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Nitrate production beneath a High Arctic glacier, Svalbard

Wynn, P.M^{a*}, Hodson, A.J^a, Heaton, T.H.E^b, and Chenery, S.R.^b

^a Department of Geography, University of Sheffield, Winter St., Sheffield S10 2TN, UK ^b British Geological Survey. Keyworth, Nottingham NG12 5GG, UK

Abstract

8 Natural environmental isotopes of nitrate and ammonium are used in conjunction with major 9 ion chemistry and hydrological data to establish controls upon the biogeochemical cycling of 10 nitrogen beneath a High Arctic polythermal glacier (Midtre Lovénbreen). Here, high nitrate 11 concentrations in subglacial meltwaters suggest that the subglacial environment may be 12 furnishing nitrate in excess of that released from the snowpack and glacier ice. Isotopic values of $\delta^{18}O_{NO3}$ suggest the provenance of such excess nitrate to be microbial in origin and $\delta^{15}N_{NO3}$ 13 indicates the source nitrogen compounds to have high $\delta^{15}N$ values relative to supraglacial 14 15 runoff. We address the nitrification of supraglacial ammonium, the mineralization of organic 16 nitrogen and the oxidation of geologic ammonium as potential sources of this additional nitrate. Mass fluxes of N compounds in a subglacial river and their δ^{15} N ratios indicate that 17 18 the nitrification of supraglacial ammonium delivered to the glacier bed can account for much, 19 but not all, of the excess nitrate. The additional source most likely involves the mineralization of organic nitrogen, although δ^{15} N values in rock samples suggest that the dissolution of rock-20 21 derived ammonium cannot be discounted if large fractionation effects occur during 22 dissolution. Our results therefore agree with previous catchment scale mass balance studies at the site, which report a major internal loss of NH_4^+ from the snowpack following melt. 23 24 However, at the catchment scale, the NH_4^+ loss is greater than the excess of NO_3^- observed in 25 runoff, indicating that microbial assimilation of ammonia into organic matter in a range of other habitats is also likely. The identification of NH₄⁺ assimilation and nitrification further 26 highlights the non-conservative behaviour of nitrogen in glacial environments and testifies to 27 28 the importance of microbially-mediated reactions in the biogeochemical cycling of nitrogen in 29 an environment that has, until recently, been regarded as biologically inert. 30

^{*} Corresponding Author: School of Geography, Earth and Environmental Sciences, University of Birmingham, Edgbaston, Birmingham, B15 2TT. Tel: +44 121 414 5544; Fax: +44 121 414 5528; e-mail: <u>p.m.wynn@bham.ac.uk</u>

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Microbiological activity

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34 **1. Introduction**

35 Solute acquisition by glacial meltwaters takes place at the bed of glaciers during transit 36 through distributed and channelised drainage systems (eg. Raiswell, 1984; Tranter et al. 1993, 37 1996, 1997). The distributed system pertains to hydrological flowpaths conveying water 38 under high pressure with long residence times and high rock/water contact ratios. Channelised 39 drainage networks however follow discrete flowpaths, represent low pressure hydrological 40 systems and evacuate large volumes of water rapidly from beneath the glacier. The spatial and 41 temporal evolution between the two systems depends upon the flux of meltwater to the glacier 42 bed, whereby high inputs of supraglacial meltwater raise the basal water pressure and 43 encourage a spatial re-organisation of the drainage system such that the channelised system 44 evolves at the expense of the distributed configuration (e.g. Richards et al. 1996; Nienow et 45 al. 1998). Such hydrological forcing of subglacial drainage evolution at predominantly cold, 46 polythermal glaciers can be rapid and closely coupled to the dynamics of glacier movement 47 (Copland et al. 2003; Nuttall and Hodgkins, 2005; Rippin et al., 2005; Bingham et al. 2006). 48 A concomitant change in the hydrochemistry of subglacial runoff accompanies this transition 49 and can be manifest as a rapid switch in the chemical properties of the melt water (Copland et 50 al. 2003; Wynn et al. 2006). Due to extensive rock/water contact and weathering reactions, 51 high pressure distributed drainage configurations encourage high total dissolved ion loads and 52 low concentrations of atmospheric gases in solution. However, during the hydrological 53 evolution of the subglacial drainage system, the development of channelised flow paths 54 conveying large volumes of meltwater subjected to only limited rock/water contact means that 55 waters are more dilute and dissolved gases in solution exceed the capacity of oxygen 56 consuming reactions at the glacier bed (e.g. Tranter et al, 2002). However, since the presence 57 of microbiological activity within ice and subglacial sediments is now widely acknowledged 58 (Bhatia et al. 2006; Foght et al. 2004; Mader et al. 2006; Sharp et al. 1999, Skidmore et al. 59 2000; Tranter et al. 2005; Welker et al. 2002), researchers are increasingly aware that 60 acquisition of solute in sediments at distance from such channels may be effectively 61 decoupled from inorganic controls of gas supply and rock/water contact ratios. Atmospheric 62 gas supplies thus become depleted in the distributed drainage system, and so the use of 63 alternative oxidising agents by microbial populations is required to drive further solute

acquisition under anoxic conditions (Bottrell and Tranter, 2002; Wadham et al. 2004; Wynn
et al. 2006). Despite this increased understanding, the degree to which such biotic solute
acquisition is governed by the evolution of the subglacial hydrological system remains largely
unknown (Tranter et al. 2005; Wynn et al. 2006; Hodson et al. In Press).

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69 The NO₃⁻ ion represents one of the more readily available electron donors under reducing 70 conditions, and so should be highly sensitive to the redox evolution of the subglacial 71 hydrological system (Tranter et al. 1994; Wynn et al. 2006; Hodson et al. In Press). Thus, 72 during the early stages of summer ablation, subglacial meltwater in the distributed drainage 73 configuration includes pockets of local anoxia that are characterised by nitrogen isotopic 74 values that are diagnostic of microbial denitrification (Wynn et al. 2006). However, later in 75 the summer, when most meltwaters are conveyed through the aerated low pressure 76 channelised system, concentrations of nitrate and NO_3^{-1}/CI^{-1} ratios in the subglacial meltwaters 77 increase significantly, implying the presence of an additional source of nitrate to the 78 subglacial drainage system. Hodson et al. (2005a) and Hodson (2006), show that some form 79 of NO₃ production must take place, because annual NO₃ yields exceed inputs in a number of 80 glacial environments, (including Midtre Lovénbreen). Such "excess NO₃" is also implicit in 81 meltwater hydrochemical studies of temperate glacier basins in the European Alps (Tockner et al. 2002) and cold-based glaciers of the maritime Antarctic (Caulkett and Ellis-Evans, 82 83 1997; Hodson, 2006) although its precise source has yet to be identified.

