




DATA NOTE

The genome sequence of the Green-brindled Crescent, *Allophyes oxyacanthae* (Linnaeus, 1758) [version 1; peer review: awaiting peer review]

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Abstract

We present a genome assembly from an individual male *Allophyes oxyacanthae* (the Green-brindled Crescent; Arthropoda; Insecta; Lepidoptera; Noctuidae). The genome sequence is 458 megabases in span. The whole assembly is scaffolded into 31 chromosomal pseudomolecules, including the assembled Z sex chromosome. The mitochondrial genome has also been assembled and is 15.3 kilobases in length. Gene annotation of this assembly on Ensembl has identified 17,301 protein coding genes.

Keywords

Allophyes oxyacanthae, Green-brindled Crescent, genome sequence, chromosomal, Lepidoptera



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Open Peer Review

Approval Status AWAITING PEER REVIEW

Any reports and responses or comments on the article can be found at the end of the article.

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Species taxonomy

Eukaryota; Metazoa; Ecdysozoa; Arthropoda; Hexapoda; Insecta; Pterygota; Neoptera; Endopterygota; Lepidoptera; Glossata; Ditrysia; Noctuoidea; Noctuidae; Amphipyridae; *Allophyes*; *Allophyes oxyacanthae* (Linnaeus, 1758) (NCBI:txid988056).

Background

The Green-brindled Crescent *Allophyes oxyacanthae* is a moth in the family Noctuidae with an autumn flight period in the UK. The typical form of the moth is unmistakable, with a dense scattering of shimmering metallic green scales on the forewings, contrasting against a deep brown ground colour, pale marginal band and pale orbicular and reniform stigmata (oval and kidney marks). A distinct colour variant is also encountered in the UK, denoted form *capucina*, with uniform brown colouration and fewer green scales. Breeding experiments suggest that the difference between the two forms is controlled by a single genetic locus, with the f. *capucina* allele dominant to the wild type allele (Steward, 1977). It has been suggested that f. *capucina* moths may have had a selective advantage in areas of industrial pollution, although the data are unclear on this issue (Ford, 1967). The genetic locus and molecular basis of the polymorphism have not been identified.

A. oxyacanthae has been recorded across most of UK, including the north of Scotland, through Scandinavia and across mainland Europe although there are few records from Italy and southern Spain (GBIF Secretariat, 2022). The species is attracted to light and can be found in woodlands and gardens where the larval food plants of hawthorn, blackthorn, rowan and fruit trees are found (Robinson *et al.*, 2010).

A genome sequence for *A. oxyacanthae* will facilitate study of the genetic basis of colour polymorphism and the molecular adaptations underpinning polyphagy, and contribute to a growing data set of resources for understanding lepidopteran biology.

Genome sequence report

The genome was sequenced from one male *Allophyes oxyacanthae* specimen (Figure 1) collected in Wytham Woods (latitude 51.77, longitude -1.34). A total of 71-fold coverage in Pacific Biosciences single-molecule HiFi long reads was generated. Primary assembly contigs were scaffolded with chromosome conformation Hi-C data. Manual assembly curation corrected three missing or mis-joins, reducing the scaffold number by 8.82%.

The final assembly has a total length of 458.5 Mb in 31 sequence scaffolds with a scaffold N50 of 16.7 Mb (Table 1). The whole assembly sequence was assigned to 31 chromosomal-level scaffolds, representing 30 autosomes and the Z sex chromosome. Chromosome-scale scaffolds confirmed by the Hi-C data have been named in order of size (Figure 2–Figure 5; Table 2). The assembly has a BUSCO v5.3.2 (Manni *et al.*, 2021) completeness of 99.2% (single 98.8%, duplicated 0.4%) using the lepidoptera_odb10 reference set ($n = 5286$). While not fully phased, the assembly deposited is of one haplotype. Contigs



Figure 1. Photograph of the *Allophyes oxyacanthae* (ilAlIOxya1) specimen used for genome sequencing.

corresponding to the second haplotype have also been deposited.

