



Article (refereed) - postprint

Wang, Jinyang; Hayes, Felicity; Chadwick, David R.; Hill, Paul W.; Mills, Gina; Jones, Davey L.. 2019. **Short-term responses of greenhouse gas emissions and ecosystem carbon fluxes** to elevated ozone and N fertilization in a temperate grassland. *Atmospheric Environment*, 211. 204-213. https://doi.org/10.1016/j.atmosenv.2019.05.027

© 2019 Elsevier Ltd

This manuscript version is made available under the CC-BY-NC-ND 4.0 license http://creativecommons.org/licenses/by-nc-nd/4.0/

(CC) BY-NC-ND

This version available http://nora.nerc.ac.uk/id/eprint/523603/

NERC has developed NORA to enable users to access research outputs wholly or partially funded by NERC. Copyright and other rights for material on this site are retained by the rights owners. Users should read the terms and conditions of use of this material at http://nora.nerc.ac.uk/policies.html#access

NOTICE: this is the author's version of a work that was accepted for publication in *Atmospheric Environment*. Changes resulting from the publishing process, such as peer review, editing, corrections, structural formatting, and other quality control mechanisms may not be reflected in this document. Changes may have been made to this work since it was submitted for publication. A definitive version was subsequently published in *Atmospheric Environment*, 211. 204-213. https://doi.org/10.1016/j.atmosenv.2019.05.027

www.elsevier.com/

Contact CEH NORA team at noraceh@ceh.ac.uk

The NERC and CEH trademarks and logos ('the Trademarks') are registered trademarks of NERC in the UK and other countries, and may not be used without the prior written consent of the Trademark owner.

1	Short-term responses of greenhouse gas emissions and ecosystem carbon fluxes to
2	elevated ozone and N fertilization in a temperate grassland
3	Jinyang Wang ^{a,*} , Felicity Hayes ^b , David R. Chadwick ^a , Paul W. Hill ^a , Gina Mills ^b , Davey L.
4	Jones ^{a,c}
5	^a School of Natural Sciences, Environment Centre Wales, Bangor University, Bangor, Gwynedd
6	LL57 2UW, UK
7	^b Centre for Ecology and Hydrology, Environment Centre Wales, Bangor, Gwynedd LL57 2UW,
8	UK
9	^c UWA School of Agriculture and Environment, University of Western Australia, Crawley, WA
10	6009, Australia
11	*Corresponding author.
12	E-mail address: jy.wang@bangor.ac.uk (J. Wang)
13	
14	
15	
16	

Abstract

17

18

19

20

21

22

23

24

25

26

27

28

29

30

31

32

33

34

35

36

Growing evidence suggests that tropospheric ozone has widespread effects on vegetation, which can contribute to alter ecosystem carbon (C) dynamics and belowground processes. In this study, we used intact soil mesocosms from a semi-improved grassland and investigated the effects of elevated ozone, alone and in combination with nitrogen (N) fertilization on soil-borne greenhouse gas emissions and ecosystem C fluxes. Ozone exposure under fully open-air field conditions was occurred during the growing season. Across a one-year period, soil methane (CH₄) and nitrous oxide (N₂O) emissions did not differ between treatments, but elevated ozone significantly depressed soil CH₄ uptake by 14% during the growing season irrespective of N fertilization. Elevated ozone resulted in a 15% reduction of net ecosystem exchange of carbon dioxide, while N fertilization significantly increased ecosystem respiration during the growing season. Aboveground biomass was unaffected by elevated ozone during the growing season but significantly decreased by 17% during the non-growing season. At the end of the experiment, soil mineral N content, net N mineralization and extracellular enzyme activities (i.e., cellobiohydrolase and leucine aminopeptidase) were higher under elevated ozone than ambient ozone. The short-term effect of single application of N fertilizer was primarily responsible for the lack of the interaction between elevated ozone and N fertilization. Therefore, results of our short-term study suggest that ozone exposure may have negative impacts on soil CH₄ uptake and C sequestration and contribute to accelerated rates of soil Ncycling.

- 37 Keywords: Air pollutant; Fertilizer management; Ground level O₃; Plant-soil feedbacks; Soil
- 38 nutrient cycling

1. Introduction

40

41

42

43

44

45

46

47

48

49

50

51

52

53

54

55

56

57

58

59

Owing to the global increase in ozone precursor emissions (i.e., nitrogen oxides, carbon monoxide and volatile organic compounds), further increases of background ozone concentrations in the Northern Hemisphere may occur over this century unless precursor emissions are effectively controlled (Fowler et al., 2008; Meehl et al., 2007). Tropospheric ozone is not only the third-most-important contributor to the human-induced greenhouse gas (GHG) effect after carbon dioxide (CO₂) and methane (CH₄) (IPCC, 2007), but also the important gaseous air pollutant in terms of its effects on net primary production, soil carbon (C) sequestration, as well as other ecosystem services (Ainsworth et al., 2012; Sicard et al., 2017). Ozone may have important indirect effects on global change through its effects on CH₄ and nitrous oxide (N2O) emissions. In the past decade, the effects of elevated ozone on CH4 and N₂O emissions, especially from agroecosystems, have been increasingly studied. For example, studies in rice fields conducted under either free-air open conditions or the open-top chambers (OTC) have consistently shown that CH₄ emissions were substantially less under elevated ozone than under ambient or charcoal-filtered air conditions (Bhatia et al., 2011; Tang et al., 2015; Zheng et al., 2011). The impacts of elevated ozone on N₂O emissions have been examined in a range of agricultural systems, including rice fields (Bhatia et al., 2011; Kou et al., 2015; Tang et al., 2015), a soybean field (Decock et al., 2012), an annual grassland (Sánchez-Martín et al., 2017) and a meadow ecosystem (Kanerva et al., 2007). However, the results from these experiments are conflicting, showing either positive (Sánchez-Martín et al.,

60 2017), negative (Bhatia et al., 2011; Kanerva et al., 2007) or no significant (Decock et al., 2012; Kou et al., 2015; Tang et al., 2015) responses of N₂O emissions to ozone exposure. Such 62 inconsistencies may stem from the complex impacts of other factors (e.g. exposure 63 methodology, exposure duration and plant type etc.) on the responses of N₂O emissions to 64 elevated ozone (Tang et al., 2015).

61

65

66

67

68

69

70

71

72

73

74

75

76

77

78

79

On the other hand, ozone may also have important indirect effects on global change through its effect on terrestrial C sequestration. At the global scale, Sitch et al. (2007) used a global land C cycle model by including the ozone deposition effect and demonstrated that the negative impact of elevated ozone on plant productivity may result in a significant suppression of the global land-C sink. Such modeling studies (Sitch et al., 2007), however, are based on the effects of ozone on photosynthetic rates and aboveground growth, and do not consider its effects on soil C fluxes. Furthermore, studies now support the view that ozone may also have a profound impact on belowground processes because of altered C allocation to the roots and associated rhizosphere (Andersen, 2003; Fuhrer et al., 2016; Grantz et al., 2006; Wang et al., 2019). To better understand how increasing tropospheric ozone will affect global C fluxes, studies addressing ozone effects on soil C fluxes are therefore required.

Net ecosystem exchange (NEE) of CO₂ represents the balance between gross ecosystem productivity (GEP) and ecosystem respiration (R_{eco}) (Niu et al., 2010). It has been suggested that directly measuring NEE can be used to evaluate the response of ecosystem C sequestration to climatic factors (Luyssaert et al., 2007). To date, divergent responses of ecosystem C fluxes to elevated ozone have been reported in an annual grassland (Calvete-Sogo et al., 2014), peatlands (Haapala et al., 2011; Toet et al., 2011), and a subalpine grassland (Volk et al., 2011). These contrasting results of studies from different ecosystem types under distinct ozone exposure conditions signify a limited understanding of ozone effects on ecosystem C fluxes.

Despite considerable awareness of the potential interaction between elevated ozone and nitrogen (N) fertilization, very little attention has been paid to investigate their combined effects on ecosystem production and function (Mills et al., 2016). In a seven-year study from a subalpine grassland, plant biomass was positively affected by N addition but neither ozone nor its interaction with N addition (Volk et al., 2014). In contrast, results of a recent meta-analysis show that elevated ozone generally tends to exacerbate the negative impact of limiting N on plant growth (Yendrek et al., 2013). Similarly, in an annual grassland ozone-induced reductions in N content and C/N ratio of plant biomass can be counterbalanced by N addition (Sánchez-Martín et al., 2017), despite the absence of their interaction on GHG emissions and ecosystem C fluxes. Altogether, these conflicting results suggest that the direction and magnitude of the combined effect of elevated ozone and N fertilization on ecosystem processes remains unpredictable.