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85 An appreciation of the manner in which glaciers regulate the biogeochemical cycling of 86 nutrients and thereby impact upon the productivity of neighbouring freshwater and marine 87 ecosystems demands a better understanding of the non-conservative behaviour of nitrogen 88 and other nutrients in glacial environments on a sub-annual basis. Here, we build upon earlier 89 annual mass balance studies of nitrogen and trace the provenance and dynamics of 'excess' 90 NO₃ production in the subglacial environment of a high Arctic glacier at a seasonal time scale. In so doing, major ion chemistry (NO_3^- , NH_4^+ and CI^-) and environmental isotopes of 91 $\delta^{15}N_{NO3}$, $\delta^{18}O_{NO3}$ and $\delta^{15}N_{NH4}$ in pre-melt snow, meltwater, organic matter and whole rock 92 93 samples are presented from Midtre Lovénbreen, Svalbard.

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95 2. Methodology

96 2.1. Field site and sampling

97 Midtre Lovénbreen is a high Arctic polythermal glacier situated on the Brøggerhalvøya 98 peninsula in North West Spitsbergen (78.53°N and 12.04°E). The geology of the peninsula 99 includes basement rocks belonging to the Lower, Middle and Upper Proterozoic that are 100 situated beneath the glacier and predominantly composed of phyllites and beds of quartzite 101 (Hjelle, 1993). To the south of the phyllites are more strongly metamorphosed rocks that 102 include mica schists and beds of marble beneath the accumulation area of the glacier (Hjelle, 103 1993).

104 The glacier is up to 180m thick and two thirds of the ice is at the pressure melting point, 105 thereby supporting an extensive subglacial drainage system (Rippin et al. 2003). Bulk 106 meltwater runoff is conveyed towards the adjacent fjord of Kongsfjorden via two main 107 proglacial melt channels draining the lateral margins of the glacier (Hodson et al. 2005b) 108 (Figure 1). Meltwater accesses the bed of the glacier via moulins in the accumulation area, 109 later emerging as a pressurised subglacial upwelling (MLSG) at the terminus of the glacier 110 (Irvine-Fynn et al. 2005) (Figure 1), the dynamics of which are described by Rippin et al. 111 (2003) and Hodson et al. (2005a). Since subglacial meltwaters are characteristically sub-oxic 112 early in the ablation season, but well aerated thereafter, we have separated our observation 113 periods accordingly (see Wynn et al. 2006). Hereafter, early (sub-oxic) and later (oxic) 114 subglacial runoff will be referred to as the "initial" and "principal" runoff phases respectively, 115 reflecting the relative importance of the runoff fluxes associated with each period. During 116 summer 2002, samples of subglacial runoff were collected directly from the upwelling, 117 although during summer 2003, the upwelling changed location during the melt season causing 118 a small supraglacial stream to enter the subglacial runoff before sampling was possible, thus 119 confounding direct geochemical characterisation (Figure 1 and Table 1). Major ion 120 characterisation and data sourced from gauging stations installed on the two major bulk 121 meltwater channels draining the proglacial zone during 2002 (namely MLE and MLW, Table 122 1, Figure 1) are presented and discussed with regard to the impact subglacial processes may 123 have on the total runoff and solute budgets from the entire catchment. Meltwater samples 124 have been collected at these stream sites for major ion characterisation as part of an ongoing 125 monitoring programme that began in 1997 (Hodson et al. 2000; 2004; 2005a). On the glacier 126 surface, snowpack samples were collected using a depth-integrating 0.5 litre PVC tube 127 (Hodson et al. 2005a). Lysimeters of a similar design to those described in Hodson (2006), 128 were inserted into the base of the snowpack during summer 2003 to document the

hydrochemistry during snowpack ablation and major ion samples collected from one of the
main supraglacial streams during 2003 (Figure 1) enabled solute characterisation of bulk
supraglacial discharge.

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133 All meltwater samples for major ion composition were filtered immediately in the field using 134 a handheld Nalgene vacuum unit and 0.45µm cellulose nitrate filters to prevent further 135 reaction with suspended sediment. Filters were pre-rinsed in the field using aliquots of sample 136 water and field blanks have failed to indicate any potential contamination. Samples were 137 stored airtight in pre-rinsed 60ml polyethylene Nalgene bottles (pre-rinsed with filtered 138 sample water) and refrigerated to await further analysis. Snow samples were treated in the 139 same manner as runoff samples after being melted in a 25 °C water bath (see Hodson et al. 140 2005a).

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142 During 2002 and 2003 our geochemical sampling campaign was augmented by the determination of stable isotope ratios of ${}^{15}N/{}^{14}N$ and ${}^{18}O/{}^{16}O$ in NO₃⁻ and ${}^{15}N/{}^{14}N$ in NH₄⁺. 143 144 For the snowpack this involved collecting a c. 60 kg section of snow down to the glacier ice surface, in polyethylene bags, and for aqueous samples the collection of 20-40 L samples in 145 146 polyethylene jerry cans. Details of sample processing are reported elsewhere (Wynn, 2004; Heaton et al. 2004; Wynn et al. 2006). The ${}^{15}N/{}^{14}N$ ratios of glacial till and fresh bedrock 147 were determined on material from the glacier forefield. In the case of the glacial till, samples 148 149 were collected from the immediate ice margin, representing young, former subglacial tills that 150 have been exposed by the glacier's recent retreat. The rock samples included metamorphic 151 basement rocks from the glacier's upper accumulation area (phyllite, schist, sandstones and 152 quartzite) and younger sequences (chert) outcropping in the proximal parts of the glacier 153 forefield.

