

1 **Organic phosphorus in the terrestrial environment: A perspective on the state of the art and future**
2 **priorities**

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62 **Abstract**

63 **Background:** The dynamics of phosphorus (P) in the environment is important for regulating nutrient cycles in
64 natural and managed ecosystems and an integral part in assessing biological resilience against environmental
65 change. Organic P (P_o) compounds play key roles in biological and ecosystems function in the terrestrial
66 environment, being critical to cell function, growth and reproduction.

67 **Scope:** We asked a group of experts to consider the global issues associated with P_o in the terrestrial environment,
68 methodological strengths and weaknesses, benefits to be gained from understanding the P_o cycle, and to set
69 priorities for P_o research.

70 **Conclusions:** We identified seven key opportunities for P_o research including: the need for integrated, quality
71 controlled and functionally based methodologies; assessment of stoichiometry with other elements in organic
72 matter; understanding the dynamics of P_o in natural and managed systems; the role of microorganisms in
73 controlling P_o cycles; the implications of nanoparticles in the environment and the need for better modelling and
74 communication of the research. Each priority is discussed and a statement of intent for the P_o research community
75 is made that highlights there are key contributions to be made toward understanding biogeochemical cycles,
76 dynamics and function of natural ecosystems and the management of agricultural systems.

77 **Keywords**

78 , Ecosystems services, Method development, Microbiome, Modelling, Organic Phosphorus, Stoichiometry.

79 **Abbreviations**

80 $\delta^{18}\text{OP}$ – oxygen-18 isotope ratio

81 16S rRNA = 16S ribosomal Ribonucleic acid

82 Al = Aluminium

83 ATP = Adenosine triphosphate

- 84 C = Carbon
- 85 DNA = Deoxyribonucleic acid
- 86 Fe = Iron
- 87 N = Nitrogen
- 88 P = Phosphorus
- 89 Pho = Pho regulon transcription factors
- 90 P_i = Inorganic orthophosphate
- 91 P_o = Organic phosphate compounds
- 92 S = Sulphur

93 **The Importance of Phosphorus and Organic Phosphorus**

94 The dynamics of phosphorus (P) in the terrestrial environment is critical for regulating nutrient cycling in both
95 natural and managed ecosystems. Phosphorus compounds fundamentally contribute to life on earth: being
96 essential to cellular organization as phospholipids, as chemical energy for metabolism in the form of ATP, genetic
97 instructions for growth, development and cellular function as nucleic acids, and as intracellular signalling
98 molecules (Butusov and Jernelöv 2013). Plant growth is limited by soil P availability, so turnover of organic
99 phosphorus (P_o) represents a source of P for ecosystem function and, critically, P supply affects crop production
100 (Runge-Metzger 1995). Phosphorus deficiency constrains the accumulation and turnover of plant biomass and
101 dictates community assemblages and biodiversity in a range of natural ecosystems (Attiwill and Adams 1993;
102 McGill and Cole 1981).

103 Chemically, P is a complex nutrient that exists in many inorganic (P_i) and organic (P_o) forms in the environment.
104 Through the utilization of orthophosphate, plants and other organisms drive the conversion of P_i to P_o . Death,
105 decay and herbivory facilitate the return of both P_o and P_i in plant materials to soil. Inputs of P to soil through
106 these processes may contribute P_o directly to soil or indirectly, following decomposition, accumulation, and
107 stabilization of P_o by microorganisms (Harrison 1982; Lang et al. 2016; Magid et al. 1996; McGill and Cole 1981;
108 Stewart and Tiessen 1987; Tate and Salcedo 1988). In its simplest definition, P_o is any compound that contains
109 an organic moiety in addition to P, while a wider definition would include phosphate which is associated with

110 organic matter. Such discrete P_o compounds are categorized into similarly structured forms and these forms and
111 their relative lability in soil is shown in Figure 1, taken from Darch et al. (2014). The P_o compounds, which are
112 considered to be biologically relevant include monoesters, inositol phosphates, diesters and phosphonates. The
113 relative lability and accumulation of these different groups varies in the environment, but overall the labile
114 monoesters and diesters tend to be less prevalent and the inositol phosphates tend to be less labile and accumulate
115 in the environment (Darch et al. 2014). In general, soil organic P forms have a smaller affinity to the soil solid
116 phase than inorganic P forms and a large proportion of the P forms found in leachate are found to be in organic
117 forms (Chardon & Oenema, 1995; Chardon et al. 1997; Espinosa et al. 1999) and can therefore have large impacts
118 on ecosystem function (Sharma et al. 2017; Toor et al. 2003). All P_o compounds have a range of chemical bonds,
119 and all require specific catalytic enzymes to make them biologically available in the form of orthophosphate. The
120 hydrolysis of P_o is mediated by the action of a suite of phosphatase enzymes which may have specificity for single
121 compounds or broad specificity to a range of compounds (George et al. 2007). Unlike for organic nitrogen, there
122 is no evidence for direct uptake of dissolved P_o compounds by biology, apart from the uptake of phosphonates by
123 bacteria in marine systems (Dyhrman et al. 2006). Plants and microbes possess a range of phosphatases that are
124 associated with various cellular functions, including; energy metabolism, nutrient transport, metabolic regulation
125 and protein activation (Duff et al. 1994). However, it is the extracellular phosphatases released into the soil that
126 are of particular importance for the mineralisation of soil P_o . Extracellular phosphatase activity is induced under
127 conditions of P deficiency and is either associated with root cell walls or released directly into the rhizosphere
128 (Richardson et al. 2009).

129 There have been a number of important advances in our understanding of P_o dynamics at the ecosystem and
130 rhizosphere scale in the past decade, with particular advancement in understanding of plant-soil-microorganism
131 interactions and concomitant advances in techniques used to assess these dynamics. It is now timely to start to
132 consider how to integrate this information and extract further understanding of the dynamics of P_o in the managed
133 and natural environment and this will have a number of potentially important impacts on how we tackle some of
134 the most pressing global issues of today. Here we summarise the state of the art of P_o research and identify
135 priorities for future research, which will help meet these goals.

136 **Establishing Priorities for Organic Phosphorus Research**

137 There has been a large increase in the number of publications in the P_o research field in the last two decades, with
138 ~400 publications in 2016, compared to 150 in 2000. In September 2016 a workshop on Organic Phosphorus was

139 held (<https://op2016.com>), gathering together 102 experts in the field of P_o research from 23 countries to identify
140 research priorities. Contributors were asked, in five groups, to consider the global issues associated with P_o,
141 methodological strengths and weaknesses, benefits to be gained from understanding the P_o cycle, and priorities
142 for P_o research. The information from the five groups was collected and the concepts, where consensus between
143 at least two of the groups was reached, are summarized in Table 1. It is clear from this that research into P_o has
144 the potential to have impacts on global biogeochemical cycles of P both in natural and managed systems and will
145 therefore potentially impact food security, agricultural sustainability, environmental pollution of both the aquatic
146 and atmospheric environments and will be profoundly affected by environmental change both in geopolitical terms
147 and through man-made climate change. We are well placed to tackle these as there are a number of strengths in
148 the way the research is performed and the weaknesses are well understood. It was considered that P_o research will
149 have a range of impactful outcomes on our understanding of how natural and agricultural systems work and has
150 the potential to give society a number of important tools to help manage the environment more effectively to either
151 prevent or mitigate against some of the major global threats. A number of research priorities were identified and
152 grouped into specific opportunities which are detailed below. The key opportunities to improve the effectiveness
153 of P_o research identified here are similar to those highlighted in Turner et al. (2005), although it is clear that some
154 progress has been made since that set of recommendations were made. However, the similarities and consistency
155 between the outcomes of these two studies suggests we still have some progress to make. A number of new priority
156 areas were identified here that were not identified in Turner et al. (2005), including the need for greater
157 understanding of the metagenomics and functional microbial genes involved in organic P turnover, greater
158 understanding of the impact of nanoparticles in the environment on organic P turnover and the need to integrate
159 the system more effectively in the form of models. It is clear that P_o research field is evolving, but some of the
160 issues of a decade ago still persist.

161 **1) Opportunities in organic phosphorus analytical methodologies**

162 The core analytical tools for the P_o discipline are ³¹P NMR spectroscopy (Cade-Menun and Liu 2014; Cade-Menun
163 2005; Cade-Menun et al. 2005; Turner et al. 2005), which is used to identify P_o compounds in several
164 environmental matrices, along with more traditional soil extraction methods, such as those to measure total P_o and
165 the fractionation method developed by Hedley et al. (Condon and Newman 2011; Hedley et al. 1982; Negassa
166 and Leinweber 2009). There is discussion and debate focused around the suitability of these analytical
167 methodologies for characterizing P_o in soil and terrestrial systems (Liu et al. 2014; Doolette and Smernik, 2011)

168 and this debate revolves around the identity of the broad base of the inositol hexaphosphate peak on NMR spectra,
169 which some contest is resolved and other suggest is unidentified (Jarosch et al. 2015). Despite this, research into
170 P_o is still limited methodologically and many methods are operationally-defined. Importantly, there is a need to
171 link the results from these methods to biological and biogeochemical processes in the environment. In the process
172 of achieving this, there is debate over the benefits of (i) standardization or homogenization of analytical methods,
173 versus the merits of (ii) promoting diversity of analytical procedures.

