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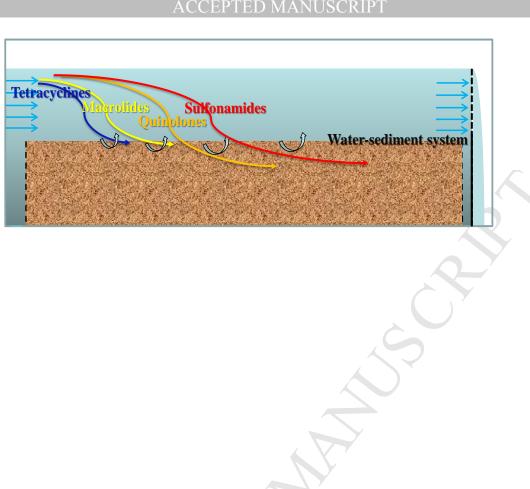
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- 1 Persistence and migration of tetracycline, sulfonamide, fluoroquinolone, and
- 2 macrolide antibiotics in streams using a simulated hydrodynamic system
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- Abstract: The potential persistence and migration of 14 antibiotics comprising sulfonamides, 18 fluoroquinolones, macrolides and tetracyclines were conducted using a 50-d recirculating 19 flume study supported by batch attenuation experiments with spiked concentrations. The 20 study demonstrated that photodegradation was the dominant attenuation process for these 21 antibiotics in the water environment. The half-lives of 2 to 26 d were in order of 22 sulfadiazine > sulfadimethoxine > sulfamerazine > sulfamethoxazole > sulfamethazine > 23 sulfathiazole > ofloxacin > enrofloxacin > norfloxacin > ciprofloxacin > erythromycin > 24 tetracycline > roxithromycin > oxytetracycline. These modest half-lives meant that the 25 antibiotics were predicted to travel 30-400 km down a typical river before half the 26 concentration would be lost. All antibiotics were detected on the surface sediment in the 27 flume study. Under hyporheic exchange, some of them continually migrated into the deeper 28 sediment and also the sediment pore water. All fluoroquinolones were detected in the 29 sediments. The sulfonamides were detected in the pore water with relatively high 30 concentrations and frequencies. Sulfadiazine, sulfamethazine and sulfathiazole in the upper 31 layer pore water were found to be approaching equilibrium with the surface water. The high 32 presence of sulfonamides in the pore water indicated that their high mobility and persistence 33 potentially pose a risk to hyporheic zone. 34 Keywords: attenuation; hyporheic exchange; conservative tracer; water-sediment system; 35
- 37 Capsule: Natural attenuation of antibiotics in streams.
- 38 1. Introduction

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flume experiment

39	Antibiotics are now widely used to treat human and animal infections and to promote animal
40	growth in China. The current usage of antibiotics in China recently was estimated at 162 000
41	tons. The usage with daily doses per 1000 inhabitants per day (DID) is almost six times
42	greater than each of those in European countries and America (Ying et al., 2017). Around half
43	of the un-metabolized human-sourced antibiotics enter waterways following partially
44	effective removal in municipal sewage treatment plants (Kümmerer, 2009a; Qiao et al., 2018).
45	Animal-sourced antibiotics move to waterways via surface runoff from manure applied to
46	land (Zhang et al., 2015). Therefore, rivers can become a major sink for antibiotics, and
47	antibiotics have been widely detected in surface waters across the world (Kümmerer, 2009b;
48	Qiao et al., 2018). Although antibiotics in the water environment rarely pose an acute toxicity
49	risk to aquatic organisms (Johnson et al., 2015), the levels may still induce transfer and
50	selection of antibiotic resistance genes (Lopatkin et al., 2016; Wang et al., 2016). Spread of
51	antibiotic resistance genes via the food chain could have consequences for the safety and
52	health of humans (Verraes et al., 2013).
53	In order to assess the risk caused by water borne antibiotics, the determination of the
54	dominant attenuation processes and overall attenuation rates including biodegradation,
55	photodegradation, adsorption and hydrolysis are needed. Currently, most research on the fate
56	of antibiotics in water and sediment have mostly focused on individual attenuation processes
57	based on batch experiments (Baena-Nogueras et al., 2017; Conde-Cid et al., 2018; Li et al.,
58	2018; Kaeseberg et al., 2018). However, in the real-world water environment, multiple
59	processes are occurring simultaneously and the key challenges are to assess which one

- dominates and to obtain an overall attenuation rate from the multiple processes. Although
- Luo et al. (2011) provided information on the occurrence and overall attenuation rates of 12
- antibiotics in rivers, this still did not distinguish which were the dominant mechanisms.
- Additionally, the exchange of shallow groundwater and surface water (hyporheic exchange)
- 64 might cause antibiotics to move into groundwater. Li et al. (2015) has studied
- sulfamethoxazole fate in the hyporheic zone. Little is known about the transport of other
- antibiotics in the hyporheic zone.
- This study used a recirculating flume to provide hydrodynamic simulation system to mimic
- 68 material and energy transfer in streams. To complement the flume work, a series of batch
- 69 experiments were conducted to investigate the major loss mechanism. Fourteen antibiotics
- 70 which are frequently detected in Chinese rivers including the sulfonamides sulfadiazine
- 71 (SDZ), sulfamerazine (SMR), sulfamethazine (SMZ), sulfadimethoxine (SDM),
- sulfamethoxazole (SMX), sulfathiazole (STZ), the fluoroquinolones enrofloxacin (EFC),
- ofloxacin (OFC), norfloxacin (NFC), ciprofloxacin (CFC), the tetracyclines oxytetracycline
- 74 (OTC), tetracycline (TC), and the macrolides erythromycin (ETM) and roxithromycin (RTM)
- 75 (Table S1) (Bu et al., 2013; Li et al., 2018) were selected for this study. It is believed EFC is
- only used in animal husbandry; the other antibiotics are used both in human health and
- animal welfare in China (Zhang et al., 2015). The objectives of this study were:
- Assess overall attenuation rates using a flume with local Chinese river water and
- 79 sediment.