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Organic matter present within small melt pools known as cryoconite holes on the glacier surface was collected during summer 2002 and 2003 from three locations on the glacier surface and air-dried prior to analysis back in the UK.

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159 2.2. Laboratory analysis

Anions of chloride and nitrate were determined using a Dionex DX100 ion chromatograph and soluble ammonium was analysed using a FOSS-Tecator FIAstar 5000 flow injection analyser. Based on repeat analysis of reference standard materials of comparable concentration to the samples being analysed, precision was calculated as 3.57, 1.16 and 2.54% RSD (relative standard deviation) for chloride, nitrate and ammonium respectively.

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Snow, supraglacial and subglacial meltwaters collected for isotopic analysis were gravity-fed through cation and anion exchange resins, with nitrate processed to silver nitrate (Chang et al. 1999; Hwang et al. 1999; Silva et al. 2000; Heaton et al. 2004) and ammonium processed to ammonium sulphate (Sigman et al. 1997; Heaton, 2001).

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171 2.3. Mass spectrometric analysis

172 Product silver nitrate, ammonium sulphate and cryoconite organic matter was analysed using 173 ThermoFinnigan elemental analysers linked to a Delta + XL continuous flow mass 174 spectrometer. Reductive pyrolysis of silver nitrate at 1400°C yielded CO and N₂ for determination of ¹⁸O/¹⁶O and N/O ratios, and oxidative combustion of silver nitrate, 175 ammonium sulphate or organic matter at 900°C yielded N2 and CO2 for determination of 176 ¹⁵N/¹⁴N and C/N ratios. Yield of N₂ from organic matter samples was determined by 177 178 comparison of sample peak area with those from known weights of acetanilide. Ratios were converted to δ^{18} O values versus VSMOW: 179

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181
$$\delta^{18}O_{\text{sample}}$$
, in ‰ = {[(¹⁸O/¹⁶O)_{sample} / (¹⁸O/¹⁶O)_{VSMOW})] -1} x 1000 1)

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183 and $\delta^{15}N$ values versus atmospheric N₂:

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$$\delta^{15}N_{\text{sample}}$$
, in ‰ = {[(¹⁵N/¹⁴N)_{sample} / (¹⁵N/¹⁴N)_{atmos})] -1} x 1000 2)

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by comparison with within-run laboratory standards calibrated against IAEA-N3 (δ^{18} O value = +25.6 ‰ versus VSMOW (IAEA, 2004)), or against IAEA-N1 (δ^{15} N value = +0.4 ‰ versus atmos. N₂ (IAEA, 2004)). Precision on replicates of within-run laboratory standards was typically better than ±0.7‰ for δ^{18} O and ±0.3‰ for δ^{15} N (1 SD).

192 2.4. Correction of $\delta^{18}O_{NO3}$ for organic matter

If δ^{15} N and δ^{18} O values of nitrate are to be used as indicators of provenance, the silver nitrate 193 samples must be pure, i.e. free from any potential N- or O-bearing contaminants (Kendal, 194 195 1998; Haberhauer and Blochberger, 1999; Heaton et al. 2004). We found this to be a major problem in the case of contamination derived from dissolved organics in low ionic strength 196 197 samples from the glacier surface during the first field campaign (2002). The amount of contaminant organic matter in the final silver nitrate was reduced, but not eliminated by use of 198 ISOLUTE[®] Env⁺ resin during sample preparation. Analysis of this organic matter, eluted 199 from the Env⁺ resins with methanol and dichloromethane, failed to detect significant amounts 200 of nitrogen, but indicated large proportions of oxygen with average δ^{18} O values of +22‰ (1 201 SD = 2.4, n = 16). During the 2003 field campaign, Amberlite[®] XAD-7 and ISOLUTE[®] Env⁺ 202 resins were placed upstream of the cation and anion exchange resins during sample 203 204 concentration, thereby screening the ion exchange resins from extensive organic matter 205 contamination.

The fractional contribution of organic contaminant oxygen to the total oxygen in an impure silver nitrate sample, f_{organic} , may be calculated from the sample's measured atomic N/O ratio ((N/O)_{measured}), assuming pure nitrate has a ratio of 0.33 and the organic contaminant a ratio of 0:

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$$f_{\text{organic}} = 1 - [(\text{N/O})_{\text{measured}}/0.33]$$
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213 If f_{organic} has a δ^{18} O value of +22 ‰, then the isotopic composition of the pure nitrate, δ^{18} O_{NO3}, 214 may be calculated from that of the measured impure silver nitrate sample, δ^{18} O_{measured}, from: 215

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$$\delta^{18}O_{NO3} = \left(\delta^{18}O_{\text{measured}} - f_{\text{organic}} \cdot 22\right) / \left(1 - f_{\text{organic}}\right)$$
(4)

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All δ^{18} O-nitrate signatures quoted in the remainder of this paper represent values corrected for organic oxygen in the above manner. Obviously we cannot be totally certain that the δ^{18} O value of the organic matter recovered from the Env⁺ resin is identical to the value for the organic matter contaminating the sample. However, the values we measured are within the defined range for organic oxygen and we believe that any differences which do occur will besmall enough not to have a significant effect on the correction procedure.

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225 2.5. Geologic nitrogen analysis

226 The term 'geologic nitrogen' is taken to represent nitrogen located within the surrounding 227 bedrock (Holloway et al. 1998; 1999; Holloway and Dahlgren, 2002). In sedimentary rocks 228 such 'geologic nitrogen' is derived predominantly from decomposed organic matter which 229 thermally degrades during diagenesis and substitutes for potassium in minerals such as illite, 230 muscovite, biotite, and feldspars, forming fixed ammonium in metamorphic and igneous 231 rocks (Mingram and Brauer, 1998). The total nitrogen content of whole rock samples was 232 therefore determined using a Hydrofluoric acid / sulphuric acid digest (Honma and Itihara, 233 1981; Haendel et al. 1986). Precise experimental procedures followed were adapted from 234 Bradley (1992) and resultant ammonium/sulphuric acid solution was processed to ammonium sulphate by ammonium diffusion (Sigman et al. 1997; Heaton, 2001) and analysed for $\delta^{15}N$. 235 236 Measurement precision based on replicates of standards was typically better than $\pm 0.3\%$ for δ^{15} N (1 S.D) whilst experimental precision based on the repeat analysis of rock samples was 237 238 <2% (1 S.D). Yield of N₂ was determined by comparison of sample peak area with those of 239 known weights of acetanilide and blank contamination from the combined digest and 240 ammonium diffusion procedure was calculated as $2.5\mu g$ N per analysis (n=12, 1 S.D = 0.75).

