

DATA NOTE

The genome sequence of the Lunar Hornet, *Sesia*bembeciformis (Hübner 1806) [version 1; peer review: 2

approved, 1 approved with reservations]

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v1

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Abstract

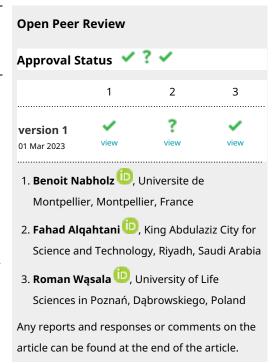
We present a genome assembly from an individual male *Sesia bembeciformis* (the Lunar Hornet; Arthropoda; Insecta; Lepidoptera; Sesiidae). The genome sequence is 477.1 megabases in span. Most of the assembly is scaffolded into 31 chromosomal pseudomolecules, including the Z sex chromosome. The mitochondrial genome has also been assembled and is 16.1 kilobases in length. Gene annotation of this assembly on Ensembl has identified 15,843 protein coding genes.

Keywords

Sesia bembeciformis, Lunar Hornet, genome sequence, chromosomal, Lepidoptera



This article is included in the Tree of Life gateway.



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Author roles: Boyes D: Investigation, Resources; Langdon WBV: Writing – Original Draft Preparation, Writing – Review & Editing;

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Species taxonomy

Eukaryota; Metazoa; Ecdysozoa; Arthropoda; Hexapoda; Insecta; Pterygota; Neoptera; Endopterygota; Lepidoptera; Glossata; Ditrysia; Sesioidea; Sesiidae; Sesiinae; Sesiin; Sesia; Sesia bembeciformis (Hübner 1806) (NCBI:txid287197).

Background

The Lunar Hornet, Sesia bembeciformis (Hübner 1806), is a moth of the family Sesiidae, commonly known as clearwings. This group are largely diurnal and named for their mostly scale-less, elongated wings which together with the banded, elongated abdomens of many species, contribute to their Batesian mimicry of aposematic Hymenoptera. S. bembeciformis is a relatively large species, with a wingspan of 31 to 48 mm (Lastuvka & Lastuvka, 2001) and a broad abdomen with alternating yellow and black bands that enable it to mimic hornets (Vespa spp).

Sesia bembeciformis feeds mainly on willows (Salix spp.) particularly S. caprea and S. cinerea (Goossens, 2020; Henwood et al., 2020; Lastuvka & Lastuvka, 2001). Some authors also state that the species is occasionally found on Populus spp. (Henwood et al., 2020; St John, 1890), which are the main foodplants of the related S. apiformis (which is also occasionally found on Salix spp.) (Lastuvka & Lastuvka, 2001). S. bembeciformis can be encountered in a variety of habitats where these foodplants occur, and may prefer more open areas with scattered trees, in sunny positions (Goossens, 2020). In the past it has occasionally been considered a pest of commercial willow crops – (Johnson, 1874) wrote of it 'doing considerable injury to the osier beds'.

In common with other clearwing species, S. bembeciformis larvae are rather nondescript, with creamy-white bodies, brown heads and prothoraic plates, and internal development on the foodplants (Henwood et al., 2020). Eggs are laid in small batches on the bark near the base of the tree, and larvae enter the trunk, often through a wound, where they feed initially on the sap (Goossens, 2020). They can take several years to develop, hibernating over winter and moving deeper into the trunk, often down into the roots. Before their final hibernation, larvae prepare an exit hole in the trunk (leaving a thin layer of bark to cover it) before spinning a loose cocoon above it in spring and emerging in mid-summer. Adults are on the wing in June and July (Randle et al., 2019) and usually emerge early in the morning where they can be found resting on trunks above their emergence hole before engaging in active flight by day.

Males can be attracted to synthetic lures which mimic the sex pheromones produced by females, mostly during the morning before midday (Pühringer, 2021). These lures were only developed for this species relatively recently, being released commercially in 2020. They provide a quick and efficient method for recording the species and have revolutionised understanding of its distribution, as have other lures developed for other clearwing species, e.g. (Burman et al., 2016). Many clearwing species are rarely observed as adults without lures,

due to their similarity to the Hymenoptera they mimic, and the fact that they fly by day, but many (like *S. bembecifo*rmis) have reduced mouth-parts and do not visit flowers.