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• Use the same flume set-up to study migration in the water-pore-sediments

- Through batch studies, to identify the dominant attenuation mechanism causing these antibiotics to be lost from the water column.
- Assess the consequences of these processes in likely transport of these antibiotics in
 Chinese rivers
- 2. Materials and methods
- 2.1. Chemicals and reagents

Target standards Sulfadiazine (SDZ), Sulfamerazine (SMR), Sulfamethazine (SMZ), 87 Sulfadimethoxine (SDM), Sulfamethoxazole (SMX), Sulfathiazole (STZ) and Enrofloxacin 88 (EFC) were purchased from Sigma-Aldrich (St. Louis, MO, USA). Target standards 89 Ofloxacin (OFC), Norfloxacin (NFC), Ciprofloxacin (CFC) hydrochloride, Oxytetracycline 90 (OTC) hydrochloride, Tetracycline (TC) hydrochloride, Erythromycin (ETM) and 91 Roxithromycin (RTM) were obtained from Dr. Ehrenstorfer GmbH (Augsburg, Germany). 92 Internal standards Sulfamethazine-¹³C₆ (SMZ-¹³C₆) and Enrofloxacin-D₅ (EFC-D₅) 93 hydrochloride were purchased from Witrga Laboratorien Berlin-Adlershof GmbH (Berlin, 94 Germany); Sulfamethoxazole-D₄ (SMX-D₄) was obtained from Dr. Ehrenstorfer GmbH 95 (Augsburg, Germany); Erythromycin-¹³C, D₃ (ETM-¹³C, D₃) and Tetracycline-D₆ (TC-D₆) 96 were obtained from Toronto Research Chemicals (North York, ON, Canada). Each antibiotic 97 standard was dissolved in methanol as standard stock solutions (100 mg L⁻¹). The standard 98 stock solutions were stored at -18°C and were used within three months of purchase to reduce 99 error caused by antibiotic degradation. Working standard mixtures (5, 10, 20, 50, 100 $\mu g \; L^{-1}$) 100

- were freshly prepared by serial dilution of the stock solutions with acetonitrile and Milli-Q water with 0.1% formic acid (5/95, v/v) at each batch analysis. A mixture of internal standards including SMZ- 13 C₆, EFC-D₅ hydrochloride, SMX-D₄, ETM- 13 C, D₃ and TC-D₆ were prepared in methanol (2 mg L $^{-1}$).
- HPLC-grade methyl alcohol and acetone were purchased from Tedia Company (Fairfield, OH, USA). HPLC-grade acetonitrile was obtained from Merck (Darmstadt, Germany). HPLC-grade formic acid (purity of 99%) was purchased from Anaqua Chemicals Supply (Wilmington, USA). The other analytical reagents were obtained from Sinopharm Chemical Reagent Co. Ltd (Shanghai, China).
- 2.2. Water and sediment collection and characterization

Water and sediment used in the experiment were taken from the upstream tributary of Tong-yang River in October 2rd 2017 (31°43'14"N; 117°35'29"E; Fig. S1). The Tong-yang River, connecting to the Chaohu Lake in eastern China, was less influenced by human and farming activities. No background antibiotics were found in these samples using sample pre-treatment and instrumental analysis described in detail in Section 2.5 and 2.6. Water was collected in amber containers. Physico-chemical parameters of water (pH, dissolved oxygen, temperature, conductivity, and Oxidation-reduction potential (ORP)) were measured using handheld water quality monitor (Ultrameter IITM 6P, Myronl, US) in situ and in the lab-scale test. Sediment was collected from the top sediment (20-30 mm), and wet sieved (2-mm mesh) in situ. The particle sizes of the sieved sediment were analyzed by a Laser Particle Sizer

- (Mastersizer 2000, Malvern, UK). The total organic carbon contents (TOC) in water and sediment samples were determined with the TOC Analyser (Multi N/C 3100, Analytikjena, Gemany). The cooled water and sediment samples (4 °C) were transported to the lab located in Hefei City within 8 hours for use in the flume and batch experiments.
- 2.3. Hydrodynamic simulation system using flumes