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3. Results

Figure 2 depicts the bulk hydrochemical outputs from the glacier catchment in major proglacial streams (MLE and MLW) and the subglacial runoff (MLSG) during the 2002 observation period. Time series of discharge, Cl⁻ and NO₃⁻ show the typical early season development of the hydrological system from a period dominated by snowmelt to one of icemelt, supplemented by the release of subglacial water. Data from 2003 are not shown because the position of the subglacial upwelling changed during the season, leading to complex time series.

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Table 2 shows summary statistics of the ionic and isotopic composition of NO_3^- and NH_4^+ in pre-melt snow, snowmelt, supraglacial runoff and subglacial runoff during initial and principal flow phases. The sharp boundary between the initial and principal flow phases occurred just prior to sample collection on DOYs 184 and 193 during 2002 and 2003 respectively and is described in detail by Wynn et al. (2006).

During 2002, average NO₃⁻ concentrations in pre-melt snow, were 1.6 μ M. Concentrations of NO₃⁻ in the subglacial waters of the principal runoff phase increased markedly to ca. 4 μ M and represent concentrations much greater than those observed during the initial runoff phase (average 1.12 μ M). During 2003, a strong elution phase produced very high average NO₃⁻ concentrations in snowmelt relative to the pre-melt snowpack. However, concentrations of NO₃⁻ were still greater in subglacial runoff of the principal runoff phase than the initial runoff phase, although the effect was smaller than in 2002.

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Average NH4⁺ concentrations were greatest in the pre-melt snowpack and snowmelt, 265 266 especially in 2003, (5.1 and 4.6 µM respectively), but lower in supraglacial streams (2.0 µM 267 in 2003), and subglacial runoff (0.17 μ M at first, then 0.13 μ M). In direct contrast to NO₃, average NH₄⁺ concentrations were lowest in subglacial runoff during the principal runoff 268 phase (Table 2). Table 2 also shows NO₃⁻/Cl⁻ ratios for all sample types and mean δ^{15} N and 269 δ^{18} O values for NO₃⁻ in pre-melt snow, supraglacial runoff and subglacial meltwater. The 270 271 NO₃⁻/Cl⁻ ratios were 0.015 in the pre-melt snowpack during 2002, whilst subglacial waters 272 showed average ratios that were lower during the initial flow phase (ca. 0.009), and markedly 273 higher during the principal runoff phase (0.038). During 2003, NO₃⁻/Cl⁻ ratios in the 274 subglacial runoff during the initial runoff phase ($NO_3^{-}/Cl^{-} = 0.028$) were similar to those 275 reported in supraglacial samples, although higher ratios did appear during the principal runoff 276 phase (NO₃/Cl⁻ = 0.056). Therefore NO₃/Cl⁻ ratios in subglacial runoff show a clear tendency 277 to increase across the boundary of the initial and principal flow phases.

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The isotope values in Table 2 and depicted in Figures 3 and 4 show significant differences between sample types. Nitrate in pre-melt snow has low δ^{15} N values (-9.9 ‰ and -9.8 ‰ for summer 2002 and 2003 respectively) and high δ^{18} O values (+57 ‰ to +72 ‰); the subglacial waters of the principal runoff phase have higher δ^{15} N (-5.5 ‰ to -4.5 ‰) and lower δ^{18} O (+20 ‰), whilst subglacial waters during the initial flow phase have the highest δ^{15} N values (+2.3 ‰ to +4.0 ‰) (Figures 3 and 4). For ammonium there was no clear distinction between different samples, with all δ^{15} N values in the range of -6 ‰ to -2 ‰.

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287 Geologic nitrogen concentrations and δ^{15} N signatures for rocks collected from the catchment 288 are given in Table 3. Concentrations of nitrogen were greatest in the phyllite, a major rock type in the subglacial system, and pyritic chert, which lies amongst the chert sequences beneath the lower ablation area of the glacier and its forefield. Phyllite and chert also demonstrated the highest organic carbon contents (0.11 to 0.29 %). With the exception of the low δ^{15} N value for the pyritic chert, which is a minor component of the chert, all other rock types had δ^{15} N in the range +4.8 ‰ to +7.7 ‰.

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Table 4 shows the δ^{15} N signature, N content and C/N ratio of the cryoconite organic matter sampled during the two field campaigns. Nitrogen contents are at least an order of magnitude greater than those recovered in the rock samples owing to the abundance of micro-algae, bacteria, viruses and a number of other microorganisms in these active microbial habitats (Säwström et al. 2002).

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301 4. Discussion

302 *4.1. Hydrochemical regime*

303 The hydrochemical regime of Midtre Lovénbreen during the 2002 summer melt season is 304 depicted in Figure 2. A steady increase in discharge in both the bulk meltwater proglacial 305 streams began when the glacier was completely snow covered on DOY 168. At MLE, 306 discharge continued to rise after DOY 182 due to the subglacial outburst, whilst discharge 307 driven principally by icemelt stabilised at MLW. By the end of the observation period shown 308 in Figure 2, icemelt was dominating runoff production across the entire glacier, although 309 snowmelt continued to emerge from the subglacial upwelling due to runoff from the 310 accumulation area entering the subglacial drainage system through stable crevasses and 311 moulins at approximately 400m altitude (Irvine-Fynn et al. 2005). The persistent input of 312 snowmelt is indicated by the high Cl⁻ in waters marked MLSG in Figure 2, whilst more dilute 313 Cl⁻ concentrations rapidly appear at MLE and MLW when icemelt dominates runoff delivery 314 to other flowpaths (i.e. supraglacial streams and lateral channels: cf Tranter et al. 1996). The 315 decline in Cl⁻ concentrations at MLE and MLW therefore reflects this dilution. However, they 316 also reflect the elution mechanism that removes solute rapidly from the snowpack, which 317 explains why Cl⁻ concentrations also gradually decline in the subglacial runoff (Tranter et al. 318 1996; Wadham et al. 1998). This elution process also appears to have governed changes in 319 NO_3^- in the proglacial rivers, because these too show a significant decline prior to the 320 emergence of subglacial runoff. Concentrations of NO₃⁻ then increase disproportionately 321 compared to Cl⁻ in the MLE stream two days after the emergence of the subglacial runoff.