The main method for recording many species before the advent of lures has therefore been to search for the well-hidden larvae, and dedicated surveys for *S. bembeciformis* larval exit holes have revealed it from areas where it was previously entirely unknown (Crégu, 2015; Goossens, 2020; Lastuvka & Lastuvka, 2014), and in some cases showed it to be very common. In the UK, it is the most widespread clearwing species, known from all areas (Waring *et al.*, 2017). Outside the UK it has a limited, mostly western European distribution (Lastuvka & Lastuvka, 2001).

The genome of *Sesia bembeciformis* was sequenced as part of the Darwin Tree of Life Project, a collaborative effort to sequence all named eukaryotic species in the Atlantic Archipelago of Britain and Ireland. Here we present a chromosomally complete genome sequence for *Sesia bembeciformis*, based on one male specimen from Wytham Woods, Oxfordshire, UK.

Genome sequence report

The genome was sequenced from one male *S. bembeciformis* (Figure 1) collected from Wytham Woods, Oxfordshire (latitude 51.775, longitude –1.315). A total of 27-fold coverage in Pacific Biosciences single-molecule HiFi long was generated. Primary assembly contigs were scaffolded with chromosome conformation Hi-C data. Manual assembly curation corrected four missing joins or mis-joins and removed ten haplotypic duplications, reducing the scaffold number by 18.52%.

The final assembly has a total length of 477.1 Mb in 44 sequence scaffolds with a scaffold N50 of 17.4 Mb (Table 1). Most (99.99%) of the assembly sequence was assigned to 31 chromosomal-level scaffolds, representing



Figure 1. Photograph of the Sesia bembeciformis (ilSesBemb1) specimen used for genome sequencing.

30 autosomes and the Z sex chromosome. Chromosome-scale scaffolds confirmed by the Hi-C data are named in order of size. (Figure 2–Figure 5; Table 2). The assembly has a BUSCO v5.3.2 (Manni *et al.*, 2021) completeness of 98.7%

(single 98.2%, duplicated 0.5%) using the lepidoptera_odb10 reference set. While not fully phased, the assembly deposited is of one haplotype. Contigs corresponding to the second haplotype have also been deposited.

Table 1. Genome data for Sesia bembeciformis, ilSesBemb1.1.

Project accession data			
Assembly identifier	ilSesBemb1.1		
Species	Sesia bembeciformis		
Specimen	ilSesBemb1		
NCBI taxonomy ID	287197		
BioProject	PRJEB52659		
BioSample ID	SAMEA10978929		
Isolate information	male, ilSesBemb1 (PacBio and Hi-C) male, ilSesBemb2 (RNA-Seq)		
Assembly metrics*		Benchmark	
Consensus quality (QV)	64.7	≥ 50	
k-mer completeness	100%	≥ 95%	
BUSCO**	C:98.7%[S:98.2%,D:0.5%], F:0.3%,M:1.0%,n:5,286	C ≥ 95%	
Percentage of assembly mapped to chromosomes	99.99%	≥ 95%	
Sex chromosomes	Z chromosome	localised homologous pairs	
Organelles	Mitochondrial genome assembled	complete single alleles	
Raw data accessions			
PacificBiosciences SEQUEL II	ERR9793194		
Hi-C Illumina	ERR9710923		
PolyA RNA-Seq Illumina	ERR10123699		
Genome assembly			
Assembly accession	GCA_943735995.1		
Accession of alternate haplotype	GCA_943735985.1		
Span (Mb)	477.1		
Number of contigs	51		
Contig N50 length (Mb)	17.1		
Number of scaffolds	44		
Scaffold N50 length (Mb)	17.4		
Longest scaffold (Mb)	23.1		
Genome annotation			
Number of protein-coding genes	15,843		
Number of gene transcripts	16,010		

^{*} Assembly metric benchmarks are adapted from column VGP-2020 of "Table 1: Proposed standards and metrics for defining genome assembly quality" from (Rhie et al., 2021).

^{**} BUSCO scores based on the lepidoptera_odb10 BUSCO set using v5.3.2. C = complete [S = single copy, D = duplicated], F = fragmented, M = missing, n = number of orthologues in comparison. A full set of BUSCO scores is available at https://blobtoolkit.genomehubs.org/view/ilSesBemb1.1/dataset/CALSEX01/busco.