As the dominant form of agriculture by land area, grasslands represent over two thirds of utilized agricultural land area in the UK (Defra, 2016). Semi-natural vegetation is well known to be adversely affected by elevated ozone concentrations across Europe (Hayes et al., 2007; Mills et al., 2011). The aforementioned studies have mainly focused on croplands and annual

grasslands, but less is known about how elevated ozone and its interaction with N fertilization affect ecosystem production and function (e.g. fodder production, trace gas emissions and soil properties) in temperate grasslands (Wang et al., 2019). Our objectives for this study were therefore (i) to determine the effects of elevated ozone on CH₄ and N₂O fluxes from a temperate grassland in the UK throughout a one-year period; (ii) to investigate how elevated ozone would affect ecosystem C fluxes during the growing season (i.e., ozone exposure period); and (iii) to test whether the responses of these fluxes to elevated ozone would be interacted with N fertilization. To this end, we carried out a field experiment with intact soil mesocosms from a temperate semi-improved grassland and exposed them to elevated ozone for one growing season using a ozone free-air controlled enrichment (O₃-FACE) platform.

2. Materials and methods

2.1 Experimental design

The experiment was conducted in 2017 in a O₃-FACE system at CEH Bangor Air Pollution Facility, Abergwyngregyn, North Wales, UK (13 m asl, 53°15'N, 4°01'W). The climate at the site is classed as temperate-oceanic with a mean annual soil temperature of 11°C at 10 cm depth and a mean annual rainfall of 1250 mm. Experimental plots consisted of 16 intact soil mesocosms (31 cm diameter × 25 cm deep), which were excavated in the early spring of 2017 from a semi-improved upland grassland located at the Henfaes Research Station, Abergwyngregyn, North Wales, UK (53°13'N, 4°0'W). The upland site is located at approximately 270 m altitude with a mean growing season temperature of 10 °C and a mean

annual precipitation of 1200 mm. This semi-improved grassland site had not received any fertilizer applications and is grazed at low stocking densities (1-2 ewes ha⁻¹). The vegetation was classified as *Cynosurus cristatus-Centaurea nigra* grassland (NVC MG5; Rodwell, 1992). The soil is classified as an Orthic Podzol (FAO, 1981). The soil of this site had a slightly acidic pH of 5.39±0.04, total soil C of 8.45±0.09%, and total N of 0.81±0.01%.

Four treatments were established to determine the effects of elevated ozone, N fertilization and their potential interaction. The low and high ozone treatments from the O₃-FACE system as described below (hereafter called the ambient and elevated ozone treatments, respectively) were used in this study. The exposure period of ozone was from May 26 to October 9, 2017. Each quarter of these mesocosms was randomly assigned to one of the four combinations of ozone and N fertilizer treatments. Nitrogen was added as NH₄NO₃ at a rate of 100 kg N ha⁻¹ dissolved in 200 mL of deionized water per mesocosm on May 26, 2017. For the control treatment, an equal amount of deionized water was added. To ensure all other macro-nutrients were not limiting pasture growth, and that any nutrient response was the result of the N addition, calcium superphosphate for P and potassium chloride for K were also used and applied to all treatments at a rate of 10 and 50 kg ha⁻¹, respectively. The fertilizer addition rates were based on national guidelines.

The O_3 -FACE system was established in the spring of 2014, consisting of nine rings of 4 m diameter as described previously (Wang et al., 2019). Breifly, the rings were arranged in a replicated 3 \times 3 Latin square with 10 m between the centers of each ring. Low (ambient air)

and high ozone (ambient air + 20 ppb) treatments were used in this study. Ozone was generated from oxygen concentrated from air (Integra 10, SeQual) using a G11 ozone generator (Pacific Ozone). Small fans (200 mm, Explair) were used to push the ozone through the delivery pipe (65 mm, with 3 mm holes every 10 cm). Ozone delivery was achieved via computer controlled (LabView version 2012) solenoid valves operating using pulse width modulation. Wind speed was monitored continuously (WindSonic, Gill Instruments Ltd, UK) and was used to instantaneously adjust solenoid operation and thus ozone delivery. Ozone release was reduced at wind speeds below 16 m s⁻¹ and did not occur when wind speeds fell below 2 m s⁻¹. At very high wind speeds the ozone concentrations may not be well controlled and thus did not reach the target maximum concentrations. Despite this, we still got elevated ozone with the 'high' ozone treatment compared to the low ozone treatment as the solenoid valves were <1 m from the O₃-FACE rings, the response time of ozone delivery to track windspeed was fast. Ozone was sampled adjacent to the plants in each ring at a height of 30 cm for approximately 3.5 min in every 30-min using an ozone analyzer (ThermoScientific, Model 49i, Reading, UK).

2.2 Soil GHG emissions and ecosystem C fluxes

140

141

142

143

144

145

146

147

148

149

150

151

152

153

154

155

156

157

158

159

Soil GHG fluxes were measured throughout the experimental year. Gas samples were collected by placing a handmade opaque chamber over the intact soil mesocosms and sealing with a wide rubber band to ensure that the headspace inside the chamber was air-tight. Three gas samples were taken within a 40-min enclosure time and immediately transferred into pre-evacuated 20-mL screw-cap vials (QUMA Elektronik & Analytik GmbH, Germany). Gas samples were

stored under positive pressure and analyzed using a Perkin Elmer Clarus 580 gas chromatograph equipped with a Turbo Matrix 110 autosampler (PerkinElmer Inc., Shelton, CT, USA). CH₄ and CO₂ were detected with a flame ionization detector connected to a methanizer and N₂O with a ⁶³Ni electron-capture detector. Three standard gases were used for calibration with concentrations of 1.42, 2.92, and 10.4 ppm CH₄; 258, 496 and 1276 ppm CO₂; and 308, 641, and 1500 ppb N₂O (BOC Gases Ltd, Guildford, UK). Gas fluxes were calculated from the concentration change in the chamber versus time and were adjusted for air temperature and atmospheric pressure at the time of sampling. The annual fluxes of these gases were approximated by applying the trapezoid rule assuming constant flux rates per day.

Ecosystem C fluxes were measured during the growing season using a static-chamber method, which has been used and validated in previous studies (Niu et al., 2010; Volk et al., 2011). The measurements were made from June to September in 2017 and was taken between 8:00 and 11:00 am on sunny days. If it is rained or cloudy on a scheduled sampling date, we postponed the measurements and selected the next sunny day. In total, complete sets of measurements were made seven times during the growing season. A portable infrared gas analyzer (EGM-4 Environmental Gas Monitor for CO₂; PP Systems Ltd, Hitchin, UK), which was attached to a handmade transparent plastic chamber, was employed to sample and measure CO₂ concentration ([CO₂]) *in situ* as described elsewhere (Williamson et al., 2016). A small fan created moderate turbulence inside the chamber to facilitate air mixing. A temperature probe was also installed inside to determine the chamber air temperature. During the measurement, the chamber was tightly sealed to the rim of the mesocosm using a cell foam band. The chamber

[CO₂] were consecutively recorded at 20 s intervals during a 2-min measurement period per mesocosm. The chamber [CO₂] did not drop below 340 ppm or rise above 500 ppm. After the net ecosystem exchange (NEE) of CO₂ measurement, the chambers were opened to allow the chamber [CO₂] to return to the ambient atmospheric concentration. Subsequently, the chamber was again placed over the mesocosm and covered with a shade cloth, and the [CO₂] measurement was repeated over a 2-min period (i.e., dark respiration). Due to the light removal inside the chamber, the data of the chamber [CO₂] were considered to represent ecosystem respiration ($R_{\rm eco}$). To eliminate the disturbance effect during measurement, the first 20 s of data was omitted in subsequent analysis to allow for initial adjustment of the chamber [CO₂]. The quality of the measurement was considered acceptable if a linear regression of [CO₂] versus time during the following 100 s yielded $R^2 \ge 0.9$, indicating strictly linear changes in the chamber [CO₂]. Gross ecosystem productivity (GEP) was computed by the sum of absolute values of NEE and $R_{\rm eco}$. By convention, negative NEE values refer net C uptake by the ecosystem, while positive NEE values represent net C loss from the ecosystem.

2.3 Soil abiotic and biotic properties and plant analysis

181

182

183

184

185

186

187

188

189

190

191

192

193

194

195

196

197

198

199

200

Air temperature and rainfall were recorded hourly on site with an automated weather station. Parallel to gas sampling and ecosystem C flux measurements, soil temperature and volumetric water content (v/v, %) at a depth of 10 cm in each mesocosm were measured by a handheld thermometer (HI98509 Checktemp[®]1; Hanna Instruments Ltd, Leighton Buzzard, UK) and moisture sensor (ML3 ThetaProbe; Delta-T, Cambridge, UK), respectively. Water-filled pore

space (WFPS) was calculated by dividing volumetric water content by total porosity (Linn and Doran, 1984). Total porosity was calculated as [1-(bulk density/particle density)] \times 100% and using a particle density of 2.65 g cm⁻³.