The two day delay in increasing NO_3^- concentrations was likely caused by the displacement of "old" subglacial water during the initial flow phase. This component of the subglacial runoff was depleted in both oxygen and NO_3^- prior to the transition to the principal runoff phase (Wynn et al. 2006).

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327 Later in the principal runoff phase, the subglacial drainage system developed an efficient, 328 channelised configuration that conveyed runoff with residence times of just 2-4 hours 329 following descent into moulins in the uppermost part of the glacier (Irvine-Fynn et al. 2005). 330 During this phase, runoff via the subglacial upwelling increased to ca. 25 - 33 % of the total 331 catchment runoff (Hodson et al. 2005b). Intuitively, one would therefore expect the 332 proportion of atmospheric-derived NO_3^- (i.e. snow pack NO_3^-) transported by the subglacial 333 river to dominate during this period. However, the following characteristics confound this 334 interpretation and require an alternative explanation:

- 335 1) High NO₃⁻/Cl⁻ ratios and NO₃⁻ concentrations develop in subglacial runoff relative to
 336 all other streams flowing at the same time and to subglacial runoff during the initial
 337 flow phase earlier in the summer (Figure 2, Table 3);
- 338 2) At the same time, the subglacial runoff nitrate displays higher δ^{15} N values and lower 339 δ^{18} O values than the snowpack and supraglacial streams.
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Waters from the initial flow phase, when subglacial runoff is dominated via delayed flow pathways, have lower NO₃⁻ concentrations and NO₃⁻/Cl⁻ ratios due to denitrification (Wynn et al. 2006). Thus their mixing with surface-derived meltwaters would reduce the NO₃⁻ levels of subglacial runoff relative to surface melt waters – which is opposite to the observed pattern (Table 2). An alternative source of NO₃⁻, with higher δ^{15} N and lower δ^{18} O values than those of the surface runoff, must be added to these supraglacial meltwaters during their passage through the glacier.

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349 4.2. Nitrification as a mechanism for additional NO₃⁻ production

Nitrification has been invoked as a source of NO_3^- in a number of glacial and periglacial environments without actually being observed directly. For example, very significant excess NO_3^- production in two cold-based glacier basins has been proposed by Hodson et al. (2005a) and Hodson (2006). Further, studies of talus waters within rock glaciers have identified high NO₃⁻ waters that are also thought to betray bacterial activity and nitrification (Bieber et al. 1998). However, two studies have failed to stimulate nitrification in supraglacial snowpacks amended with NH_4^+ (Williams et al. 1996; Wynn, Unpublished Data), suggesting that the process takes place in alternative environments. The possibility that nitrification takes place at the bed of Midtre Lovénbreen is therefore considered below using $\delta^{18}O_{NO3}$. data from 2003.

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The microbial nitrification of ammonia utilises both H₂O and O₂ as the source of oxygen during the production of NO₃⁻. On the basis that two thirds of the oxygen molecules are sourced from H₂O and the remainder obtained from atmospheric O₂ (Kumar et al. 1983; Anderson and Hooper, 1983), it has been proposed that the expected δ^{18} O value for microbially-produced nitrate can be calculated (Equation 5) (eg. Amberger and Schmidt, 1987; Kendall, 1998; Mayer et al. 2001).

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$$\delta^{18}O_{NO3} = (0.33 \times \delta^{18}O_{Air}) + (0.66 \times \delta^{18}O_{water})$$
 (5)

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As the average $\delta^{18}O_{water}$ value of the subglacial discharge during the principal flow phase was 369 -12.3‰ (Table 2), and assuming $\delta^{18}O_{Air} = +23.7$ ‰ (value for atmospheric O₂; Horibe, et al. 370 1973), the theoretically expected $\delta^{18}O_{NO3}$ value for microbially produced nitrate is -0.3 ‰. 371 Taking an average $\delta^{18}O_{NO3}$ value for snow to be +64 ‰, a mixture of 68 % microbial nitrate 372 plus 32 % snowmelt nitrate would produce the $\delta^{18}O_{NO3}$ value of +20.3 %, similar to the value 373 374 for NO₃⁻ observed in the 2003 principal subglacial runoff phase (20.3 $\% \pm 6.1 \%$ (1 S.D)). As 375 there is no isotopic evidence for the presence of microbially mediated nitrate in the lysimeter melt waters and supraglacial streams (average $\delta^{18}O_{NO3} = +63.8$ %), these data therefore imply 376 377 that microbial nitrification must be occurring within or at the bed of the glacier, accounting 378 for much of the nitrate present in subglacial runoff during the principal runoff phase. Using 379 these same proportions for nitrate from snow and microbial sources, we can then estimate the $\delta^{15}N_{NO3}$ value of the microbial nitrate. Thus, if the subglacial nitrate outflow during the 380 principal runoff phase, ($\delta^{15}N_{NO3}$ value = -5.5 ‰ in 2003: Table 2) was made up of 32 % 381 snowmelt nitrate with a $\delta^{15}N_{NO3}$ value between -9.8 ‰ and -7.8 ‰ (range for snow and 382 supraglacial waters), then the 68 % of nitrate sourced from microbial activity in the 2003 383 subglacial runoff would have had a $\delta^{15}N_{NO3}$ value of -4.4 % to -3.5 %. 384

- 387 Given the above, the following potential sources of nitrogen might be driving a nitrification388 process:
- 1) NH₄⁺ from snowpack and glacier ice;

390 2) Rock-derived NH_4^+ liberated following dissolution reactions; and

391 3) Biologically-derived NH_4^+ produced following mineralisation of organic N.