Genome annotation report

The *A. oxyacanthae* genome assembly (GCA_932294325.1) was annotated using the Ensembl rapid annotation pipeline (Table 1). The resulting annotation includes 17,485 transcribed mRNAs from 17,301 protein-coding genes.

Methods

Sample acquisition and nucleic acid extraction

An individual male *A. oxyacanthae* specimen (ilAlIOxya1) was collected in Wytham Woods, Oxfordshire (biological vice-county: Berkshire), UK (latitude 51.77, longitude -1.34) on 8 October 2020 using a light trap. The specimens were collected and identified by Douglas Boyes (University of Oxford) and snap-frozen on dry ice. This specimen was used for DNA and Hi-C sequencing.

DNA was extracted at the Tree of Life laboratory, Wellcome Sanger Institute (WSI). The ilAlIOxya1 sample was weighed and dissected on dry ice with head tissue set aside for Hi-C sequencing. Thorax tissue was disrupted using a Nippi Powermasher fitted with a BioMasher pestle. High molecular weight (HMW) DNA was extracted using the Qiagen MagAttract HMW DNA extraction kit. HMW DNA was sheared into an average fragment size of 12–20 kb in a Megaruptor 3 system with speed setting 30. Sheared DNA was purified by solid-phase reversible immobilisation using AMPure PB beads with a 1.8X ratio of beads to sample to remove the shorter fragments and concentrate the DNA sample. The concentration of the sheared and purified DNA was assessed using a Nanodrop spectrophotometer and Qubit Fluorometer and Qubit dsDNA High Sensitivity Assay kit. Fragment size distribution was evaluated by running the sample on the FemtoPulse system.

Table 1. Genome data for *Allophyes oxyacanthae*, iAllOxya1.1.

Project accession data		
Assembly identifier	iAllOxya1.1	
Species	<i>Allophyes oxyacanthae</i>	
Specimen	iAllOxya1	
NCBI taxonomy ID	988056	
BioProject	PRJEB50741	
BioSample ID	SAMEA8603204	
Isolate information	iAllOxya1; male: thorax (PacBio), head (Hi-C)	
Assembly metrics*		Benchmark
Consensus quality (QV)	67.9	≥ 50
k-mer completeness	100%	≥ 95%
BUSCO**	C:99.2%[S:98.8%,D:0.4%], F:0.1%,M:0.7%,n:5,286	C ≥ 95%
Percentage of assembly mapped to chromosomes	100%	≥ 95%
Sex chromosomes	Z chromosome	localised homologous pairs
Organelles	Mitochondrial genome assembled	complete single alleles
Raw data accessions		
PacificBiosciences SEQUEL II	ERR8575376, ERR8575377	
Hi-C Illumina	ERR8571663	
Genome assembly		
Assembly accession	GCA_932294325.1	
Accession of alternate haplotype	GCA_932294395.1	
Span (Mb)	458.5	
Number of contigs	38	
Contig N50 length (Mb)	16.4	
Number of scaffolds	31	
Scaffold N50 length (Mb)	16.7	
Longest scaffold (Mb)	20.3	
Genome annotation		
Number of protein-coding genes	17,301	
Number of transcripts	17,485	

* Assembly metric benchmarks are adapted from column VGP-2020 of "Table 1: Proposed standards and metrics for defining genome assembly quality" from (Rhie *et al.*, 2021).

** BUSCO scores based on the lepidoptera_odb10 BUSCO set using v5.3.2. C = complete [S = single copy, D = duplicated], F = fragmented, M = missing, n = number of orthologues in comparison. A full set of BUSCO scores is available at https://blobtoolkit.genomehubs.org/view/iAllOxya1_1/dataset/iAllOxya1_1/busco.