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The migration and occurrence of antibiotics in natural water environment was simulated in a recirculating flume with a 5-m long, 30-cm wide and 50-cm deep rectangular channel (Fig. S2). A water pump with 0.75 kw was used to derive water flow. The structure and parameters of the flume were based on descriptions by Elliott and Brooks (1997) and Jin et al. (2010), which are described in the SI and Fig. S3. The sediment was put into the rectangular channel with an approximately 18-cm thickness and 0.3-m³ sediment. Approximate 0.9-m³ water was poured into the tank of the flume system with 17-cm water depth in the rectangular channel. Flow velocity was controlled by a valve at 0.2 m s⁻¹, and was monitored by a flow meter (Flowatch, Switzerland). The system was settled beside a window to allow natural sunlight to shine on the apparatus. The system was equilibrated for a week and then run from October 3rd to November 21st 2017. The amount of water lost from the system by evaporation was quantified by using a water gauge every two days. The same amount of the evaporated water was added into the system to maintain water balance. A one hundred-milliliter liter standard solution containing the 14 target antibiotics (9 mg L⁻¹ of each compound) was added into water column to obtain an initial concentration of approximately 10 µg L⁻¹ for each compound to simulate a high exposure level for a natural water environment level (der Beek

et al., 2016).

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2.4. Batch experiments

In parallel to the flume experiment, batch experiments were carried out to explore the dominant attenuation process of each compound. These used water and sediment freshly collected from the Tong-yang River. The experiments were divided into four groups: 1) the sterile water-only group to quantify photodegradation and hydrolysis; 2) sterile water-only group in the dark for hydrolysis only; 3) non-sterile water-sediment group in the dark for biotransformation, adsorption and hydrolysis); 4) sterile water-sediment group in the dark for adsorption and hydrolysis. Each group was set up in duplicate. A one-liter quantity of water was transferred into each glass bottle for the water-only experiments. In the water-sediment groups, 400g (wet) of sediment was put into each glass bottle and then 950-mL water was added (similar to OECD 308). The steady-state photo-degradation test was conducted in an illuminated incubator with sunlight simulators (SPX-250B-G, Boxun, China). The illumination intensity and wavelength were set as 140 W m⁻² and 300–800 nm, repectively; the photoperiod was eight hours per day. The annual average values of the intensity and photoperiod for Hefei region during October to December were from the NASA Atmospheric Science Data Center (https://eosweb.larc.nasa.gov/). In the sterile test, water was autoclaved at 121 °C for 20 min, and sodium azide (NaN₃, final concentration 0.1%) was added into water-sediment system of groups 1, 2, 4 for inhibiting microbial activity. The initial concentration 50 µg L⁻¹ of target compounds was spiked with the antibiotic standards solution. All the systems were incubated at 25 °C over a 30 d period. The percentages of

- methanol spiked in the batch and flume experiments were approximate 0.05% and 0.011%, respectively, thus there was negligible impact on microbial growth (Ramil et al., 2010).
- 2.5. Sampling and sample pretreatment
- The surface water, surface sediment and pore water were sampled at hour 2, day 1, 3, 5, 7, 10, 166 15, 20, 30, 40, 50 after antibiotic spiking in the flume. At each sampling point, three 20 mL of 167 surface water and three 2 g samples of surface sediment at 2-3 cm depth were collected at the 168 front, middle and rear position of the flume. After flushing out residual water in the sampling 169 pipes, three 20 mL of pore water sample from the upper, middle and lower sediment layers 170 were collected in amber glass bottles at the front (pore 1), middle (pore 2) and rear position 171 (pore 3) of the flume (Fig. S3). In addition, three 2 g of sediment samples from the upper, 172 middle and lower sediment layers were collected through three column sampling pores (Fig. 173 S3) at day 10, 20, and 30. The sediments from the different layers were sampled using a steel 174 grooved sampler inserted into sediment sampling pore, which did not impede the system 175 operation for the layer sediment sampling. The used pore was filled with fresh sediment and 176 not sampled again. Meanwhile, water samples from the batch experiments were collected at 177 day 1, 3, 5, 7, 10, 15, 20, and 30 in duplicate. A 975 µL sample of water filtered through 178 syringe filters (0.2-µm PTFE, 13 mm, Agilent, US) was transferred to a vial with 25 µL of 2 179 mg L⁻¹ internal standards of deuterated and isotope labelling antibiotic analogues, and stored 180 at -18 °C prior to analysis. 181
- 182 The water samples taken from the flume were spiked with 25 μL of 2 mg L^{-1} internal

standards, filtered through glass fiber filters (0.7-\mu GF/F, Whatman, UK). The pH of
filtered samples were adjusted to 3 using H_2SO_4 (30%, v/v). Na_2EDTA (0.2 g) was added into
the sample to minimize interference from Ca^{2+} and Mg^{2+} . An Oasis Hydrophile-Lipophile
Balance (HLB) cartridge (200 mg, 6 mL, Waters, US) preconditioned with 5 mL methanol
and 5 mL Milli-Q water was used to extract and clean up each water sample. The water
samples were passed through the HLB cartridges at a flow rate of 5-10 mL min ⁻¹ . The HLB
cartridges were washed by 5 mL Milli-Q water and were dried by vacuum pump for at least
10 min. The target compounds were eluted with 3 mL methanol/acetone (85:15, v/v) twice
(Hou et al., 2015). The 6 mL extract was blown to near dryness under a gentle stream of
nitrogen (37 °C) and dissolved in 1 mL of acetonitrile and Milli-Q water with 0.1% formic
acid (5:95, v/v). The final extracts were mixed by vortex mixer, ultrasonicated for 5 min, and
finally filtered through $0.22\text{-}\mu m$ PTFE syringe filters. The filtered extracts were stored at
-18°C prior to instrumental analysis.
The sediment samples were freeze-dried, ground, and then passed through a sieve (120 mesh).
Two-grams of sediment was transferred into centrifuge tubes, and then spiked with the
mixture of internal standards containing 50 ng of each compound. The spiked samples were
placed at 4 °C overnight. The extraction processes followed that of Zhou et al. (2012) with
some small modifications. Ten milliliter of acetonitrile and 10 mL of 0.1 M EDTA-Mcllvaine
buffer (Na ₂ EDTA:citric acid monohydrate:Na ₂ HPO ₄ ·12H ₂ O = 12.4:4.3:9.2, pH = 4) was
added to each centrifuge tube. The mixture was mixed by vortex mixer, ultrasonicated for 10
min, and then centrifuged for 5 min at 6000 rpm. This extraction step was repeated twice, and