174 It is critical to develop non-destructive methods to analyse soil pools and their dynamics without the need for
175 extraction. Some solid-state methods, such as solid-state NMR or P-XANES (X-ray Adsorptive Near Edge
176 Structure) spectroscopy are limited by the naturally low concentrations of P_o forms in soils (Liu et al. 2013; 2014;
177 2015). Visible Near-Infrared Reflectance Spectroscopy (VNIRS) has shown some promise for determining total
178 P_o in soils (Abdi et al, 2016), but further testing is needed. Another priority for P_o methodologies is the
179 development of standard analytical quality controls through the use of standardized reference materials for cross-
180 comparison and checks on analytical methods. These standardized reference materials will include reference soils
181 and chemicals. There is a need for the community to identify standardized natural reference materials such as soils
182 and manures, but a large amount of effort would be needed to put together a collection of appropriate materials as
183 well as a means to share them internationally. Standardization of P_o compounds could be achieved through the
184 use of simple, relatively pure, and inexpensive P_o compounds (e.g. Na-phytate, glucose 1-P) purchased from a
185 single supplier operating in many countries with a guaranteed long-term production commitment. And there is a
186 need to develop a commercial supply of other commonly identified P_o compounds in soils, such as scyllo-inositol
187 hexakisphosphate, to allow the use of appropriate substrates for research fully understand the biological and
188 chemical processes controlling the behaviour of this and other P_o compounds in the environment. It is a priority
189 for researchers to further develop methods, while also refining existing P_o methods and standards, to generate
190 useful and comparable datasets and to build a consensus with respect to P_o dynamics and function in agricultural
191 and natural ecosystems.

192 **2) Opportunities from understanding stoichiometry – interactions of organic phosphorus with other** 193 **element cycles**

194 Comparing element ratios of living organisms and their non-living environment has been at the centre of scientific
195 debate for many years. In oceans, planktonic biomass is characterized by similar C:N:P ratios as marine water
196 (106:16:1) (Redfield 1958). While similar characteristic element ratios also exist for terrestrial ecosystems with

197 much greater heterogeneity across a range of spatial scales (Cleveland and Liptzin 2007). The comparison of
198 C:N:P ratios in the microbial biomass of soils with that of soil organic matter (SOM) may therefore help to identify
199 the nutrient status of the soil (Redfield 1958). Following this concept, the stoichiometric ratios of resources (e.g.,
200 SOM) over the microbial biomass has been calculated as a proxy for nutrient imbalances (Cleveland and Liptzin
201 2007). An understanding of stoichiometric ratios in soils and their relationship to those in crop plants and for the
202 decomposition of litter and SOM will provide an important indicator of nutrient status in terrestrial ecosystems
203 and better management of systems.

204 Until now, the large temporal and spatial heterogeneity of soil systems and the heterogeneous distribution of SOM
205 constituents have made the analysis and interpretation of ecosystem stoichiometry a challenge because for
206 microbial decomposers the elemental composition of micro-sites in soils might be more relevant than the overall
207 element ratio of the soil. For example, by analysing the C:N:P ratio of bulk soils only, information on relevant
208 and spatially-dependent processes may be lost (e.g., rhizosphere, soil horizons). The most obvious reason for soil-
209 specificity and heterogeneity among stoichiometric ratios is that part of the SOM is separated from
210 microorganisms and roots via physical and physicochemical barriers. By re-analysing the results of C:N:P:Sulphur
211 (S) analyses of SOM obtained from 2000 globally distributed soil samples, Tipping et al. (2016) demonstrated
212 that there is both nutrient-poor and nutrient-rich SOM, with the latter being strongly sorbed by soil minerals
213 (Tipping et al. 2016). This may be explained by the incorporation of SOM into aggregates (Stewart and Tiessen
214 1987) or the adsorption of P-containing organic and inorganic molecules to mineral surfaces (Celi et al. 2003;
215 Giaveno et al. 2010). Clay and metal (oxy)hydroxide minerals can sequester P_o and P_i released by microbial- or
216 plant-driven processes and/or affect enzyme activities, while limiting P biocycling (Celi and Barberis 2005). This
217 highlights the need to understand the tight interrelationship between chemical, physical and biological processes
218 and the potential for stoichiometric assessment as an indicator of P and organic matter availability in soils. Modern
219 analytical techniques which enable to analyse the stoichiometry of the soil constituents at a high resolution might
220 help provide this knowledge (Mueller et al. 2012).

221 There are many known mechanisms by which organisms can improve access to P_o (Richardson et al. 2011), but
222 there are several novel mechanisms being identified that target key components of SOM, such as polyphenols and
223 tannins, to mobilise P (Kohlen et al. 2011). A priority will be to understand the plant and microbial mechanisms
224 involved in the accumulation and mobilization of P from organic matter. It is important to attempt to determine
225 the optimal stoichiometry between C:N:P, and understand the role P_o plays in this, to allow sustainable

226 management of P in arable soils and to identify anthropogenic nutrient imbalances in natural, agricultural and
227 forest ecosystems (Frossard et al. 2015).

228 **3) Opportunities from understanding interactions of organic phosphorus with land management**

229 An ability to utilise P_o to sustain agronomic productivity with declining conventional fertiliser inputs drives
230 research into interactions among P_o, land use and management (Nash et al. 2014; Stutter et al. 2012). The
231 conditions to better utilise P_o may bring benefits for other soil quality factors (e.g., SOM status and microbial
232 cycling), but may require management of potentially adverse effects on wider biological cycles and water quality
233 (Dodd and Sharpley 2015). Societal drivers for food and timber production underpin much of the research into P_o
234 speciation, biological turnover and integration with agronomic systems. Numerous studies have reported P_o stocks
235 and changes associated with management; fewer have studied the time-course of transformations and turnover
236 with management change, linked with soil chemical and biological processes. The interactions between P
237 speciation, (bio)availability and SOM are of prime importance since land management greatly affects SOM in
238 space and time (in beneficial or detrimental ways) and exert strong geochemical and microbial controls on P_o
239 cycling.

240 The interactions of land cover, use and management are important for understanding the role of P_o across
241 ecosystems. In agricultural systems, the information on soil P_o stocks is well represented have been quantified by
242 numerous studies in North America (Abdi et al. 2014; Cade-Menun et al. 2015; Liu et al. 2015; Schneider et al.
243 2016), Europe (Ahlgren et al. 2013; Annaheim et al. 2015; Keller et al. 2012; Stutter et al. 2015), China (Liu et
244 al. 2013), South America (de Oliveira et al. 2015), and Australia (Adeloju et al. 2016). In forestry, such
245 information is available in tropical (Zaia et al. 2012) and temperate systems (Slazak et al. 2010) and orchards (Cui
246 et al. 2015). However, an important improvement will be to better understand the reasons as to why particular
247 stocks exist under certain geoclimatic-land cover combinations. Key opportunities exist to understand P_o dynamics
248 for sustainable P use in tropical systems and for forests growing on marginal soils, both of which depend on
249 effective management of P_o resources.

250 It is known that both land cover and management factors (tillage, fertilizer type, application rate and timing)
251 interact with abiotic factors in controlling P_o stocks and cycling, such as SOM, stabilizing surfaces [e.g., Fe- and
252 aluminium (Al)-oxides, calcium (Ca) forms, clays] and soil moisture, (Adeloju et al. 2016; Cade-Menun et al.
253 2015; Stutter et al. 2015). Chemical fractionation studies of P_o stocks provide a snap-shot in time, missing
254 temporal aspects of cycling associated with management-induced change at seasonal or to longer term

255 management. As a result, short periods of rapid change in P speciation and turnover may not be appreciated. The
256 utilization of ‘legacy P’ (Haygarth et al. 2014; Powers et al. 2016), following declining fertiliser inputs or altered
257 cropping practices, has been studied following long-duration manipulations. Often these look at the end point of
258 change (Cade-Menun et al. 2015), but have not ‘followed’ the dynamic. Although powerful methods for P_o
259 assessment are developing rapidly, studies that preceded these have the opportunity to incorporate them with
260 archived samples or control soils (Keller et al. 2012; Liu et al. 2015). Long-term understanding of P_o dynamics in
261 management systems should be pursued, while short-term seasonal observations (for example Ebuele et al. 2016)
262 will be needed to understand the influence of microbial dynamics on P speciation and turnover under various land-
263 use and management scenarios. If studies of short-term perturbations (via management, climate etc) can show
264 benefits for providing greater P_o resources into available pools then these processes may be beneficially
265 incorporated in future land management.

266 ‘Organic’ farming brings a commercial stimulus to substitute agro-chemicals (including chemical P fertilisers)
267 with sustainable management, such as use of organic amendments, for example enhancing soil P cycling with the
268 aim of better utilizing P already present and moving towards a ‘closed’ system (Annaheim et al. 2015; Gaiind and
269 Singh 2016; Schneider et al. 2016). The same approaches can be applied to less intensive, or developing,
270 agricultural systems. Canadian pastures managed under an organic regime, had a greater abundance of P_o (65%
271 vs 52% of total P) compared to conventional pastures and were able to maintain yield without inorganic fertilisers
272 (Schneider et al. 2016). These authors concluded that plants were using P_i rather than P_o and supported by other
273 studies showing no indication that the greater microbial activity under organic farming caused utilization of
274 stabilized P_o forms (Keller et al. 2012). Therefore, the management conditions and actions required to promote
275 better acquisition of P_o pools remain elusive.