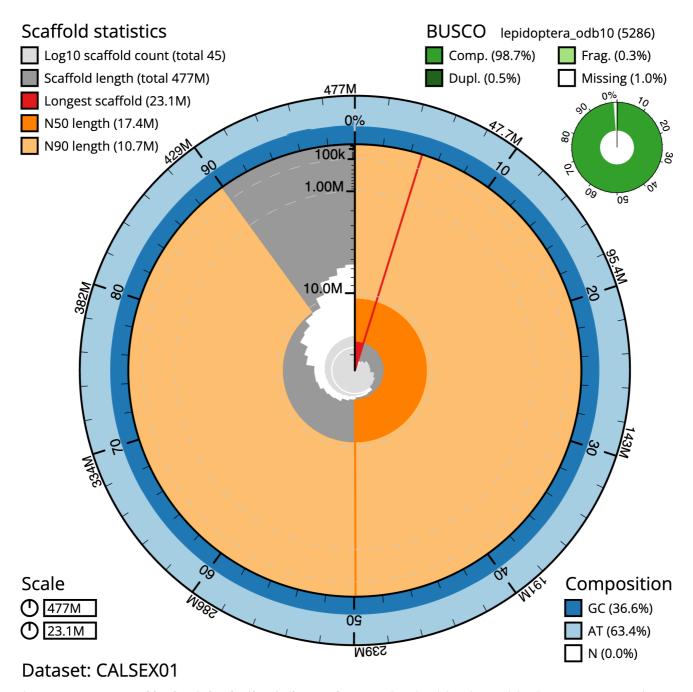


Figure 2. Genome assembly of *Sesia bembeciformis*, **ilSesBemb1.1: metrics.** The BlobToolKit Snailplot shows N50 metrics and BUSCO gene completeness. The main plot is divided into 1,000 size-ordered bins around the circumference with each bin representing 0.1% of the 477,151,446 bp assembly. The distribution of scaffold lengths is shown in dark grey with the plot radius scaled to the longest scaffold present in the assembly (23,076,235 bp, shown in red). Orange and pale-orange arcs show the N50 and N90 scaffold lengths (17,397,286 and 10,737,896 bp), respectively. The pale grey spiral shows the cumulative scaffold count on a log scale with white scale lines showing successive orders of magnitude. The blue and pale-blue area around the outside of the plot shows the distribution of GC, AT and N percentages in the same bins as the inner plot. A summary of complete, fragmented, duplicated and missing BUSCO genes in the lepidoptera_odb10 set is shown in the top right. An interactive version of this figure is available at https://blobtoolkit.genomehubs.org/view/ilSesBemb1.1/dataset/CALSEX01/snail.

Genome annotation report

The S. bembeciformis GCA_943735995.1 genome assembly was annotated using the Ensembl rapid annotation pipeline

(Table 1; https://rapid.ensembl.org/Sesia_bembeciformis_GCA_943735995.1/). The resulting annotation includes 16,010 transcribed mRNAs from 15,843 protein-coding genes.