201

202

203

204

205

206

207

208

209

210

211

212

213

214

215

216

217

218

219

220

221

In May 2018, two soil cores (10-cm deep; 5-cm diameter) were collected from each mesocosm. Fresh soils were passed through a 2-mm sieve to remove visible plant material and small stones, and then placed at 4 °C to await analysis. Soil water content was determined gravimetrically by drying soil in an oven (24 h, 105 °C). Available soil C and N pools were quantified by extracting soil subsamples with 0.5 M K_2SO_4 (1:5 w/v). The concentrations of ammonium (NH₄⁺) and nitrate (NO₃⁻) in the extracts were determined via the colorimetric salicylate procedure of Mulvaney (1996) and the vanadate method of Miranda et al. (2001), respectively. For soil microbial biomass, additional subsamples were fumigated for 24 h with chloroform and similarly extracted with 0.5 M K₂SO₄ (1:5 w/v) (Vance et al., 1987). Dissolved organic C (DOC) and total dissolved N (TDN) in the 0.5 M K₂SO₄ extracts (fumigation and non-fumigation samples) were quantified using a Multi N/C 2100 TOC analyzer (AnalytikJena, Jena, Germany). Dissolved organic N (DON) was calculated as the difference between TDN and soil mineral N. Microbial biomass C and N concentrations were corrected using correction factors of 0.45 for C and 0.54 for N (Brookes et al., 1985; Wu et al., 1990). Total C (TC) and N (TN) contents of ground soil and plant samples were determined with a TruSpec® elemental analyzer (Leco Corp., St Joseph, MI). Net N mineralization and nitrification rates were determined by the aerobic incubation of soil samples for 14 days at 10 °C in the dark (Hart et al., 1994), followed by extraction with 0.5 M K₂SO₄ and analyzing for soil mineral N as

described above. To assess the effects of elevated ozone and N fertilization on hydrolytic extracellular enzyme activities, the potential activities of four extracellular enzymes: β glucosidase, cellobiohydrolase, N acetyl glucosaminidase and leucine aminopeptidase were measured according to the fluorometric protocol (DeForest, 2009; Saiya-Cork et al., 2002).

Plant biomass was measured twice by cutting all plants at 2 cm above the ground at the end of growing season (early November of 2017) and in May 2018, respectively. For root biomass measurement, two additional soil cores (10-cm deep; 5-cm diameter) was sampled from each mesocosm. The harvested soil cores were rinsed thoroughly and passed through a 0.5-mm aperture sieve and the root fragments remaining in the sieve recovered. All plant samples were oven-dried at 65 °C and weighed to determine their dry biomass.

232 2.4 Statistical analysis

Each parameter was tested for normal distribution (using Shapiro-Wilk's test) and equality of variance (using Levene's test), and parameters with non-normal distributions or unequal variances were either logarithmically or square-root transformed when necessary. For analysis of time-series data (i.e., repeated measurements of soil gas fluxes and soil factors), we used linear mixed effects models (LME, *package* LME4; Bates et al. 2014) to test for the fixed effects of ozone, N fertilization and their possible interaction. The spatial replication and time (sampling days) were included as random effects. If the Akaike's information criterion (AIC) showed an improvement in the LME models, we included a first-order temporal autoregressive function to account for the decreasing correlation of the measurements with increasing time

and/or a variance function (varIdent) to account for heteroscedasticity in the fixed-factor variances (Crawley, 2012). We used linear mixed effects models to test the fixed effects of ozone, N fertilization and their interaction on soil properties, extracellular enzyme activities and plant biomass. Multiple comparisons were made using the Tukey HSD test.

To assess the relationships between soil GHG emissions or ecosystem C fluxes and soil factors (temperature and WFPS), we used the mean values of the four replicate mesocosms on each sampling day, and conducted Pearson correlation tests over the entire sampling period. In all statistical tests, differences among treatments were considered significant at $P \le 0.05$ and marginally significant at $P \le 0.09$. Statistical analyses were performed in R version 3.2.2 (R Development Core Team, 2015).

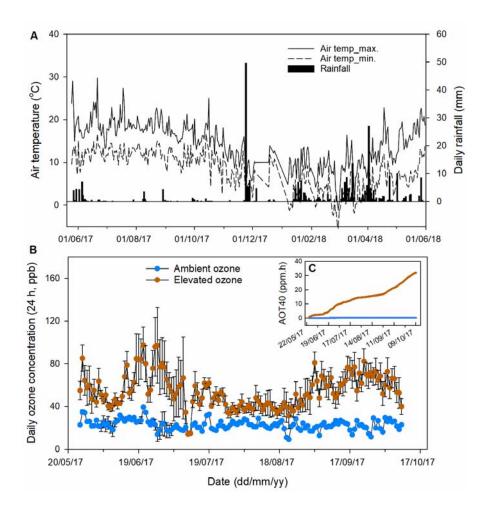


Figure 1. Daily maximum and minimum air temperature and daily rainfall (A) over the period of May 25, 2017 to May 31, 2018, daily ozone concentration (B) and AOT40 during daylight hours (C) from ambient and elevated ozone treatments in the 2017 growing season. Values represent mean \pm SEM (n = 3).

3. Results

3.1 Climatic conditions and ozone exposure

Over the experimental period, air temperature ranged from -5 °C to 30 °C, with an annual average of 11 °C (Fig. 1A). The total rainfall was 324 mm, of which only 13% occurred during

the growing season (May–September 2017). Intact soil mesocosms were exposed to ozone in the field from May 25 to October 9, 2017, with a total of 136 days effective exposure (Fig. 1B). During the exposure period, the 24-h mean ozone concentrations were 22.9±0.6 ppb and 54.9±6.1 ppb in the ambient and elevated ozone treatments, respectively. As shown in Fig. 1C, AOT40 (accumulated ozone exposure over a threshold of 40 ppb) in the elevated ozone treatment was 31.9±10.1 ppm.h being markedly higher than that (< 0.5 ppm.h) in the ambient treatment.

3.2 Seasonal variability of soil microclimate and GHG emissions

Soil temperature followed a clear seasonal pattern with an annual mean of 13.7 °C across all treatments (Fig. 2A). Soil temperatures were highest in July 2017 and lowest during January 2018. Soil WFPS varied seasonally in response to rainfall and ranged from 12.6% to 87.6% (Fig. 2B). Soil WFPS periodically increased following summer rainfall. Soil water contents steadily increased during the winter season when a large proportion of the annual rainfall occurred. Neither soil temperature nor WFPS differed between treatments (all P > 0.1).

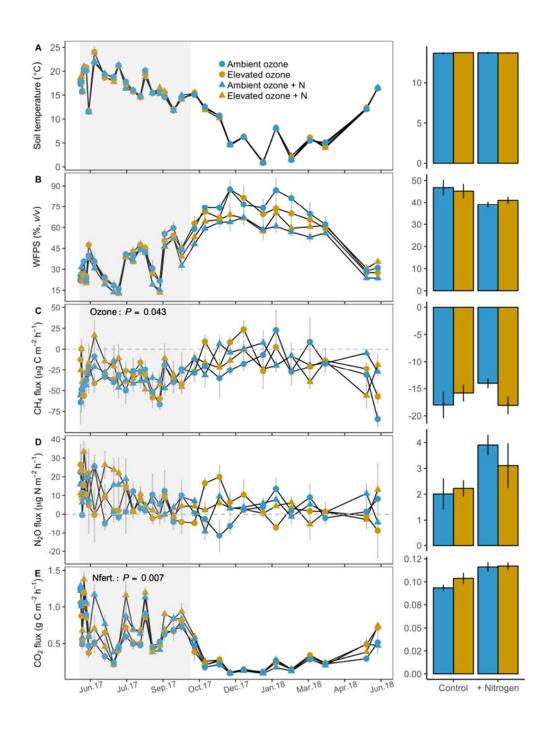


Figure 2. Soil temperature (A), water-filled pore space (WFPS) (B) at a depth of 10 cm, and year-round fluxes of CH_4 (C), N_2O (D) and night-time CO_2 (E) from a temperate semi-improved grassland exposed to different levels of ozone and N. The averages of soil microclimate parameters and gas fluxes over the experimental year are shown in the right panel. Values represent mean \pm SEM (n = 4). The horizontal dashed line marks the zero flux. Grey shading

marks the ozone exposure period during the growing season. Results from the linear mixed effects model at $P \le 0.05$ are shown.

This semi-improved grassland was a small sink of atmospheric CH₄. Across all treatments, CH₄ fluxes ranged from -84.1 to $23.9 \,\mu g$ C m⁻² h⁻¹ (Fig. 2C). During the phase of ozone exposure, CH₄ uptake rates were significantly affected by elevated ozone with a 14% reduction (P = 0.043) regardless of N fertilizer. A marginally significant interaction between elevated ozone and N fertilizer was detected during the winter season (P = 0.077). In parallel with high soil WFPS, CH₄ fluxes tended to be high and occasionally positive during the winter season. Overall, CH₄ fluxes were negatively correlated with soil temperature and positively correlated with soil WFPS (r = -0.41 and 0.51, both P < 0.001; Table 1), indicative of an increase in CH₄ uptake rates with an increase in soil temperature but a decrease of that with an increase in soil water content. Because of the large spatial and temporal variations, neither elevated ozone nor N fertilization had a significant effect on CH₄ emissions throughout the experimental period.