276 The consensus is that a key question remains: How long could the turnover of P_o sustain crop yields under
277 scenarios of reduced P inputs and maintained or increased outputs and thus contribute to agricultural production
278 and feed supplies? The mechanistic understanding required to answer this question lies in the role of biota (in the
279 context of their abiotic setting) in P_o turnover and the potential pathways of P_o loss to be managed (e.g. runoff).
280 In order to progress, a systems approach is needed to fully assess the opportunities and role of P_o , as well as the
281 interactions of soil chemical, physical and biological processes and impacts of land use change that control P
282 availability.

283 **4a) Opportunities from understanding microbial P_o : functional genes and metagenomics**

284 As our abilities to analyse and interpret the complexity inherent in the soil microbiome improves, interest is
285 burgeoning around the functional ecology of microorganisms. Organic P dynamics across ecosystems, along with
286 development of many techniques that will aid in this understanding, are beginning to emerge. Scavenging of P
287 from P-containing organic compounds by soil microbes is tightly controlled by intracellular P availability through
288 the Pho pathway in yeast (Secco et al. 2012) and the Pho regulon in bacteria. In both cases, transcription of
289 phosphatase and phytase, which act to release orthophosphate from phosphate esters, and high affinity transporters
290 which transport P_i into the cell, are up-regulated under P_i limitation, affecting the organisms' ability to utilise P_o .
291 The Pho regulon also acts as a major regulator of other cellular processes, including N assimilation and ammonium
292 uptake (Santos-Beneit 2015). The C:N:P elemental ratios of the soil bacterium *Bacillus subtilis* range between
293 $C_{53-125}:N_{12-29}:P_1$ under N- and P-limited culture conditions (Dauner et al. 2001), although environmental
294 assemblages may exhibit greater stoichiometric flexibility (Godwin and Cotner 2015). Given this regulatory cross-
295 talk, nutrient stoichiometry will be important to cellular and community metabolism meaning that the cycling of
296 P must be considered within the context of other biogeochemical cycles, as highlighted earlier.

297 Soil type, nutrient inputs, and plant species have been shown to determine microbiota species composition and
298 function (Alegria-Terrazas et al. 2016). However, plant root exudation drives recruitment of specific microbes
299 and microbial consortia to the rhizosphere and may outweigh the impacts of soil and its management in shaping
300 community composition and function (Tkacz et al. 2015). As yet, there is only limited understanding of how
301 specific root exudates affect microbial recruitment (Neal et al. 2012), let alone specific microbiota responsible for
302 phosphatase expression and production. A better understanding of interactions between plants and microbes would
303 facilitate identification of functional redundancy among them, which could ultimately help manage the availability
304 of P in soils and sediments by selection of the optimal plant rhizosphere complement.

305 Alkaline phosphatase and phytase genes are distributed across a broad phylogenetic range and display a high
306 degree of microdiversity (Jaspers and Overmann 2004; Lim et al. 2007; Zimmerman et al. 2013), where closely
307 related organisms exhibit different metabolic activities. It is therefore not possible to determine community
308 functional potential from 16S rRNA gene abundance – functional gene abundance information is required and
309 this can be provided by employing sequencing techniques to assess the soil metagenome. In marine systems, there
310 is evidence from metagenomic sequencing of environmental DNA that alkaline phosphatase genes *phoD* and
311 *phoX* are more abundant than *phoA* (Luo et al. 2009; Sebastian and Ammerman 2009) and the β -propeller phytase
312 is the most abundant phytase gene (Lim et al. 2007). The dominant alkaline phosphatase gene in terrestrial

313 ecosystems is also *phoD* (Tan et al. 2013), which is more abundant in soils than other environments (Courty et al.
314 2010; Ragot et al. 2015; Fraser et al. 2017). From a functional standpoint, abundance of *phoD*-like sequences
315 correlate well with estimates of potential alkaline phosphatase activity (Fraser et al. 2015), although this is not
316 always the case (Ragot et al. 2015). Moreover, in soils there is little information regarding other phosphatases and
317 little is known about the distribution and abundance of bacterial acid phosphatases, but there is some information
318 related to *phoX* (Ragot et al. 2016). In contrast, fungi are well known for their capacity to secrete acid phosphatases
319 (Plassard et al. 2011; Rosling et al. 2016), especially ectomycorrhizal fungi. Since only a small percentage of soil
320 microorganisms are cultivable, research will need to rely upon culture-independent approaches to generate a
321 thorough understanding of the abundance and diversity of genes associated with P_o turnover. Environmental
322 metagenomic sequencing can form the basis of an efficient molecular toolkit for studying microbial gene
323 dynamics and processes relevant to P_o mineralization (Neal et al., 2017). Such an approach will need to prioritize
324 generating comprehensive understanding of the distribution of alkaline and acid phosphatase and phytase genes
325 within soils, coupled with activity measurements, and a sense of their relative sensitivities to edaphic factors. This
326 will allow explicit incorporation of microbial P_o turnover in the new generation of soil models, as well as allowing
327 rapid assessment of a soil's capabilities for P_o cycling. Improved knowledge will allow the exploitation of
328 microbial activity to sustain and improve soil fertility and allow the tailoring of new fertilizers based upon the
329 capacity of microbes to exploit P_o .

330 **4b) Opportunities from understanding microbial P_o : measuring stocks, mineralisation and dynamics of** 331 **turnover**

332 The apparently large diversity of genes associated with P_o -hydrolysing enzymes suggests that changes in
333 community composition are unlikely to result in a loss of ecosystem function. This confers resilience to P -cycling
334 processes, although many of these genes have very specific functions intracellularly. However, trait differences
335 are likely to have significant implications for community function in soils, e.g., the contrasting effects of
336 arbuscular and ectomycorrhizal fungi upon the cycling of P in forest soils, where it has been shown that P_o is more
337 labile in ectomycorrhizal dominated systems than arbuscular mycorrhizal systems (Rosling et al. 2016). The fact
338 that enzyme activity in soil appears to be disconnected from soil P status is at odds with the apparent influence of
339 the *Pho* regulon or pathway upon gene expression and indicates that much of the observed activity derives from
340 multiple enzyme sources, which have been stabilised by soil colloids (Nannipieri et al. 2011). This also suggests
341 that soil enzyme activity does not directly represent microbial activity or simply reflects the complexity in current

342 P requirements of different microbial species. However, visualization of acid and alkaline phosphatase activity
343 associated with roots by zymography (Spohn and Kuzyakov 2013) does provide an exciting means to determine
344 regulation of soil phosphatase activity with P availability and illustrates the clear spatial separation among the
345 activities of physiologically different enzymes. It is a priority to develop and couple techniques that resolve the
346 distribution of active enzymes in soil with estimates of gene expression derived from functional genes or meta-
347 transcriptomic studies.

348 The stock of microbial P is an easy-to-determine component in soils, which is widely used to characterize the P
349 status of microbial communities and ecosystems (Brookes et al. 1982; 1984). Nevertheless, its analysis relies on
350 many different protocols (Bergkemper et al. 2016). Building on the previous work, further insights into both
351 microbial-mediated and enzyme-mediated P transformations in soils may now be gained from measurement of
352 the isotopic composition of oxygen associated with phosphate ($\delta^{18}\text{O}_\text{P}$) (Tamburini et al. 2014; von Sperber et al.
353 2014) and the use of radiolabelled (^{32}P or ^{33}P) P_o compounds to measure mineralisation and immobilisation rates
354 directly (Harrison 1982). A powerful tool for quantifying soil P pools and transformation rates is the isotope
355 dilution technique [reviewed in Bünemann 2015; Di et al. 2000; Frossard et al. 2011]. The decrease in
356 radioactivity with time is caused by the exchange of the added radiolabelled P (either ^{32}P or ^{33}P) with ^{31}P from the
357 sorbed/solid phase and by the release of inorganic ^{31}P from the organic pool via hydrolysing enzymes (Bünemann
358 2015). Determination of gross P_o mineralization rates from P_o to P_i remains a critical approach, helping understand
359 the processes and rates of P cycling in different soils and under different environmental conditions (Frossard et
360 al. 2011). These techniques present new opportunities to link P cycling to other biogeochemical cycles, such as C
361 and N.

362 **5) Opportunities in the emerging area of interactions between P_o dynamics and nanoparticles**

363 Reactive nanoparticles can take the form of natural soil colloids or man-made particles and are potential P_o
364 carriers, sources and sinks in ecosystems. Up to 90% of P in stream water and runoff is present in nano- and
365 colloidal sized materials (Borda et al. 2011; Gottselig et al. 2014; Uusitalo et al. 2003; Withers et al. 2009).
366 Colloidal P may comprise nano-sized aggregates (Jiang et al. 2015) bound to Fe, Al and SOM (Celi and Barberis
367 2005; Celi and Barberis 2007), including inositol phosphates. However, the influence of nanoparticles on the
368 dynamics and bioavailability of P in soil-plant systems is unclear (Bol et al. 2016). Nanoparticles such as C-
369 magnetite, which adsorb and retain P_i and P_o , are used to enhance the recovery and recycling of P from P-rich
370 wastes (Magnacca et al. 2014; Nisticò et al. 2016). It may also be possible to enhance soil enzyme activity with

371 amendments containing mesoporous nanoparticle materials (Zhou and Hartmann 2012). Phytase encapsulated in
372 nanoparticles was shown to be resistant to inhibitors and proteases and to promote the hydrolysis of phytate for P
373 uptake by *Medicago truncatula* (Trouillefou et al. 2015). Nanotechnology has also been used to develop new
374 fertilizers and plant-growth-enhancing materials (Liu and Lal 2015), representing one potentially effective option
375 for enhancing global food production. A better understanding of the P_o nanoparticle interaction may improve our
376 understanding on P fluxes in natural and agricultural systems, and provide innovative technologies for fertilizer
377 production and environmental remediation.

