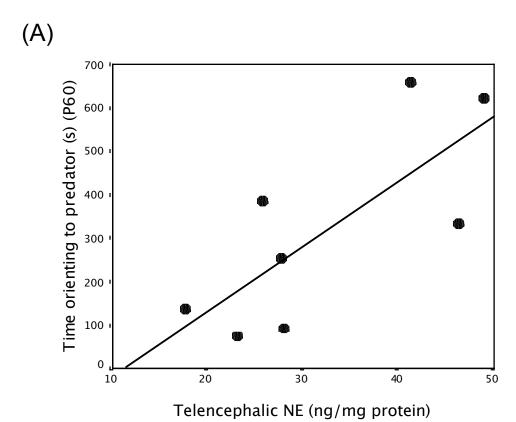


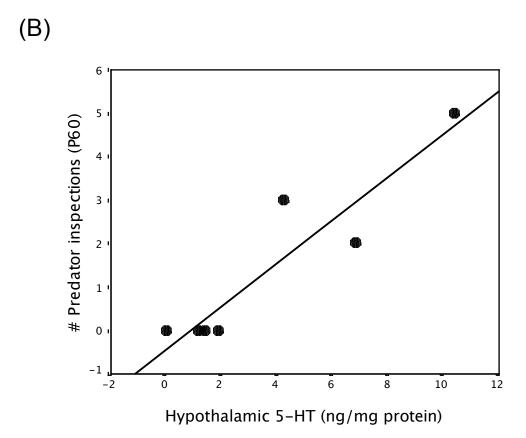




Telencephalic DOPAC (ng/mg protein)

Figure 4





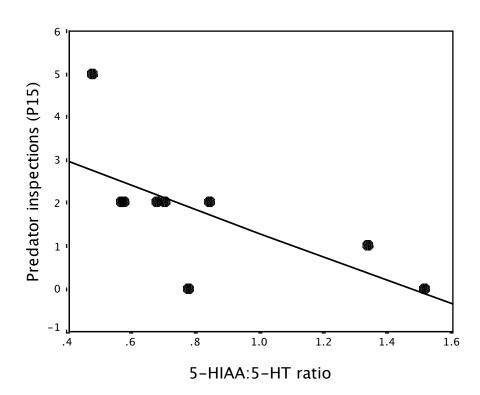


Table 1. Concentrations (ng/mg protein) of monoamines in the different brain regions for the different treatments. Statistics are presented as mean \pm sd. Sample sizes are in parentheses.

	NE	DOPAC	5-HIAA	DA	HVA	5-HT
HYPOTHALAM	US					
Control	10.48 ± 12.91(10)	$4.04 \pm 3.76(6)$	3.16 ± .737(4)	2.27 ± 1.57(10)	$0.84 \pm 0.50(10)$	5.83 ± 4.32(10
Conspecific						
15 min	1.16 ± 0.59(7)	3.60 ± 4.71(3)	und	3.16 ± 1.82(8)	0.78 ± .55(5)	und
30 min	0.84 ± 0.83(6)	3.03 ± 3.40(3)	und	2.04 ± 1.91(5)	1.00 ± 1.13(6)	2.27 ± 2.54(3)
60 min	1.44 ± 2.47(8)	2.65 ± 3.19(3)	und	4.43 ± 5.01(4)	2.68 ± 3.38(7)	4.26 ± 4.56(8)
Predator						
15 min	25.57 ± 15.63(10)	2.62 ± .94(6)	3.96 ± 1.77(9)	4.11 ± 4.07(10)	1.13 ± 1.34(10)	4.67 ± 2.48(10
30 min	21.36 ± 10.53(8)	4.24 ± .56(5)	2.07 ± 0.74(8)	2.92 ± 3.09(8)	0.21 ± 0.14(7)	4.81 ± 3.45(8)
60 min	27.86 ± 8.21(8)	1.07 ± 1.07(4)	3.33 ± 1.56(8)	3.36 ± 2.73(8)	0.58 ± 0.41(8)	3.75 ± 3.44(8)
RETICULAR FO	ORMATION					
Control	9.04 ± 11.20(10)	6.10 ± 2.73(6)	2.57 ± 1.41(4)	1.84 ± 1.08(9)	0.41 ± 0.17(10)	3.19 ± 2.45(10
Conspecific						
15 min	1.32 ± .84(6)	2.54 ± 2.47(8)	und	3.27 ± 1.58(6)	0.84 ± .49(5)	20.30 ± 0(1)
30 min	0.83 ± .56(6)	4.38 ± 3.24(5)	und	1.98 ± 0.80(7)	0.95 ± 0.53(7)	1.80 ± 1.54(3)
60 min	1.08 ± 0.71(8)	4.49 ± 4.18(7)	und	1.57 ± 1.21(4)	0.87 ± 0.69(8)	2.07 ± 2.36(8)
Predator						
15 min	21.51 ± 10.78(10)	und	2.06 ± .79(8)	2.26 ± 1.04(10)	1.45 ± 1.90(10)	2.11 ± 1.37(10
30 min	15.53 ± 6.40(8)	und	1.46 ± .66(8)	2.18 ± 2.14(8)	0.15 ± 0.14(7)	2.10 ± 1.83(8)
60 min	19.88 ± 8.26(9)	und	1.98 ± .70(8)	1.32 ± 1.23(9)	0.69 ± .96(8)	2.58 ± 1.45(8)
TELENCEPHAI	LON					
Control	11.44 ± 14.59(10)	3.60 ± 1.80(6)	3.43 ± 1.59(5)	2.63 ± 1.29(10)	0.84 ± 0.49(10)	6.36 ± 4.46(10
Conspecific						
15 min	1.36 ± 0.68(8)	2.14 ± 2.23(8)	14.39 ± 19.61(2)	2.11 ± 0.72(8)	0.64 ± .82(3)	und
30 min	7.60 ± 16.94(8)	3.41 ± 2.74(6)	2.12 ± 0(1)	1.31 ± 1.13(6)	0.75 ± 0.53(8)	5.15 ± 2.10(2)
60 min	1.54 ± 2.17(8)	6.48 ± 2.88(6)	1.24 ± 0(1)	1.68 ± 2.37(4)	0.45 ± 0.71(8)	1.92 ± 3.56(10
Predator		. ,	•			•
	24.18 ± 12.70(10)	53.64 ± 0(1)	3.01 ± 2.40(9)	4.71 ± 3.61(10)	2.20 ± 2.59(10)	4.75 ± 3.42(9)
	21.07 ± 14.45(8)	und	2.53 ± 2.79(8)	2.20 ± 1.41(8)	0.42 ± 0.26(8)	3.52 ± 3.93(8)
	29.24 ± 14.30(10)	0.37 ± 0(1)	4.04 ± 2.38(9)	3.87 ± 2.77(10)	0.86 ± 0.71(10)	6.59 ± 4.18(9)