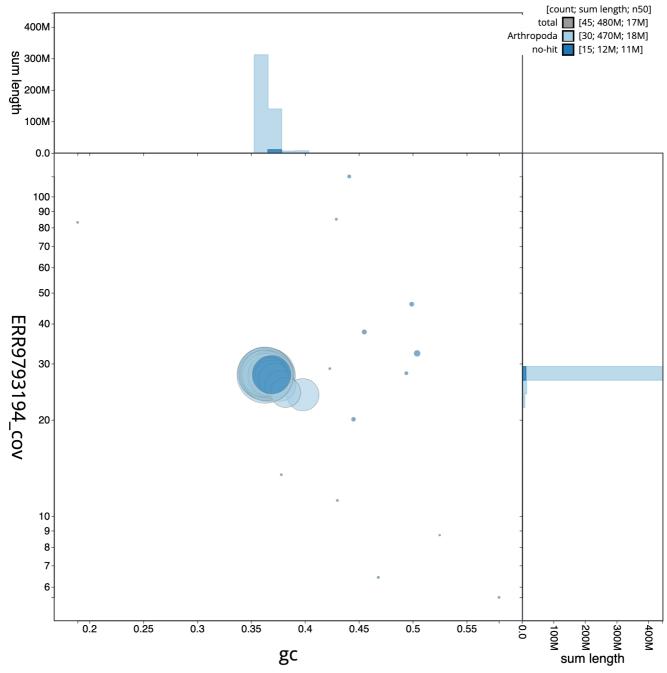


Figure 3. Genome assembly of *Sesia bembeciformis***, ilSesBemb1.1: GC coverage.** BlobToolKit GC-coverage plot. Scaffolds are coloured by phylum. Circles are sized in proportion to scaffold length. Histograms show the distribution of scaffold length sum along each axis. An interactive version of this figure is available at https://blobtoolkit.genomehubs.org/view/ilSesBemb1.1/dataset/CALSEX01/blob.

Methods

Sample acquisition and nucleic acid extraction

Two male *Sesia bembeciformis* specimens (ilSesBemb1 and ilSesBemb2) were collected from Wytham Woods, Oxfordshire (biological vice-county Berkshire) (latitude 51.775, longitude –1.315) on 17 July 2021. The specimens were taken from an orchard by Douglas Boyes (University of Oxford)

using a *pheromone* lure. The specimens were identified by Douglas Boyes and snap-frozen on dry ice.

DNA was extracted at the Tree of Life laboratory, Wellcome Sanger Institute (WSI). The ilSesBemb1 sample was weighed and dissected on dry ice with tissue set aside for Hi-C sequencing. Abdomen tissue was cryogenically disrupted

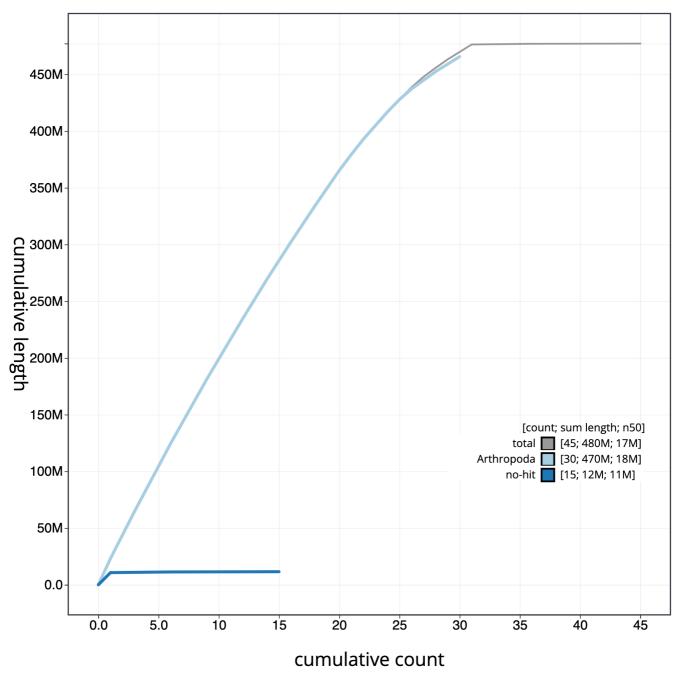


Figure 4. Genome assembly of *Sesia bembeciformis, ilSesBemb1.1: cumulative sequence.* BlobToolKit cumulative sequence plot. The grey line shows cumulative length for all scaffolds. Coloured lines show cumulative lengths of scaffolds assigned to each phylum using the buscogenes taxrule. An interactive version of this figure is available at https://blobtoolkit.genomehubs.org/view/ilSesBemb1.1/dataset/CALSEX01/cumulative.

to a fine powder using a Covaris cryoPREP Automated Dry Pulveriser, receiving multiple impacts. High molecular weight (HMW) DNA was extracted using the Qiagen MagAttract HMW DNA extraction kit. HMW DNA was sheared into an average fragment size of 12–20 kb in a Megaruptor 3 system with speed setting 30. Sheared DNA was purified by

solid-phase reversible immobilisation using AMPure PB beads with a 1.8X ratio of beads to sample to remove the shorter fragments and concentrate the DNA sample. The concentration of the sheared and purified DNA was assessed using a Nanodrop spectrophotometer and Qubit Fluorometer and Qubit dsDNA High Sensitivity Assay kit. Fragment size