378 **6) Opportunities to use modelling of P_o in soil and ecosystems**

379 The use of all types of modelling approaches to study P_o is generally overlooked and there is a dearth of P_o based
380 models, but development of such models would be extremely beneficial. Modelling should facilitate the
381 development of a systems-based perspective and help to identify knowledge gaps in the current understanding of
382 P_o. Models of all types are needed including those that are conceptual, mechanistic or empirical in nature and in
383 general there is a lack of focus on all the types of models that exist for P_o. The potential benefits of advances in
384 modelling for P_o include:

- 385 • Prediction of the relationship between soil P_o and plant uptake, which should be developed in both conceptual
386 and mechanistic models of P dynamics in the environment.
- 387 • Application at different scales to determine the relationship between P_o with land use and management should
388 be possible by building empirical models based on existing data.
- 389 • Application of modelling to help understand the role of microbial traits in soil (Wieder et al. 2015), which
390 may determine the effects of gene expression, enzyme activities and the stoichiometric ratio of C:N:P in the
391 microbial biomass relative to that of SOM
- 392 • Application of complete Life-Cycle Analysis for relying of the run-down of soil P_o as a replacement to
393 inorganic fertilisers will help us develop adequate conceptual models for management of the system.
- 394 • Modelling could also be used to help in the quantification of soil P pools for estimating flow among P_o pools.

395 In general, there is a great opportunity for the development of modelling in all areas of P_o research and this will
396 be of considerable benefit to the subject if this can be developed and integrated with all areas. The cooperation of
397 modellers and empiricists is essential for building models with great potential use to predict changes in P_o
398 bioavailability due to land-use and management change and to infer the sustainability of the system as a whole.

399

400 **7) Opportunities to better communicate and translate research**

401 Organic P represents a small, albeit critical component of biogeochemical research. The marginal nature of the
402 subject to date creates a need to communicate the importance of this science for the future of P sustainability. As
403 for other scientific disciplines, communication priorities include (1) strengthening communication among
404 scientists within and outside of the P_o research community; (2) engagement with stakeholders; and (3)
405 dissemination of knowledge to the public and specific end-users.

406 Conferences and workshops on the topic of organic P promote the exchange of ideas and forging of new research
407 partnerships (Sharpley et al. 2015; Turner et al. 2015). Online platforms are also powerful tools to connect
408 researchers and stakeholders on issues of global P sustainability (e.g., European Sustainable Phosphorus Platform,
409 www.phosphorusplatform.eu, North America Partnership for Phosphorus Sustainability) (Rosemarin and Ekane
410 2015). The ‘Soil Phosphorus Forum’ (www.soilpforum.com) provides a platform for the exchange of information
411 relating to P_o. Specific protocols and conference presentations are also featured in archived YouTube channels
412 (<https://www.youtube.com/channel/UCtGI3eUZscCgByewafsQKdw>). A central platform for P_o research and
413 communications is still needed, to connect existing forums to global research networks and would include features
414 such as researcher membership, methodological resources, links to relevant organizations and platforms, and a
415 clearing house of P_o data for future meta-analysis and modelling efforts.

416 Key stakeholder groups such as land managers, farmers and extension services are a natural link between industry,
417 government, and academia (FAO 2016). These key groups hold traditional knowledge on sustainable farming
418 techniques, which serve as a potential basis for future P_o research. Industry initiatives such as the 4R Nutrient
419 Stewardship framework provide feedback from end users and practitioners on research priorities associated with
420 the management of agricultural nutrients (Vollmer-Sanders et al. 2016). The engagement of P_o researchers with
421 existing nutrient initiatives such as these will be critical for bolstering public understanding of P_o and its important
422 role in global P dynamics.

423

424 **Conclusion - Statement of intent for the P_o research community**

425 Organic P research has a critical role to play in tackling a number of important global challenges and there are
426 key contributions to be made toward understanding biogeochemical cycles, dynamics and function of natural
427 ecosystems and the management of agricultural systems. In particular, we must reduce our reliance on inorganic
428 P fertilisers and strategies to do this will increase the relevance of soil P_o for plant nutrition. Secondly, there is a

429 need to develop a circular P economy and close the P cycle which will likely lead to an increase in the amounts
430 of organic P “waste” products being recycled to land shifting the P_o/P_i balance in the soil. To address these global
431 environmental changes and challenges, we should concentrate our efforts on understanding the biological
432 significance of P_o by considering its interactions with other elements in SOM, soil microorganisms and active soil
433 surfaces. We should consider these interactions with respect to changes in land use and management and as a
434 function of geochemical conditions in the wider biophysical and socio-economic environment. We need to
435 integrate this understanding through the production of models for P_o , which capture both whole systems and fine-
436 scale mechanisms. In addition, we need to develop novel and standardised methodologies that can integrate the
437 dynamics and function of P_o on appropriate scales in a non-invasive manner. To achieve a step-change in the
438 impact of P_o research, we need to engage with researchers outside of the discipline, align the research with pressing
439 societal issues, and become more global, collaborative, inclusive, interdisciplinary, and longer-term in nature. The
440 key to fostering this change will depend on logically communicating the importance of P_o to society at large,
441 engaging with stakeholders on important global issues, and ultimately pushing this important area of research up
442 the agenda of policy makers and funding bodies on a global scale.

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451 **References**

- 452 Abdi D, Cade-Menun BJ, Ziadi N, Parent L-É (2014) Long-Term Impact of Tillage Practices and Phosphorus
453 Fertilization on Soil Phosphorus Forms as Determined by ^{31}P Nuclear Magnetic Resonance
454 Spectroscopy. *J Environ Qual* 43: 1431-1441. doi: 10.2134/jeq2013.10.0424.
- 455 Abdi D, Cade-Menun BJ, Ziadi N, Tremblay GF, Parent L-É (2016) Visible near infrared reflectance
456 spectroscopy to predict soil phosphorus pools in chernozems of Saskatchewan, Canada. *Geoderma*
457 *Region 7*: 93-101.

458 Adeloju S, Webb B, Smernik R (2016) Phosphorus Distribution in Soils from Australian Dairy and Beef
459 Rearing Pastoral Systems. *Appl Sci* 6: 31.

460 Ahlgren J, Djodjic F, Börjesson G, Mattsson L (2013) Identification and quantification of organic phosphorus
461 forms in soils from fertility experiments. *Soil Use and Management* 29: 24-35. doi:
462 10.1111/sum.12014.

463 Alegria-Terrazas R, Giles CD, Paterson E, Robertson-Albertyn S, Cesco S, Mimmo T, Pii Y, Bulgarelli D
464 (2016) Plant-Microbiota Interactions as a Driver of the Mineral Turnover in the Rhizosphere. *Adv
465 Appl Microbiol.* Springer.

466 Annaheim KE, Doolette AL, Smernik RJ, Mayer J, Oberson A, Frossard E, Bünemann EK (2015) Long-term
467 addition of organic fertilizers has little effect on soil organic phosphorus as characterized by ³¹P NMR
468 spectroscopy and enzyme additions. *Geoderma* 257–258: 67-77. doi:
469 <http://dx.doi.org/10.1016/j.geoderma.2015.01.014>.

470 Attiwill PM, Adams MA (1993) Nutrient cycling in forests. *New Phytol* 124: 561-582. doi: 10.1111/j.1469-
471 8137.1993.tb03847.x.

472 Bergkemper F, Bünemann EK, Hauenstein S, Heuck C, Kandeler E, Krüger J, Marhan S, Mészáros É, Nassal D,
473 Nassal P, Oelmann Y, Pistocchi C, Schloter M, Spohn M, Talkner U, Zederer DP, Schulz S (2016) An
474 inter-laboratory comparison of gaseous and liquid fumigation based methods for measuring microbial
475 phosphorus (P_{mic}) in forest soils with differing P stocks. *J Microbiol Methods* 128: 66-68. doi:
476 <http://dx.doi.org/10.1016/j.mimet.2016.07.006>.

477 Bol R, Julich D, Brödlin D, Siemens J, Kaiser K, Dippold MA, Spielvogel S, Zilla T, Mewes D, von
478 Blanckenburg F, Puhlmann H, Holzmann S, Weiler M, Amelung W, Lang F, Kuzyakov Y, Feger K-H,
479 Gottselig N, Klumpp E, Missong A, Winkelmann C, Uhlig D, Sohr J, von Wilpert K, Wu B, Hagedorn
480 F (2016) Dissolved and colloidal phosphorus fluxes in forest ecosystems—an almost blind spot in
481 ecosystem research. *J Plant Nutr Soil Sci* 179: 425-438. doi: 10.1002/jpln.201600079.

482 Borda T, Celi L, Zavattaro L, Sacco D, Barberis E (2011) Effect of agronomic management on risk of
483 suspended solids and phosphorus losses from soil to waters. *J Soils Seds* 11: 440-451. doi:
484 10.1007/s11368-010-0327-y.

485 Brookes PC, Powlson DS, Jenkinson DS (1982) Measurement of microbial biomass phosphorus in soil. *Soil
486 Biol Biochem* 14: 319-329. doi: [http://dx.doi.org/10.1016/0038-0717\(82\)90001-3](http://dx.doi.org/10.1016/0038-0717(82)90001-3).