Sequencing

Pacific Biosciences HiFi circular consensus DNA sequencing libraries were constructed according to the manufacturers'

instructions. DNA sequencing was performed by the Scientific Operations core at the WSI on Pacific Biosciences SEQUEL II (HiFi) instrument. Hi-C data were also generated from head

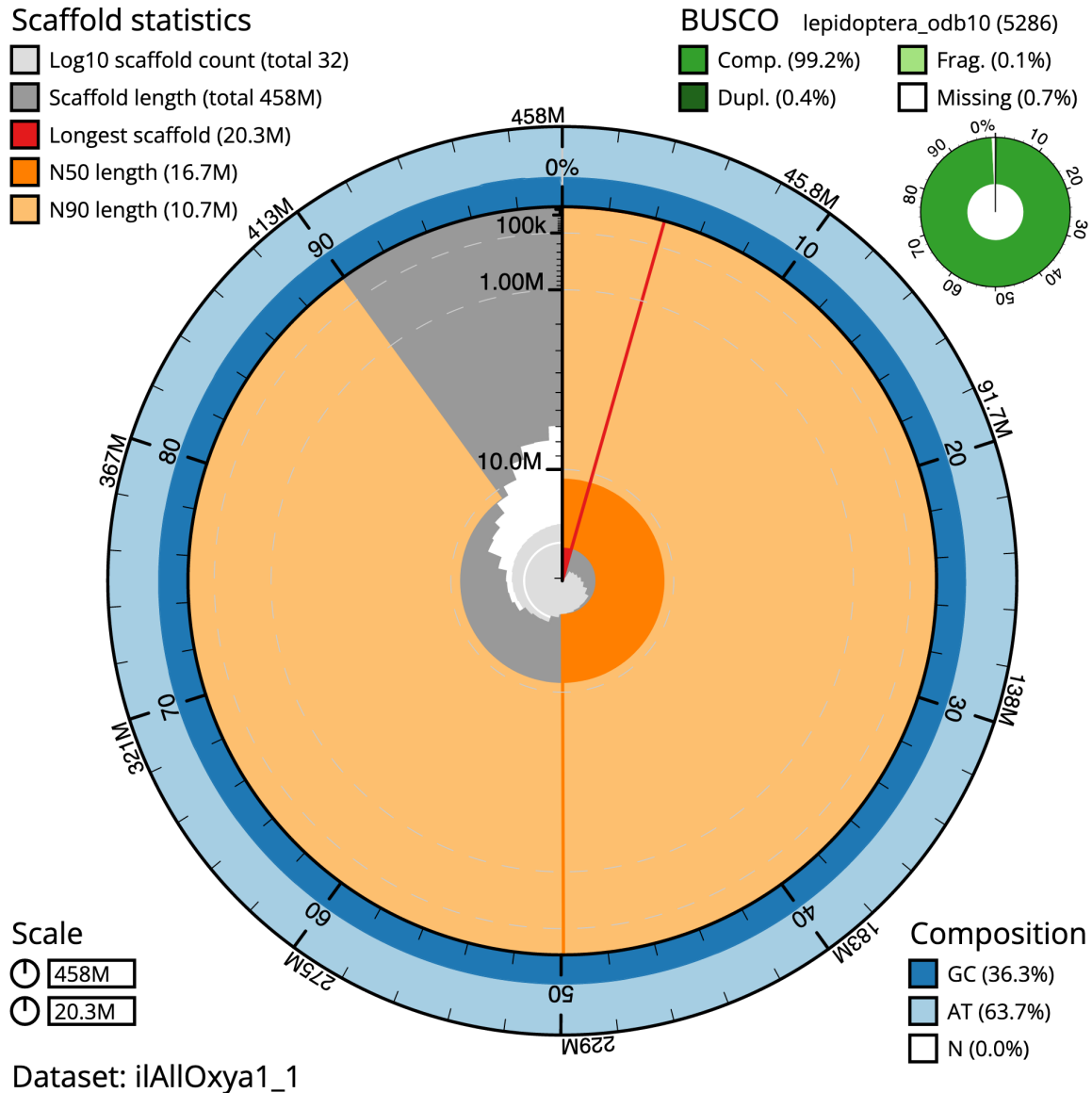


Figure 2. Genome assembly of *Allophyes oxyacanthae*, ilAllOxya1.1: metrics. The BlobToolKit Snailplot shows N50 metrics and BUSCO gene completeness. The main plot is divided into 1,000 size-ordered bins around the circumference with each bin representing 0.1% of the 458,479,537 bp assembly. The distribution of scaffold lengths is shown in dark grey with the plot radius scaled to the longest scaffold present in the assembly (20,278,968 bp, shown in red). Orange and pale-orange arcs show the N50 and N90 scaffold lengths (16,683,655 and 10,717,387 bp), respectively. The pale grey spiral shows the cumulative scaffold count on a log scale with white scale lines showing successive orders of magnitude. The blue and pale-blue area around the outside of the plot shows the distribution of GC, AT and N percentages in the same bins as the inner plot. A summary of complete, fragmented, duplicated and missing BUSCO genes in the lepidoptera_odb10 set is shown in the top right. An interactive version of this figure is available at https://blobtoolkit.genomehubs.org/view/ilAllOxya1_1/dataset/ilAllOxya1_1/snail.

tissue of ilAllOxya1 using the Arima v2 kit and sequenced on the Illumina NovaSeq 6000 instrument.

Genome assembly

Assembly was carried out with Hifiasm (Cheng *et al.*, 2021) and haplotypic duplication was identified and removed with

purge_dups (Guan *et al.*, 2020). The assembly was scaffolded with Hi-C data (Rao *et al.*, 2014) using YaHS (Zhou *et al.*, 2022). The assembly was checked for contamination as described previously (Howe *et al.*, 2021). Manual curation was performed using HiGlass (Kerpedjiev *et al.*, 2018) and Pretext (Harry, 2022). The mitochondrial genome was assembled using MitoHiFi