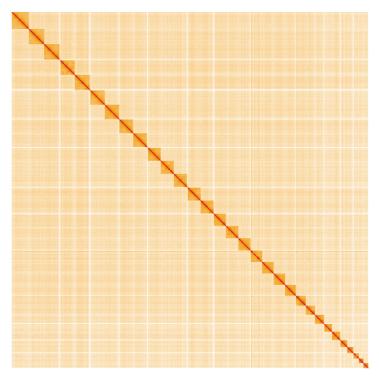


Figure 5. Genome assembly of *Sesia bembeciformis***, ilSesBemb1.1: Hi-C contact map.** Hi-C contact map of the ilSesBemb1.1 assembly, visualised using HiGlass. Chromosomes are shown in order of size from left to right and top to bottom. An interactive version of this figure may be viewed at https://genome-note-higlass.tol.sanger.ac.uk/l/?d=UU22HbFPTsSHjXIUIZh37w.

Table 2. Chromosomal pseudomolecules in the genome assembly of *Sesia bembeciformis*, ilSesBemb1.

INSDC accession	Chromosome	Size (Mb)	GC%
OX031025.1	1	20.93	36.6
OX031026.1	2	20.62	36.3
OX031027.1	3	20.06	36.5
OX031028.1	4	19.97	36.4
OX031029.1	5	19.91	36.2
OX031030.1	6	19.17	36.2
OX031031.1	7	18.84	36.1
OX031032.1	8	18.69	36.3
OX031033.1	9	17.98	36.5
OX031034.1	10	17.83	36.2
OX031035.1	11	17.81	36.2
OX031036.1	12	17.4	36.3
OX031037.1	13	17.14	36.4
OX031038.1	14	16.6	36.4
OX031039.1	15	16.42	36.2
OX031040.1	16	16.21	36.3

INSDC accession	Chromosome	Size (Mb)	GC%
OX031041.1	17	15.71	36.7
OX031042.1	18	15.5	36.7
OX031043.1	19	15.44	36.6
OX031044.1	20	14.08	36.3
OX031045.1	21	13.46	36.7
OX031046.1	22	12.15	37.2
OX031047.1	23	11.89	36.6
OX031048.1	24	10.94	36.7
OX031049.1	25	10.74	36.9
OX031050.1	26	9.36	36.8
OX031051.1	27	7.49	39.8
OX031052.1	28	7.8	37.2
OX031053.1	29	6.74	37.7
OX031054.1	30	6.45	38.2
OX031024.1	Z	23.08	36.3
OX031055.1	MT	0.02	19.8
-	unplaced	0.75	47.5

distribution was evaluated by running the sample on the FemtoPulse system.

RNA was extracted from abdomen tissue of ilSesBemb2 in the Tree of Life Laboratory at the WSI using TRIzol, according to the manufacturer's instructions. RNA was then eluted in 50 μl RNAse-free water and its concentration assessed using a Nanodrop spectrophotometer and Qubit Fluorometer using the Qubit RNA Broad-Range (BR) Assay kit. Analysis of the integrity of the RNA was done using Agilent RNA 6000 Pico Kit and Eukaryotic Total RNA assay.

Sequencing

Pacific Biosciences HiFi circular consensus and 10X Genomics read cloud DNA sequencing libraries were constructed according to the manufacturers' instructions. Poly(A) RNA-Seq libraries were constructed using the NEB Ultra II RNA Library Prep kit. DNA and RNA sequencing were performed by the Scientific Operations core at the WSI on Pacific Biosciences SEQUEL II (HiFi) and Illumina NovaSeq 6000 (RNA-Seq). Hi-C data were also generated from abdomen tissue of ilSesBemb1 using the Arima v2 kit and sequenced on the Illumina NovaSeq 6000 instrument.

Genome assembly

Assembly was carried out with Hifiasm (Cheng et al., 2021). The assembly was scaffolded with Hi-C data (Rao et al., 2014) using YaHS (Zhou et al., 2023). The assembly was checked for contamination as described previously (Howe et al., 2021). Manual curation was performed using HiGlass (Kerpedjiev et al., 2018) and Pretext (Harry, 2022). The mitochondrial genome was assembled using MitoHiFi (Uliano-Silva et al., 2022), which performed annotation using MitoFinder (Allio et al., 2020). The genome was analysed and BUSCO scores were generated within the BlobToolKit environment (Challis et al., 2020). Table 3 contains a list of all software tool versions used, where appropriate.