487 Brookes PC, Powlson DS, Jenkinson DS (1984) Phosphorus in the soil microbial biomass. *Soil Biol Biochem*
488 16: 169-175. doi: [http://dx.doi.org/10.1016/0038-0717\(84\)90108-1](http://dx.doi.org/10.1016/0038-0717(84)90108-1).

489 Bünemann EK (2015) Assessment of gross and net mineralization rates of soil organic phosphorus – A review.
490 *Soil Biology Biochem* 89: 82-98. doi: 10.1016/j.soilbio.2015.06.026.

491 Butusov M, Jernelöv A (2013) Phosphorus in the Organic Life: Cells, Tissues, Organisms. *Phosphorus: An*
492 *Element that could have been called Lucifer*. Springer New York, New York, NY.

493 Cade-Menun B, Liu CW (2014) Solution phosphorus-31 nuclear magnetic resonance spectroscopy of soils from
494 2005 to 2013: A review of sample preparation and experimental parameters. *Soil Sci Soc Am J* 78: 19-
495 37. doi: 10.2136/sssaj2013.05.0187dgs.

496 Cade-Menun BJ (2005) Characterizing phosphorus in environmental and agricultural samples by 31 P nuclear
497 magnetic resonance spectroscopy. *Talanta* 66: 359-371.

498 Cade-Menun BJ, He Z, Zhang H, Endale DM, Schomberg HH, Liu CW (2015) Stratification of Phosphorus
499 Forms from Long-Term Conservation Tillage and Poultry Litter Application. *Soil Sci Soc Am J* 79:
500 504-516. doi: 10.2136/sssaj2014.08.0310.

501 Cade-Menun BJ, Turner B, Frossard E, Baldwin D (2005) Using phosphorus-31 nuclear magnetic resonance
502 spectroscopy to characterize organic phosphorus in environmental samples. *Organic phosphorus in the*
503 *environment*: 21-44.

504 Celi L, Barberis E (2005) Abiotic stabilization of organic phosphorus in the environment. *Organic phosphorus*
505 *in the environment*. CABI Pub pp 113-132.

506 Celi L, Barberis E (2007) Abiotic reactions of inositol phosphates in soils. In: BL Turner, AE Richardson, EJ
507 Mullaney (eds) *Inositol Phosphates: Linking Agriculture and the Environment*. CAB International,
508 Oxfordshire, UK.

509 Celi L, De Luca G, Barberis E (2003) Effects of interaction of organic and inorganic P with ferrihydrite and
510 kaolinite-iron oxide systems on iron release. *Soil Sci* 168: 479-488.

511 Chardon WJ, Oenema O (1995) Leaching of dissolved organically bound phosphorus. DLO Research Institute
512 for Agrobiological and Soil Fertility.

513 Chardon WJ, Oenema O, del Castilho P, Vriesema R, Japenga J, Blaauw D (1997) Organic phosphorus in
514 solutions and leachates from soils treated with animal slurries. *J. Environ. Q.* 26: 372-378.

515 Cleveland CC, Liptzin D (2007) C:N:P stoichiometry in soil: is there a “Redfield ratio” for the microbial
516 biomass? *Biogeochem* 85: 235-252. doi: 10.1007/s10533-007-9132-0.

517 Condon LM, Newman S (2011) Revisiting the fundamentals of phosphorus fractionation of sediments and
518 soils. *J Soils Seds* 11: 830-840. doi: 10.1007/s11368-011-0363-2.

519 Courty P-E, Franc A, Garbaye J (2010) Temporal and functional pattern of secreted enzyme activities in an
520 ectomycorrhizal community. *Soil Biol Biochem* 42: 2022-2025. doi: 10.1016/j.soilbio.2010.07.014.

521 Cui H, Zhou Y, Gu Z, Zhu H, Fu S, Yao Q (2015) The combined effects of cover crops and symbiotic microbes
522 on phosphatase gene and organic phosphorus hydrolysis in subtropical orchard soils. *Soil Biology and*
523 *Biochemistry* 82: 119-126. doi: 10.1016/j.soilbio.2015.01.003.

524 Darch T, Blackwell MSA, Hawkins JMB, Haygarth PM, Chadwick D (2014) A Meta-Analysis of Organic and
525 Inorganic Phosphorus in Organic Fertilizers, Soils, and Water: Implications for Water Quality. *Crit Rev*
526 *Environ Sci Technol* 44: 2172-2202. doi: 10.1080/10643389.2013.790752.

527 Dauner M, Storni T, Sauer U (2001) *Bacillus subtilis* Metabolism and Energetics in Carbon-Limited and
528 Excess-Carbon Chemostat Culture. *J Bacteriol* 183: 7308-7317. doi: 10.1128/JB.183.24.7308-
529 7317.2001.

530 de Oliveira CMB, Erich MS, Gatiboni LC, Ohno T (2015) Phosphorus fractions and organic matter chemistry
531 under different land use on Humic Cambisols in Southern Brazil. *Geoderma Regional* 5: 140-149. doi:
532 <http://dx.doi.org/10.1016/j.geodrs.2015.06.001>.

533 Di HJ, Cameron KC, McLaren RG (2000) Isotopic dilution methods to determine the gross transformation rates
534 of nitrogen, phosphorus, and sulfur in soil: a review of the theory, methodologies, and limitations. *Soil*
535 *Res* 38: 213-230. doi: <http://dx.doi.org/10.1071/SR99005>.

536 Dodd RJ, Sharpley AN (2015) Recognizing the role of soil organic phosphorus in soil fertility and water quality.
537 *Res Conserv Recycl* 105, Part B: 282-293. doi: 10.1016/j.resconrec.2015.10.001.

538 Doolette AL, Smernik RJ. (2011) Soil organic phosphorus speciation using spectroscopic techniques. In
539 *Phosphorus in action*, Springer Berlin Heidelberg pp. 3-36

540 Duff SM, Sarath G, Plaxton WC (1994) The role of acid phosphatases in plant phosphorus metabolism. *Physiol.*
541 *Plant*. 90: 791-800.

542 Dyhrman ST, Chappell PD, Haley ST, Moffett JW, Orchard ED, Waterbury JB, Webb EA. (2006) Phosphonate
543 utilization by the globally important marine diazotroph *Trichodesmium*. *Nature*. 439: 68.

544 Ebuele VO, Santoro A, Thoss V (2016) Phosphorus speciation by ³¹P NMR spectroscopy in bracken (*Pteridium*
545 *aquilinum* (L.) Kuhn) and bluebell (*Hyacinthoides non-scripta* (L.) Chouard ex Rothm.) dominated
546 semi-natural upland soil. *Sci Tot Environ* 566–567: 1318-1328. doi: 10.1016/j.scitotenv.2016.05.192.

547 Espinosa M, Turner B, Haygarth P (1999) Preconcentration and separation of trace phosphorus compounds in
548 soil leachate. *J. Environ Q* 28: 1497-1504.

549 Food and Agricultural Organization of the United Nations (2016). Research and Extension.
550 <http://www.fao.org/nr/research-extension-systems/res-home/en/>. Date Accessed: 13 October 2016.

551 Fraser T, Lynch DH, Entz MH, Dunfield KE (2015) Linking alkaline phosphatase activity with bacterial *phoD*
552 gene abundance in soil from a long-term management trial. *Geoderma* 257–258: 115-122. doi:
553 10.1016/j.geoderma.2014.10.016.

554 Fraser TD, Lynch DH, Gaiero J, Khosla K, Dunfield KE. (2017) Quantification of bacterial non-specific acid
555 (*phoC*) and alkaline (*phoD*) phosphatase genes in bulk and rhizosphere soil from organically managed
556 soybean fields. *Applied Soil Ecology* 111:48-56.

557 Frossard E, Achat DL, Bernasconi SM, Bünemann EK, Fardeau J-C, Jansa J, Morel C, Rabeharisoa L,
558 Randriamanantsoa L, Sinaj S, Tamburini F, Oberson A (2011) The Use of Tracers to Investigate
559 Phosphate Cycling in Soil–Plant Systems. In: E Bünemann, A Oberson, E Frossard (eds) *Phosphorus in*
560 *Action: Biological Processes in Soil Phosphorus Cycling*. Springer Berlin Heidelberg, Berlin,
561 Heidelberg.

562 Frossard E, Buchmann N, Bünemann EK, Kiba DI, Lompo F, Oberson A, Tamburini F, Traoré OY. (2015) Soil
563 properties and not inputs control carbon, nitrogen, phosphorus ratios in cropped soils in the long-term.
564 *Soil Discuss.* 2:995-1038.

565 Gaind S, Singh YV (2016) Soil organic phosphorus fractions in response to long-term fertilization with
566 composted manures under rice–wheat cropping system. *J Plant Nutri* 39: 1336-1347. doi:
567 10.1080/01904167.2015.1086795.

568 George TS, Simpson RJ, Gregory PJ, Richardson AE (2007) Differential interaction of *Aspergillus niger* and
569 *Peniophora lycii* phytases with soil particles affects the hydrolysis of inositol phosphates. *Soil Biol.*
570 *Biochem.* 39: 793-803.

571 Giaveno C, Celi L, Richardson AE, Simpson RJ, Barberis E (2010) Interaction of phytases with minerals and
572 availability of substrate affect the hydrolysis of inositol phosphates. *Soil Biol Biochem* 42: 491-498.
573 doi: 10.1016/j.soilbio.2009.12.002.

574 Godwin CM, Cotner JB (2015) Aquatic heterotrophic bacteria have highly flexible phosphorus content and
575 biomass stoichiometry. *ISME J* 9: 2324-2327. doi: 10.1038/ismej.2015.34.

576 Gottselig N, Bol R, Nischwitz V, Vereecken H, Amelung W, Klumpp E (2014) Distribution of Phosphorus-
577 Containing Fine Colloids and Nanoparticles in Stream Water of a Forest Catchment. *Vadose Zone J* 13.
578 doi: 10.2136/vzj2014.01.0005.

579 Harrison AF (1982) 32P-method to compare rates of mineralization of labile organic phosphorus in woodland
580 soils. *Soil Biol Biochem* 14: 337-341. doi: 10.1016/0038-0717(82)90003-7.

581 Haygarth PM, Jarvie HP, Powers SM, Sharpley AN, Elser JJ, Shen J, Peterson HM, Chan NI, Howden NJ, Burt
582 T, Worrall F, Zhang F, Liu X (2014) Sustainable phosphorus management and the need for a long-term
583 perspective: the legacy hypothesis. *Environ Sci Technol* 48: 8417-8419. doi: 10.1021/es502852s.

584 Hedley MJ, Stewart JWB, Chauhan BS (1982) Changes in inorganic and organic soil phosphorus fractions
585 induced by cultivation practices and by laboratory incubations. *Soil Sci Soc Am J* 46: 970-976.

586 Jarosch KA, Doolette AL, Smernik RJ, Tamburini F, Frossard E, Bünemann EK.(2015) Characterisation of soil
587 organic phosphorus in NaOH-EDTA extracts: a comparison of 31 P NMR spectroscopy and enzyme
588 addition assays. *Soil Biology and Biochemistry* 91:298-309.

589 Jaspers E, Overmann J (2004) Ecological Significance of Microdiversity: Identical 16S rRNA Gene Sequences
590 Can Be Found in Bacteria with Highly Divergent Genomes and Ecophysologies. *Appl Environ*
591 *Microbiol* 70: 4831-4839. doi: 10.1128/AEM.70.8.4831-4839.2004.

592 Jiang X, Bol R, Willbold S, Vereecken H, Klumpp E (2015) Speciation and distribution of P associated with Fe
593 and Al oxides in aggregate-sized fraction of an arable soil. *Biogeosci* 12: 6443-6452. doi: 10.5194/bg-
594 12-6443-2015.

595 Keller M, Oberson A, Annaheim KE, Tamburini F, Mäder P, Mayer J, Frossard E, Bünemann EK (2012)
596 Phosphorus forms and enzymatic hydrolyzability of organic phosphorus in soils after 30 years of
597 organic and conventional farming. *Journal of Plant Nutrition and Soil Science* 175: 385-393. doi:
598 10.1002/jpln.201100177.

599 Kohlen W, Charnikhova T, Liu Q, Bours R, Domagalska MA, Beguerie S, Verstappen F, Leyser O,
600 Bouwmeester H, Ruyter-Spira C (2011) Strigolactones are transported through the xylem and play a
601 key role in shoot architectural response to phosphate deficiency in nonarbuscular mycorrhizal host
602 *Arabidopsis*. *Plant physiol* 155: 974-987. doi: 10.1104/pp.110.164640.

603 Lang F, Bauhus J, Frossard E, George E, Kaiser K, Kaupenjohann M, Krüger J, Matzner E, Polle A, Prietzel J,
604 Rennenberg H, Wellbrock N (2016) Phosphorus in forest ecosystems: New insights from an ecosystem
605 nutrition perspective. *J Plant Nutri Soil Sci* 179: 129-135. doi: 10.1002/jpln.201500541.

606 Lim BL, Yeung P, Cheng C, Hill JE (2007) Distribution and diversity of phytate-mineralizing bacteria. *ISME J*:
607 321-330. doi: 10.1038/ismej.2007.40.

608 Liu J, Hu Y, Yang J, Abdi D, Cade-Menun BJ (2015) Investigation of soil legacy phosphorus transformation in
609 long-term agricultural fields using sequential fractionation, P K-edge XANES and solution P NMR
610 spectroscopy. *Environ Sci Technol* 49: 168-176. doi: 10.1021/es504420n.

611 Liu J, Yang J, Cade-Menun BJ, Liang X, Hu Y, Liu CW, Zhao Y, Li L, Shi J (2013) Complementary
612 Phosphorus Speciation in Agricultural Soils by Sequential Fractionation, Solution ^{31}P Nuclear
613 Magnetic Resonance, and Phosphorus K-edge X-ray Absorption Near-Edge Structure Spectroscopy. *J*
614 *Environ Qual* 42: 1763-1770. doi: 10.2134/jeq2013.04.0127.

615 Liu J, Hu Y, Yang J, Abdi D, Cade-Menun BJ (2014) Investigation of soil legacy phosphorus transformation in
616 long-term agricultural fields using sequential fractionation, P K-edge XANES and solution P NMR
617 spectroscopy. *Environ Sci & Tech*. 49:168-76.

618 Liu R, Lal R (2015) Potentials of engineered nanoparticles as fertilizers for increasing agronomic productions.
619 *Science of The Total Environment* 514: 131-139. doi: 10.1016/j.scitotenv.2015.01.104.

620 Luo H, Benner R, Long RA, Hu J (2009) Subcellular localization of marine bacterial alkaline phosphatases.
621 *PNAS* 106: 21249-21223.

622 Magid J, Tiessen H, Condron LM (1996) Humic substances in terrestrial ecosystems. In: A Piccolo (ed)
623 Dynamics of organic phosphorus in soils under natural and agricultural ecosystems. Elsevier Science,
624 Amsterdam.

625 Magnacca G, Allera A, Montoneri E, Celi L, Benito DE, Gagliardi LG, Gonzalez MC, Mártire DO, Carlos L
626 (2014) Novel Magnetite Nanoparticles Coated with Waste-Sourced Biobased Substances as
627 Sustainable and Renewable Adsorbing Materials. *ACS Sustainable Chemistry & Engineering* 2: 1518-
628 1524. doi: 10.1021/sc500213j.

629 McGill WB, Cole CV (1981) Comparative aspects of cycling of organic C, N, S and P through soil organic
630 matter. *Geoderma* 26: 267-286.

631 Mueller CW, Kölbl A, Hoeschen C, Hillion F, Heister K., Herrmann AM, Kögel-Knabner I (2012). Submicron
632 scale imaging of soil organic matter dynamics using NanoSIMS—from single particles to intact
633 aggregates. *Org. Geochem*. 42: 1476-1488.

634 Nannipieri P, Giagnoni L, Landi L, Renella G (2011) Role of Phosphatase Enzymes in Soil. In: E Bünemann, A
635 Oberson, E Frossard (eds) Phosphorus in Action: Biological Processes in Soil Phosphorus Cycling.
636 Springer Berlin Heidelberg, Berlin, Heidelberg.

637 Nash DM, Haygarth PM, Turner BL, Condon LM, McDowell RW, Richardson AE, Watkins M, Heaven MW
638 (2014) Using organic phosphorus to sustain pasture productivity: A perspective. *Geoderma* 221: 11-19.
639 doi: 10.1016/j.geoderma.2013.12.004.

640 Neal AL, Ahmad S, Gordon-Weeks R, Ton J (2012) Benzoxazinoids in root exudates of maize attract
641 *Pseudomonas putida* to the rhizosphere. *PLoS One* 7: e35498. doi: 10.1371/journal.pone.0035498.

642 Neal AL, Rossman M, Brearley C, Akkari E, Guyomar C, Clark IM, Allen E, Hirsch PR (2017) Land-use
643 influences phosphatase gene microdiversity. *Environ. Microbiol.* (in press doi:10.1111/1462-
644 2920.13778)

645 Negassa W, Leinweber P (2009) How does the Hedley sequential phosphorus fractionation reflect impacts of
646 land use and management on soil phosphorus: A review. *J Plant Nutr Soil Sci-Z Pflanzenernahr*
647 *Bodenkd* 172: 305-325. doi: 10.1002/jpln.200800223.

648 Nisticò R, Evon P, Labonne L, Vaca-Medina G, Montoneri E, Francavilla M, Vaca-Garcia C, Magnacca G,
649 Franzoso F, Negre M (2016) Extruded Poly(ethylene-co-vinyl alcohol) Composite Films Containing
650 Biopolymers Isolated from Municipal Biowaste. *ChemistrySelect* 1: 2354-2365. doi:
651 10.1002/slct.201600335.

652 Plassard C, Louche J, Ali MA, Duchemin M, Legname E, Cloutier-Hurteau B (2011) Diversity in phosphorus
653 mobilisation and uptake in ectomycorrhizal fungi. *Ann Forest Sci* 68: 33-43. doi: 10.1007/s13595-010-
654 0005-7.

655 Powers SM, Bruulsema TW, Burt TP, Chan NI, Elser JJ, Haygarth PM, Howden NJK, Jarvie HP, Lyu Y,
656 Peterson HM, Sharpley AN, Shen J, Worrall F, Zhang F (2016) Long-term accumulation and transport
657 of anthropogenic phosphorus in three river basins. *Nature Geosci* 9: 353-356. doi: 10.1038/ngeo2693

658 Ragot SA, Kertesz MA, Bünemann EK (2015) *phoD* Alkaline Phosphatase Gene Diversity in Soil. *Appl*
659 *Environ Microbiol* 81: 7281-7289. doi: 10.1128/aem.01823-15.

660 Ragot SA, Kertesz MA, Mészáros É, Frossard E, Bünemann EK. (2016) Soil *phoD* and *phoX* alkaline
661 phosphatase gene diversity responds to multiple environmental factors. *FEMS microbiology ecology*.
662 93:fiw212.

663 Redfield AC (1958) The biological control of chemical factors in the environment *American Scientist* 46: 230A-
664 221.

665 Richardson AE, Hocking PJ, Simpson RJ, George TS (2009) Plant mechanisms to optimise access to soil
666 phosphorus. *Crop Past. Sci.* 60: 124-143.

667 Richardson AE, Lynch JP, Ryan PR, Delhaize E, Smith FA, Smith SE, Harvey PR, Ryan MH, Veneklaas EJ,
668 Lambers H, Oberson A, Culvenor RA, Simpson RJ (2011) Plant and microbial strategies to improve
669 the phosphorus efficiency of agriculture. *Plant Soil* 349: 121-156. doi: 10.1007/s11104-011-0950-4.

670 Rosemarin A, Ekane N (2015) The governance gap surrounding phosphorus. *Nutri Cycl Agroecosys*: 1-15. doi:
671 10.1007/s10705-015-9747-9.

672 Rosling A, Midgley MG, Cheeke T, Urbina H, Fransson P, Phillips RP (2016) Phosphorus cycling in deciduous
673 forest soil differs between stands dominated by ecto- and arbuscular mycorrhizal trees. *New Phytol*
674 209: 1184-1195. doi: 10.1111/nph.13720.

675 Runge-Metzger A (1995) Closing the cycle: obstacles to efficient P management for improved global food
676 security. *Scope-Scientific Committee on Problems of the Environment International Council of*
677 *Scientific Unions* 54: 27-42.

678 Santos-Beneit F (2015) The Pho regulon: a huge regulatory network in bacteria. *Frontiers in Microbiology* 6.
679 doi: 10.3389/fmicb.2015.00402.

680 Schneider KD, Cade-Menun BJ, Lynch DH, Voroney RP (2016) Soil Phosphorus Forms from Organic and
681 Conventional Forage Fields. *Soil Sci Soc Am J* 80: 328-340. doi: 10.2136/sssaj2015.09.0340.

682 Sebastian M, Ammerman JW (2009) The alkaline phosphatase PhoX is more widely distributed in marine
683 bacteria than the classical PhoA. *ISME* 3: 563-572. doi: 10.1038/ismej.2009.10.

684 Secco D, Wang C, Shou H, Whelan J (2012) Phosphate homeostasis in the yeast *Saccharomyces cerevisiae*, the
685 key role of the SPX domain-containing proteins. *FEBS letters* 586: 289-295. doi:
686 10.1016/j.febslet.2012.01.036.

687 Sharma R, Bella RW, Wong MTF (2017) Dissolved reactive phosphorus played a limited role in phosphorus
688 transport via runoff, throughflow and leaching on contrasting cropping soils from southwest Australia.
689 *Sci. Tot. Env.* 577: 33-44.

690 Sharpley AN, Bergström L, Aronsson H, Bechmann M, Bolster CH, Börling K, Djodjic F, Jarvie HP,
691 Schoumans OF, Stamm C, Tonderski KS, Ulén B, Uusitalo R, Withers PJA (2015) Future agriculture

692 with minimized phosphorus losses to waters: Research needs and direction. *AMBIO* 44: 163-179. doi:
693 10.1007/s13280-014-0612-x.

694 Slazak A, Freese D, da Silva Matos E, Hüttl RF (2010) Soil organic phosphorus fraction in pine–oak forest
695 stands in Northeastern Germany. *Geoderma* 158: 156-162.

696 Spohn M, Kuzyakov Y (2013) Distribution of microbial- and root-derived phosphatase activities in the
697 rhizosphere depending on P availability and C allocation – Coupling soil zymography with ¹⁴C
698 imaging. *Soil Biol Biochem* 67: 106-113. doi: <http://dx.doi.org/10.1016/j.soilbio.2013.08.015>.

699 Stewart JWB, Tiessen H (1987) Dynamics of soil organic phosphorus. *Biogeochem* 4: 41-60. doi:
700 10.1007/bf02187361.

701 Stutter MI, Shand CA, George TS, Blackwell MSA, Bol R, MacKay RL, Richardson AE, Condon LM, Turner
702 BL, Haygarth PM (2012) Recovering Phosphorus from Soil: A Root Solution? *Environ Sci Technol* 46:
703 1977-1978. doi: 10.1021/es2044745.

704 Stutter MI, Shand CA, George TS, Blackwell MSA, Dixon L, Bol R, MacKay RL, Richardson AE, Condon
705 LM, Haygarth PM (2015) Land use and soil factors affecting accumulation of phosphorus species in
706 temperate soils. *Geoderma* 257–258: 29-39. doi: 10.1016/j.geoderma.2015.03.020.

707 Tamburini F, Pfahler V, von Sperber C, Frossard E, Bernasconi SM (2014) Oxygen Isotopes for Unraveling
708 Phosphorus Transformations in the Soil–Plant System: A Review. *Soil Sci Soc Am J* 78: 38-46. doi:
709 10.2136/sssaj2013.05.0186dgs.

710 Tan H, Barret M, Mooij MJ, Rice O, Morrissey JP, Dobson A, Griffiths B, O’Gara F (2013) Long-term
711 phosphorus fertilisation increased the diversity of the total bacterial community and the phoD
712 phosphorus mineraliser group in pasture soils. *Biol Fertil Soils* 49: 661-672. doi: 10.1007/s00374-012-
713 0755-5.

714 Tate KR, Salcedo I (1988) Phosphorus control of soil organic matter accumulation and cycling. *Biogeochem* 5:
715 99-107. doi: 10.1007/bf02180319.

716 Tipping E, Somerville CJ, Luster J (2016) The C:N:P:S stoichiometry of soil organic matter. *Biogeochem* 130:
717 117-131. doi: 10.1007/s10533-016-0247-z.

718 Tkacz A, Cheema J, Chandra G, Grant A, Poole PS (2015) Stability and succession of the rhizosphere
719 microbiota depends upon plant type and soil composition. *ISME J* 9: 2349-2359. doi:
720 10.1038/ismej.2015.41.

721 Toor GS, Condrón LM, Di HJ, Cameron KC, Cade-Menun BJ (2003) Characterization of organic phosphorus in
722 leachate from a grassland soil. *Soil Biol. Biochem.* 35:1317-23.

723 Trouillefou CM, Le Cadre E, Cacciaguerra T, Cunin F, Plassard C, Belamie E (2015) Protected activity of a
724 phytase immobilized in mesoporous silica with benefits to plant phosphorus nutrition. *J Sol-Gel Sci*
725 *Technol* 74: 55-65. doi: 10.1007/s10971-014-3577-0.

726 Turner BL, Cade-Menun BJ, Condrón LM, Newman S (2005) Extraction of soil organic phosphorus. *Talanta*
727 66: 294-306. doi: 10.1016/j.talanta.2004.11.012.

728 Turner BL, Cheesman AW, Condrón LM, Reitzel K, Richardson AE (2015) Introduction to the special issue:
729 Developments in soil organic phosphorus cycling in natural and agricultural ecosystems. *Geoderma*
730 257–258: 1-3. doi: <http://dx.doi.org/10.1016/j.geoderma.2015.06.008>. Turner BL, Frossard E, Baldwin
731 DS, editors. (2005) *Organic phosphorus in the environment*. CABI Pub.pp 377-380.

732 Uusitalo R, Turtola E, Puustinen M, Paasonen-Kivekas M, Uusi-Kamppa J (2003) Contribution of particulate
733 phosphorus to runoff phosphorus bioavailability. *J Environ Qual* 32: 2007-2016.

734 Vollmer-Sanders C, Allman A, Busdeker D, Moody LB, Stanley WG (2016) Building partnerships to scale up
735 conservation: 4R Nutrient Stewardship Certification Program in the Lake Erie watershed. *J Great*
736 *Lakes Res.* doi: <http://dx.doi.org/10.1016/j.jglr.2016.09.004>.