392

Given that the NH_4^+ abundance in this glacial environment is low (Table 2), a negligible 393 394 fractionation effect is likely during the nitrification process (Fogel and Ciffuentes, 1993; Kendall, 1998; Heaton, 1986). We would therefore expect the $\delta^{15}N_{NH4+}$ value of the 395 ammonium to be very similar to the average product $\delta^{15}N_{NO3-}$ of ca. –4.4 ‰ to –3.5 ‰. 396 Snowpack NH₄⁺ has already been constrained by our measurements ($\delta^{15}N_{NH4+}$ of -1.7 % + 1.6 397 398 ‰ in 2002 to -2.8 ‰ in 2003: Table 2) and appears slightly higher than what is required if the NO_3^- were derived from nitrification of this source. Bulk snowmelt $\delta^{15}N_{NH4}$ values sampled 399 from the lysimeters were lower than the snowpack composition ($\delta^{15}N_{NH4+}$ of -5.2 % during 400 summer 2003) and thus closer to the principal phase subglacial runoff $\delta^{15}N_{NO3-}$ and $\delta^{15}N_{NH4+}$ 401 values observed during summer 2003. The difference between the high parent snowpack 402 $\delta^{15}N_{NH4+}$ value and the low lysimeter and supraglacial stream runoff values is unclear. 403 404 However, it is also uncertain on account of just one snowpack sample being available for 405 2003. We therefore believe that the larger data sets used to constrain the isotopic composition 406 of snowmelt in the lysimeters (n = 8) and mixtures of snowmelt and supraglacial streams (n = 8)407 6) provide the best indication of surface inputs to the subglacial drainage system during the 408 principal runoff phase.

409

In contrast to the supraglacial samples described above, rock δ^{15} N values were far higher than the snowpack and stream samples, being +7.2 ‰ for phyllite, the most dominant N-containing rock type beneath the glacier, and +7.7 ‰ for subglacial till. It therefore appears that the nitrification of ammonia within crushed rock and glacial till beneath the glacier is unlikely to furnish a product δ^{15} N_{NO3}. composition of ca. -4.4 ‰ to -3.5 ‰ unless there are very significant fractionation effects during the mineral dissolution process. Presently there are no data with which to assess the magnitude of such effects.

417 During the overall conversion of organic nitrogen to nitrate, mineralization represents the rate

- 418 determining step when the size of the ammonium substrate pool is small (Equation 6)
- 419 (Heaton, 1986).

421	Organic-N \rightarrow NH ₄ ⁺	$\rightarrow NO_2^- \rightarrow NO_3^-$	6	5)
422	mineralisation	nitrification		

423

424 Under these conditions, we would again expect the nitrate formed from the mineralization and nitrification of organic N to have a δ^{15} N-value very similar to that of the source organic 425 matter. Organic material lies upon the glacier surface in cryoconite holes, characterised by an 426 427 organic N content of 1 to 2 mg/g and C/N ratios (10.6 to 11.7) typical of microbial matter 428 (Takeuchi et al, 2002). Importantly, since cryoconite holes cover up to 6% of the glacier 429 surface (Hodson et al. 2005a) they offer much potential for ammonia assimilation into the organic phase and subsequent mineralisation. Further, the ¹⁵N values observed for organic 430 matter (-3 to -5 ‰) (Table 4) lie closer to the inferred product $\delta^{15}N_{NO3}$ range of -4.4 ‰ to -431 432 3.5 % than any of the other potential sources (ie other pathways of nitrification from surface 433 melt and rocks). The mineralization of surface derived microbial organic matter transported to the glacier bed may thus provide a reliable source of NH_4^+ for nitrification in addition to the 434 NH_4^+ contained within snow melt and supraglacial runoff. However, since their isotopic 435 436 compositions are very similar, no further distinctions can be made using isotopic data and we 437 instead use time series of NO₃⁻ production inferred from our ionic data.

438

439 4.4. Temporal dynamics of NO_3^- production and NH_4^+ loss

440 Our observations suggest that nitrification of NH_4^+ in the glacial catchment is significant 441 following the onset of snowmelt, a finding that is commensurate with the ammonia loss 442 identified in catchment-scale annual mass balances presented by Hodson et al. (2005a) and 443 Wynn (2004). Here we use the 2002 major ion data set to establish the seasonal (sub-annual) 444 dynamics of NO_3^- production and NH_4^+ loss in proglacial streamflow. The terms $excess}NO_3^-$ 445 and $deficitNH_4^+$ are defined in the following manner:

446

447
$$excessNO_3^- = totalNO_3^- - (0.021Cl^-)$$
 7)

448
$$^{\text{deficit}}NH_4^+ = ^{\text{total}}NH_4^+ - (0.014Cl^-)$$
 8)

449

450 The constants 0.021 and 0.014 represent the average NO_3^-/Cl^- and NH_4^+/Cl^- ratios respectively

451 (in μ M) in 2002 lysimeter melt waters, and thus enable removal of snowmelt NO₃⁻ and NH₄⁺

452 under the assumption that Cl⁻ is a conservative tracer of snowmelt. We used the lysimeter

453 data, rather than bulk snowpack data, because the number of pre-melt snow samples was too

- 454 small (see Table 2) and because the ratios could then be estimated using a wide range of
- solute concentrations similar to those observed in the streams. Our ratios were not influenced
- 456 by preferential elution effects, because time series of NO_3^{-}/Cl^{-} ratios in supraglacial streams
- 457 do not indicate that the process occurs on the glacier surface.
- 458

Figure 5 shows time series of the $^{\text{deficit}}NH_4^+$ and $^{\text{excess}}NO_3^-$ at MLE, MLW and in the initial and 459 460 principal phases of subglacial runoff during 2002. Concentrations of dissolved Si are also 461 shown to indicate the presence of waters from high rock-water contact environments (no Si 462 could be detected in snowmelt). Prior to DOY 173 streamflow was dominated by runoff from the glacier forefield and margins. During this time ^{excess}NO₃⁻ was negative at MLW and 463 464 coincident with high dissolved silica concentrations suggesting meltwater passage through 465 low redox environments conducive to denitrification in the proglacial sediments. Subsequent 466 to DOY 173, ^{excess}NO₃⁻ values remained remain close to zero for the remainder of the ablation 467 season at MLW, thereby providing no evidence of NO_3^- production in the western parts of the glacier, its margins and forefield during 2002. In contrast, significant ^{excess}NO₃⁻ levels at MLE 468 469 were co-incident with the emergence of subglacial meltwaters on DOY 184, indicating a 470 major input of NO₃⁻ that cannot be accounted for by the delayed release of concentrated 471 snowpack waters. The first fractions of subglacial outflow during the initial flow phase carried a distinct signature of negative ^{excess}NO₃⁻ associated with denitrification (see Wynn et 472 al. 2006). Maximum ^{excess}NO₃⁻ occurred on DOY 187 and for the rest of the monitoring period 473 ^{excess}NO₃⁻ concentrations remain stable, as do dissolved silica levels. Subglacial runoff is 474 therefore responsible for all ^{excess}NO₃⁻ in the stream at MLE, as ^{excess}NO₃⁻ concentrations here 475 476 were negligible prior to DOY 184.

477

Figure 5 also shows that the $^{\text{deficit}}NH_4^+$ was greatest at the start of the ablation period, 478 479 implying that catchment-wide loss of ammonium is greatest when the snowpack is isothermal and begins to produce runoff. The ^{deficit}NH₄⁺ was identical at MLE and MLW prior to DOY 480 481 184, suggesting rates of ammonium loss might be uniform across the catchment during the 482 early ablation period. However, the release of subglacial runoff on DOY 184 (the start of the principal flow phase) coincided with the release of very NH₄⁺-deficient runoff from beneath 483 the glacier, suggesting snowpack-derived NH_4^+ is very likely to undergo nitrification within 484 485 the subglacial environment.

Table 5 shows estimates of Cl⁻, NH₄⁺, NO₃⁻, ^{excess}NO₃⁻ and ^{deficit}NH₄⁺ mass fluxes for the main 487 rivers shown in Figure 1. The fluxes have been estimated from the product of daily discharge 488 489 and concentration pairs, using linear interpolation to fill any gaps in the latter series. These data show that for subglacial runoff, NO_3^- production (i.e. the ^{excess} NO_3^- flux) exceeded NH_4^+ 490 loss (the ^{deficit}NH₄⁺ flux) during 2002. These mass balance calculations therefore suggest that 491 the nitrification of snow and icemelt NH_4^+ only explained 83 % of the excess NO_3^- , 492 suggesting that an additional substrate for excessNO3 production within the subglacial 493 494 environment must also exist. Isotope data suggest the mineralisation of surface-derived 495 organic matter is the most likely source. However, when all rivers are considered, the mass balance calculations show that the $^{\text{deficit}}NH_4^+$ flux exceeds $^{\text{excess}}NO_3^-$ at the catchment scale. 496 497 Thus it is very likely that ammonia assimilation into organic matter (most likely within 498 cryoconite holes on the surface of the glacier) is important before streams enter the glacier 499 and undergo ammonia loss through processes of nitrification.

500

501 The processes responsible for $excessNO_3$ and $deficitNH_4^+$ within the glacial system can thus be 502 summarised as follows:

503

 $504 = \frac{\text{excess}}{\text{NO}_3} = \text{nitrification of ammonia} + \text{mineralization and nitrification of Organic N}$

505 deficit NH_4^+ = nitrification of ammonia + assimilation into organic matter

506

Mass flux calculations in Table 5 show that the excess NO₃⁻ flux in the principal subglacial 507 runoff phase accounts for 100 % of its catchment-wide production and is equivalent to 25 % 508 509 of the entire NO_3^- exported by the proglacial rivers. However, the total ^{deficit} NH_4^+ is even more striking, implying an 82 % reduction in the total meltwater transport of NH₄⁺ prior to leaving 510 511 the catchment. In this particular year, the net effect is for NH₄⁺ assimilation into the organic phase to reduce the export of dissolved inorganic nitrogen (i.e. DIN or $NO_3^- + NH_4^+$) by 27 %. 512 These figures therefore serve to demonstrate the significant reactivity of DIN in the glacial 513 514 ecosystem following the onset of melt.

515

516 **5.** Conclusions

517 The dissolved inorganic nitrogen content of glacial meltwaters, NO₃⁻/Cl⁻ ratios and δ^{15} N and

- 518 δ^{18} O isotopic ratios have been used to try to understand the capacity for glacial runoff to
- 519 export NO_3^- fluxes that are in excess of those entering the catchment via atmospheric

520	deposition. In so doing, we have been able to characterise significant rates of NO ₃ ⁻ production
521	beneath a high Arctic glacier in Svalbard, European High Arctic. Our data show that much of
522	the NO ₃ ⁻ present within the subglacial meltwaters has most likely been produced following
523	the microbial nitrification of snowpack NH_4^+ and mineralisation of organic nitrogen.
524	Presently, we are unable to identify the exact substrate(s) used, due to similar $\delta^{15}N$ end
525	member compositions. Mass fluxes indicate that loss of NH_4^+ is widespread across the entire
526	glacier basin and often exceeds the excess nitrate produced in the subglacial environment.
527	This suggests there must be an additional sink for ammonia within the catchment and is likely
528	represented through the assimilation of ammonia into organic matter such as that contained
529	within cryoconite holes on the surface of the glacier. Overall, NO ₃ ⁻ production, which is
530	responsible for enhancing the NO_3^- fluxes leaving the catchment by 25%, was far outweighed
531	by NH_4^+ assimilation occurring elsewhere and causing an 82% reduction in the NH_4^+ content
532	of runoff.
533	
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538	Hodson.
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775	Figure legends
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Tables

Table 1: Hydrological components sampled at Midtre Lovénbreen during summer 2002 and