- the supernatant at each step was merged into one bottle. The extracts were diluted with Milli-Q water to 200 mL. The solution was extracted and cleaned up by the same processes as those of water sample described above.
- 207 2.6. Instrumental analysis
- The target antibiotic compounds were determined by an Agilent 1290 rapid resolution liquid chromatography tandem an Agilent 6460 Triple Quadrupole mass spectrometer (RRLC-MS/MS, Agilent, US). The separation of target antibiotics compounds was accomplished by Agilent Zorbax Eclipse plus-C18 column (RRHD, 2.1 × 100 mm, 1.8 μm, Agilent, US). The mass spectrometer was operated in multiple-reaction monitoring (MRM) mode with positive ionization (ESI+). The instrumental conditions for the target compounds analysis are shown in Table S2.
- 2.7. Quality control and quality assurance
- The quantification of 14 antibiotics was achieved by using internal standard method with 216 calibration of working standard solutions. The correlation coefficients (R2) of calibration 217 curve were between 0.99 and 0.9999. The recoveries were performed by spiking 1 L water 218 samples and 2 g sediment samples with standard solutions to three concentrations of 10 ng 219 L⁻¹, 20 ng L⁻¹, 50 ng L⁻¹ and 10 ng g⁻¹, 20 ng g⁻¹, 50 ng g⁻¹, 100 ng g⁻¹, respectively. The 220 recoveries of 14 antibiotics in the water samples ranged from $56 \pm 1\%$ to $117 \pm 11\%$ and the 221 sediment samples ranged from $57 \pm 0.1\%$ to $127 \pm 5\%$, with relative standard deviation 222 (RSD) less than 15% (Table S3 and Table S4). The method detection limits (MDLs) of 14 223

- antibiotics ranged from 0.23 5.88 ng L^{-1} for water samples (Table S3) and from 0.25 2.94 ng g^{-1} for the sediment samples (Table S4). The MDLs were determined by spiking 1 L water samples with the mixed standard solution to 5 ng L^{-1} , and then performing the whole pre-treatment processes. The extracts were gradually diluted until the signal-to-noise ratio was equal to 3.
 - 2.7. Attenuation rates calculation for the flume study
 - After the antibiotics were spiked into the flume system, the concentrations would naturally decrease due to dilution and mixing with sediment and pore water. Therefore, in order to obtain the true attenuation rates without the influence of mixing dilution, concentration corrections were conducted by using a conservative tracer bromide as a reference compound (Eq. S1 in SI). One liter potassium bromide solution (3 g L^{-1}) was added into water phase of the flume system simultaneously with the antibiotics. Bromide concentrations in surface water and pore water samples were measured by ion chromatography (881 Compact, Metrohm, Switzerland) at the same intervals as the antibiotic determination. Attenuation rate constants (k) and half-life time ($t_{1/2}$) of antibiotics in the surface water were calculated by fitting a first-order kinetic decay model to these corrected concentrations (Eq. S2 and Eq.S3).
- 3. Results and discussion

- 3.1. Simulation system operation performance
- During the whole operating period, pH was around 7 and ORP was within 150-170 mV (Table S5). The conductivity was stable at around 300 µs cm⁻¹ (Table S5) in the flume, which

was higher than the on-site value of $80~\mu s~cm^{-1}$. However, the conductivities in this flume were not beyond the range values in the natural water (50-500 $\mu s~cm^{-1}$), which would not influence the experiments. The most likely explanation for the elevated conductivity comes from the addition of the potassium bromide tracer. The temperature was maintained around $25~^{\circ}C$ and the values of DO were between 7.2 to 9.6 mg L⁻¹ (Table S5). The TOCs in water and sediment measured at the initial period and end of the experiment did not show significant changes (Table S6). Therefore, the operational performance of this system was relatively stable, and close to water quality conditions of a natural river.