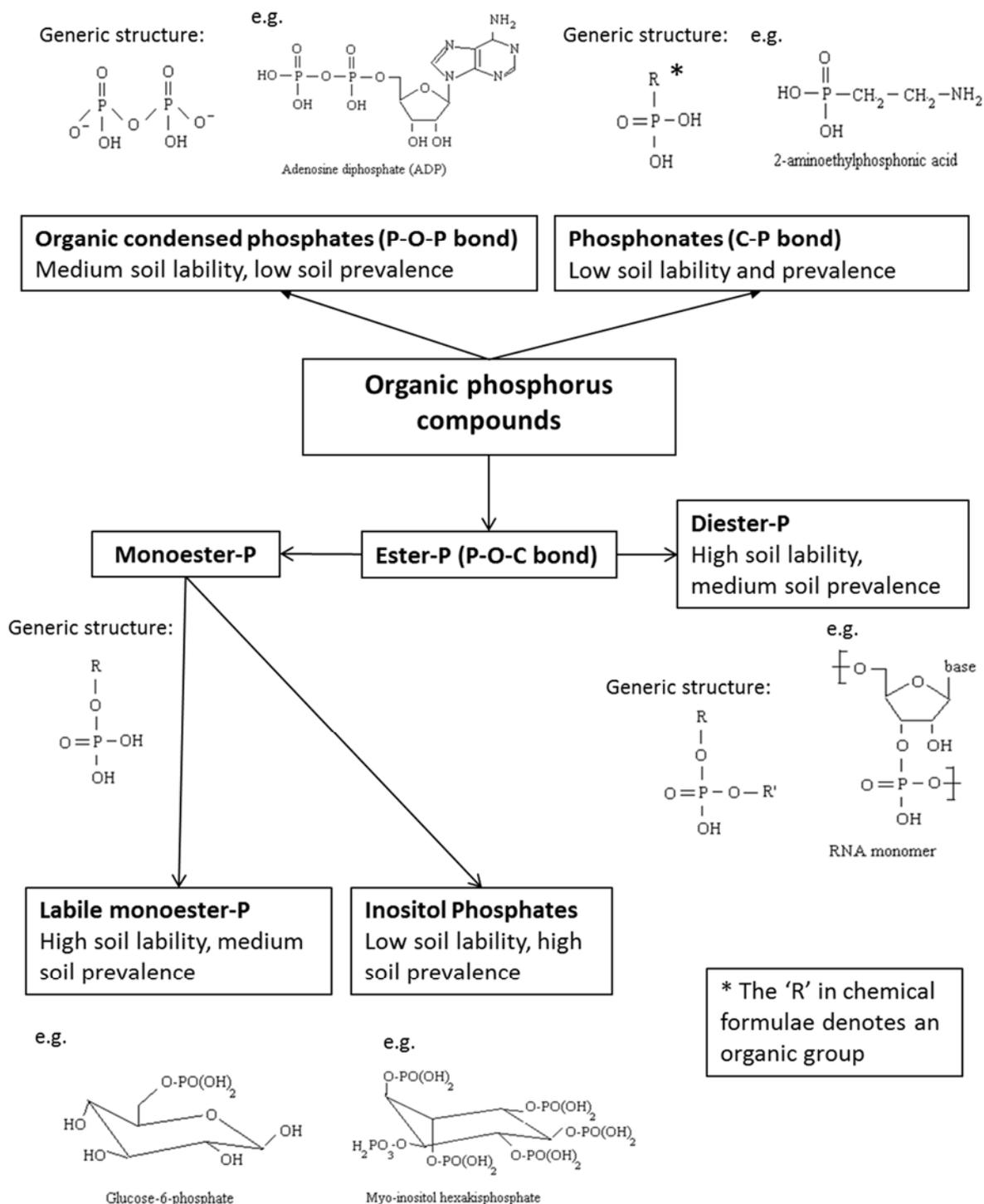
737 von Sperber C, Kries H, Tamburini F, Bernasconi SM, Frossard E (2014) The effect of phosphomonoesterases
738 on the oxygen isotope composition of phosphate. *Geochimica et Cosmochimica Acta* 125: 519-527.
739 doi: 10.1016/j.gca.2013.10.010.

740 Wieder WR, Grandy AS, Kallenbach CM, Taylor PG, Bonan GB (2015) Representing life in the Earth system
741 with soil microbial functional traits in the MIMICS model. *Geosci Model Dev* 8: 1789-1808. doi:
742 10.5194/gmd-8-1789-2015.

743 Withers PJA, Hartikainen H, Barberis E, Flynn NJ, Warren GP (2009) The effect of soil phosphorus on
744 particulate phosphorus in land runoff. *Euro J Soil Sci* 60: 994-1004. doi: 10.1111/j.1365-
745 2389.2009.01161.x.

746 Zaia FC, Gama-Rodrigues AC, Gama-Rodrigues EF, Moço MKS, Fontes AG, Machado RCR, Baligar VC
747 (2012) Carbon, nitrogen, organic phosphorus, microbial biomass and N mineralization in soils under
748 cacao agroforestry systems in Bahia, Brazil. *Agroforest Sys* 86: 197-212. doi: 10.1007/s10457-012-
749 9550-4.

750 Zhou Z, Hartmann M (2012) Recent Progress in Biocatalysis with Enzymes Immobilized on Mesoporous Hosts.
 751 Topics Catalysis 55: 1081-1100. doi: 10.1007/s11244-012-9905-0.
 752 Zimmerman AE, Martiny AC, Allison SD (2013) Microdiversity of extracellular enzyme genes among
 753 sequenced prokaryotic genomes. ISME 7: 1187-1199. doi: 10.1038/ismej.2012.176.



754

755 FIGURE 1. Organic phosphorus forms with generic and example structures and information on the relative lability
756 and prevalence in soil. (Adapted from Darch et al. (Darch et al. 2014))

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760 **Table Legend**

761

762 Table 1: Synthesis of expert opinions on the global issues associated with organic phosphorus, how the research
763 community can potentially contribute to solutions to such issues, and identification of opportunities for research
764 to allow this to happen.

765

What are the global issues associated with P _o ?	What are the methodological strengths and weaknesses?	What are benefits of understanding dynamics of P _o ?	What are the priorities for P _o research?	Opportunities in P _o research
<p>Food Security and agricultural sustainability P_o has a role as a source of P for agricultural crops</p>	<p>Strengths</p> <p>Strong collection of well-developed methods</p>	<p>Management of plant P nutrition</p> <p>Assessment of soil P availability</p>	<ul style="list-style-type: none"> • Use existing datasets more effectively • Avoid repeating experiments by being aware of past research • Better access to shared facilities • Training programmes in P_o related techniques and concepts • Interdisciplinary and long term research 	<p>General advances in the research model</p>
<p>Nutrient cycling in natural ecosystems P_o buffers ecosystem function with effects on ecosystem resilience and biodiversity</p>	<p>Wide range of techniques</p> <p>Capacity for multi-disciplinarity</p>	<p>Understanding biological system function</p> <p>Input into climate and biogeochemical models</p>	<ul style="list-style-type: none"> • Link operationally-defined pools with biological processes • Some standardisation of protocols • Development of in situ, non-destructive techniques for P_o • Develop a minimum dataset and an accessible database 	<p>Opportunities in organic phosphorus analytical methodologies</p>
<p>Renewable resources Use of wastes containing P_o as fertilisers to close the loop</p>	<p>Strong international networks</p> <p>Potential for commercialisation of techniques</p>	<p>Potential to close the P cycle</p> <p>Manage ecosystem services and resilience</p>	<ul style="list-style-type: none"> • Link the P_o cycle with other biogeochemical cycles • Optimise stoichiometry between P_o and other elements for system function • Integrate soil physics, chemistry and biology to understand P_o and how it fits with wider soil fertility 	<p>Opportunities from understanding stoichiometry – interactions with other element cycles</p>
<p>C storage in soils Utilisation of soil P_o may be counter to our need to store C in organic matter</p>	<p>Range of field based applications</p>	<p>Understand the role of soil biology – fungal vs bacterial dominated systems</p> <p>Assess stability of P forms in soil</p>	<ul style="list-style-type: none"> • Design tailored systems for specific managed environments that optimise use of P_o • Optimise P_o utilisation over loss • Improve soil P testing • Develop a P credits system • Utilise P_o more effectively by using what's in soil, what's added to soil and what's lost 	<p>Opportunities from understanding interactions with land management</p>
<p>Environmental pollution</p>	<p>Weaknesses</p>			

<p>Need to manage the balance of food security vs environmental P pollution</p> <p>Environmental change Warmer temperatures will shift the biogeochemical cycle of P_o</p> <p>Biogeochemical cycling from global to cellular scales P_o compounds are vital for cell function and are moved globally as part of biogeochemical cycles and in the food chain</p> <p>Geopolitical stability P_o as an alternative to mined P resources</p>	<p>'Snap-shot' rather than dynamic techniques</p> <p>Operational methodologies lack biological relevance</p> <p>Lack of standardisation and quality control</p> <p>Methodological limitations (matrix issues)</p> <p>Loss of training/education in soil science</p> <p>Lack of replication and appropriate statistical approaches</p> <p>Limited access to advanced techniques for all</p>	<p>Identify mechanisms from natural systems that can be applied in managed systems</p> <p>Separate plant and microbial contributions to soil functions</p> <p>Develop indicators for tipping points in ecosystem function – identify conditions of resistance, resilience and “points of no return”</p> <p>Allow scaling up in time and space through input to models</p> <p>Extend our understanding of global nutrient dynamics beyond what can be ascertained empirically</p>	<ul style="list-style-type: none"> • Understand which genes and transcripts control the microbial response to P_o • Understand microbial impacts on P_o cycles • Understand the P limits to plants and microbes • Produce a molecular toolkit for studying microbial structure and function 	<p>Opportunities from understanding Microbial Po: Function and dynamics</p>
			<ul style="list-style-type: none"> • Understand P_o interaction with natural and manmade nanoparticles • Assess the utility of nanoparticles to help manage the system 	<p>Opportunities from interactions with nanoparticles</p>
			<ul style="list-style-type: none"> • Model P dynamics in the environment • Develop conceptual models of cycling at a range of scales • Build empirical models using existing data • Produce a life cycle analysis of P_o 	<p>Opportunities to use modelling of Po in soil and ecosystems</p>
			<ul style="list-style-type: none"> • Promote discussion of P_o within the scientific community • Better communication with stakeholders and the public on the importance of P_o • Develop a central platform for knowledge exchange • Understand the needs and motivations of land managers and policy makers with respect to P_o • Emphasise educating the public in issues associated with P_o • Understand the socio-economic factors influencing P_o dynamics • Improve the translation of research in P_o to impactful outcomes 	<p>Opportunities to better communicate and translate research</p>