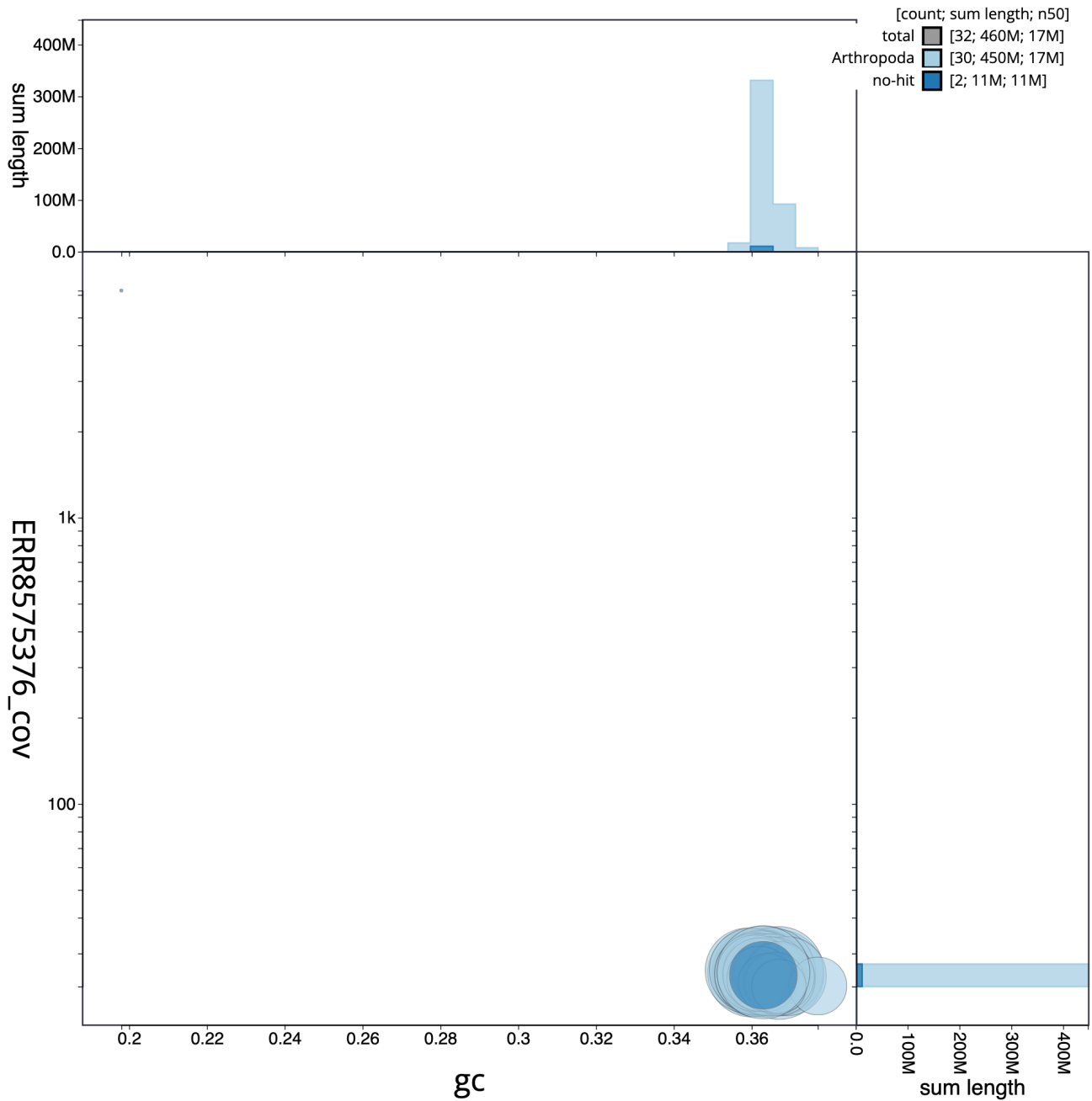


Figure 3. Genome assembly of *Allophyes oxyacanthae*, ilAllOxya1.1: GC coverage. BlobToolKit GC-coverage plot. Scaffolds are coloured by phylum. Circles are sized in proportion to scaffold length. Histograms show the distribution of scaffold length sum along each axis. An interactive version of this figure is available at https://blobtoolkit.genomehubs.org/view/ilAllOxya1_1/dataset/ilAllOxya1_1/blob.

(Uliano-Silva *et al.*, 2022), which performed annotation using MitoFinder (Allio *et al.*, 2020). The genome was analysed and BUSCO scores generated within the BlobToolKit environment (Challis *et al.*, 2020). Table 3 contains a list of all software tool versions used, where appropriate.

Genome annotation

The BRAKER2 pipeline (Brûna *et al.*, 2021) was used in the default protein mode to generate annotation for the *Allophyes oxyacanthae* assembly (GCA_934047225.1) in Ensembl Rapid Release.