3.2. Dominant attenuation processes

The relative importance of the different attenuation processes for these antibiotics was investigated with batch studies. These used river water and sediment from the Tong Yang River, which had a pH of 7.8, conductivity of 80 µs cm⁻¹ and sediment TOC of 28.3 g kg⁻¹. Photodegradation proved to be the most important of these processes for all the antibiotics apart from OTC (Fig. 1). Photodegradation was a particularly important loss (estimated > 70% of the total) for SDZ, SMZ, STZ, EFC, and OFC. The half-lives controlled by photodegradation alone ranged from 4.40 to 32.9 d (Fig. S4). The importance of photodegradation for antibiotics has also been noted by others (Baena-Nogueras et al., 2017; Batchu et al., 2014; Conde-Cid et al. 2018; Li et al., 2018). Biodegradation and adsorption accounted for the rest of the losses from the water column. Adsorption was the most important process for OTC. Hydrolysis, judged on the basis of the dark sterile control, was not found to be important for any of the antibiotics over 30 d (< 1%).

3.3. Attenuation of target antibiotics in the surface water flume study

266	After 50 d, most of the macrolides, tetracyclines and fluoroquinolones had been lost from the
267	water column whilst the more persistent sulfonamides had around 20% remaining (Fig. 2).
268	The losses of the 14 antibiotics in the surface water were corrected for dilution by
269	comparison with the conservative tracer bromide which was simultaneously measured in the
270	surface water (Fig. S5). The k and $t_{1/2}$ of SMR, SMZ, SDM, SMX, STZ, CFC, and OTC were
271	calculated using a first-order kinetic model and these fitted well with the observations (R^2 >
272	0.8; p < 0.01); whilst the fitting for the others presented relatively weakly correlated fitting
273	$(0.6 < R^2 < 0.8; p < 0.01)$ (Fig. S6). The order of attenuation rate was OTC > RTM > TC >
274	$ETM > CFC > NFC > EFC > OFX > STZ > SMZ > SMX > SMR > SDM > SDZ \ (Table \ 1).$
275	Among these antibiotics, SDZ was the most persistent with 25.6-d $t_{1/2}$. The fluoroquinolones
276	presented moderate attenuation rates with k ranging from 0.06 d ⁻¹ and 0.13 d ⁻¹ . OTC and
277	RTM had the shortest half-lives (Table 1), and were completely removed after 15 d (Fig. 2).
278	Overall, the attenuation rates for these antibiotics are rather low and would allow them to
279	travel considerable distances down river. This persistence would increase on cloudy days due
280	to the low contribution of photodegradation.
281	Another way of examining the relevance of these loss rates is to consider the distances
282	travelled down a river after which 50% would be lost. Thus, for these antibiotics half would
283	be lost only following a river travel of 31 km to 444 km at 0.2 m s ⁻¹ velocity (Table 1). In the
284	case of the Nanfei River (a typical unban river close to the sampling sites) in winter which
	has a flow velocity 0.15 m s ⁻¹ , half of the antibiotics loss will take place following a travel
285	has a now velocity 0.15 m/s, han of the antibiotics loss will take place following a traver

distance of 23 km to 333 km. In summer, with a 3.5 m s⁻¹ flow velocity, the travel distance would range from 55 km to 777 km before half the antibiotics would be lost. In fact, for the Nanfei River, the distance from the urban discharge to Chaohu Lake is only 25 km. Thus, a considerable portion of the antibiotic discharge from Hefei City would reach this lake without dissipation.

3.4. Binding and movement of the antibiotics within pore waters and sediment

All antibiotics were detected in the surface sediments, which was attributed to their migration and adsorption to sediment from the water phase (Fig. 3). This is consistent with the adsorption mechanism playing a role in the removals of all antibiotics from the water phase in the batch experiments. For EFC, NFC, TC, OTC and RTM, adsorption played an important role its loss from the water column. This is related to the relatively high hydrophobicity of these compounds (150 < K_d < 889 L kg⁻¹) (Table S7). Adsorption made a relatively small contribution to the dissipation of sulfonamides (Fig. 1) due to the low adsorption affinity (K_d < 80 L kg⁻¹). Sulfonamides, as acidic compounds, have a declining sorption capacity with increasing alkalinity due to electrostatic repulsion from sediment (Gothwal and Shashidhar, 2015). Therefore, their adsorption capacity decrease in the weakly-alkaline water environment of the flume (Table S5).

The sulfonamides concentration in sediment continuously increased until around day 20 when it stabilized to 10 - 50 ng g⁻¹ at day 50. Similarly, the concentrations of the other antibiotics in the surface sediments increased rapidly before day 10, and then quickly

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declined until leading to their disappearance at day 50. The two types of behavior were attributed to the difference in their persistence. The sulfonamides have relatively low attenuation rates, thus the part transported to sediment only slowly dissipated compared with the tetracyline, macrolide and fluoroquinolone antibiotics which have higher attenuation rates. For OTC adsorption was shown to be particularly important in the batch studies (58.7%) (Fig. 1), which may explain why OTC had the highest concentration in sediment (Fig. 3). In addition, due to its relatively high biodegradation rate ($k = 0.0469 \text{ d}^{-1}$) (Fig. S4), OTC in sediment was rapidly dissipated after day 10 and almost disappeared at day 20 (Fig. 3), thus it could not be detected in the lower layer sediments (Fig. 4). All fluoroquinolones including EFC, NFC, OFC, and CFC were detected in the layer sediments at 5 cm, 10 cm and 15 cm depth (Fig. 4), which might be attributed to the relatively strong adsorption to sediment (Fig. 1) and the low attenuation rates (Fig. 1 and Table 1). Most of these antibiotics were detected in the pore water from the upper layer (Fig. 5 and Fig. S7). The sulfonamides were present in the upper layer pore water at the highest concentrations. SDZ, SMZ, SDM and STZ were found to be approaching equilibrium with the surface water until day 50, which might be attributed to the higher mobility caused by the low adsorption (Fig. 1). However, in the middle and lower layer pore water, only a few antibiotics were detected, which may have two explanations. One is the limited exchange of surface and pore water caused by the low flow velocity (0.2 m s⁻¹). Another is that most antibiotics were completely retained and dissipating in the upper layer due to the relatively high organic contents and fine texture of sediments (Table S6 and Table S8). Currently,