As with CH₄, N₂O emissions were temporally and spatially variable (Fig. 2D). Across all treatments, N₂O fluxes ranged from -11.6 to 33.3 μ g N m⁻² h⁻¹. On a few occasions, fluxes were negative indicating the occurrence of N₂O consumption by the soil. In contrast to CH₄, N₂O fluxes correlated positively with soil temperature and negatively with soil WFPS (r = 0.36 and -0.33, both P < 0.001; Table 1). For both seasons, neither elevated ozone nor N fertilizer affected N₂O emissions.

Table 1 Pearson correlation coefficients between greenhouse gas (CH₄, N₂O and night-time CO_2) fluxes or ecosystem C fluxes and soil environmental factors (soil temperature and water-filled pore space (WFPS) at a depth of 10 cm). NEE, net ecosystem exchange of CO_2 ; R_{eco} , ecosystem respiration; GEP, gross ecosystem productivity.

Parameter	n	Soil temperature	WFPS				
Greenhouse gas fluxes							
CH ₄	123	-0.41***	0.51***				
N_2O	123	0.36***	-0.33***				
Night-time CO ₂	123	0.69***	-0.53***				
Ecosystem C fluxes							
NEE	28	0.04	-0.67***				
$R_{ m eco}$	28	0.45^{*}	0.44^*				
GEP	28	0.26	0.67***				

*, **, *** Values are significantly different from zero at P < 0.05, 0.01, and 0.001, respectively

The variation in seasonal night-time CO_2 fluxes was more pronounced as compared with CH_4 and N_2O emissions (Fig. 2E). Greatest fluxes occurred during the growing season despite a few occasions when small fluctuations occurred. Lowest emissions occurred after the autumn harvest of aboveground biomass in 2017 and remained relatively stable throughout the winter season. In addition, night-time CO_2 fluxes were best correlated with soil temperature and WFPS (r = 0.69 and -0.53, both P < 0.001; Table 1). Overall, night-time CO_2 fluxes were negatively and positively correlated with CH_4 and N_2O emissions, respectively (r = -0.41 and 0.46, both P < 0.001).

3.3 Seasonal variability of ecosystem C fluxes

During the growing season, ecosystem C fluxes were measured seven times in total (Fig. 3). Marked fluctuations in ecosystem C fluxes occurred in July 2017, which may have resulted from substantial variations in soil WFPS (Fig. 3). Ecosystem C fluxes were strongly correlated with soil WFPS (r = 0.44 to 0.67, P < 0.05–0.001; Table 1), while only the seasonal pattern of R_{eco} had a positive relationship with soil temperature (r = 0.45, P < 0.05).

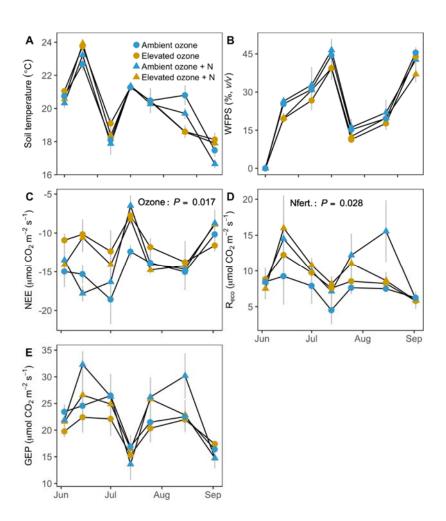


Figure 3 Soil temperature (A), water-filled pore space (WFPS) (B) at a depth of 10 cm, net ecosystem carbon exchange of CO_2 (NEE, C), ecosystem respiration (R_{eco} , D) and gross ecosystem productivity (GEP, E) during the growing season from a temperate semi-improved

grassland exposed to different levels of ozone and N. Values represent mean \pm SEM (n=4). Results from the linear mixed effects model at $P \le 0.05$ are shown.

Across the growing season, elevated ozone significantly decreased NEE (P = 0.017), whereas no effects of N fertilization (P = 0.823) or its interaction with elevated ozone (P = 0.161) were detected (Fig. 3C). Elevated ozone resulted in a 15% reduction of seasonal mean NEE. By contrast, $R_{\rm eco}$ was significantly increased by N fertilization (P = 0.028) but not by elevated ozone or their interaction (P = 0.315 and 0.155; Fig. 3D). Irrespective of ozone exposure, N fertilization significantly increased mean $R_{\rm eco}$ by 11%. Unlike NEE and $R_{\rm eco}$, none of elevated ozone, N fertilization, or their interaction affected GEP (Fig. 3E).

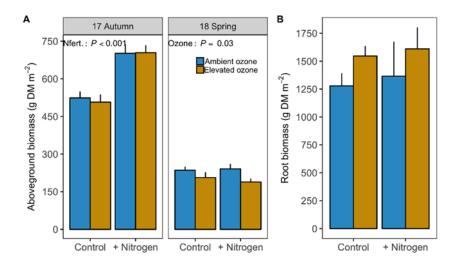


Figure 4 Aboveground biomass (A) and root biomass (B) in a temperate semi-improved grassland exposed to different levels of ozone and N. Values represent mean \pm SEM (n=4). Results from the linear mixed effects model at $P \le 0.05$ are shown.

3.4 Aboveground and root biomass

Harvested aboveground biomass differed significantly between cut events (P < 0.001; Fig. 4A). In the 2017 autumn harvest, aboveground biomass from the N fertilized treatment (mean: 703 ± 23 g DM m⁻²) was higher than from the control treatment (mean: 515 ± 20 g DM m⁻², P < 0.001), but there was no difference between N treatments at the 2018 spring harvest (P = 0.736). Aboveground biomass was affected by elevated ozone in the 2018 spring harvest but not in the 2017 autumn harvest. In the 2018 spring harvest, aboveground biomass decreased from 238±12 g DM m⁻² in the ambient ozone treatment to 197±13 g DM m⁻² in the elevated ozone treatment (P = 0.03). The average root biomass was 1450 g DM m⁻² and did not differ between either elevated ozone or N fertilization treatments (Fig. 4B).

3.5 Soil properties, net N-cycling rates and enzyme activities

Towards the end of the experiment, neither elevated ozone or N fertilization nor their interaction affected NH₄⁺ concentrations, DON or microbial biomass C (Table 2). Nitrate concentrations and microbial biomass N were higher in the elevated ozone plots than in the ambient ozone plots (P = 0.003 and 0.012, respectively), but there was no difference for both parameters between control and N-fertilized plots. A marginally significant interaction between ozone and N fertilizer was detected for DOC (P = 0.081). Microbial biomass C:N ratio was lower in the elevated ozone plots than in the ambient ozone plots (P = 0.018). Net N mineralization, which is often used as an index of plant available mineral N, was higher in the elevated ozone treatment than in the ambient ozone treatment (P = 0.056) but did not differ between the control

and N fertilized treatments. Further, net nitrification was higher in the elevated ozone than the ambient ozone treatment (P = 0.011).

Potential activities in two of studied soil extracellular enzymes were not affected by elevated ozone, N fertilization or their interaction (Table 2). Elevated ozone and N fertilization significantly increased cellobiohydrolase activity by 26% and 73%, respectively (P = 0.04 and 0.003). Elevated ozone also marginally stimulated leucine aminopeptidase activity by 15% as compared to under ambient ozone (P = 0.069).

4. Discussion

In the past decade, many conclusions about the impacts of elvated ozone have been drawn from croplands (Bhatia et al., 2011; Decock et al., 2012; Kou et al., 2015; Tang et al., 2015; Zheng et al., 2011), peatlands (Toet et al., 2017, 2011) and subalpine grasslands (Volk et al., 2011). The effects of elevated ozone and N fertilization alone or in combination on GHG emissions and ecosystem C fluxes have been rarely reported, especially in temperate grasslands (Sánchez-Martín et al., 2017; Wang et al., 2019). This is to our knowledge the first study to report how GHG emissions, coupled with ecosystem C fluxes, in response to elevated ozone and N fertilization alone or in combination from a temperate semi-improved grassland under fully open-air field conditions.

Table 2 Soil properties, net N-cycling rates and potential enzyme activities measured in May 2018 from a temperate semi-improved grassland exposed to different levels of ozone and N. Values represent mean \pm SEM (n = 4).

	Ambient ozone			Elevated ozone				
	Control	+ Nitrogen		Control	+ N	+ Nitrogen		
$NH_4{}^+ (mg\ N\ kg^{-1})$	4.48±0.54	6.25	±0.54	5.30±0.27	5.12	5.12±0.90		
NO_3^- (mg N kg ⁻¹)	$0.43 \pm 0.03^{\mathrm{B}*}$	0.46	±0.21 ^B	1.28 ± 0.26^{A}	1.01	1.01±0.50 A		
$DOC (mg C kg^{-1})$	$336\pm13^{\dagger}$	295±	:14	314±11	318	318±8		
DON (mg N kg ⁻¹)	59.7±1.8	54.6	±3.1	55.6±2.4	57.1	57.1±1.0		
Microbial biomass								
$C (mg C kg^{-1})$	2445±67	2371	±103	2548 ± 172	246	1±73		
Microbial biomass								
$N (mg N kg^{-1})$	227 ± 3^{B}	229±	:15 ^B	259±24 ^A	253	253±9 ^A		
Microbial biomass								
C:N	10.8±0.3 ^A	10.4	±0.3 ^A	9.9 ± 0.3^{B}	9.7±0.1 ^B			
Net N								
mineralization								
$(mg N kg^{-1} day^{-1})$	$-0.03\pm0.11^{\ddagger}$	-0.10	5±0.09 0.22±0.15		0.13 ± 0.15			
Net nitrification								
$(mg N kg^{-1} day^{-1})$	$0.04\pm0.04^{\mathrm{B}}$	0.06	±0.09 ^B	0.33 ± 0.06^{A} 0.19 ± 0.08^{A}		9±0.08 ^A		
ß-glucosidase								
$(nmol g^{-1} h^{-1})$	126±24	121±	±15 113±2		132±16			
Cellobiohydrolase								
$(nmol g^{-1} h^{-1})$ 16.7±2.8 ^{Bb§}		27.1	±5.7 ^{Ba}	19.7 ± 3.0^{Ab}	36.0 ± 3.5^{Aa}			
N-acetyl-glucosaminidase								
$(nmol g^{-1} h^{-1})$	83.6±6	.2	94.3±7.5		83.5±7.8	84.3±9.9		
Leucine aminopeptidase								
$(nmol g^{-1} h^{-1})$	42.7±3	.4 [‡]	37.8±1.7		45.1±2.3	47.1±3.8		

^{376 *} Values with different uppercase letters indicate significant differences between ambient and

378

380

373

374

375

³⁷⁷ elevated ozone treatments (linear mixed effects model with the Tukey HSD test at $P \le 0.05$)

[†] Marginally significant interaction between ozone and N fertilized treatments (linear mixed

³⁷⁹ effects model at $P \le 0.09$)

[‡] Marginally significant effect of elevated ozone (linear mixed effects model at $P \le 0.09$)

- § Values with different lowercase letters indicate significant differences between control and N-fertilized treatments (linear mixed effects model with the Tukey HSD test at $P \le 0.05$)
- 383 4.1 Response of CH₄ fluxes to elevated ozone and N fertilization

381

382

384

385

386

387

388

389

390

391

392

393

394

395

396

397

398

399

400

The fluxes of CH₄ from our studied grasslands were generally negative throughout the experimental period, which agrees with the argument that temperate grassland soils generally act as an atmospheric CH₄ sink (Hörtnagl et al., 2018; Smith et al., 2000). We found that elevated ozone tended to decrease the magnitude of CH₄ uptake (a slightly significant effect) compared to the ambient plots during the growing season but not the winter period, suggesting a negative impact on the CH₄ uptake capacity. This is unlikely to be related to a direct effect of ozone on soil methanogens and methanotrophs since ozone does not penetrate far into the soil (Blum and Tingey, 1977; Toet et al., 2009). The reduced CH₄ uptake during the growing season could be possibly due to altered methanotroph activity at elevated ozone. Across the one-year period CH₄ emissions were not affected by elevated ozone, which agrees with findings of several studies under OTC conditions (Kanerva et al., 2007; Sánchez-Martín et al., 2017). For example, studies in an annual Mediterranean grassland reveal that ozone exposure for a shortterm period (49 days) did not affect CH₄ emissions (Sánchez-Martín et al., 2017). Similarly, the three-year field study of Kanerva et al. (2007) reported no overall effect of ozone exposure on CH₄ fluxes from ground-planted meadow mesocosms. By contrast, studies in rice fields under either OTC or O₃-FACE conditions show that elevated ozone can decrease CH₄ emissions during the rice growing season (Bhatia et al., 2011; Tang et al., 2015; Zhang et al., 2016). Either a negative or transient effect of elevated ozone on CH_4 emissions from peatlands is also reported (Mörsky et al., 2008; Toet et al., 2011).

401

402

403

404

405

406

407

408

409

410

411

412

413

414

415

416

417

418

419

420

Although it has been suggested that N addition decreases CH₄ uptake in upland ecosystems (Liu and Greaver, 2009), our results show that neither N fertilization nor its interaction with elevated ozone affected CH₄ uptake. This is supported by the findings of a meta-analysis where they found that soil CH₄ uptake in natural or with short-term N fertilization sites showed a nonsignificant change when N is added (Aronson and Helliker, 2010). Thus, the lack of CH₄ uptake in response to N addition could be attributed to a one-time application of N fertilizer into this semi-improved grassland. On the other hand, our result contradicts with their finding that N addition effects on soil CH₄ uptake may be switched from stimulation to inhibition when N addition exceeds a threshold value of 100 kg N ha⁻¹ yr⁻¹ (Aronson and Helliker, 2010). This implies that a shift from stimulation to inhibition of soil CH₄ uptake might be related to not only the amount of N added but also the inherent soil N status of the studied system. We are aware of only one previous field study where they examined the interaction effect between elevated ozone and N fertilization on GHG emissions from a simplified annual grassland (Sánchez-Martín et al., 2017). Consistent with our findings, the response of CH₄ emission to ozone exposure from their experiment site was not N fertilization dependent. While we found either elevated ozone or N fertilization may have a negligible effect on soil CH4 uptake in this temperate grassland, future multiple-year studies are needed to reveal the inter-annual variability and its underlying mechanisms.

4.2 Response of N_2O fluxes to elevated ozone and N fertilization

421

422

423

424

425

426

427

428

429

430

431

432

433

434

435

436

437

438

439

440

This semi-improved grassland acted as a source of N₂O, although negative N₂O fluxes were recorded occasionally, especially during the winter season (Fig. 2D). Our results showed that neither elevated ozone or N fertilization nor their interaction had an appreciable effect on N₂O emission over the experimental year. The lack of responsiveness of N₂O emissions to N addition suggests that a one-time application of 100 kg N ha⁻¹ (the first fertilizer N application to this grassland for >25 years) at the beginning of the experiment would not have been sufficient to produce significant N₂O emissions in this semi-improved grassland. Annual N₂O emissions from this study were much higher (0.18–0.34 kg N ha⁻¹ yr⁻¹) than that (ca. 0.06 kg N ha⁻¹ yr⁻¹) from the field study of Marsden et al. (2018) in the same grassland. This is likely due to the differences in microclimate between the two study sites. In their study, the large amount of N applied in the form of sheep urine (ca. 2000 kg N ha⁻¹ yr⁻¹) had no marked effect on N₂O fluxes (Marsden et al., 2018), which partly supports our finding. Furthermore, N fertilization significantly increased aboveground biomass during the growing season, but had minor resultant effects on plant yield and soil characteristics at the end of the experimental year (Fig. 4A; Table 2). Therefore, these findings collectively point to the consumption of applied N fertilizer tightly coupled to plant uptake and microbial immobilization in this semi-improved grassland.

To date, the divergent responses of N_2O fluxes to elevated ozone have been reported. For example, some studies showed a decrease in N_2O emissions under elevated ozone (Bhatia et al.,

2011; Kou et al., 2015), other reported stimulated emissions (Sánchez-Martín et al., 2017) or no marked effects (Decock et al., 2012; Kanerva et al., 2007) compared to the ambient ozone concentrations. As soil nitrification and denitrification processes are major pathways of N₂O emissions, changes in the substrate availability due to elevated ozone are expected to alter N_2O emissions. We found that elevated ozone resulted in a significant decline of aboveground biomass at the 2018 spring harvest, which may, in turn, contribute to accelerating soil N-cycling (i.e., a build-up of soil nitrate, higher rates of net N mineralization and nitrification; Table 2). The unchanged N₂O emissions but stimulated N-cycling processes could be explained by the low intensity of soil N2O emission and/or other pathways being altered which might be primarily responsible for N loss from this system. Indeed, in a soybean agroecosystem under fully open-air conditions, Decock et al. (2012) also demonstrated that elevated ozone stimulated soil N-cycling but had no effect on soil N₂O emissions. In contrast, in an annual Mediterranean grassland, ozone exposure reduced the fertilization effect of N on plant growth and thereby resulted in a significant reduction of soil N₂O emissions (Sánchez-Martín et al., 2017). Clearly, this was not the case for our study because of the absence of an interaction between elevated ozone and N fertilization on plant yield, especially during the growing season (Fig. 4). Nevertheless, it should be noted that the single N application in our study might counterbalance the negative impact of ozone exposure on plant growth during the growing season.

441

442

443

444

445

446

447

448

449

450

451

452

453

454

455

456

457

458

459

4.3 Effects of elevated ozone and N fertilization on ecosystem C fluxes

A growing number of studies have documented the detrimental effects of elevated ozone on grassland plant species, ranging from visible injury of leaves to substantially decreased productivity (Hayes et al., 2007; Volk et al., 2006). In our study, elevated ozone resulted in a small reduction (3.2% for the control plots) but a significant reduction of 17.1% in aboveground biomass during the growing and non-growing seasons, respectively (Fig. 4A). During the growing season, elevated ozone caused a significant reduction of NEE (14.9%), which is consistent with other studies in an annual temperate grassland (Calvete-Sogo et al., 2014; Sánchez-Martín et al., 2017) and peatland microcosms (Haapala et al., 2011). Because of the unresponsiveness of total aboveground biomass, the reduced NEE by elevated ozone at the canopy level is likely due to the changed photosynthetic and/or respiration rates (Andersen, 2003). We noted that these semi-improved grassland mesocosms had a higher growth rate, in terms of stimulated fluxes of trace gas and night-time CO₂ (Fig. 2C-E), under the O₃-FACE field condition than in situ on the upland site (Marsden et al., 2018). Thus, our results support the view that plant communities with fast-growing rates (i.e., annual and newly established systems) would be more responsive to elevated ozone than these perennial and well established systems that are characterized by slower growth rates (Grantz et al., 2006; Grime, 2000). In contrast, we are aware of a transient effect of elevated ozone on NEE in peatlands (Haapala et al., 2011; Niemi et al., 2002; Rinnan et al., 2003). On a subalpine grassland, Volk et al. (2011) found no effect of elevated ozone on NEE measured during the third growing season. Taken together, our results and those of previous findings suggest that further long-term investigations are warranted to evaluate the inter-annual response of NEE to elevated ozone in grasslands.

460

461

462

463

464

465

466

467

468

469

470

471

472

473

474

475

476

477

478

479

480

Ecosystem respiration as the sum of the respiration from plants and heterotrophs was not affected by elevated ozone during the growing season (Figs. 2E and 3D). This is in agreement with findings from other studies in subalpine and annual grasslands (Calvete-Sogo et al., 2014; Volk et al., 2011). Similarly, studies from peatlands suggested no effect of elevated ozone on ecosystem respiration, especially during the first experimental year (Haapala et al., 2011; Kanerva et al., 2007; Toet et al., 2011). In our study, the unchanged ecosystem respiration might be due to the response of heterotrophic respiration to elevated ozone which was unresponsive or masked by the larger proportion of plant respiration (Volk et al., 2011). As expected, stimulation of plant productivity provides more substrate for plant and soil respiration, leading to increases in ecosystem respiration following N fertilization (Figs. 2E and 3D). The positive response of ecosystem respiration to N addition is consistent with that in a subalpine grassland (Volk et al., 2011). In contrast, the lack of N effects on ecosystem respiration in an annual grassland was attributed to the fact that the low N doses were only enough to meet plant N demand but had no effect on the yield or gas exchange rates (Calvete-Sogo et al., 2014). Since the assimilation processes was primarily N-limited, we expected to see the positive response of GEP to N fertilization in this semi-improved grassland. However, the differential responses of NEE and ecosystem respiration to elevated ozone and N fertilization resulted in non-significant changes in GEP, suggesting N fertilization counterbalanced the effect of elevated ozone on GEP in our studied grassland.

481

482

483

484

485

486

487

488

489

490

491

492

493

494

495

496

497

498

499

500

4.4 Short-term effects of elevated ozone and N fertilization on belowground processes

While the intact grassland mesocosms were exposed to ozone only for one growing season, our results indeed showed that soil N dynamics were altered at the end of the experimental year. Increased soil mineral N availability under elevated ozone may be due to the significant reduction of aboveground biomass during the non-growing season, which may have contributed to reduced plant N uptake. On the other hand, we found that the activities of cellobiohydrolase and leucine aminopeptidase were higher in the elevated ozone plots than in the ambient plots, suggesting that elevated ozone stimulates microbes in the soil to produce enzymes that degrade cellulose and peptide-containing moieties. Parallel to the marginal increase of net N mineralization, our results suggest an increased soil N mineralization by elevated ozone in this grassland soil. Under elevated ozone, the unchanged soil NH₄⁺ is likely related to increased microbial N immobilization, whereas stimulated soil net nitrification may have resulted in an accumulation of soil NO₃⁻. Consistent with our findings, other studies in the O₃-FACE systems have reported the increased soil mineral N availability under elevated ozone in both soybean and wheat fields (He et al., 2014; Wu et al., 2016). Note that our soil sampling was carried out in early spring of 2018, which probably contributes to masking the possible N fertilization effect or its interaction with ozone, especially during the growing season. As soil mineral N availability is mainly regulated by plant uptake and microbial-mediated N transformation processes (Schimel and Bennett, 2004), our results collectively point to a positive feedback of soil N transformation in this semi-improved grassland to short-term of ozone exposure.

5. Conclusions

501

502

503

504

505

506

507

508

509

510

511

512

513

514

515

516

517

518

519

520

To our knowledge, this study is the first to report the effects of elevated ozone on GHG and ecosystem C fluxes in temperate grasslands under fully open-air field conditions. Our results demonstrate that elevated ozone may reduce atmospheric CH₄ uptake and net C uptake during the growing season in this semi-improved grassland. Given that the reported responses of CH₄ emission and NEE to elevated ozone are still inconsistent and controversial yet, we speculate that the depressed CH₄ uptake under elevated ozone could be transient, which warrants further investigation. The lack of any interaction between elevated ozone and N fertilization is likely due to the one-time N fertilization at the beginning of experiment, which probably was not synchronized with the occurrence of ozone injury of the plants. Further, our results suggest that short-term of ozone treatment may have contributed to accelerated soil N cycling in this grassland. Further investigations are warranted to examine the long-term impact of elevated ozone on ecosystem production and function.

Acknowledgements

This work was supported by the European Commission under Horizon 2020 for a Marie Skłodowska-Curie Actions COFUND Fellowship (663830-BU-048); and the Welsh Government and Higher Education Funding Council for Wales through the Sêr Cymru National Research Network for Low Carbon, Energy and Environment. We thank Aled Williams for technical support in running the O₃-FACE facility.

Reference

540 Ainsworth, E., Yendrek, C.R., Sitch, S., Collins, W.J., Emberson, L.D., 2012. The effects of 541 tropospheric ozone on net primary productivity and implications for climate change. Annu. 542 Rev. Plant Biol. 63, 637–61. doi:10.1146/annurev-arplant-042110-103829 Andersen, C.P., 2003. Source-sink balance and carbon allocation below ground in plants 543 544 exposed to ozone. New Phytol. 157, 213–228. doi:10.1046/j.1469-8137.2003.00674.x Aronson, A.E.L., Helliker, B.R., 2010. Methane flux in non-wetland soils in response to 545 546 nitrogen addition: a meta-analysis Linked references are available on JSTOR for this 547 article: Methane flux in non-wetland soils in response to nitrogen addition: a metaanalysis. Ecology 91, 3242–3251. doi:10.2307/20788157 548 549 Bates, D., Mächler, M., Bolker, B., Walker, S., 2014. Fitting linear mixed-effects models using lme4. arXiv Prepr. arXiv1406.5823. 550 Bhatia, A., Ghosh, A., Kumar, V., Tomer, R., Singh, S.D., Pathak, H., 2011. Effect of elevated 551 tropospheric ozone on methane and nitrous oxide emission from rice soil in north India. 552 Agric. Ecosyst. Environ. 144, 21–28. doi:10.1016/j.agee.2011.07.003 553 554 Blum, U., Tingey, D.T., 1977. A study of the potential ways in which ozone could reduce root growth and nodulation of soybean. Atmos. Environ. 11, 737-739. doi:10.1016/0004-555 556 6981(77)90182-2 Brookes, P.C., Landman, A., Pruden, G., Jenkinson, D.S., 1985. Chloroform fumigation and 557 558 the release of soil nitrogen: A rapid direct extraction method to measure microbial 559 biomass nitrogen in soil. Soil Biol. Biochem. 17, 837-842. doi:10.1016/0038-

- Calvete-Sogo, H., Elvira, S., Sanz, J., González-Fernández, I., García-Gómez, H., Sánchez-
- Martín, L., Alonso, R., Bermejo-Bermejo, V., 2014. Current ozone levels threaten gross
- primary production and yield of Mediterranean annual pastures and nitrogen modulates
- the response. Atmos. Environ. 95, 197–206. doi:10.1016/j.atmosenv.2014.05.073
- 565 Crawley, M.J., 2012. The R book. John Wiley & Sons.
- Defra, 2016. Agriculture in the United Kingdom 2015. Department for Environment, Food and
- Rural Affairs, London, UK.
- Decock, C., Chung, H., Venterea, R., Gray, S.B., Leakey, A.D.B., Six, J., 2012. Elevated CO₂
- and O₃ modify N turnover rates, but not N₂O emissions in a soybean agroecosystem. Soil
- 570 Biol. Biochem. 51, 104–114. doi:10.1016/j.soilbio.2012.04.015
- 571 DeForest, J.L., 2009. The influence of time, storage temperature, and substrate age on potential
- soil enzyme activity in acidic forest soils using MUB-linked substrates and 1-DOPA. Soil
- 573 Biol. Biochem. 41, 1180–1186. doi:10.1016/j.soilbio.2009.02.029
- 574 FAO, 1981. FAO-UNESCO Soil Map of the World 1:5000000. Volume V: Europe, UNESCO,
- 575 Paris, France.
- Fowler, D., Amann, M., Anderson, F., Ashmore, M., Cox, P., Depledge, M., Derwent, D.,
- Grennfelt, P., Hewitt, N., Hov, O., Jenkin, M., Kelly, F., Liss, P., Pilling, M., Pyle, J., J.
- 578 S., Stevenson, D., 2008. Ground level Ozone in the 21st Century: Future Trends, Impacts
- and Policy Implications. R. Soc. Sci. Policy Rep. 15(08).

- Fuhrer, J., Val Martin, M., Mills, G., Heald, C.L., Harmens, H., Hayes, F., Sharps, K., Bender,
- J., Ashmore, M.R., 2016. Current and future ozone risks to global terrestrial biodiversity
- and ecosystem processes. Ecol. Evol. 6, 8785–8799. doi:10.1002/ece3.2568
- Grantz, D.A., Gunn, S., Vu, H.B., 2006. O₃ impacts on plant development: A meta-analysis of
- root/shoot allocation and growth. Plant, Cell Environ. 29, 1193–1209.
- 585 doi:10.1111/j.1365-3040.2006.01521.x
- 586 Grime, J.P., 2000. The Response of Two Contrasting Limestone Grasslands to Simulated
- 587 Climate Change. Science (80-.). 289, 762–765. doi:10.1126/science.289.5480.762
- Haapala, J.K., Mörsky, S.K., Rinnan, R., Saarnio, S., Martikanen, P.J., Holopainen, T., Silvola,
- J., 2011. Long-term effects of ozone on CO₂ exchange in peatland microcosms. Atmos.
- 590 Environ. 45, 4002–4007. doi:10.1016/j.atmosenv.2011.04.057
- Hart, S.C., Stark, J.M., Davidson, E.A., Firestone, M.K., 1994. Nitrogen mineralization,
- immobilization, and nitrification. Methods Soil Anal. Part 2. Microbiol. Biochem. Prop.
- 593 985–1018. doi:10.2136/sssabookser5.2.c42
- Hayes, F., Jones, M.L.M., Mills, G., Ashmore, M., 2007. Meta-analysis of the relative
- sensitivity of semi-natural vegetation species to ozone. Environ. Pollut. 146, 754–762.
- 596 doi:10.1016/j.envpol.2006.06.011
- He, Z., Xiong, J., Kent, A.D., Deng, Y., Xue, K., Wang, G., Wu, L., Van Nostrand, J.D., Zhou,
- J., 2014. Distinct responses of soil microbial communities to elevated CO₂ and O₃ in a
- soybean agro-ecosystem. ISME J. 8, 714–26. doi:10.1038/ismej.2013.177

- Hörtnagl, L., Barthel, M., Buchmann, N., Eugster, W., Butterbach-Bahl, K., Díaz-Pinés, E.,
- Zeeman, M., Klumpp, K., Kiese, R., Bahn, M., Hammerle, A., Lu, H., Ladreiter-Knauss,
- T., Burri, S., Merbold, L., 2018. Greenhouse gas fluxes over managed grasslands in
- 603 Central Europe. Glob. Chang. Biol. 24, 1843–1872. doi:10.1111/gcb.14079
- 604 IPCC, 2007. Summary for Policymakers, in: Solomon, S., Qin, D., Manning, M., Chen, Z.,
- Marquis, M., Averyt, K.B., M.Tignor, Miller, H.L. (Eds.), Climate Change 2007: The
- Physical Science Basis. Contribution of Working Group I to the Fourth Assessment
- Report of the Intergovernmental Panel on Climate Change. Cambridge University Press,
- 608 Cambridge, United Kingdom and New York, NY, USA.
- 609 Kanerva, T., Regina, K., Rämö, K., Ojanperä, K., Manninen, S., 2007. Fluxes of N2O, CH4 and
- 610 CO₂ in a meadow ecosystem exposed to elevated ozone and carbon dioxide for three years.
- Environ. Pollut. 145, 818–828. doi:10.1016/j.envpol.2006.03.055
- Kou, T.J., Cheng, X.H., Zhu, J.G., Xie, Z.B., 2015. The influence of ozone pollution on CO₂,
- 613 CH₄, and N₂O emissions from a Chinese subtropical rice-wheat rotation system under
- 614 free-air O₃ exposure. Agric. Ecosyst. Environ. 204, 72–81.
- doi:10.1016/j.agee.2015.02.013
- 616 Linn, D.M., Doran, J.W., 1984. Aerobic and Anaerobic Microbial Populations in No-till and
- 617 Plowed Soils1. Soil Sci. Soc. Am. J. 48, 794–799.
- doi:10.2136/sssaj1984.03615995004800040019x
- 619 Liu, L., Greaver, T.L., 2009. A review of nitrogen enrichment effects on three biogenic GHGs:

- the CO₂ sink may be largely offset by stimulated N₂O and CH₄ emission. Ecol. Lett. 12,
- 621 1103–1117. doi:10.1111/j.1461-0248.2009.01351.x
- Luyssaert, S., Inglima, I., Jung, M., Richardson, A.D., Reichstein, M., Papale, D., Piao, S.L.,
- Schulze, E.D., Wingate, L., Matteucci, G., Aragao, L., Aubinet, M., Beer, C., Bernhofer,
- 624 C., Black, K.G., Bonal, D., Bonnefond, J.M., Chambers, J., Ciais, P., Cook, B., Davis,
- K.J., Dolman, A.J., Gielen, B., Goulden, M., Grace, J., Granier, A., Grelle, A., Griffis, T.,
- 626 Gr??nwald, T., Guidolotti, G., Hanson, P.J., Harding, R., Hollinger, D.Y., Hutyra, L.R.,
- Kolari, P., Kruijt, B., Kutsch, W., Lagergren, F., Laurila, T., Law, B.E., Le Maire, G.,
- Lindroth, A., Loustau, D., Malhi, Y., Mateus, J., Migliavacca, M., Misson, L.,
- Montagnani, L., Moncrieff, J., Moors, E., Munger, J.W., Nikinmaa, E., Ollinger, S. V.,
- Pita, G., Rebmann, C., Roupsard, O., Saigusa, N., Sanz, M.J., Seufert, G., Sierra, C., Smith,
- M.L., Tang, J., Valentini, R., Vesala, T., Janssens, I.A., 2007. CO₂ balance of boreal,
- temperate, and tropical forests derived from a global database. Glob. Chang. Biol. 13,
- 633 2509–2537. doi:10.1111/j.1365-2486.2007.01439.x
- Marsden, K.A., Holmberg, J.A., Jones, D.L., Chadwick, D.R., 2018. Sheep urine patch N₂O
- emissions are lower from extensively-managed than intensively-managed grasslands.
- Agric. Ecosyst. Environ. 265, 264–274. doi:10.1016/j.agee.2018.06.025
- Meehl, G.A., Stocker, T.F., Collins, W.D., Friedlingstein, P., Gaye, T., Gregory, J.M., Kitoh,
- A., Knutti, R., Murphy, J.M., Noda, A., Raper, S.C.B., Watterson, I.G., Weaver, A.J.,
- Zhao, Z.C., 2007. Global climate projections, in: Solomon, S., Qin, D., Manning, M.,
- Chen, Z., Marquis, M., Averyt, K.B., Tignor, M., Miller, H.L. (Eds.), IPCC, 2007:

641 Climate Change 2007: The Physical Science Basis. Contribution of Working Group I to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change. 642 643 Cambridge, UK, Cambridge University Press, pp. 747–846. Mills, G., Harmens, H., Wagg, S., Sharps, K., Hayes, F., Fowler, D., Sutton, M., Davies, B., 644 645 2016. Ozone impacts on vegetation in a nitrogen enriched and changing climate. Environ. 646 Pollut. 208, 898–908. doi:10.1016/j.envpol.2015.09.038 Mills, G., Hayes, F., Simpson, D., Emberson, L., Norris, D., Harmens, H., Büker, P., 2011. 647 648 Evidence of widespread effects of ozone on crops and (semi-)natural vegetation in Europe 649 (1990-2006) in relation to AOT40- and flux-based risk maps. Glob. Chang. Biol. 17, 592-613. doi:10.1111/j.1365-2486.2010.02217.x 650 Miranda, K.M., Espey, M.G., Wink, D.A., 2001. A rapid, simple spectrophotometric method 651 652 for simultaneous detection of nitrate and nitrite. Nitric Oxide - Biol. Chem. 5, 62-71. 653 doi:10.1006/niox.2000.0319 Mörsky, S.K., Haapala, J.K., Rinnan, R., Tiiva, P., Saarnio, S., Silvola, J., Holopainen, T., 654 655 Martikainen, P.J., 2008. Long-term ozone effects on vegetation, microbial community and 656 methane dynamics of boreal peatland microcosms in open-field conditions. Glob. Chang. 657 Biol. 14, 1891–1903. doi:10.1111/j.1365-2486.2008.01615.x 658 Mulvaney, R.L., 1996. Nitrogen—Inorganic Forms, in: Sparks, D.L. (Ed.), Methods of Soil 659 Analysis Part 3—Chemical Methods. Soil Science Society of America, American Society 660 of Agronomy, Madison, WI, pp. 1123–1184. doi:10.2136/sssabookser5.3.c38

- Niemi, R., Martikainen, P.J., Silvola, J., Holopainen, T., 2002. Ozone effects on Sphagnum
- mosses, carbon dioxide exchange and methane emission in boreal peatland microcosms.
- 663 Sci. Total Environ. 289, 1–12. doi:10.1016/S0048-9697(01)01012-9
- Niu, S., Wu, M., Han, Y., Xia, J., Zhang, Z., Yang, H., Wan, S., 2010. Nitrogen effects on net
- ecosystem carbon exchange in a temperate steppe. Glob. Chang. Biol. 16, 144–155.
- doi:10.1111/j.1365-2486.2009.01894.x
- R Development Core Team, 2015. R: A language and environment for statistical computing.
- Vienna, Austria.
- Rinnan, R., Impiö, M., Silvola, J., Holopainen, T., Martikainen, P.J., 2003. Carbon dioxide and
- 670 methane fluxes in boreal peatland microcosms with different vegetation cover Effects of
- ozone or ultraviolet-B exposure. Oecologia 137, 475–483. doi:10.1007/s00442-003-
- 672 1366-5
- Rodwell, J.S., 1992. British plant communities. Volume 3. Grasslands and montane
- 674 communities, Cambridge University Press, Cambridge, UK. Cambridge University Press,
- 675 Cambridge, UK.
- Saiya-Cork, K., Sinsabaugh, R., Zak, D., 2002. The effects of long term nitrogen deposition
- on extracellular enzyme activity in an Acer saccharum forest soil. Soil Biol. Biochem. 34,
- 678 1309–1315. doi:10.1016/S0038-0717(02)00074-3
- 679 Sánchez-Martín, L., Bermejo-Bermejo, V., García-Torres, L., Alonso, R., de la Cruz, A.,
- Calvete-Sogo, H., Vallejo, A., 2017. Nitrogen soil emissions and belowground plant

- processes in Mediterranean annual pastures are altered by ozone exposure and N-inputs.
- 682 Atmos. Environ. 165, 12–22. doi:10.1016/j.atmosenv.2017.06.030
- Schimel, J.P., Bennett, J., 2004. Nitrogen mineralization: Challenges of a changing paradigm.
- 684 Ecology 85, 591–602. doi:10.1890/03-8002
- Sicard, P., Anav, A., De Marco, A., Paoletti, E., 2017. Projected global ground-level ozone
- impacts on vegetation under different emission and climate scenarios. Atmos. Chem. Phys.
- 687 17, 12177–12196. doi:10.5194/acp-17-12177-2017
- 688 Sitch, S., Cox, P.M., Collins, W.J., Huntingford, C., 2007. Indirect radiative forcing of climate
- change through ozone effects on the land-carbon sink. Nature 448, 791–794.
- 690 doi:10.1038/nature06059
- 691 Smith, K.A., Dobbie, K.E., Ball, B.C., Bakken, L.R., Sitaula, B.K., Hansen, S., Brumme, R.,
- Borken, W., Christensen, S., Priemé, A., Fowler, D., Macdonald, J.A., Skiba, U.,
- Klemedtsson, L., Kasimir-Klemedtsson, A., Degórska, A., Orlanski, P., 2000. Oxidation
- of atmospheric methane in Northern European soils, comparison with other ecosystems,
- and uncertainties in the global terrestrial sink. Glob. Chang. Biol. 6, 791-803.
- 696 doi:10.1046/j.1365-2486.2000.00356.x
- Tang, H., Liu, G., Zhu, J., Kobayashi, K., 2015. Effects of elevated ozone concentration on CH₄
- and N₂O emission from paddy soil under fully open-air field conditions. Glob. Chang.
- 699 Biol. 21, 1727–1736. doi:10.1111/gcb.12810
- 700 Toet, S., Ineson, P., Peacock, S., Ashmore, M., 2011. Elevated ozone reduces methane

- 701 emissions from peatland mesocosms. Glob. Chang. Biol. 17, 288–296.
- 702 doi:10.1111/j.1365-2486.2010.02267.x
- Toet, S., Oliver, V., Ineson, P., McLoughlin, S., Helgason, T., Peacock, S., Stott, A.W., Barnes,
- J., Ashmore, M., 2017. How does elevated ozone reduce methane emissions from
- 705 peatlands? Sci. Total Environ. 579, 60–71. doi:10.1016/j.scitotenv.2016.10.188
- Toet, S., Subke, J.A., D'Haese, D., Ashmore, M.R., Emberson, L.D., Crossman, Z., Evershed,
- R.P., Barnes, J.D., Ineson, P., 2009. A new stable isotope approach identifies the fate of
- 708 ozone in plant-soil systems. New Phytol. 182, 85–90. doi:10.1111/j.1469-
- 709 8137.2009.02780.x
- Vance, E.D., Brookes, P.C., Jenkinson, D.S., 1987. An extraction method for measuring soil
- 711 microbial biomass C. Soil Biol. Biochem. 19, 703-707. doi:10.1016/0038-
- 712 0717(87)90052-6
- Volk, M., Bungener, P., Contat, F., Montani, M., Fuhrer, J., 2006. Grassland yield declined by
- a quarter in 5 years of free-air ozone fumigation. Glob. Chang. Biol. 12, 74–83.
- 715 doi:10.1111/j.1365-2486.2005.01083.x
- Volk, M., Obrist, D., Novak, K., Giger, R., Bassin, S., Fuhrer, J., 2011. Subalpine grassland
- 717 carbon dioxide fluxes indicate substantial carbon losses under increased nitrogen
- deposition, but not at elevated ozone concentration. Glob. Chang. Biol. 17, 366–376.
- 719 doi:10.1111/j.1365-2486.2010.02228.x
- Volk, M., Wolff, V., Bassin, S., Ammann, C., Fuhrer, J., 2014. High tolerance of subalpine

- grassland to long-term ozone exposure is independent of N input and climatic drivers.
- 722 Environ. Pollut. 189, 161–168. doi:10.1016/j.envpol.2014.02.032
- Wang, J., Hayes, F., Turner, R., Chadwick, D.R., Mills, G., Jones, D.L., 2019. Effects of four
- years of elevated ozone on microbial biomass and extracellular enzyme activities in a
- 725 semi-natural grassland. Sci. Total Environ. 660, 260–268.
- 726 doi:10.1016/j.scitotenv.2019.01.040
- Williamson, J.L., Mills, G., Hayes, F., Jones, T., Freeman, C., 2016. How do increasing
- background concentrations of tropospheric ozone affect peatland plant growth and carbon
- 729 gas exchange? Atmos. Environ. 127, 133–138. doi:10.1016/j.atmosenv.2015.12.004
- Wu, H., Li, Q., Lu, C., Zhang, L., Zhu, J., Dijkstra, F.A., Yu, Q., 2016. Elevated ozone effects
- on soil nitrogen cycling differ among wheat cultivars. Appl. Soil Ecol. 108, 187–194.
- 732 doi:10.1016/j.apsoil.2016.08.015
- Wu, J., Joergensen, R., Pommerening, B., Chaussod, R., Brookes, P.C., 1990. Measurement of
- soil microbial biomass C by fumigation-extraction—an automated procedure. Soil Biol.
- 735 Biochem. 22, 1167–1169. doi:10.1016/0038-0717(90)90046-3
- Yendrek, C.R., Leisner, C.P., Ainsworth, E.A., 2013. Chronic ozone exacerbates the reduction
- in photosynthesis and acceleration of senescence caused by limited N availability in
- 738 Nicotiana sylvestris. Glob. Chang. Biol. 19, 3155–3166. doi:10.1111/gcb.12237
- Zhang, J., Tang, H., Zhu, J., Lin, X., Feng, Y., 2016. Divergent responses of methanogenic
- archaeal communities in two rice cultivars to elevated ground-level O₃. Environ. Pollut.

213, 127–134. doi:10.1016/j.envpol.2016.01.062
Zheng, F., Wang, X., Lu, F., Hou, P., Zhang, W., Duan, X., Zhou, X., Ai, Y., Zheng, H., Ouyang,
Z., Feng, Z., 2011. Effects of elevated ozone concentration on methane emission from a
rice paddy in Yangtze River Delta, China. Glob. Chang. Biol. 17, 898–910.
doi:10.1111/j.1365-2486.2010.02258.x