Genome annotation

The BRAKER2 pipeline (Brůna et al., 2021) was used in the default protein mode to generate annotation for the

Table 3. Software tools and versions used.

Software tool	Version	Source
BlobToolKit	3.5.2	Challis et al., 2020
Hifiasm	0.16.1-r375	Cheng <i>et al.,</i> 2021
HiGlass	1.11.6	Kerpedjiev et al., 2018
MitoHiFi	2	Uliano-Silva et al., 2022
PretextView	0.2	Harry, 2022
YaHS	yahs-1.1.91eebc2	Zhou et al., 2023

S. bembeciformis assembly (GCA_943735995.1) in Ensembl Rapid Release.

Ethics and compliance issues

The materials that have contributed to this genome note have been supplied by a Darwin Tree of Life Partner. The submission of materials by a Darwin Tree of Life Partner is subject to the Darwin Tree of Life Project Sampling Code of Practice. By agreeing with and signing up to the Sampling Code of Practice, the Darwin Tree of Life Partner agrees they will meet the legal and ethical requirements and standards set out within this document in respect of all samples acquired for, and supplied to, the Darwin Tree of Life Project. All efforts are undertaken to minimise the suffering of animals used for sequencing. Each transfer of samples is further undertaken according to a Research Collaboration Agreement or Material Transfer Agreement entered into by the Darwin Tree of Life Partner, Genome Research Limited (operating as the Wellcome Sanger Institute), and in some circumstances other Darwin Tree of Life collaborators.

Data availability

European Nucleotide Archive: *Sesia bembeciformis* (lunar hornet). Accession number PRJEB52659; https://identifiers.org/ena.embl/PRJEB52659. (Wellcome Sanger Institute, 2022)

The genome sequence is released openly for reuse. The *Sesia bembeciformis* genome sequencing initiative is part of the Darwin Tree of Life (DToL) project. All raw sequence data and the assembly have been deposited in INSDC databases. Raw data and assembly accession identifiers are reported in Table 1.

Author information

Members of the University of Oxford and Wytham Woods Genome Acquisition Lab are listed here: https://doi.org/10.5281/zenodo.4789928.

Members of the Darwin Tree of Life Barcoding collective are listed here: https://doi.org/10.5281/zenodo.4893703.

Members of the Wellcome Sanger Institute Tree of Life programme are listed here: https://doi.org/10.5281/zenodo.4783585.

Members of Wellcome Sanger Institute Scientific Operations: DNA Pipelines collective are listed here: https://doi.org/10.5281/zenodo.4790455.

Members of the Tree of Life Core Informatics collective are listed here: https://doi.org/10.5281/zenodo.5013541.

Members of the Darwin Tree of Life Consortium are listed here: https://doi.org/10.5281/zenodo.4783558.

References

Allio R, Schomaker-Bastos A, Romiguier J, et al.: MitoFinder: Efficient automated large-scale extraction of mitogenomic data in target enrichment phylogenomics. Mol Ecol Resour. 2020; 20(4): 892-905. PubMed Abstract | Publisher Full Text | Free Full Text

Brůna T, Hoff KJ, Lomsadze A, et al.: BRAKER2: Automatic eukaryotic genome annotation with GeneMark-EP+ and AUGUSTUS supported by a protein database. NAR Genom Bioinform. 2021; 3(1): Iqaa108. PubMed Abstract | Publisher Full Text | Free Full Text

Burman J, Westerberg L, Ostrow S, et al.: Revealing hidden species distribution with pheromones: the case of Synanthedon vespiformis (Lepidoptera: Sesiidae) in Sweden. J Insect Conserv. 2016; 20(1): 11-21. **Publisher Full Text**

Challis R, Richards E, Rajan J, et al.: BlobToolKit - Interactive Quality Assessment of Genome Assemblies. G3 (Bethesda). 2020; 10(4): 1361-1374. PubMed Abstract | Publisher Full Text | Free Full Text

Cheng H, Concepcion GT, Feng X, et al.: Haplotype-resolved de novo assembly using phased assembly graphs with hifiasm. Nat Methods. 2021; 18(2):

PubMed Abstract | Publisher Full Text | Free Full Text

Créqu A: Nouvelles observations d'une espèce peu citée en France. Sesia bembeciformis (Hübner, 1806) (Lep. Sesiidae). Oreina. 2015; (30): 7–8. Reference Source

Goossens R: Research on the distribution of Sesia bembeciformis (Lepidoptera: Sesiidae) in Belgium, the Netherlands and northern France. Phegea. 2020; 48(3): 50-57.

Reference Source

Harry E: PretextView (Paired REad TEXTure Viewer): A desktop application for viewing pretext contact maps. 2022; (Accessed: 19 October 2022).

Henwood B, Sterling P, Lewington R: Field Guide to the Caterpillars of Great Britain and Ireland. London: Bloomsbury, 2020.

Howe K, Chow W, Collins J, et al.: Significantly improving the quality of genome assemblies through curation. GigaScience. Oxford University Press, 2021; **10**(1): giaa153.

PubMed Abstract | Publisher Full Text | Free Full Text

Johnson T: Illustrations of British Hawk moths and their larvae (including the Sesidae). 1874.

Reference Source

Kerpedijev P. Abdennur N. Lekschas F. et al.: HiGlass: Web-based visual exploration and analysis of genome interaction maps. Genome Biol. 2018; **19**(1): 125.

PubMed Abstract | Publisher Full Text | Free Full Text

Lastuvka Z, Lastuvka A: The Sesiidae of Europe. Stenstrup: Apollo Books, 2001.

Lastuvka Z, Lastuvka A: Sesiidae of the Iberian Peninsula, new records and distributional analysis (Insecta: Lepidoptera). SHILAP Revista de lepidopterologia. 2014; **42**(168): 559-580.

Reference Source

Manni M, Berkeley MR, Seppey M, et al.: BUSCO Update: Novel and Streamlined Workflows along with Broader and Deeper Phylogenetic Coverage for Scoring of Eukaryotic, Prokaryotic, and Viral Genomes. Mol Biol Evol. 2021; 38(10): 4647–4654. PubMed Abstract | Publisher Full Text | Free Full Text

Pühringer F: Pheromone attraction of European and North African clear wing moths (Lepidoptera: Sesiidae) II. 2021; (Accessed: 14 February 2023). **Reference Source**

Randle Z, Evans-Hill LJ, Parsons MS, et al.: Atlas of Britain & Ireland's Larger Moths. Newbury: NatureBureau, 2019.

Reference Source

Rao SSP, Huntley MH, Durand NC, et al.: A 3D map of the human genome at kilobase resolution reveals principles of chromatin looping. Cell. 2014; **159**(7): 1665-80.

PubMed Abstract | Publisher Full Text | Free Full Text

Rhie A, McCarthy SA, Fedrigo O, et al.: Towards complete and error-free genome assemblies of all vertebrate species. Nature. 2021; 592(7856): 737–746.

PubMed Abstract | Publisher Full Text | Free Full Text

St John S: Larva collecting and breeding. London: William Wesley and Son,

Reference Source

Uliano-Silva M, Ferreira JGRN, Krasheninnikova K, et al.: MitoHiFi: a python pipeline for mitochondrial genome assembly from PacBio High Fidelity reads. bioRxiv. [Preprint], 2022.

Publisher Full Text

Waring P, Townsend M, Lewington R: **Field Guide to the Moths of Great Britain and Ireland: Third Edition.** Bloomsbury Wildlife Guides, 2017.

Wellcome Sanger Institute: The genome seguence of the Lunar Hornet. Sesia bembeciformis (Hübner 1806). European Nucleotide Archive, [dataset], accession number PRJEB52659, 2022.

Zhou C, McCarthy SA, Durbin R: YaHS: yet another Hi-C scaffolding tool. Bioinformatics. Edited by C. Alkan, 2023; 39(1): btac808.

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Open Peer Review

Current Peer Review Status:







Version 1

Reviewer Report 11 October 2023

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Roman Wasala 🗓



Department of Entomology and Environment Protection, University of Life Sciences in Poznań, Dąbrowskiego, Poland

The authors present the genome sequence of the Lunar Hornet, Sesia bembeciformis (Hübner 1806) Two male Sesia bembeciformis were obtained from Wytham Woods, Oxfordshire, on 17 July 2021. The specimens were collected from the orchard using a pheromone lure. This is the next Sesiidae genome to be sequenced. The Sesia bembeciformis genome was sequenced as part of the Darwin Tree of Life project. The chromosome scale is of the same high quality as other genomes generated by DtoL. The pipeline is identical to other genomes, which is great. The resulting annotation includes 16,010 transcribed mRNAs from 15,843 protein-coding genes. The introductory text is presented in a comprehensive and enjoyable manner for the reader. The photographs of the Sesia bembeciformis specimens could have been better or properly corrected in a suitable graphics programme.

Is the rationale for creating the dataset(s) clearly described?

Are the protocols appropriate and is the work technically sound?

Are sufficient details of methods and materials provided to allow replication by others?

Yes

Are the datasets clearly presented in a useable and accessible format?

Yes

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: Broad Lepidopterology with particular emphasis on moths of the superfamily Noctuoidea

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Reviewer Report 05 October 2023

https://doi.org/10.21956/wellcomeopenres.21187.r67884

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了 💮 Fahad Alqahtani 🗓

King Abdulaziz City for Science and Technology, Riyadh, Saudi Arabia

The authors of "The genome sequence of the Lunar Hornet, Sesia bembeciformis (Hübner 1806)," have successfully reconstructed the genome sequence of the Lunar Hornet. The assembly quality was evaluated using BUSCO, leading to high scores. The genome assembly contains 31 chromosomal-level scaffolds, including 30 autosomes and the Z sex chromosome, totaling 477,100,000 base pairs. The background is well-organized, with a clear and logical flow throughout the content, showcasing a significant contribution to the field of genomics.

However, there are a few minor issues that need to be addressed.

- In the Methods section (Sequencing), the authors mentioned 10X Genomics read cloud DNA sequencing. However, the paper specified the use of Pacific Biosciences SEQUEL II, Hi-C Illumina, and PolyA RNA-Seq Illumina. Can you explain this ambiguity?
- In the "Genome Sequence Report," the authors also stated that the genome was reconstructed from one male. However, in the Methods - Sample Acquisition and Nucleic Acid Extraction, it was mentioned that two males were collected. Can you explain this?
- A third issue arises from the RNA-seq data, which was not mentioned in the genome assembly section. The genome assembly section requires more details, including the data used for Hifiasm (Pacbio data is known to be used), as I understand the manuscript should be reproducible.
- Moreover, in the Methods section (Genome Assembly, Curation, and Evaluation), it would be valuable to specify the closely related species used when MitoHiFi/Mitofinder guided the annotation of the mitochondrial genome.
- Lastly, in the Methods section (Genome Assembly, Curation, and Evaluation), the term
 "Pretext" should align with the software tools table, which is mentioned as "PretextView."

Is the rationale for creating the dataset(s) clearly described?

Yes

Are the protocols appropriate and is the work technically sound?

Yes

Are sufficient details of methods and materials provided to allow replication by others? Yes

Are the datasets clearly presented in a useable and accessible format?

Yes

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: Bioinformatics

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard, however I have significant reservations, as outlined above.

Reviewer Report 07 September 2023

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Benoit Nabholz 🗓



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The authors present the chromosome scale assembly of the Lunar Hornet, Sesia bembeciformis. It is the 16th Sesiidae genome to be sequenced (most of them by the Darwin Tree of Life project, DtoL).

The genome is of the same high quality as the other genomes produced by the DtoL project. The pipeline is the same as for the other genomes which is excellent. All the links and datasets presented are publicly available.

The text of the introduction reads well and is interesting.

The photos of the specimens could have been better, i.e. more light, a scale and a side and dorsal photo.

Is the rationale for creating the dataset(s) clearly described?

Are the protocols appropriate and is the work technically sound?

Are sufficient details of methods and materials provided to allow replication by others?

Yes

Are the datasets clearly presented in a useable and accessible format?

Yes

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: Molecular evolution

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.