antibiotics have been detected in groundwater of different regions across the world (Kivits et al., 2018; Lopez-Serna et al., 2013; Ma et al., 2015). Thus, further clarification of their transport paths to groundwater is needed.

4. Conclusions

Photodegradation was the dominant attenuation mechanism of these antibiotics in a water-sediment system. These antibiotics had a wide-range of half-lives with 1.28 d and 25.7 d in the water column, which would permit considerable travel distances to take place for many of them in rivers. The adsorption onto the surface sediment in the flume study contributed to part of antibiotic removals from the water phase. All fluoroquinolones and two sulfonamides (SDZ and SDM) migrated further to the deep layer sediments, but there was less presence in the pore water due to the strong adsorption. Adsorption made a relatively small contribution to the dissipation of sulfonamides due to the low adsorption affinity. Thus, the sulfonamides were present in the pore water and approached equilibrium with the surface water at the upper layer, which would permit their high mobility to pore water in hyporheic zone. Therefore, these sulfonamides due to their high mobility and persistence might be important candidates for groundwater contamination.

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References

353

- Baena-Nogueras, R.M., Gonzalez-Mazo, E., Lara-Martin, P.A., 2017. Degradation kinetics of
- pharmaceuticals and personal care products in surface waters: photolysis vs biodegradation.
- 356 Sci. Total Environ. 590, 643–654.
- Batchu, S.R., Panditi, V.R., O'Shea, K.E., Gardinali, P.R., 2014. Photodegradation of
- antibiotics under simulated solar radiation: Implications for their environmental fate. Sci.
- 359 Total Environ. 470-471, 299-310.
- Bu, Q., Wang, B., Huang, J., Deng, S., Yu, G., 2013. Pharmaceuticals and personal care
- products in the aquatic environment in China: A review, J. Hazard. Mater. 262, 189–211.
- Conde-Cid, M., Fernández-Calviño, D., Nóvoa-Muñoz, J.C., Arias-Estévez, M., Díaz-Raviña,
- 363 M., Fernández-Sanjurjo, M.J., Núñez-Delgado, A., Álvarez-Rodríguez, E., 2018. Biotic
- and abiotic dissipation of tetracyclines using simulated sunlight and in the dark. Sci. Total
- 365 Environ. 635, 1520–1529.
- der Beek, T.A, Weber, F.A., Bergmann, A., Hickmann, S., Ebert, I., Hein, A., Kuster, A., 2016.
- Pharmaceuticals in the environment-global occurrences and perspectives. Environ. Toxicol.
- 368 Chem. 35, 823–835.

- 369 Elliott, A.H., Brooks, N.H., 1997. Transfer of nonsorbing solutes to a streambed with bed
- forms: Theory. Water Resour. Res. 33, 123–136.
- 371 Gothwal, R., Shashidhar, T., 2015. Antibiotic pollution in the environment: A review.
- 372 Clean-Soil Air Water, 43, 479–489.
- 373 Hou, J., Wan, W.N., Mao, D.Q., Wang, C., Mu, Q.H., Qin, S.Y., Luo, Y., 2015. Occurrence
- and distribution of sulfonamides, tetracyclines, quinolones, macrolides, and nitrofurans in
- livestock manure and amended soils of Northern China. Environ. Sci. Pollut. R. 22,
- 376 4545-4554.
- Jin, G., Tang, H., Gibbes, B., Li, L., Barry, D.A., 2010. Transport of nonsorbing solutes in a
- streambed with periodic bedforms. Adv. Water Resour. 33, 1402–1416.
- Johnson, A.C., Keller, V., Dumont, E., Sumpter, J.P., 2015. Assessing the concentrations and
- risks of toxicity from the antibiotics ciprofloxacin, sulfamethoxazole, trimethoprim and
- erythromycin in European rivers. Sci. Total Environ. 511, 747–755.
- Kaeseberg, T., Zhang, J., Schubert, S., Oertel, R., Krebs, P., 2018. Abiotic, biotic and
- photolytic degradation affinity of 14 antibiotics and one metabolite batch experiments
- and a model framework. Environ. Pollut. 241, 339–350.
- Kivits, T., Broers, H.P., Beeltje, H., van Vliet, M., Griffioen, J., 2018. Presence and fate of
- veterinary antibiotics in age-dated groundwater in areas with intensive livestock farming."
- 387 Environ. Pollut. 241, 988-998.
- 388 Kümmerer, K., 2009a. Antibiotics in the aquatic environment A review Part I.
- 389 Chemosphere 75(4), 417-434.
- 390 Kümmerer, K., 2009b. Antibiotics in the aquatic environment A review Part I.