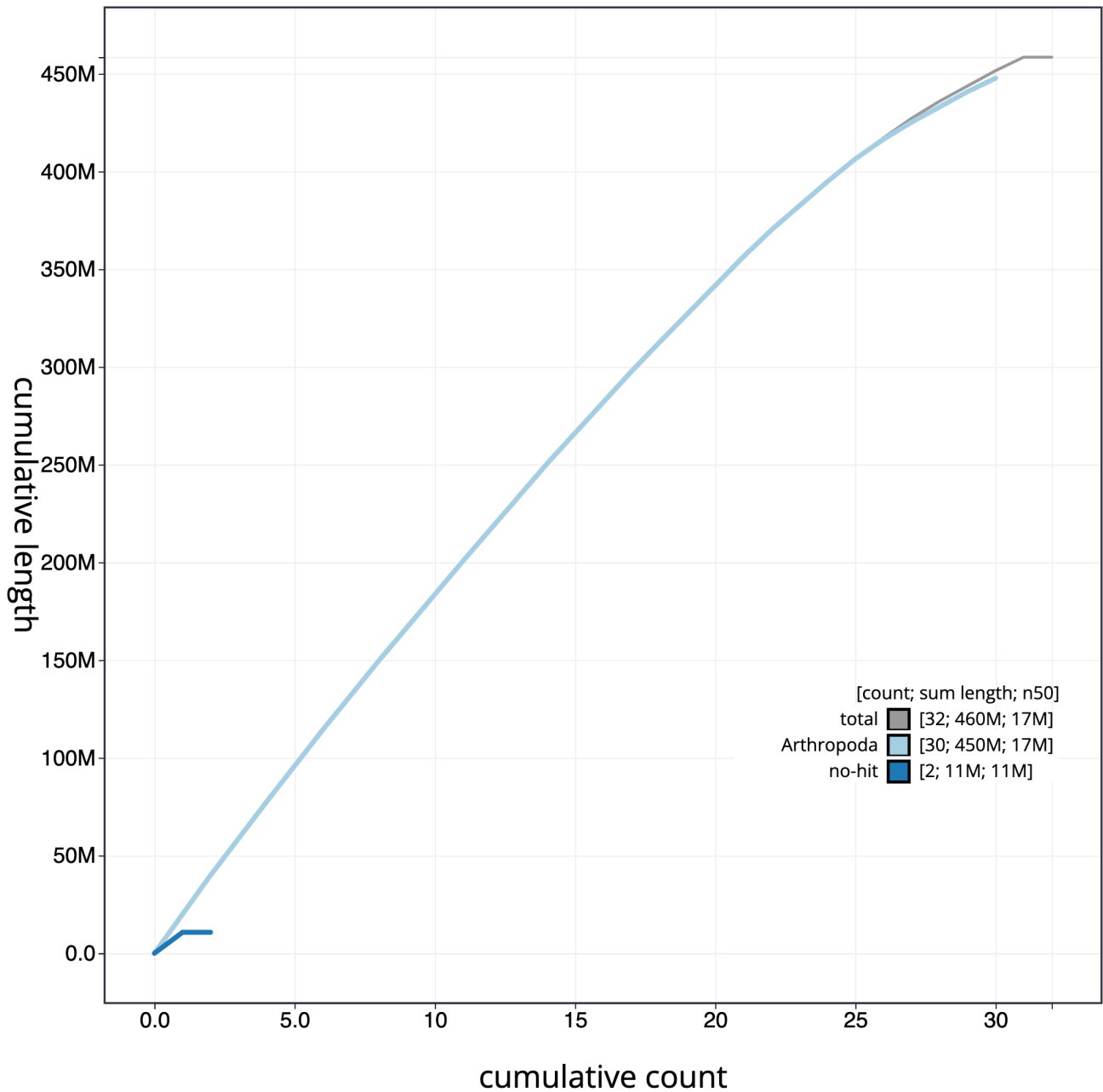


Figure 4. Genome assembly of *Allophyes oxyacanthae*, iAllOxya1.1: cumulative sequence. BlobToolKit cumulative sequence plot. The grey line shows cumulative length for all scaffolds. Coloured lines show cumulative lengths of scaffolds assigned to each phylum using the buscogenes taxrule. An interactive version of this figure is available at https://blobtoolkit.genomehubs.org/view/iAllOxya1_1/dataset/iAllOxya1_1/cumulative.

Ethics/compliance issues

The materials that have contributed to this genome note have been supplied by a Darwin Tree of Life Partner. The submission of materials by a Darwin Tree of Life Partner is subject to the [Darwin Tree of Life Project Sampling Code of Practice](#). By agreeing with and signing up to the Sampling Code of

Practice, the Darwin Tree of Life Partner agrees they will meet the legal and ethical requirements and standards set out within this document in respect of all samples acquired for, and supplied to, the Darwin Tree of Life Project. Each transfer of samples is further undertaken according to a Research Collaboration Agreement or Material Transfer Agreement entered

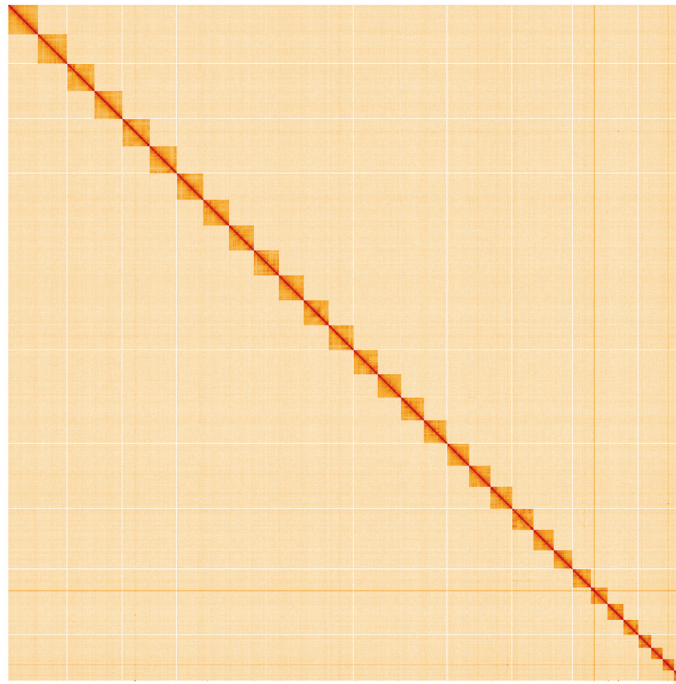


Figure 5. Genome assembly of *Allophytes oxyacanthae*, iAllOxya1.1: Hi-C contact map. Hi-C contact map of the iAllOxya1.1 assembly, visualised using HiGlass. Chromosomes are shown in order of size from left to right and top to bottom. An interactive version of this figure may be viewed at <https://genome-note-higlass.tol.sanger.ac.uk/l/?d=OzgdewECSHeyOqN4f7ptEQ>.

Table 2. Chromosomal pseudomolecules in the genome assembly of *Allophytes oxyacanthae*, iAllOxya1.

INSDC accession	Chromosome	Size (Mb)	GC%
OW028610.1	1	19.67	36.3
OW028611.1	2	18.89	36.7
OW028612.1	3	18.72	36.7
OW028613.1	4	18.51	36.5
OW028614.1	5	18.2	36
OW028615.1	6	17.92	36.1
OW028616.1	7	17.47	36.2
OW028617.1	8	17.03	36.1
OW028618.1	9	17	35.9
OW028619.1	10	16.91	36.3
OW028620.1	11	16.8	36
OW028621.1	12	16.68	36.3
OW028622.1	13	16.39	36.2
OW028623.1	14	15.76	36.1
OW028624.1	15	15.71	36.1

INSDC accession	Chromosome	Size (Mb)	GC%
OW028625.1	16	15.55	36.3
OW028626.1	17	14.93	36.5
OW028627.1	18	14.7	36.9
OW028628.1	19	14.63	36.5
OW028629.1	20	14.54	36.4
OW028630.1	21	13.74	36.1
OW028631.1	22	12.47	36.2
OW028632.1	23	12.29	36.7
OW028633.1	24	11.69	36.7
OW028634.1	25	10.72	36.3
OW028635.1	26	9.96	36.2
OW028636.1	27	8.76	36.6
OW028637.1	28	7.85	36.4
OW028638.1	29	7.81	37.7
OW028639.1	30	6.89	36.7
OW028609.1	Z	20.28	36.3
OW028640.1	MT	0.02	20.1

Table 3. Software tools and versions used.

Software tool	Version	Source
BlobToolKit	3.5.0	Challis et al., 2020
Hifiasm	0.16.1-r375	Cheng et al., 2021
HiGlass	1.11.6	Kerpedjiev et al., 2018
MitoHiFi	2	Uliano-Silva et al., 2022
PretextView	0.2	Harry, 2022
purge_dups	1.2.3	Guan et al., 2020
YaHS	yahs-1.1.91eebc2	Zhou et al., 2022

into by the Darwin Tree of Life Partner, Genome Research Limited (operating as the Wellcome Sanger Institute), and in some circumstances other Darwin Tree of Life collaborators.

Data availability

European Nucleotide Archive: *Allophyes oxyacanthae* (green-brindled crescent). Accession number [PRJEB50741](#); <https://identifiers.org/ena.embl/PRJEB50741>. (Wellcome Sanger Institute, 2022)