Sampling location	Abbreviation in text	Active monitoring period (DOY)	Description of water type
Pre-melt snowpack 2002		101-102	Winter accumulation unaffected by summer ablation
Pre-melt snowpack 2003		164	Winter accumulation unaffected by summer ablation
Lysimeter snowmelt 2002		173-189	Snowmelt collected prior to contact with glacier ice
Lysimeter snowmelt 2003		167-178	Snowmelt collected prior to contact with glacier ice
Supraglacial runoff 2003		175-203	Bulk snowmelt and glacier ice
Subglacial runoff, initial runoff phase, 2002	MLSG	182-183	Sub-oxic discharge of long residence time and high rock-water contact. Sampled directly from upwelling on eastern side of glacier margin
Subglacial runoff: principal runoff phase, 2002	MLSG	184 onwards	Aerated subglacial discharge of low residence time. Sampled directly from upwelling on eastern side of glacier margin
Subglacial runoff, initial runoff phase, 2003	MLSG	186-193	Sub-oxic discharge of long residence time and high rock-water contact. Sampled directly from upwelling on eastern side of glacier margin
Subglacial runoff, principal runoff phase, 2003	MLSG	193 onwards	Aerated subglacial discharge of low residence time. Sampled downstream from upwelling on western side of glacier margin as mixture of supraglacial and subglacial water.
Midtre Lovénbreen East proglacial stream 2002	MLE	168-207	Bulk supraglacial and subglacial runoff plus groundwater component from proglacial zone
Midtre Lovénbreen West proglacial stream 2002	MLW	168-207	Bulk supraglacial runoff plus groundwater component from proglacial zone

Table 2: Nitrogen chemistry for pre-melt snow, supraglacial and subglacial meltwater samples from Midtre Lovénbreen

	$n^* =$	NO ₃ ⁻ (µmol/l)	NH4 ⁺ (μmol/l)	NH₄ ⁺ :Cl⁻ (µmol/l)	NO3 ⁻ :Cl ⁻ (μmol/l)	δ ¹⁵ N- _{NO3} * (‰)	δ ¹⁸ O- _{NO3} * (‰)	δ ¹⁵ N- _{NH4} * (‰)	δ ¹⁸ Ο- (%
Surface water				X. /			X /		
Pre-melt snow 2002	$n^1 = 2$ $n^2 = 2$	1.6 (0.1)	0.97 (0.1)	0.009 (0.001)	0.015 (0.0001)	-9.9 (0.02)	+57.3 (2.7)	-1.7 (1.6)	-12
Pre-melt snow 2003	$n^1 = 1$ $n^2 = 1$	3.1	5.1	0.04	0.027	-9.8	+72.0	-2.8	-11
Lysimeter snowmelt 2002 (DOY 173 – 189)	$n^1 = 8$	1.4 (0.7)	1.14 (1.37)	0.014 (0.012)	0.21 (0.017)				
Lysimeter snowmelt 2003 (DOY 167 – 178)	$n^1 = 9$ $n^2 = 8$	5.7 (7.0)	4.6 (3.4)	0.067 (0.036)	0.047 (0.0078)	-8.6 (0.7)	+64.5 (2.8)	-5.2 (0.2)	-12.73
Supraglacial streams	$n^1 = 8$	1.9	2.0	0.02	0.029	-7.8	+63.1	-5.8	-11
2003	$n^2 = 6$	(2.0)	(1.7)	(0.02)	(0.017)	(2.20)	(3.81)	(1.0)	(0.8
Subglacial, initial ru	noff phas	e							
2002 DOY 182-183	$n^1 = 2$ $n^2 = 1$	1.1 (0.50)	0.52 (0.30)	0.004 (0.002)	0.0094 (0.0036)	+2.3		-2.0	-12
2003 DOY 186-193	$n^1 = 8$ $n^2 = 4$	3.0 (1.0)	0.17 (0.08)	0.001 (0.0007)	0.028 (0.0084)	+4.0 (5.5)		-6.1 (1.5)	-12
Subglacial, principal runoff phase									
2002 DOY 184 onwards	$n^1 = 11$ $n^2 = 4$	4.3 (0.6)	0.14 (0.10)	0.0009 (0.0008)	0.038 (0.002)	-4.5 (1.3)		-2.9 (2.7)	-12
2003 DOY 193 onwards	$n^1 = 8$ $n^2 = 4$	3.9 (0. 6)	0.13 (0.20)	0.001 (0.002)	0.059 (0.0031)	-5.5 (1.1)	+20.3 (6.1)	-5.8 (3.5)	-11

n represents the number of field samples. n^1 represents the number of major ion samples

collected. n^2 represents the number of isotope samples collected. 1SD is given in parentheses.

Table 3: Analysis of geological specimens from the Midtre Lovénbreen catchment for $\delta^{15}N$ - NH_4^+ and nitrogen concentration

Dook type	NH4 ⁺ -N	$\delta^{15}N_{-NH4+}$ * (‰)	Replication	Organic Carbon
коск туре	(µg/g)	Vs AIR	(1 SD) (‰)	(%)
Phyllite	198	+7.2	0.18 (<i>n</i> =3)	0.29
Green Chert	59.3	+4.8	2.00 (<i>n</i> =2)	0.11
Pyritic Chert	168	-1.6		No data
Subglacial till	52.0	+7.7	0.37 (<i>n</i> =2)	0.06
Green schist	0.00	Below detection		0.03
Basement carbonates	6.06	Below detection		0.03
Sandstone	13.2	+4.8		No data
quartz	0.23	Below detection		0.03

07	1
02	I

Table 4: Nitrogen composition of cryoconite organic matter

	<i>n</i> =	δ ¹⁵ N (‰)	N content (mg/g)	C/N ratio
Cryoconite organic matter 2002	3	-4.8 (0.31)	1.2 (1.0-1.4)	11.4 (11.05-11.63)
Cryoconite organic matter 2003	6	-3.3 (0.83)	1.9 (1.8-2.0)	11.3 (10.6-11.7)

Values in parentheses represent one standard deviation for isotopes and ranges for all otherdata.

828 Table 5: Molar flux estimates during the 2002 observation period829

	excessNO ₃ (M)	^{deficit} NH ₄ (M)	Total NO ₃ (M)	Total NH ₄ (M)	Cl (M)
East proglacial stream (MLE)	2170	-2700	6990	560	230000
West proglacial stream (MLW)	130	-990	2090	270	91000
Total Catchment	2300	-3690	9070	830	323000
Total subglacial runoff (MLSG)	2300	-1900	5350	140	148000
Subglacial delivery ratio	1.00	0.52	0.59	0.16	0.46