- 391 Chemosphere 75(4), 435-441.
- 392 Li, S., Shi, W., Liu, W., Li, H., Zhang, W., Hu, J., Ke, Y., Sun, W., Ni, J., 2018. A
- duodecennial national synthesis of antibiotics in China's major rivers and seas (2005–2016).
- 394 Sci. Total Environ. 615, 906–917.
- Li, Y., Rashid, A., Wang, H., Hu, A., Lin, L., Yu, C.P., Chen, M., Sun, Q., 2018. Contribution
- of biotic and abiotic factors in the natural attenuation of sulfamethoxazole: A path analysis
- 397 approach. Sci. Total Environ. 633, 1217-1226.
- Li, Z., Sobek, A., Radke, M., 2015. Flume experiments to investigate the environmental fate
- of pharmaceuticals and their transformation products in streams. Environ. Sci. Technol. 49,
- 400 6009-6017.
- Lopatkin, A.J., Huang, S., Smith, R.P., Srimani, J.K., Sysoeva, T.A., Bewick, S., Karig, D.K.,
- You, L., 2016. Antibiotics as a selective driver for conjugation dynamics. Nat. Microbiol. 1,
- 403 16044.
- Lopez-Serna, R., Jurado, A., Vazquez-Sune, E., Carrera, J., Petrovic, M., Barcelo, D., 2013.
- Occurrence of 95 pharmaceuticals and transformation products in urban groundwaters
- underlying the metropolis of Barcelona, Spain. Environ. Pollut. 174,305-315.
- Luo, Y., Xu, L., Rysz, M., Wang, Y., Zhang, H., Alvarez, P.J.J., 2011. Occurrence and
- Transport of Tetracycline, Sulfonamide, Quinolone, and Macrolide Antibiotics in the
- Haihe River Basin, China. Environ. Sci. Technol. 45, 1827–1833.
- Ma, Y.P., Li, M., Wu, M.M., Li, Z., Liu, X., 2015. Occurrences and regional distributions of
- 411 20 antibiotics in water bodies during groundwater recharge. Sci. Total Environ. 518,
- 412 498-506.

- 413 Qiao, M., Ying, G.G., Singer, A.C., Zhu Y.G., 2018. Review of antibiotic resistance in China
- and its environment. Environ. Int. 110, 160-172.
- Ramil, M., El Aref, T., Fink, G., Scheurer, M., Ternes, T.A. 2010. Fate of Beta Blockers in
- Aquatic-Sediment Systems: Sorption and Biotransformation. Environ. Sci. Technol. 44,
- 417 962–970.
- Verraes, C., Van Boxstael, S., Van Meervenne, E., Van Coillie, E., Butaye, P., Catry, B., de
- Schaetzen, M.A., Van Huffel, X., Imberechts, H., Dierick, K., Daube, G., Saegerman, C.,
- De Block, J., Dewulf, J., Herman L., 2013. Antimicrobial Resistance in the Food Chain: A
- Review. Int. J. Environ. Res. Public Health 10, 2643–2669.
- Wang, F., Stedtfeld, R.D., Kim, O.S., Chai, B., Yang, L., Stedtfeld, T.M., Hong S.G., Kim,
- D., Lim, H.S., Hashsham, S.A., Tiedje, J.M., Sul, W.J., 2016. Influence of soil
- characteristics and proximity to Antarctic research stations on abundance of antibiotic
- resistance genes in soils. Environ. Sci. Technol. 50, 12621–12629.
- 426 Ying, G.G., He, L.Y., Ying, A.J., Zhang, Q.Q., Liu, Y.S., Zhao, J.L., 2017. China must
- reduce its antibiotic use. Environ. Sci. Technol. 51, 1072–1073.
- Zhang, Q.Q., Ying, G.G., Pan, C.G., Liu, Y.S., Zhao, J.L., 2015. Comprehensive evaluation of
- antibiotics emission and fate in the river basins of China: source analysis, multimedia
- modeling, and linkage to bacterial resistance, Environ. Sci. Technol. 49, 6772–6782.
- Zhou, L.J., Ying, G.G., Liu, S., Zhao, J.L., Chen, F., Zhang, R.Q., Peng, F.Q., Zhang Q.Q.,
- 432 2012. Simultaneous determination of human and veterinary antibiotics in various
- environmental matrices by rapid resolution liquid chromatography–electrospray ionization
- tandem mass spectrometry. J. Chromatogr. A 1244, 123–138.