References

- Allio R, Schomaker-Bastos A, Romiguier J, et al.: **MitoFinder: Efficient automated large-scale extraction of mitogenomic data in target enrichment phylogenomics.** *Mol Ecol Resour.* 2020; **20**(4): 892–905. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- Brůna T, Hoff KJ, Lomsadze A, et al.: **BRAKER2: Automatic eukaryotic genome annotation with GeneMark-EP+ and AUGUSTUS supported by a protein database.** *NAR Genom Bioinform.* 2021; **3**(1): lqaa108. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- Challis R, Richards E, Rajan J, et al.: **BlobToolKit - interactive quality assessment of genome assemblies.** *G3 (Bethesda).* 2020; **10**(4): 1361–1374. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- Cheng H, Concepcion GT, Feng X, et al.: **Haplotype-resolved de novo assembly using phased assembly graphs with hifiasm.** *Nat Methods.* 2021; **18**(2): 170–175. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- Ford EB: **Moths.** London: New Naturalist, Collins. 1967. [Reference Source](#)
- GBIF Secretariat: **Allophyes oxyacanthae (Linnaeus, 1758).** *GBIF Backbone Taxonomy.* 2022; (Accessed: 18 January 2023). [Reference Source](#)
- Guan D, McCarthy SA, Wood J, et al.: **Identifying and removing haplotypic duplication in primary genome assemblies.** *Bioinformatics.* 2020; **36**(9): 2896–2898. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- Harry E: **PretextView (Paired REad TEXTure Viewer): A desktop application for viewing pretext contact maps.** 2022; (Accessed: 19 October 2022). [Reference Source](#)
- Howe K, Chow W, Collins J, et al.: **Significantly improving the quality of genome assemblies through curation.** *GigaScience.* Oxford University Press. 2021; **10**(1): gaa153. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- Kerpedjiev P, Abdennur N, Lekschas F, et al.: **HiGlass: Web-based visual exploration and analysis of genome interaction maps.** *Genome Biol.* 2018; **19**(1): 125. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- Manni M, Berkeley MR, Seppay M, et al.: **BUSCO Update: Novel and Streamlined Workflows along with Broader and Deeper Phylogenetic Coverage for Scoring of Eukaryotic, Prokaryotic, and Viral Genomes.** *Mol Biol Evol.* 2021; **38**(10): 4647–4654. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- Rao SSP, Huntley MH, Durand NC, et al.: **A 3D map of the human genome at kilobase resolution reveals principles of chromatin looping.** *Cell.* 2014; **159**(7): 1665–1680. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- Rhie A, McCarthy SA, Fedrigo O, et al.: **Towards complete and error-free genome assemblies of all vertebrate species.** *Nature.* 2021; **592**(7856): 737–746. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- Robinson GS, Kitching I, Hernández L, et al.: **HOSTS - A Database of the world's Lepidopteran Hostplants.** London: Natural History Museum. 2010; (Accessed: 18 January 2023). [Reference Source](#)
- Steward RC: **Genetic control of the melanistic forms of the moths *Diurnea fagella* and *Allophyes oxyacanthae*.** *Heredity.* 1977; **39**: 235–241. [Publisher Full Text](#)
- Uliano-Silva M, Ferreira, JG, Krashennikova K, et al.: **MitoHiFi: a python pipeline for mitochondrial genome assembly from PacBio High Fidelity reads.** *bioRxiv.* [Preprint], 2022. [Publisher Full Text](#)
- Wellcome Sanger Institute: **The genome sequence of the Green-brindled Crescent, *Allophyes oxyacanthae* (Linnaeus, 1758).** European Nucleotide Archive. [dataset], accession number [PRJEB50741](#), 2022.
- Zhou C, McCarthy SA, Durbin R: **YaHS: yet another Hi-C scaffolding tool.** *bioRxiv.* [Preprint], 2022. [Publisher Full Text](#)

The genome sequence is released openly for reuse. The *Allophyes oxyacanthae* genome sequencing initiative is part of the Darwin Tree of Life (DTOL) project. All raw sequence data and the assembly have been deposited in INSDC databases. Raw data and assembly accession identifiers are reported in [Table 1](#).

Author information

Members of the University of Oxford and Wytham Woods Genome Acquisition Lab are listed here: <https://doi.org/10.5281/zenodo.4789928>.

Members of the Darwin Tree of Life Barcoding collective are listed here: <https://doi.org/10.5281/zenodo.4893703>.

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