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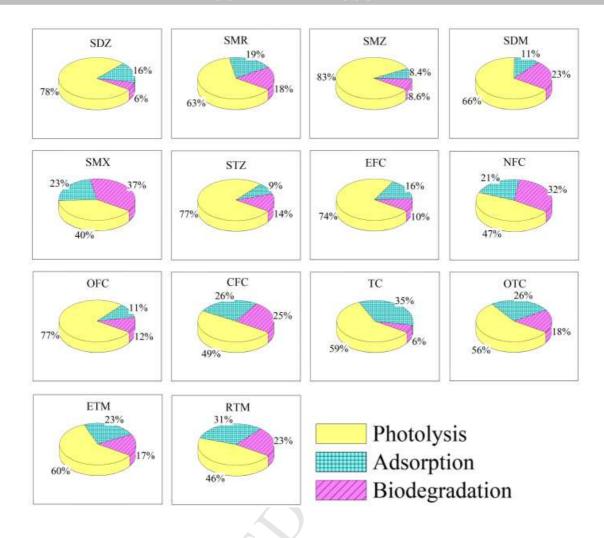
- 437 Figure captions
- 438 Fig. 1. Contribution ratio of each individual attenuation processes (including
- 439 photodegradation, adsorption and biodegradation) in the batch experiments carried out at
- 440 25 °C over 30 d periods.
- 441 Fig. 2. Temporal profiles of four classes of antibiotics in surface water of the flume study
- over a 50 d period (mean \pm standard deviation).
- Fig. 3. Temporal profiles of four classes of antibiotics in surface sediment of the flume study
- over a 50 d period (mean \pm standard deviation).
- Fig. 4. Antibiotics in the upper, middle and lower layer sediment at the three sampling time
- (day 10, day 20 and day 30). The depth of the upper, middle and lower layers was 5 cm,
- 447 10cm and 15 cm, respectively.
- 448 Fig. 5. Dynamic equilibrium relationships of 14 antibiotics in the pore water with those in the
- surface water at pore 1. The depth of the upper, middle and lower layers was 5 cm, 10 cm and
- 450 15 cm, respectively.

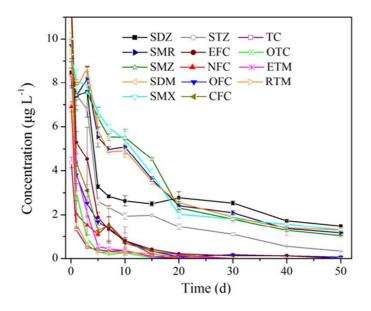
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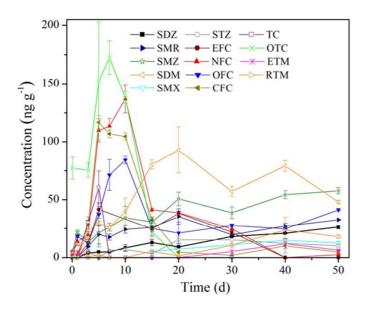
Table 1 Attenuation rate constants (k), half-life time ($t_{1/2}$) and half-life distance (d_h)* of 14 antibiotics in surface water for the flume experiment.

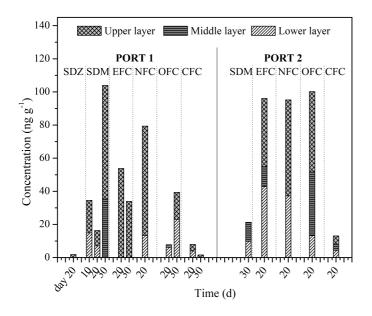
Compound	$k (d^{-1})$	t _{1/2} (d)	d _h (km)	Compound	$k (d^{-1})$	t _{1/2} (d)	d _h (km)
SDZ	0.027	25.7	444	NFC	0.123	5.64	97
SMZ	0.040	17.3	299	OFC	0.062	11.1	192
SMX	0.039	17.8	308	CFC	0.130	5.33	92
SDM	0.038	18.2	314	TC	0.167	4.15	72
SMR	0.039	17.9	309	отс	0.380	1.82	31
STZ	0.052	13.3	230	ETM	0.164	4.22	73
EFC	0.079	8.78	152	RTM	0.251	2.76	48

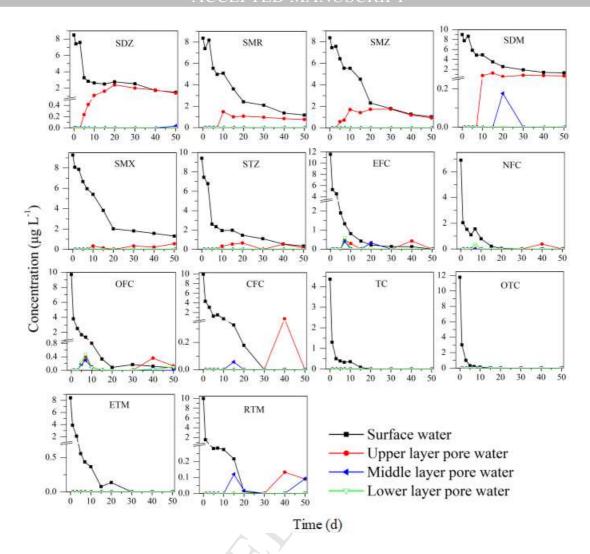
 d_h was calculated by the average flow velocity (0.2 m $\ensuremath{s^{-1}}\xspace$) multiplying $t_{1/2}$











Highlight:

- Attenuation of 14 antibiotics was studied for 50 d in a simulated stream.
- Persistence was in order of sulfonamides > quinolones > macrolides > tetracyclines.
- Photodegradation was the dominant attenuation mechanism.
- All quinolones were detected in the lower layer sediments at 15 cm depth.
- Sulfonamides were present in the sediment pore water with high concentrations.

Declaration of interests
oxtimes The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.
□The authors declare the following financial interests/personal relationships which may be considered as potential competing interests: